

Draft Genome Sequence of *Herbaspirillum lusitanum* P6-12, an Endophyte Isolated from Root Nodules of *Phaseolus vulgaris*

Vinícius Almir Weiss,^a Helisson Faoro,^a Michelle Zibbetti Tadra-Sfeir,^a Roberto Tadeu Raittz,^b Emanuel Maltempi de Souza,^a Rose Adele Monteiro,^a Rodrigo Luis Alves Cardoso,^a Roseli Wassem,^a Leda Satie Chubatsu,^a Luciano Fernandes Huergo,^a Marcelo Müller-Santos,^a Maria Berenice Reynaud Steffens,^a Liu Un Rigo,^a Fábio de Oliveira Pedrosa,^a and Leonardo Magalhães Cruz^a

Department of Biochemistry and Molecular Biology, Federal University of Paraná, Curitiba, Paraná, Brazil,^a and Technological and Professional Education Sector, Federal University of Paraná, Curitiba, Paraná, Brazil^b

Herbaspirillum lusitanum strain P6-12 (DSM 17154) is, so far, the only species of *Herbaspirillum* isolated from plant root nodules. Here we report a draft genome sequence of this organism.

Perbaspirillum lusitanum strain P6-12 was isolated from root nodules of *Phaseolus vulgaris* plants in northeastern Portugal (7). Members of the genus *Herbaspirillum* are nitrogen-fixing betaproteobacteria capable of endophytically colonizing cereals of economic relevance (3, 5).

The genome sequence of *H. lusitanum* P6-12 was determined by using a combination of fragment and mate-paired libraries on a SOLiD4 sequencer (Life Technologies), producing a total of 4,460,595 fragment and 107,836,914 mate-paired reads 50 bp in length. These libraries were used for independent *de novo* genome assembly using Velvet v.1.2.03 (8). Gap closure was achieved by combination of the two assemblies.

The draft genome of *H. lusitanum* P6-12 contains 37 scaffolds and has an estimated size of 4.9 Mb with a coverage of 214-fold and 60.2% G+C content. Automatic annotation using RAST (2) revealed 5,240 open reading frames covering 84% of the chromosome, 38 tRNAs, and a single 16S-23S-5S rRNA operon. This annotation was curated by using an in-house-developed platform named GAAT (Genome Analysis and Annotation Tool) available at www.genopar.org.

H. lusitanum was originally determined to be a nitrogen-fixing bacterium by means of pellicle formation on nitrogen-free medium and nifD gene amplification (7). However, no nitrogenfixing (nif) genes were found in the genome. Furthermore, in our study, this strain was incapable of reducing acetylene in semisolid medium. Type I and II protein secretion systems were found. The type III secretion system, suggested to be involved in plant-bacterial interaction and present in the endophytes H. seropedicae SmR1 and H. rubrisubalbicans M1 (4), is absent from the H. lusitanum P6-12 genome. A nodD-like gene is present, although the nodulation (nod) genes were not found. The lack of nif and nod genes suggests that H. lusitanum is an opportunistic bacterium capable of colonizing root nodules, as well as other plant tissues. H. lusitanum P6-12 has the complete Entner-Doudoroff, pentose phosphate, and tricarboxylic acid cycle pathways. All of the genes coding for the Embden-Meyerhof-Parnas pathway are also present, although it lacks the classical 6-phosphofructokinase (EC 2.7.1.11) gene, as in *H. seropedicae* SmR1. An interesting finding is the presence of a gene coding for a protein very similar (83%) to the large ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO) subunit, a key enzyme involved in photosynthetic carbon fixation (6). RuBisCO-like proteins have been reported in

other heterotrophic bacteria (e.g., *Bacillus subtilis*) and were suggested to participate in a methionine salvage pathway (1). The gene coding for 1-aminocyclopropane-1-carboxylate (ACC) deaminase was found, indicating probable contributions to plant development under stress conditions. Finally, the lack of *nif* genes raises questions regarding the ability of *H. lusitanum* P6-12 to fix nitrogen. Further analysis of the *H. lusitanum* P6-12 genome will help to improve the understanding of how bacteria may associate and interact with plants.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under accession no. AJHH00000000. The version described in this paper is the first version, AJHH01000000.

ACKNOWLEDGMENTS

This genome sequencing project was supported by the Brazilian Program of National Institutes of Science and Technology-INCT/Brazilian Research Council-CNPq/MCT, a grant to the INCT of Biological Nitrogen Fixation.

REFERENCES

- Ashida H, Saito Y, Nakano T, Tandeau de Marsac N. 2008. RuBisCO-like proteins as the enolase enzyme in the methionine salvage pathway: functional and evolutionary relationships between RuBisCO-like proteins and photosynthetic RuBisCO. J. Exp. Bot. 59:1543–1554.
- Aziz RK, Bartels D, Best AA, Dejongh M. 2008. The RAST Server: rapid annotations using subsystems technology. BMC Genomics 9:75. doi: 10.1186/1471-2164-9-75.
- James EK, Olivares FL, Baldani JI, Döbereiner J. 1997. Herbaspirillum, an endophytic diazotroph colonizing vascular tissue in the leaves of Sorghum bicolor. J. Exp. Bot. 48:785–798.
- Monteiro RA, Balsanelli E, Tuleski T, Faoro H. 2012. Genomic comparison of the endophyte Herbaspirillum seropedicae SmR1 and the phytopathogen Herbaspirillum rubrisubalbicans M1 by suppressive subtractive hybridization and partial genome sequencing. FEMS Microbiol. Ecol. 80: 441–451.
- 5. Pedrosa FO, et al. 2011. Genome of Herbaspirillum seropedicae strain

Received 18 April 2012 Accepted 21 May 2012

Address correspondence to Leonardo Magalhaes Cruz, leonardo@ufpr.br. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/JB.00657-12 SmR1, a specialized diazotrophic endophyte of tropical grasses. PLoS Genet. 7:e1002064. doi:10.1371/journal.pgen.1002064.

- 6. Tabita FR, Satagopan S, Hanson TE, Kreel NE. 2008. Distinct form I, II, III, and IV RubisCO proteins from the three kingdoms of life provide clues about RubisCO evolution and structure/function relationships J. Exp. Bot. 59:1515–1524.
- 7. Valverde A, Velazquez E, Gutierrez C, Cervantes E. 2003. *Herbaspirillum lusitanum* sp. nov., a novel nitrogen-fixing bacterium associated with root nodules of *Phaseolus vulgaris*. Int. J. Syst. Evol. Microbiol. 53:1979–1983.
- 8. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 18:821–829.