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Itk Functions to Control Actin Polymerization at the Immune Synapse through Localized Activation of Cdc42 and WASP

Christine M. Labno¹, Carol M. Lewis³, Daoqi You⁴, Daisy W. Leung⁵, Ana Takesono³, Natalie Kamberos¹, Abhinav Seth⁵, Lisa D. Finkelstein³, Michael K. Rosen⁵, Pamela L. Schwartzberg³, and Janis K. Burkhardt^{1,2,*},⁶

¹Department of Pathology, University of Chicago, Chicago, Illinois 60637

²Committee on Immunology, University of Chicago, Chicago, Illinois 60637

³National Human Genome Research Institute, National Institutes of Health, Bethesda, Maryland 20892

⁴Cellular Biochemistry and Biophysics Program, Memorial Sloan Kettering Cancer Center, New York, New York 10021

⁵Department of Biochemistry, University of Texas Southwestern Medical Center, Dallas, Texas 75390

Summary

Actin polymerization at the immune synapse is required for T cell activation and effector function; however, the relevant regulatory pathways remain poorly understood. We showed previously that binding to antigen presenting cells (APCs) induces localized activation of Cdc42 and Wiskott-Aldrich Syndrome protein (WASP) at the immune synapse [1]. Several lines of evidence suggest that Tec kinases could interact with WASP-dependent actin regulatory processes [2]. Since T cells from *Rlk*^{-/-}, *Itk*^{-/-}, and *Rlk*^{-/-} × *Itk*^{-/-} mice have defects in signaling and development [3], we asked whether Itk or Rlk function in actin polymerization at the immune synapse. We find that *Itk*^{-/-} and *Rlk*^{-/-} × *Itk*^{-/-} T cells are defective in actin polymerization and conjugate formation in response to antigen-pulsed APCs. Itk functions downstream of the TCR, since similar defects were observed upon TCR engagement alone. Using conformation-specific probes, we show that although the recruitment of WASP and Arp2/3 complex to the immune synapse proceeds normally, the localized activation of Cdc42 and WASP is defective. Finally, we find that the defect in Cdc42 activation likely stems from a requirement for Itk in the recruitment of Vav to the immune synapse. Our results identify Itk as a key element of the pathway leading to localized actin polymerization at the immune synapse.

Results and Discussion

Itk Is Required for Actin Remodeling during T Cell-APC Interactions

T cells from mice deficient in the Tec family kinases Itk and Rlk show graded defects in Ca²⁺ flux, proliferation, and cytokine production [3]. Since T cells deficient in the actin

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*Correspondence: jburkhar@mail.med.upenn.edu.

⁶Present address: Children's Hospital of Philadelphia and The University of Pennsylvania, 816D Abramson Research Center, 3615 Civic Center Boulevard, Philadelphia, Pennsylvania 19104.

Supplemental Data

Supplemental Data including the Experimental Procedures are available at <http://www.current-biology.com/cgi/content/full/13/18/1619/DC1/>.

regulatory proteins WASP or Vav-1 have similar defects [4, 5], and Tec kinases interact with both proteins [6–8], we hypothesized that Tec kinases regulate actin remodeling in T cells. To test this, conjugates were formed between primary T cell blasts from wild-type (wt), *Itk*^{-/-}, *Rlk*^{-/-}, or *Rlk*^{-/-} × *Itk*^{-/-} TCR transgenic mice and antigen-pulsed B cells, and the distribution of F-actin was analyzed after labeling with rhodamine phalloidin. As expected, conjugates formed with wt T cells exhibited a sharp band of F-actin at the immune synapse in the presence of antigen (Figure 1B). In contrast, no antigen-dependent actin accumulation was observed in either *Itk*^{-/-} or *Rlk*^{-/-} × *Itk*^{-/-} T cells. Quantitative analysis showed that for *Itk*^{-/-} and *Rlk*^{-/-} × *Itk*^{-/-} T cells, the frequency of conjugates showing actin polymerization at the immune synapse was at background levels (defined by conjugates in the absence of antigen), while actin responses were normal for *Rlk*^{-/-} T cells (Figure 1C). The actin defect in *Itk*-deficient T cells was accompanied by defects in the formation of stable conjugates with APCs (Figure 1D). These defects are not attributable to differences in surface expression levels of the AND Tg TCR; expression of Vα11 on all mutant T cell blasts was as high as on wt controls (Figure 1A).

Since Tec kinases mediate signaling through both TCR and CD28 [9, 10], we used anti-TCR-coated beads to determine whether actin defects occur in the absence of CD28 engagement. Polymerization of actin at the bead interface was observed in wt T cells, but not in *Rlk*^{-/-} × *Itk*^{-/-} T cells (Figure 2). Moreover, costimulation with anti-CD28 under conditions that increase proliferation did not rescue the actin defect (data not shown). Thus, *Itk* is required for actin polymerization even in the absence of signaling through CD28. Since it has been reported that T cell activation by anti-TCR beads prepared in this way involves coligation of integrins by serum attachment factors [11], T cells were stimulated by crosslinking of soluble anti-CD3 in the absence of serum, and total F-actin content was assessed by flow cytometry. F-actin content in wt T cells increased by about 2-fold at 5 min of stimulation; this response was substantially blunted in the *Rlk*^{-/-} × *Itk*^{-/-} T cells (Figure 2C). Thus, while *Itk* may also function downstream of other costimulatory or integrin signaling pathways, we conclude that *Itk* functions downstream of TCR ligation in the pathway leading to net actin polymerization during conjugate formation. These results confirm and extend previous studies in *Itk*-deficient murine T cells [12] and Jurkat cells expressing kinase-inactive *Itk* [13].

Adhesive defects in *Itk*-deficient T cells were detectable only in the presence of antigen, indicating that “inside-out” signaling pathways required for high-avidity binding of the β2 integrin LFA-1 are *Itk* dependent. Indeed, we find that *Rlk*^{-/-} × *Itk*^{-/-} T cells exhibit defective TCR-activated adhesion to immobilized ICAM-1 (L.D.F. and P.L.S., unpublished data). Similarly, overexpression of dominant *Itk* mutants inhibits TCR-activated adhesion to fibronectin mediated by β1 integrins [13]. Since high-avidity integrin binding requires interaction with cortical actin, it seems likely that the defects in adhesion via β1 and β2 integrins stem from the failure to appropriately remodel F-actin.

***Itk* Is Required for Localized Cdc42 Activation and Downstream Activation of WASP**

To investigate the molecular basis for the actin defects, we asked whether *Itk* functions in the cascade leading to WASP activation. This cascade involves localized activation of Cdc42, which binds to and induces large conformational changes in WASP, which then activates the formation of branched actin structures by Arp2/3 complex [14, 15]. We have shown that WASP recruitment and activation are separable, with recruitment occurring via interactions with SLP76 and Nck, and activation occurring via binding to Cdc42-GTP accumulated at the same site [1, 16]. As shown in Figure 3, WASP and Arp2/3 complex were recruited efficiently to the immune synapse in conjugates formed with both wt and *Itk*-deficient T cells (either *Rlk*^{-/-} × *Itk*^{-/-} or *Itk*^{-/-}).

The recruitment of Arp2/3 complex to the immune synapse is not necessarily a reliable indicator of WASP activation. Though several studies indicate that Arp2/3 complex binds only to active WASP [14, 17], there is also evidence for constitutive association [18]. Moreover, Arp2/3 complex could be recruited by other proteins. To directly address the activation state of WASP, we generated an antibody against an epitope that is masked by intramolecular interactions in the autoinhibited conformation and thus specifically recognizes active WASP (D.W.L., C.M. Labno, D.Y., J.K.B., and M.K.R., unpublished data). Using this antibody, we observed antigen-dependent WASP activation at the immune synapses in wt T cells, but only background levels of activation in *Rlk*^{-/-} × *Itk*^{-/-} T cells (Figures 3E and 3F). WASP activation in *Itk*^{-/-} T cells was also significantly lower than in wt controls. Since no defect was evident in *Rlk*^{-/-} T cells, the presence of *Itk* appears to be of primary importance for localized WASP activation.

To determine if the failure to activate WASP results from a defect in Cdc42 activation, conjugates were labeled with recombinant GFP-WASP-GBD to detect Cdc42-GTP [1]. Wt and *Rlk*^{-/-} T cells showed strong accumulation of Cdc42-GTP at the immune synapse. In contrast, both *Itk*^{-/-} and *Rlk*^{-/-} × *Itk*^{-/-} T cells showed profound defects in Cdc42 activation (Figures 4A and 4B). Double-label analysis showed that Cdc42-GTP and active WASP colocalize at the immune synapse (Figure 4C). For both wt and *Rlk*^{-/-} × *Itk*^{-/-} T cells, every conjugate showing Cdc42-GTP accumulation also showed accumulation of active WASP (n = 50 conjugates for each genotype). This was not the case when the conformation-unspecific WASP antibody was used to compare overall WASP recruitment with Cdc42 activation; 24% of *Rlk*^{-/-} × *Itk*^{-/-} T cells showed recruitment of WASP in the absence of Cdc42-GTP (n = 50 conjugates). Thus, in the absence of *Itk*, WASP can be recruited to the immune synapse without undergoing the conformational changes associated with Cdc42-induced activation.

Itk Is Required for Vav Recruitment to the Immune Synapse

The results above suggest that *Itk* functions upstream of the relevant guanine exchange factor (GEF) for Cdc42. Although the role of Vav-1 as a direct GEF for Cdc42 is controversial [19, 20], we recently found that Vav-1-deficient Jurkat T cells are defective in Cdc42 activation at the immune synapse [16]. We therefore asked whether *Itk* is required for Vav recruitment. Although the background labeling of Vav was high in the absence of antigen, antigen-dependent recruitment of Vav was readily measurable in both wt and *Rlk*^{-/-} T cells (Figures 4D and 4E). In contrast, antigen-dependent Vav recruitment was completely abolished in T cells lacking *Itk*. Similar defects were obtained when Jurkat T cells were cotransfected with an epitope-tagged Vav-1 construct and siRNA oligonucleotides to inhibit *Itk* expression. As in *Itk*^{-/-} T cells, actin responses were defective and the efficiency of conjugate formation was diminished (D. Dombroski, C.M. Labno., J.K.B., and P.L.S., unpublished data). We conclude that *Itk* is required for Vav recruitment to the immune synapse. Since *Itk*-dependent phosphorylation of LAT can generate docking sites for Vav [21], LAT is a likely intermediary in this process. *Itk*-dependent phosphorylation of SLP-76 may also be required, since Vav recruitment requires tyrosine phosphorylation of SLP-76 [16].

Taken together, our findings show that *Itk* is an essential component of the actin regulatory cascade induced in T cells upon interaction with APCs. Unlike other aspects of T cell signaling in which loss of *Rlk* contributes to phenotypic severity [3], the actin defects are solely attributable to *Itk* deficiency. While this manuscript was in preparation, Grasis and coworkers reported that *Itk*^{-/-} T cells have defects in antibody-induced actin remodeling [12]. Our work confirms and extends this finding, showing that these cells have defects in APC-induced actin responses and defining activation defects in key actin regulatory molecules. Importantly, the actin defects in *Itk*^{-/-} T cells are not secondary to

developmental defects; we find similar defects in Jurkat cells in which *Itk* expression has been depressed by using siRNA, and both Grasis et al. [12] and Woods et al. [13] have observed actin defects in Jurkat cells overexpressing *Itk* mutants.

At the molecular level, we conclude that *Itk* is required for recruitment and activation of Cdc42 and WASP at the immune synapse. To our knowledge, this represents the first study to demonstrate in situ the colocalization of an activated Rho GTPase and its activated effector protein and thereby provides experimental evidence supporting the broadly accepted paradigm that localized actin responses are controlled by localized activation of Rho GTPases. We find that in *Itk*-deficient T cells, WASP is recruited to the immune synapse but is not activated, consistent with our finding that WASP recruitment and activation are independent events [1]. We have recently described a minimal model for WASP activation at the immune synapse involving signaling through TCR, Lck, ZAP-70, SLP-76, Nck, Vav, and Cdc42 [16]. We now show that *Itk* is also a key component in this cascade.

Interestingly, the phenotype of *Itk*-deficient T cells with respect to WASP recruitment and activation is identical to that of Vav-1-deficient Jurkat T cells [16]. Thus, it is attractive to speculate that the actin defects in *Itk*-deficient T cells stem from dysregulation of Vav function. Mislocalization of Vav could, in itself, lead to the observed defects; however, phosphorylation of Vav by *Itk* could also be required to regulate GEF activity. We previously reported that gross Vav phosphorylation proceeds normally in *Rlk*^{-/-} × *Itk*^{-/-} thymocytes [22], and we have obtained similar results in mature T lymphocytes from these mice. However, since Vav is phosphorylated at multiple sites [5], changes at specific sites associated with activation might not have been detected. There is conflicting evidence about the requirement for *Itk* kinase activity in actin regulatory pathways [12, 13, 23]. Thus, Vav activity may be regulated in a phosphorylation-independent manner, perhaps as part of the membrane recruitment mechanism. Finally, though we show here that *Itk* is required for Vav recruitment, there is also evidence that Vav influences *Itk* function [24]. Thus, it is possible that *Itk* functions as part of an interdependent group of signaling molecules, as proposed for Btk [25]. Additional studies will be required to resolve the mechanism(s) by which *Itk* and Vav interact.

In addition to regulating WASP via Cdc42, *Itk* could directly phosphorylate WASP at tyrosine 291 [7], a site that is revealed upon activation by Cdc42-GTP [26]. This raises the interesting possibility that *Itk* regulates WASP activity through a “feed-forward loop,” in which *Itk* first serves to activate Cdc42, revealing this phosphorylation site so that subsequent phosphorylation can modify WASP activity [26, 27]. Finally, we note that the defects in actin remodeling and conjugate formation in *Itk*-deficient T cells are much more profound than those in WASP-deficient T cells [28, 29]. This finding indicates that in addition to its role in WASP activation, *Itk* functions in other actin regulatory pathways. These could include Vav-dependent pathways and/or pathways dependent upon PLC γ and Ca²⁺ flux. The profound cytoskeletal defects in *Itk*-deficient T cells are likely to contribute to the defects in signaling, development, proliferation, and cytokine production in these cells.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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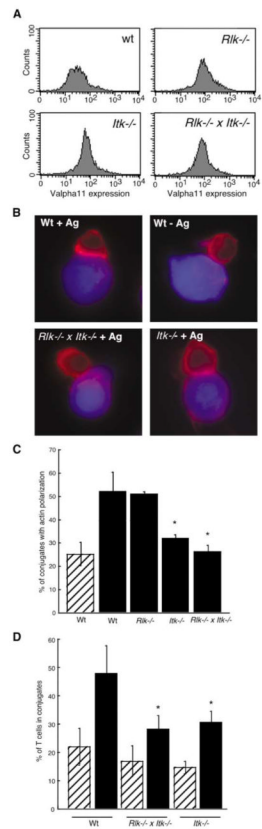


Figure 1. Actin Polymerization at the Immune Synapse and Conjugate Formation Are Reduced in *Rlk*^{-/-} × *Itk*^{-/-} and *Itk*^{-/-} T Cells

(A) Primary T cell blasts from AND TCR Tg mice were analyzed by flow cytometry to verify comparable surface expression of the Tg TCR.

(B) Primary T cell blasts were allowed to conjugate to CMAC (blue)-dyed B cells ± Ag for 10 min, were fixed, and were stained with rhodamine phalloidin to detect F-actin (red). Note the lack of F-actin enrichment at the immune synapse in conjugates formed with the mutant T cells.

(C) Conjugates formed in the absence (hatched bar) or presence (solid bars) of Ag were selected at random and were scored for accumulation of F-actin at the immune synapse. Data are mean ± SD from at least 3 independent experiments, with 50 conjugates each (the asterisk indicates a significant difference from wt + Ag, $p < 0.001$).

(D) Conjugation efficiency in the absence (hatched bars) or presence (solid bars) of Ag was analyzed by flow cytometry as described in the Experimental Procedures. Data are mean ± SD from at least three independent experiments (the asterisk indicates a significant difference from wt + Ag, $p < 0.05$).

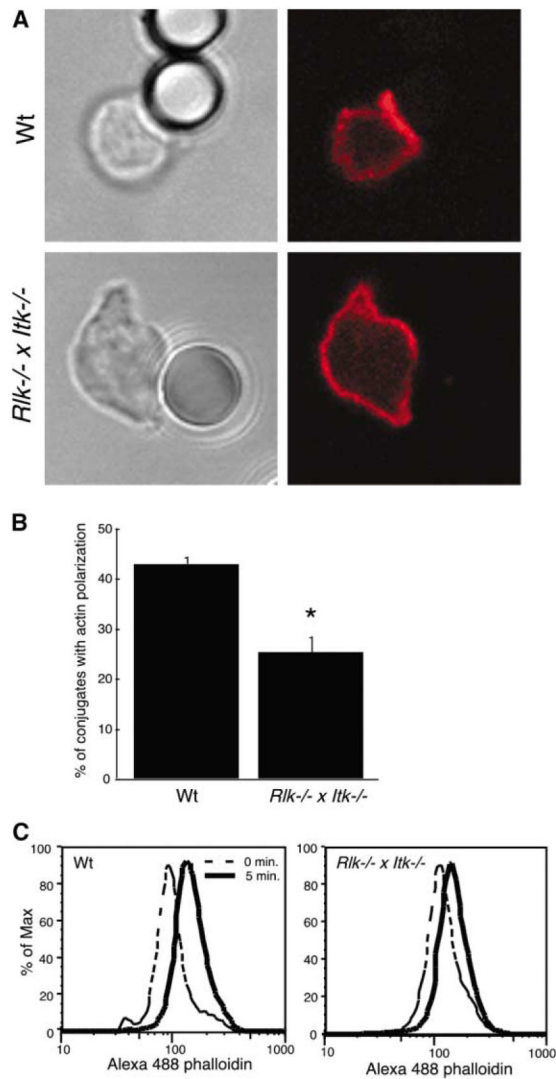


Figure 2. Defects in Actin Polymerization Occur Downstream of TCR Engagement

(A) Splenic T cells were allowed to conjugate to anti-TCR-coated beads, were fixed, and were stained with Alexa 594-phalloidin to detect F-actin (red). Note the lack of F-actin enrichment at the bead interface with the *Rlk*^{-/-} × *Itk*^{-/-} T cell.

(B) Bead conjugates formed as in (A) were scored for accumulation of F-actin. Data are mean ± SD from six independent experiments (the asterisk indicates a significant difference from wt, $p < 0.0001$).

(C) Splenic T cells were stimulated for 5 min by soluble anti-CD3 crosslinking and were fixed, and F-actin content was assessed after labeling with Alexa 488-phalloidin.

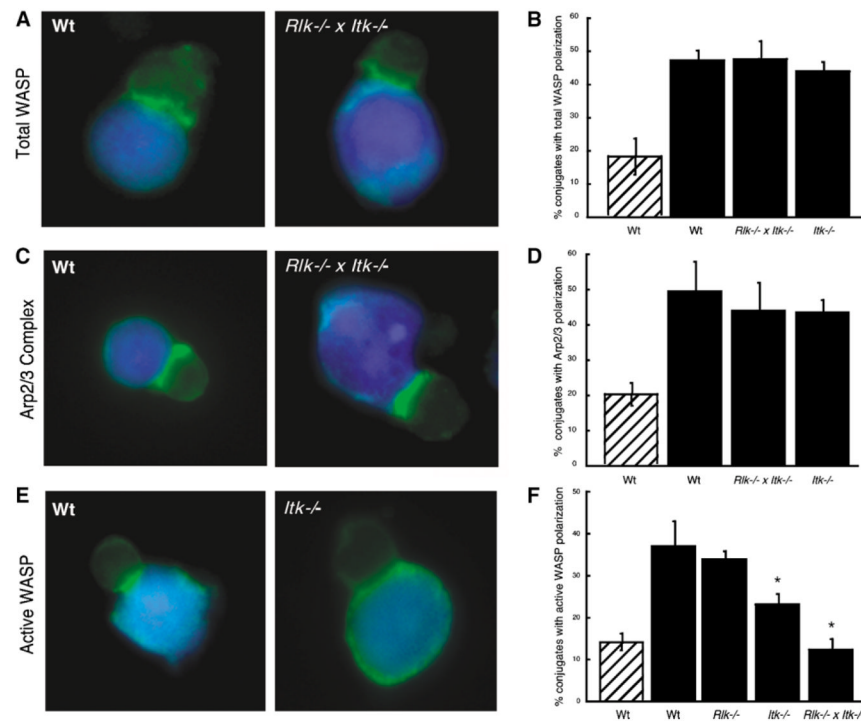


Figure 3. WASP and Arp2/3 Are Recruited to the Immune Synapse, but WASP Remains in an Inactive State

(A) Conjugates were labeled with an antibody that recognizes WASP in a conformation-independent manner. Both wt and *Rlk*^{-/-} × *Itk*^{-/-} T cells show recruitment of WASP to the immune synapse.

(B) Randomly chosen conjugates formed in the absence (hatched bar) or presence (solid bars) of Ag were scored for WASP polarization.

(C) Conjugates were stained with an anti-Arp3 antibody. Both wt and *Rlk*^{-/-} × *Itk*^{-/-} T cells show accumulation of Arp2/3 complex at the interface.

(D) Randomly chosen conjugates formed in the absence (hatched bar) or presence (solid bars) of Ag were scored for Arp2/3 polarization.

(E) Conjugates were stained with a monoclonal antibody that preferentially recognizes the open, activated form of WASP. Note that active WASP is not enriched at the immune synapse in the *Itk*^{-/-} T cell.

(F) Randomly chosen conjugates formed in the absence (hatched bar) or presence (solid bars) of Ag were scored for accumulation of open WASP at the immune synapse.

Data in (B), (D), and (F) represent means from at least three independent experiments ± SD (the asterisk indicates a significant difference from wt + Ag, $p < 0.01$ for *Itk*^{-/-}, and $p < 0.001$ for *Rlk*^{-/-} × *Itk*^{-/-} T cells).

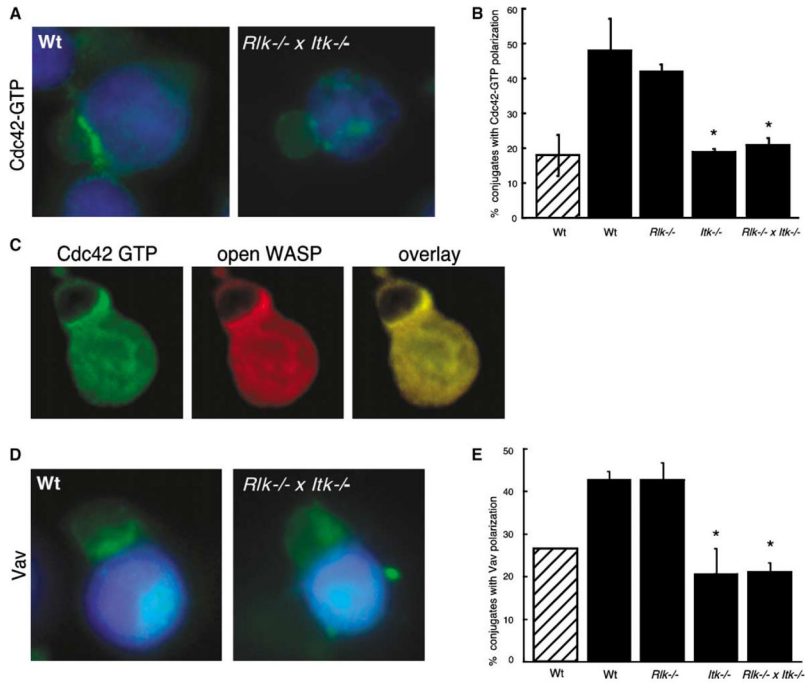


Figure 4. Itk Is Required for Vav Recruitment and Cdc42 Activation at the Immune Synapse

(A) Conjugates were labeled with recombinant GFP-WASP-GBD to detect Cdc42-GTP. Note the enrichment of activated Cdc42 at the immune synapse of the conjugate formed with the wt T cell, but not the *Rlk*^{-/-} × *Itk*^{-/-} T cell.

(B) Randomly chosen conjugates formed in the absence (hatched bar) or presence (solid bars) of Ag were scored for accumulation of Cdc42-GTP at the immune synapse.

(C) Double labeling was performed by using the GFP-WASP-GBD reagent to detect Cdc42-GTP (green) and mAb 26E6 to detect the open, active conformation of WASP (red). The overlaid image shows extensive colocalization of activated Cdc42 and WASP at the immune synapse (yellow).

(D) Conjugates were labeled with anti-Vav antibody. Note the enrichment of Vav at the immune synapse of the conjugate formed with the wt T cell, but not the *Rlk*^{-/-} × *Itk*^{-/-} T cell.

(E) Randomly chosen conjugates formed in the absence (hatched bar) or presence (solid bars) of Ag were scored for accumulation of Vav at the immune synapse. Data in (B) and (F) represent means from at least three independent experiments ± SD (the asterisk indicates a significant difference from wt + Ag, p < 0.001).