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Issue: *The Brain and Obesity***Interacting epidemics? Sleep curtailment, insulin resistance, and obesity**Eliane A. Lucassen,^{1,2} Kristina I. Rother,³ and Giovanni Cizza⁴

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In the last 50 years, the average self-reported sleep duration in the United States has decreased by 1.5–2 hours in parallel with an increasing prevalence of obesity and diabetes. Epidemiological studies and meta-analyses report a strong relationship between short or disturbed sleep, obesity, and abnormalities in glucose metabolism. This relationship is likely to be bidirectional and causal in nature, but many aspects remain to be elucidated. Sleep and the internal circadian clock influence a host of endocrine parameters. Sleep curtailment in humans alters multiple metabolic pathways, leading to more insulin resistance, possibly decreased energy expenditure, increased appetite, and immunological changes. On the other hand, psychological, endocrine, and anatomical abnormalities in individuals with obesity and/or diabetes can interfere with sleep duration and quality, thus creating a vicious cycle. In this review, we address mechanisms linking sleep with metabolism, highlight the need for studies conducted in real-life settings, and explore therapeutic interventions to improve sleep, with a potential beneficial effect on obesity and its comorbidities.

Keywords: sleep; obesity; insulin resistance; diabetes; appetite

Introduction

According to the National Health and Nutrition Examination Survey (NHANES) from 2009 to 2010, 33% of adult Americans were overweight (body mass index (BMI) of 25.0–29.9 kg/m²), and an even larger proportion (36%) were obese (BMI \geq 30.0 kg/m²).¹ The report contained good and bad news: for more than a decade, the age-adjusted prevalence of obesity had not changed in women overall, but there were marked ethnic, gender, and socioeconomic differences. Mexican American and Black women and men continued to show a significant rise in the prevalence of obesity. These figures are reflected in important obesity-related complications, such as hypertension, hyperlipidemia, and type 2 diabetes, all leading causes of cardiovascular disease.² Type 2 diabetes affects 8.3% of the U.S.

population (27% of U.S. residents aged 65 years and older), whereas 35% of U.S. adults have fasting glucose and glycosylated hemoglobin (HbA1c) levels in the prediabetic range.³ Thus, it is of great importance to characterize contributing etiological factors in order to effectively counteract the obesity and diabetes epidemics. The exact mechanisms explaining the relationship between short sleep, obesity, and diabetes remain unclear.

Accumulating evidence has pointed to an association between sleep curtailment and obesity,^{4,5} type 2 diabetes,^{6–9} and possibly mortality (see Ref. 10 for a review). In the last 50 years, self-reported sleep duration in the United States has decreased by 1.5–2 hours due to lifestyle changes in our society.¹¹ Worldwide, secular trends in adult sleep duration may be more variable. A recent review article summarizing 12 studies from 15 countries from 1960 to 2000 did not find a consistent decrease in self-reported adult sleep duration: self-reported sleep duration had increased in seven countries, decreased in six, and results were inconsistent for

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the United States and Sweden.¹² Imprecision in assessing sleep duration, as well as socioeconomic and demographic factors may play a role in this heterogeneity.

In this paper, we will first present the epidemiologic evidence for these relationships; we will then focus on sleep physiology and its role in metabolic functions. Then, we will critically evaluate the studies of sleep manipulation on glucose metabolism and energy expenditure (EE). We will highlight endocrine parameters that may influence these processes, including thyroid hormones, growth hormone (GH), cortisol, sympathovagal balance, and the immune system. We will also delineate the relationships between short sleep and some of the hormones that control appetite, including leptin, ghrelin, orexin, and neuropeptide Y (NPY). In addition, we will present evidence for the reverse effect: of obesity/diabetes on sleep. Finally, we will describe emerging evidence for a role of melatonin in the sleep–metabolism relationship and its therapeutic potential in metabolic disturbances.

Epidemiologic evidence for an association among sleep duration, obesity, and diabetes

A meta-analysis of 45 cross-sectional studies, including 604,509 adults and 30,002 children, confirmed the relationship between short sleep (generally fewer than five hours per night in adults and fewer than 10 hours per night in children) and obesity, and quantified the associated risk (OR 1.55; 1.43–1.68; OR 1.89; 1.46–2.43, for adults and children, respectively).¹³ A similar OR of 1.58 (1.26–1.98) was found in children in another meta-analysis.¹⁴ Most of the included studies used actigraphy or self-reported sleep duration. A polysomnography study, not included in these meta-analyses, of 2,700 men above the age of 65 years revealed an inverse relationship between slow-wave sleep (SWS) duration and BMI or waist circumference.¹⁵ Another study reported that obese adults (BMI 41 ± 1 kg/m²) without sleep apnea slept 88 min fewer than lean subjects.¹⁶ The relationship between short sleep and BMI was also confirmed by actigraphy in 612 participants (35–50 years old).¹⁷

In principle, causality is best addressed in prospective randomized controlled studies. This ideal approach may not be practical in this case: studies of sleep extension or curtailment cannot be blinded, and participants may not behave ac-

ording to the allocation group (i.e., control subjects changing sleep duration or, vice versa, intervention subjects not changing sleep duration). For these reasons, the existing prospective studies have all been observational in nature. Short sleepers experience greater weight increases over time according to most,^{4,18–22} but not all studies.^{17,23,24} However, average weight gain was modest and of relative clinical meaning. Short sleepers (fewer than five to six hours) gained 2 kg more in a 6-year study,¹⁹ and 0.4 kg more in another 16-year study.¹⁸ In addition, subjects had a 35% and 31% greater chance of gaining more than 5 kg in 6 years, and gaining more than 15 kg in 16 years, respectively.^{18,19} Thus, there is a large variability among studies in weight gain associated with short sleep for unknown reasons: the magnitude of this effect may be gender dependent or depend on other yet-to-be identified genetic and environmental factors. For example, short sleep may affect metabolism differently in natural “short sleepers” versus chronically sleep-deprived individuals. A large study including 35,247 subjects found that lean men, but not lean women, who slept fewer than five hours had increased odds of becoming overweight within one year (OR 1.91; 1.36–2.67).²⁵ In contrast, in a cohort of 3576 elderly subjects, women, but not men, who slept fewer than five hours, had a higher risk of gaining 5 kg in two years (OR 3.41; 1.31–8.69).²⁰ Interestingly, in this study, BMI was also increased in women sleeping more than eight hours. A similar U-shaped association has been reported in other studies.^{19,25} Collectively, these studies indicate that self-reported short or long sleep is associated with increased BMI. The amount of weight gain over time, however, varied greatly, ranging from less than 1 kg to several kg, and age may be one of the parameters modulating this association.

Many mechanisms may be responsible for the frequently observed insulin resistance in obese individuals, including increased release of free fatty acids, leptin, and TNF- α from adipose tissue.²⁶ We suggest that short sleep (fewer than five to six hours) may now qualify as an additional clinical factor in the development of insulin resistance and diabetes, as shown in several cross-sectional studies.^{27–29} A recent study of 174,542 middle-aged subjects showed that short sleepers had an OR of 1.46 (1.31–1.63) for developing diabetes after 3–10 years of follow-up.³⁰ A meta-analysis of 10 prospective studies

reported a pooled OR of 1.28 (1.03–1.60) for diabetes in short sleepers.³¹ Interestingly, this correlation was only present in men (OR 2.07; 1.16–3.72). Both long sleep (OR 1.48; 1.13–1.96) and decreased sleep efficiency also resulted in greater chances of developing diabetes.³¹ Difficulty in falling asleep resulted in a greater incidence of diabetes after 4.2–14.8 years of follow up.^{32–34} In smaller studies, the association between sleep duration and diabetes is more variable.^{7–9,35} In addition, this variability may be real and differ on the basis of ethnic group and socioeconomic status: a study found increased risk of developing diabetes only in Whites and Hispanics who slept fewer than seven hours.³⁶ Besides sleep-duration, poor sleep quality relates to insulin resistance. Frequency of nocturnal awakenings, as measured by actigraphy, correlated positively with HbA1c levels in patients with type 2 diabetes.³⁷ Furthermore, sleep disordered breathing was related to worse blood glucose control.³⁸ Taken together, there is solid epidemiologic evidence for an interconnection between sleep duration (too short or too long), insulin resistance, and obesity.

Physiological changes of metabolism during normal sleep

Sleep architecture, the structure and pattern of sleep, is best described by a combined approach that uses electroencephalographic, electromyographic,

and other criteria³⁹ (Fig. 1). Many metabolic parameters display diurnal rhythms, which are the resultant of both homeostatic and circadian mechanisms. From a homeostatic point of view, sleep pressure increases with sustained wakefulness. Circadian rhythms are endogenously generated in the suprachiasmatic nucleus of the hypothalamus by feedback loops of transcripts of a group of “core clock” genes, including the CLOCK gene.^{40,41} Time-keeping signals are passed on from the suprachiasmatic nucleus to oscillators in peripheral tissues also containing core clock genes. Endogenous rhythms, in turn, are synchronized to the outside world by various environmental clues, mostly by light via the retina.

During sleep, energy requirements are lower as metabolic needs for processes such as breathing, gut motility, heart rate and muscular activity, are decreased.⁴² Indeed, resting EE, as determined by indirect calorimetry, was lower in healthy volunteers during sleep compared to wakefulness.⁴³ During sleep glucose levels remain stable or fall only minimally in spite of fasting, mostly because of decreased energy needs.⁴⁴ In contrast, if subjects are kept awake in bed while fasting for a comparable number of hours, glucose levels do fall, approximately 15 mg/dl.⁴⁵ During the first part of the night, when most slow wave sleep (SWS) occurs, glucose production is diminished⁴⁴ (Fig. 2). Brain

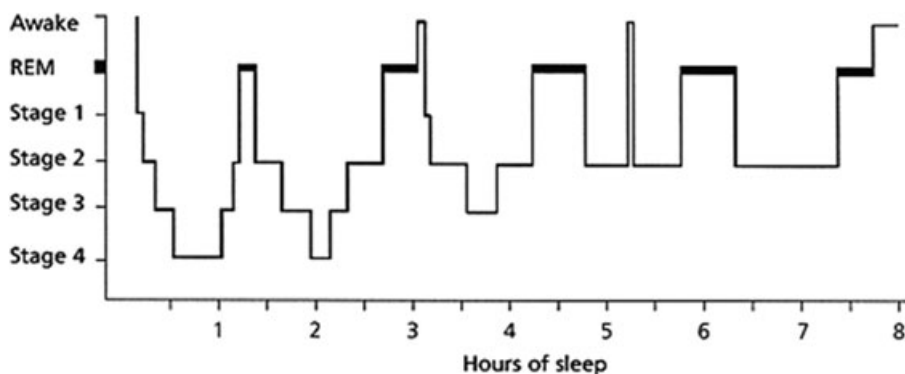


Figure 1. Representative sleep architecture during eight hours of uninterrupted sleep (adapted from Ref. 39). Sleep can be divided into rapid eye movement (REM) and nonrapid eye movement (NREM) sleep. On average, adults spend 75–80% in NREM sleep, which can be further subdivided into stages 1 to 4. Stages 3 and 4 are often referred to as slow-wave sleep (SWS) because of the appearance of well-defined waves of 0.5–4.0 Hz. In REM sleep, a phasic and a tonic phase can be distinguished. Skeletal muscles are atonic or hypotonic during REM sleep (except for the diaphragm, extraocular, and sphincter muscles), but bursts of muscle activity can occur during the phasic phase. Furthermore, dreams mostly take place during REM sleep and are more complex during this stage. Eye movement is only present during the phasic phase. Throughout a normal night of sleep, there are three to five sleep cycles, from NREM (stage 1 to 4) to REM, each cycle taking 90–120 minutes. Humans display less SWS and more REM sleep toward the end of the night.

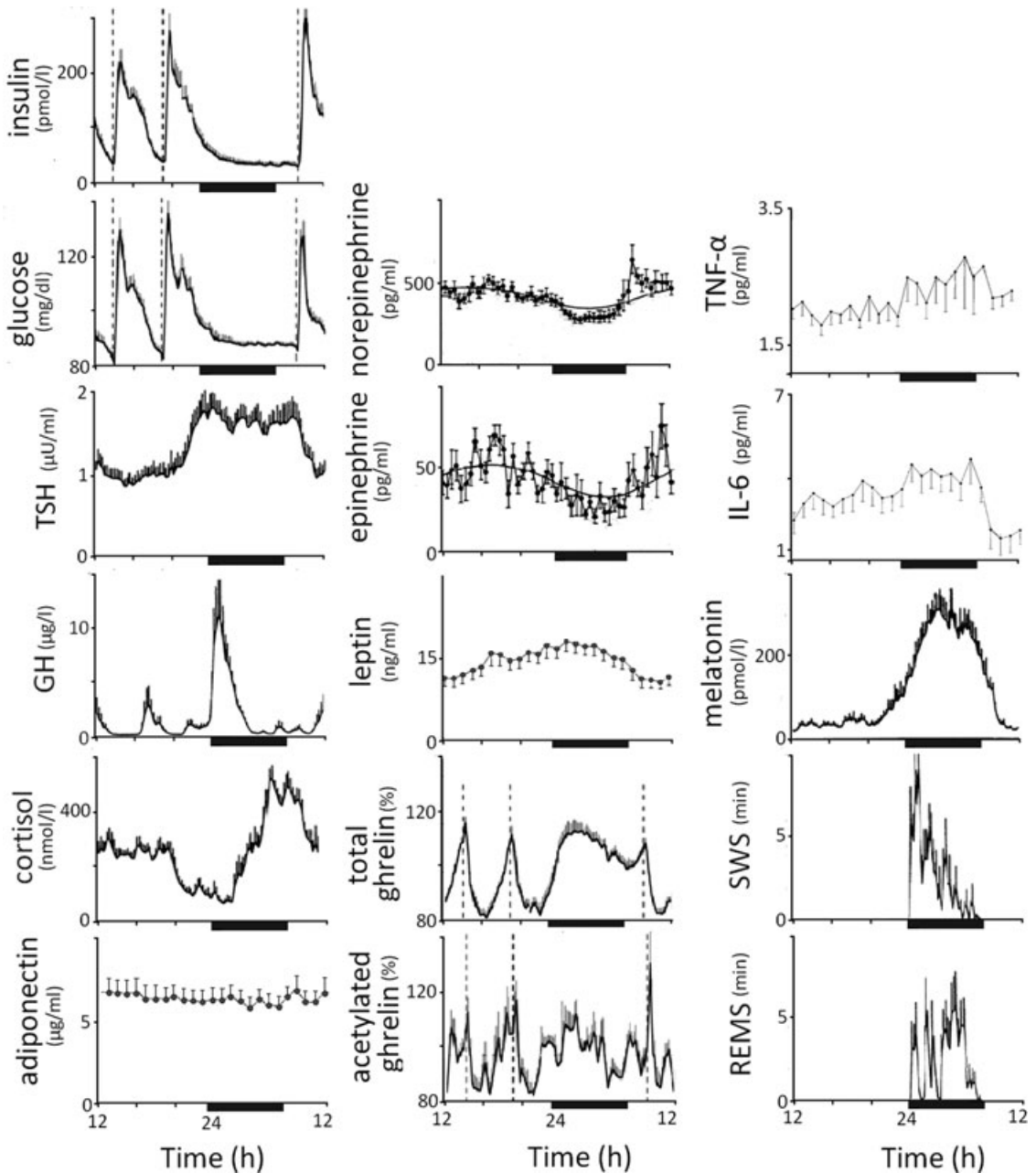


Figure 2. Mean levels of metabolic parameters and sleep stages in lean young men ($n = 8$ for TSH, cortisol, GH, melatonin, SWS, and REM sleep;⁴⁸ $n = 14$ for insulin, glucose, total and acetylated ghrelin;⁷⁶ $n = 8$ for catecholamines⁷³); in 23 lean women for leptin and adiponectin;⁶⁹ and in 25 individuals (13 females, 12 males) for IL-6 and TNF- α .⁸³ Ghrelin levels are indicated as a percent of mean 24-h values (1027 pg/mL for total ghrelin; 80 pg/mL for acetylated ghrelin); leptin levels are shown as percent change from levels at 08:00. Dark bars below each plot indicate bedtimes. Sleep was monitored polysomnographically during all measurements. Striped bars indicate meal times. Modified, with permission Refs. 48, 69, 73, 76, and 83

glucose metabolism, as measured by PET, decreases by 30–40%.^{46,47} During the second half of the night, glucose consumption increases due to increased REM sleep requiring more energy.^{46,47}

EE and glucose metabolism are modulated by a number of hormones. Thyroid hormones are important determinants of EE; cortisol and GH are powerful modulators of circadian rhythms of glucose metabolism. The sympathovagal system and adiponectin influence both EE and glucose metabolism. Thyroid stimulating hormone (TSH) levels rise at night, mostly due to increased hypothalamic secretion of thyroid releasing hormone (TRH) (Fig. 2).⁴⁸

GH secretion is under dual hypothalamic control: somatostatin is inhibitory and growth hormone releasing hormone (GHRH) is stimulatory. Somatostatin and GHRH are secreted in alternation, thus generating the typical pulsatile pattern of GH secretion. GH typically peaks at sleep onset during SWS (Fig. 2).⁴⁹ GH levels were lower in SWS-deprived individuals⁵⁰ and higher in individuals with pharmacologically induced SWS.⁵¹ In addition, REM sleep deprivation did not affect plasma GH levels.⁵² Intravenous GH administration decreases SWS, suggesting a modulatory role for GH on SWS.^{53,54} The GH peak at sleep onset has insulin lowering effects.⁵⁵ GHRH injection increased glucose levels at awakening by almost 50%⁵⁶ and GH administration rapidly decreased muscular glucose uptake.^{55,57}

Cortisol concentrations reach their zenith in the morning, experience a gradual fall during the day, which is briefly interrupted by meals, and have their nadir around 3 am (Fig. 2).⁴⁸ Cortisol impairs insulin sensitivity with a latency of four to six hours.^{56,58,59} Plasma insulin levels also display a modest diurnal variation with a 10% excursion, a nadir between midnight and 6 am and a peak between noon and 6 pm.⁶⁰

Adiponectin is a hormone secreted by the adipocytes in an inverse fashion compared to fat mass.⁶¹ Its role in energy homeostasis is unclear: increases, decreases, or no changes in energy balance are derived from peripheral adiponectin administration in rodents.⁶² Adiponectin knockout mice develop insulin resistance and increased TNF- α levels when fed a high fat diet. In humans, plasma adiponectin levels correlated negatively with insulin resistance, and higher levels correlated with

future weight gain in 1,063 women.^{63,64} Individuals with type 2 diabetes typically have low adiponectin levels, even when adjusted for body weight.⁶⁵ In contrast, patients with type 1 diabetes have higher than expected adiponectin levels: an observation that is poorly understood.⁶⁶ Sleep duration was inversely related to serum adiponectin in 109 lean Japanese males.⁶⁷ Adiponectin has anti-inflammatory properties: *in vitro* exposure to adiponectin decreases macrophage activation and TNF- α production. Finally, adiponectin displays some diurnal variability, with lower levels at night (Fig. 2).^{68,69}

During sleep, vagal tone increases.⁷⁰ Sympathovagal balance is lowest during REM sleep, when the locus coeruleus, the main brain source of catecholamine biosynthesis, is silent.⁷¹ Plasma norepinephrine (NE) and epinephrine (EPI) are produced by the sympathetic system: EPI is secreted exclusively from the adrenal medulla, whereas NE can also be released from postganglionic sympathetic nerves.⁷² In a small study of eight lean males, plasma catecholamines were consistently lower during the night with no differences among sleep stages (Fig. 2).⁷³ Heart rate and mean blood pressure were lower during NREM sleep, compared to during awakenings and REM sleep.⁷⁰

Several hormones involved in appetite regulation display circadian rhythms. Leptin is secreted by the white adipose tissue in a highly pulsatile fashion.⁷⁴ Leptin reflects fat stores: its levels increase after meals and during the night and are associated with decreased appetite (Fig. 2).⁷⁴ Ghrelin, produced in the stomach, stimulates appetite and circulates mainly in a nonacetylated form.⁷⁵ Acetylated ghrelin, the active form, is necessary for most of its endocrine actions, including GH release, appetite stimulation, and gastric emptying. Total and acetylated ghrelin levels rise with fasting and during sleep (Fig. 2).⁷⁶ Leptin and ghrelin exert their effects in the arcuate nucleus of the hypothalamus, through anorexigenic proopiomelanocortin/cocaine- and amphetamine-regulated transcript (POMC/CART) and orexigenic neuropeptide Y/agouti-related protein (NPY/AgRP) neurons. Leptin activates POMC/CART neurons, which induces hunger and inhibits NPY/AgRP neurons,^{77,78} which induces satiety. Ghrelin has the opposite effect on the arcuate nucleus, stimulating appetite, and prolonging postprandial glucose responses, while

stimulating GH release.⁷⁹ Insulin, while peripherally lowering blood glucose and stimulating appetite, inhibits appetite centrally in a leptin-like manner.⁷⁸

Orexin A and B, two neuropeptides released by lateral hypothalamic neurons, stimulate appetite. Orexin neurons are active during waking and quiescent during sleep; consistently, orexin levels in the CSF are maximal at the end of the waking period.^{80,81} The orexin system is activated by ghrelin and inhibited by leptin and glucose. Projections from orexin neurons in the lateral hypothalamus activate NPY neurons in the arcuate nucleus.⁸⁰ Melatonin is a sleep-promoting hormone that is produced by the pineal gland. Melatonin is exclusively secreted during the dark, as light inhibits its production (Fig. 2).

Interleukin 6 (IL-6), interleukin 1 β (IL-1 β), TNF- α , and C-reactive protein (CRP) are pro-inflammatory factors. IL-6 displays a circadian rhythm, with a first peak around 2 am and a second peak around 5 am (Fig. 2).^{82,83} IL-1 β decreases during the night and reaches its nadir at 8 am in the morning.⁸² TNF- α also displays a circadian rhythm, peaking close to the awakening (6 am) and reaching a nadir around 3 pm (Fig. 2). IL-6 and TNF- α stimulate secretion of cortisol and, in turn, cortisol inhibits their secretion.⁸⁴ Reports on diurnal variability of CRP are few and inconsistent: some report no diurnal variance,⁸⁵ whereas others find increased morning levels.⁸⁶ In 10 healthy, lean men, white blood cell counts peaked around 11 pm, decreased throughout the night and reached a nadir at 8 am.⁸²

The relationships here described between hormone secretion and sleep stages are necessarily descriptive. In theory, as new pharmacological tools aimed at manipulating hormonal status (i.e., selective hormone agonists/antagonists) and/or sleep become available for investigation, we may be able to better understand the numerous causal links interconnecting these phenomena. In summary, many metabolic parameters involved in EE regulation, glucose metabolism, and appetite control display diurnal rhythms, reflecting the different metabolic needs during the sleep and wake states. In the next section, we will report the effect of sleep deprivation on these metabolic parameters.

Effects of sleep restriction on metabolic parameters

The relationship between sleep and metabolism has been extensively studied in human subjects under varying conditions, such as total sleep deprivation,^{87–100} partial sleep deprivation,^{101–122} and sleep fragmentation^{123–125} (Table 1). Partial sleep deprivation affects SWS less than other sleep stages. Most of the experiments on the effects of acute sleep deprivation were conducted in healthy, lean volunteers. Food intake and physical activity were often, but not always, strictly controlled (Table 1). It is important to note that experimentally induced sleep alterations may not be representative of changes occurring in clinical condition of chronic sleep deprivation. In addition, important differences exist between acute and chronic sleep deprivation. We will describe effects of sleep curtailment on important metabolic parameters that may contribute to insulin resistance and obesity, including effects on appetite and EE (Fig. 3).

Effects on glucose metabolism

In the course of the night, glucose supply to the brain must remain adequate in the setting of slightly declining plasma glucose levels. Infusion of [³H]-labeled glucose in healthy, lean subjects revealed decreased glucose usage during the night, whereas in subjects kept awake glucose usage was increased (Table 1).⁴⁴ Total and partial sleep deprivation reduced glucose tolerance in lean individuals. This has been documented by intravenous glucose tolerance tests (ivGTT),^{101,109,114,123,124} oral glucose tolerance tests (OGTT),^{87,89,109} insulin suppression tests with octreotide,⁹⁰ as well as hypoglycemic and euglycemic hypoinsulinemic clamps^{97,114} (Table 1). Impaired glucose metabolism was not only shown in healthy study participants, but also in individuals with type 1 diabetes: after a four-hour night in adults with type 1 diabetes, glucose infusion rate decreased by 21%, as assessed by hyperinsulinemic euglycemic clamp¹¹⁵ (Table 1). In addition to altered peripheral glucose metabolism during total sleep deprivation, cerebral glucose uptake decreased, especially in the prefrontal and posterior parietal cortex and in the thalamus of healthy volunteers⁹¹ (Table 1). Interestingly, most studies found no compensatory rise in plasma insulin secondary to increased insulin resistance after short sleep,^{109,114}

Table 1. Results of 37 human experimental studies assessing the influence of sleep curtailment on metabolism

Study design	Gender/sample size Age \pm SEM (years)	Experimental sleep protocol	Techniques used for blood sampling (time, frequency)	Main findings (sleep deprived vs. longer sleep)
Energy intake	Mean weight \pm SEM			
Behavioral activity	Prestudy conditions			
Benedict <i>et al.</i> ¹⁰⁰	Men ($n = 14$)	1: 1 night of 8-h bedtime ^e	Indirect calorimetry	\uparrow postprandial glucose (8%)
Randomized	22.6 \pm 0.8		pre- and	\downarrow RMR (5%) and
cross-over study	23.9 \pm 0.5 kg/m ²	2: 1 night of TSD	postbreakfast;	postprandial (20%)
Controlled,	6 weeks regular bedtimes ^c	Conditions \geq 4 weeks apart	morning VAS	metabolic rate
postintervention			hunger ratings 24 h,	\uparrow cortisol (7%) ^a
<i>ad libitum</i>			every 1.5–3 h	\uparrow NE (12%) ^a
Bed rest				\sim food intake; \uparrow hunger (60%) ^a
				\uparrow morning ghrelin (11%) \sim leptin
Born <i>et al.</i> ⁸²	Men ($n = 10$)	1: 1 night of 8-h bedtimes ^e then 1 night of <i>ad libitum</i> sleep ^e	LPS-stimulated whole blood and ELISA (TNF- α ; IL-1 β ; IL-6)	\sim IL-6 \sim IL-1 β ; \uparrow nocturnal whole blood
Randomized	24.7 (21–29)			\sim TNF- α
cross-over study	NA			\uparrow white blood cells (8%) ¹
Standardized meals	NA	2: 24 h TSD ^e	51 h, every 3 h	
Synchronous		Conditions \geq 10 days apart		
inpatient activities				
Brondel <i>et al.</i> ¹¹³	Men ($n = 12$)	1: 1 night of 8-h bedtime ^e	Actigraphy; 12 daytime VAS	\sim EE; \uparrow physical activity (13%)
Randomized	22 \pm 3 22.3 \pm 1.83 kg/m ²	and 1 night <i>ad libitum</i> sleep at home ^{c,d}	hunger ratings	\uparrow hunger; \uparrow food intake (22%)
cross-over study	2 days sleep/diet diaries ^c			
<i>Ad libitum</i>		2: 1 night of 4-h bedtime ^e and 1 night of free bedtimes at home ^{c,d}		
Free physical activity ^d		Conditions \geq 5 days apart		
Buxton <i>et al.</i> ¹¹⁴	Men ($n = 20$)	1: 3 nights of 10-h bedtimes ^d	ivGTT penultimate day; clamp last day;	\uparrow insulin resistance (20% by ivGTT; 11% by clamp)
Consecutive phases	26.8 \pm 5.2		salivary sampling	\sim fasting RMR
study	23.3 \pm 3.1 kg/m ²	2: 7 nights of 5-h bedtimes ^d	15:00–21:00 last 2 days, every hour,	\uparrow salivary cortisol (51%)
Controlled isocaloric	\geq 5 days of 10-h bedtimes ^{c,d}		24-h urine	\uparrow urinary NE (18%) and EPI (22%) ¹
(finishing meal required)			sampling last 2 days; indirect calorimetry	
Controlled sedentary living				
Clore <i>et al.</i> ⁴⁴	5 females, 12 males ($n = 17$)	1. Group 1 ($n = 11$): 1 night of 8-h, 30 min bedtime	Continuous [³ -3H] glucose infusion	\downarrow nocturnal fall in glucose use
Comparative study				\downarrow nocturnal fall in glucose production
Similar evening meals	25 \pm 1	2. Group 2 ($n = 6$): 1 night of TSD		
Bed rest	24.1 \pm 0.6 kg/m ²			
	NA			
Donga <i>et al.</i> ¹¹⁵	3 females, 4 males ($n = 9$)	1: 3 nights of 8-h, 30-min bedtimes ^d	Hyperinsulinemic euglycemic clamp	\uparrow insulin resistance (21%)
Randomized				
cross-over study	44 \pm 7			
NA	23.5 \pm 0.9 kg/m ²			

Continued

Table 1. Continued

Study design	Gender/sample size Age ± SEM (years)	Experimental sleep protocol	Techniques used for blood sampling (time, frequency)	Main findings (sleep deprived vs. longer sleep)
Energy intake	Mean weight ± SEM			
Behavioral activity	Prestudy conditions			
NA	Type 1 diabetes, HbA1c ≤ 8.5%, 1 week of regular bedtimes ^{c,d}	2: 3 nights of 4-h bedtimes ^e Conditions ≥ 3 weeks apart		
Dzaja <i>et al.</i> ⁹⁶	Men (<i>n</i> = 10)	1: 1 night of 8-h bedtime ^e	24 h, every h	↓ GH peak (71%) ^b ~ mean cortisol levels
Randomized cross-over study	28 ± 3.1 24.0 ± 2.9 kg/m ²	2: 1 night of TSD		↓ nocturnal total ghrelin increase (39%) ^b
1,800 kcal/24 h Bed rest	1 week of regular bedtimes ^c	Conditions ≥ 2 weeks apart		
Frey <i>et al.</i> ⁹⁶	9 females, 10 males (<i>n</i> = 19)	1: 3 nights of 8-h bedtimes ^e	Saliva 36 h, every h; hsELISA (IL-6; IL-1β; CRP)	~ saliva cortisol; ↓ at 13:00 (53%) ^b and 20:00 (17%) ^b ↓ daytime IL-6
Consecutive phases study	28.1 ± 8.6	2: 1 night of TSD		↑ IL-1β
Hourly controlled intake	18.5–24.5 kg/m ²		40 h, every 30 min	↓ morning hsCRP
Bed rest	3 weeks of regular bedtimes ^{c,d}			
González-Ortiz <i>et al.</i> ⁹⁰	7 females, 7 males (<i>n</i> = 14)	1: baseline “normal sleep”	Insulin suppression test with octreotide	↑ insulin resistance (18%) ^a ~ morning cortisol
Consecutive phases study	21.0 ± 2.1 22.1 ± 1.3 kg/m ²	2: 1 night of TSD	Once, morning	
Controlled isocaloric Free inpatient movement	3 days diet diary			
Haack <i>et al.</i> ¹⁰⁸	6 females, 12 males (<i>n</i> = 18)	(<i>n</i> = 8) 1: 10 nights of 8-h bedtimes ^d	24-h urine 24-h blood, every 4 h (high sensitivity ELISA IL-6 and hsCRP)	~ food intake ↑ IL-6 (62%) ^a ~ TNF-receptor 1 ↑ hsCRP (102%; <i>P</i> = 0.11) ¹
Comparative study	27.3 ± 5.8	(<i>n</i> = 10) 2: 10 nights of 4-h bedtimes ^d		
Controlled isocaloric Free inpatient movement	23.1 ± 3.3 <30 kg/m ² ≥ 10 days regular bedtimes ^c			
Hursel <i>et al.</i> ¹²⁵	Men (<i>n</i> = 15)	1. 2 nights of 8-h bedtimes ^e	Indirect calorimetry	~ total EE ↑ physical activity (8%) ^a ↑ RQ (3%) ^a
Randomized cross-over study	23.7 ± 3.5 24.1 ± 1.9 kg/m ²	2: 2 nights of sleep fragmentation hourly 2-min-long alarms ^e		
Controlled isocaloric Sedentary activity	2 days controlled diet	Conditions ≥ 2 weeks apart		
Irwin <i>et al.</i> ¹⁰⁷	13 females, 17 males (<i>n</i> = 30)	1: 3 nights of 8-h bedtimes ^e	Flow cytometry; real time PCR (<i>n</i> = 10); high-density oligonucleotide array (<i>n</i> = 5)	↑ IL-6 monocyte expression (344%) ^a ; ↑ IL-6 mRNA (250%) ^a ↑ TNF-α monocyte expression (87%) ^a ; ↑ TNF-α mRNA (100%) ^a
Consecutive phases study	37.6 ± 9.8	2: 1 night of 4-h bedtime ^e	Daytime (08:00–23:00), every 3 to 4 h	
NA	<30 kg/m ²			
No exercise	2 weeks regular bedtimes ^c			

Continued

Table 1. Continued

Study design	Gender/sample size			
Energy intake	Age \pm SEM (years)		Techniques used for	Main findings (sleep
Behavioral activity	Mean weight \pm SEM	Experimental sleep	blood sampling	deprived vs. longer
	Prestudy conditions	protocol	(time, frequency)	sleep)
Jung <i>et al.</i> ⁴³	2 females, 5 males ($n = 7$)	1: 2 nights of 8-h	Indirect calorimetry	\uparrow total EE (7%); \uparrow
Consecutive phases	22.4 \pm 4.8	bedtimes ^e		nocturnal EE (32%);
study	22.9 \pm 2.4 kg/m ²	2: 40 h of TSD		\sim daytime EE
Controlled isocaloric				
Sedentary	1 week 8-h sleep ^{c,d} ; 3 days diet			\sim RQ
Kuhn <i>et al.</i> ⁸⁷	Human subjects ($n = 28$)	1: 4–5 days “control	OGTT	\downarrow glucose tolerance after 3–4
Consecutive phases	20–30	period”	Once daily	days
study	NA	2: 72–126 h of TSD		\uparrow 17-OH corticosteroids on
Balanced diet		3: 3 days “control		Day 2 and 3, then return
NA		period”		to baseline
				\uparrow urinary catecholamines
Mullington <i>et al.</i> ⁹⁴	Men ($n = 10$)	1. 3 nights 8-h	120 h, every 90 min	\sim leptin mesor, \downarrow leptin
Consecutive phases	27.2	bedtimes ^e		circadian amplitude
study	26.1 \pm 1.9 kg/m ²	2. 3 nights of TSD		
Controlled isocaloric	1 week of regular			
(finishing not	bedtimes ^{c,d}			
required)				
Sedentary				
Nedeltcheva <i>et al.</i> ¹⁰⁹	5 females, 6 males	1: 14 nights of 8.5-h	OGTT; ivGTT 24 h,	\downarrow insulin sensitivity (18%) ^a
Kessler <i>et al.</i> ¹¹⁶	($n = 11$)	bedtimes ^e	every 15–30 min	\downarrow TSH (7%; especially in
Randomized	39 \pm 5	2: 14 nights of 5.5-h		females) ^a
cross-over study	26.5 \pm 1.5 kg/m ²	bedtimes ^e		\downarrow free T4 (8%) ^a
<i>Ad libitum</i>	NA	Conditions \geq 3		\sim GH; \downarrow during first 4-h of
Sedentary		months apart		bedtime period in males
				(31%) ^a
				\sim cortisol; \downarrow and 44 min
				later acrophase (10%) ^a ;
				71 min later nadir
				\uparrow EPI (21%) ^a
				\sim mean NE; \uparrow nocturnal
				(24%) ^a
Nedeltcheva <i>et al.</i> ¹¹⁸	3 females, 7 males	1: 14 nights of 8.5-h	Indirect calorimetry;	\sim total EE; \downarrow RMR (8%) ^a ; \uparrow
Randomized	($n = 10$)	bedtimes ^e	VAS hunger rating	fasting and
cross-over study	41 \pm 5	2: 14 nights of 5.5-h	scale before meal/at	postprandial RQ (4%) ^a
Controlled restricted	27.4 \pm 2 kg/m ²	bedtimes ^e	22:30, 24 h, every	\sim thyroid hormones
(90% RMR)	NA	Conditions \geq 3	hour	\sim GH
Sedentary		months apart		\sim cortisol
				\downarrow EPI (12%) ^a ; \sim NE
				\sim food intake; \uparrow hunger
				\sim total weight loss (3 kg), \downarrow
				fat weight loss (55%), \uparrow
				fat-free weight loss (60%)
				\uparrow mean acylated ghrelin
				(7%) ^a
				\sim leptin

Continued

Table 1. Continued

Study design	Gender/sample size			
Energy intake	Age ± SEM (years)		Techniques used for	Main findings (sleep
Behavioral activity	Mean weight ± SEM	Experimental sleep	blood sampling	deprived vs. longer
	Prestudy conditions	protocol	(time, frequency)	sleep)
Omisade <i>et al.</i> ¹¹⁷	Women (<i>n</i> = 15)	1: 1 night of 10-h	VAS hunger ratings;	↓ median morning cortisol
Consecutive phases	21.6 ± 2.2	bedtime ^d	salivary cortisol	(19%) ^a ; slower decline; ↑
study	24.5 ± 8 kg/m ²	2: 1 night of 3-h	daytime, every 1–2	afternoon/evening
Controlled	1 week regular bedtimes	bedtime ^d	h; salivary leptin	(44%) ^a
Free inpatient	and diet ^{c,d}		morning and	~ hunger
movement			afternoon	↑ morning leptin (8%) ^a
Parker <i>et al.</i> ⁸⁸	Men (<i>n</i> = 4)	1: 1 night of 8-h	72 h, every 30 min	↑ TSH; ↑ amplitude, longer
Consecutive phases	21–30	bedtime ^e		peak, later
study	NA	2: 2 nights of TSD		nadir/acrophase
Controlled	NA			
Sedentary				
Pejovic <i>et al.</i> ⁹⁹	11 females, 10 males	1: 4 nights of 8-h	VAS hunger ratings	~ cortisol
Consecutive phases	(<i>n</i> = 21)	bedtimes ^e	24 h, every 30 min	~ adiponectin
study	24.1 ± 3.1	2: 1 night of TSD, 50%		~ hunger
<i>Ad libitum</i>	24.1 ± 2.6 kg/m ²	had 2-h afternoon		↑ leptin (14%) ^a
Ambulatory	2 weeks regular	nap		
	bedtimes ^d			
Schmid <i>et al.</i> ⁹⁷	Men (<i>n</i> = 10)	1: 1 night of 8-h	Hypoglycemic clamp,	↓ glucagon levels (16%) ^a ; ↑
Randomized	25.3 ± 1.4	bedtime ^e	Two morning samples	response to hypoglycemia
cross-over study	23.8 ± 0.5 kg/m ²	2: 1 night of TSD		↓ cortisol levels (16%) ^a ;
None	Light dinner before	Conditions ≥ 2 weeks		~ ACTH
Bed rest	arrival in the lab	apart		~ catecholamines
	(21:00)			↑ hunger
Schmid <i>et al.</i> ⁹⁸	Men (<i>n</i> = 9)	1: 1 night of 7-h,	Hunger graded 0–9,	↑ hunger (129%) ^a
Randomized	24.2 ± 1.0	30-min bedtime ^e	Two morning samples	~ leptin
cross-over study	23.8 ± 0.6 kg/m ²	2: 1 night of 5-h		↑ total ghrelin (22%)
None	NA	bedtime ^e		
Bed rest		3: 1 night of TSD		
		Conditions ≥ 2 weeks		
		apart		
Schmid <i>et al.</i> ¹¹⁰	Men (<i>n</i> = 15)	1: 2 nights of 8-h	Actigraphy, buffet on	↑ insulin (40%) and glucose
Schmid <i>et al.</i> ¹¹⁹	27.1 ± 1.3	bedtimes ^e	day 2, ELISA (IL-6)	(11%) peak response to
Randomized	22.9 ± 0.3 kg/m ²	2: 2 nights of 4-h	08:00–23:00 (every h)	breakfast
cross-over study	4 weeks regular	bedtimes ^e		↓ physical activity (13%) ^a
<i>Ad libitum</i>	bedtimes ^c	Conditions ≥ 6 weeks		~ cortisol; ~ ACTH
Free outpatient		apart		~ total energy intake; ↑ fat
movement				intake (23%) ^a
				~ hunger; ~ intake
				~ leptin; ~ ghrelin
				~ IL-6
Schmid <i>et al.</i> ¹¹¹	Men (<i>n</i> = 10)	1: 1 night of 7-h	Hypoglycemic clamp	~ insulin resistance
Randomized	25.3 ± 1.4	bedtime ^e	2 morning samples	↓ glucagon (8%) ^b
cross-over study	23.8 ± 0.5 kg/m ²	2: 1 night of 4.5-h		~ GH
None	2 weeks regular	bedtime ^e		↓ ACTH (44%) ^a ; ↓ cortisol
Bed rest	bedtimes ^c	Conditions ≥ 2 weeks		(44%) ^a
		apart		~ EPI; ~ NE

Continued

Table 1. Continued

Study design	Gender/sample size			
Energy intake	Age \pm SEM (years)		Techniques used for	Main findings (sleep
Behavioral activity	Mean weight \pm SEM	Experimental sleep	blood sampling	deprived vs. longer
	Prestudy conditions	protocol	(time, frequency)	sleep)
Shearer <i>et al.</i> ⁹³	Men ($n = 42$)	($n = 21$) 1: 3 nights of	5 days, every 6 h	No changes in condition 1;
Comparative study	28.7 (21–47)	8-h bedtimes, then	(ELISA)	in condition 2: \uparrow IL-6
Controlled isocaloric	NA	4 nights of 2-h		\sim TNF- α
No exercise	2 weeks regular	bedtimes		\uparrow sTNF- α receptor I
	bedtimes ^d	($n = 21$) 2: 3 nights of		\sim sTNF- α receptor II
		8-h bedtimes, then		
		4 nights of TSD		
Simpson <i>et al.</i> ¹²⁰	67 females, 69 males	1: 2 nights of 10-h	Once, morning	\uparrow leptin (33%) ^a
Consecutive phases	($n = 136$)	bedtimes	(10:30–12:00)	
study	30.4 (22–45)	2: 5 nights of 4-h		
<i>Ad libitum</i>	24.7, 17.7–32.6 kg/m ²	bedtimes		
Free inpatient	1 week regular			
movement	bedtimes ^{c,d}			
Simpson <i>et al.</i> ¹²¹	33 females, 41 males	1: 2 nights of 10-h	Once, morning	\sim adiponectin in men; \uparrow in
Consecutive phases	($n = 74$)	bedtimes ^e	(10:30–12:00)	African American
study	29.9, 22–45	2: 5 nights of 4-h		women; \downarrow in Caucasian
<i>Ad libitum</i>	24.6, 17.7–33.1 kg/m ²	bedtimes ^e		women
Free inpatient	1 week regular			
movement	bedtimes ^{c,d}			
Spiegel <i>et al.</i> ¹⁰¹	Men ($n = 11$)	1: 6 nights of 4-h	ivGTT; 24-h heart rate	\downarrow glucose clearance (30%)
Spiegel <i>et al.</i> ¹⁰²	22 \pm 1	bedtimes ^e	variability; salivary	\downarrow TSH (26%), \uparrow fT4 (7%) ^a
Spiegel <i>et al.</i> ¹⁰³	23.4 \pm 0.5 kg/m ²	2: 6 nights of 12-h	sampling	\sim GH, different profile
Spiegel <i>et al.</i> ¹⁰⁴	\geq 1 week of 8-h	bedtimes ^e	15:00-bedtime,	\uparrow afternoon saliva/plasma
Consecutive phases	bedtimes, diet ^{d,e}		every 30 min; blood	cortisol (25%) ^a
study			sampling 24 h,	\uparrow sympathovagal balance
Controlled			every	(17%) ^a
Free outpatient			30 min	\downarrow leptin (19%); \downarrow and 2.5 h
movement				earlier acrophase (26%);
				\downarrow amplitude (20%)
				\downarrow melatonin; later onset; \downarrow
				acrophase
Spiegel <i>et al.</i> ¹⁰⁵	Men ($n = 12$)	1: 2 nights of 10 h	VAS hunger ratings	\uparrow hunger (23%; 33–45% for
Randomized	22 \pm 2	bedtimes ^e	08:00–21:00, every 20	sweets/salty foods)
cross-over study	23.6 \pm 2.0	2: 2 nights of 4-h	min	\downarrow leptin (18%)
Controlled	\geq 1 week of 8-h bedtimes	bedtimes ^e		\uparrow ghrelin (28%)
Free outpatient		Conditions \geq 6 weeks		
movement		apart		
Stamatakis <i>et al.</i> ¹²³	2 females, 9 males	1: 1 night of	ivGTT; heart rate	\uparrow insulin resistance (25%)
Consecutive phases	($n = 11$)	unfragmented	variability;	\uparrow morning cortisol (13%)
study	23.2 (18–29)	sleep ^e	enzyme-linked	\sim adiponectin
<i>Ad libitum</i>	24.3 \pm 0.9 kg/m ²	2: 2 nights of	immunosorbent	\uparrow sympathetic tone (17%)
NA	3 days diet and $>$ 7-h	fragmented sleep	assay techniques	\sim leptin
	bedtimes ^{a,b}	(\sim 31.4 audi-	(IL-6; hsCRP)	\sim IL-6; \sim hsCRP
		tory/mechanical	2 morning samples	
		stimuli per hour) ^e	(08:00 and 16:00)	

Continued

Table 1. Continued

Study design	Gender/sample size			
Energy intake	Age ± SEM (years)		Techniques used for	Main findings (sleep
Behavioral activity	Mean weight ± SEM	Experimental sleep	blood sampling	deprived vs. longer
	Prestudy conditions	protocol	(time, frequency)	sleep)
St-Onge <i>et al.</i> ¹²²	15 females; 15 males	1: 5 nights of 9-h	Double-labeled water	~ RMR; ~ EE; ~ physical
Randomized	(<i>n</i> = 30)	bedtimes ^e	(EE); indirect	activity
cross-over study	33.9 ± 4.3; 36.6 ± 5.6	2: 5 nights of 4-h	calorimetry (RMR);	~ hunger; ↑ food intake
Controlled isocaloric,	23.0 ± 1.1;	bedtimes ^e	actigraphy; VAS	(12%) ^a ; especially ↑
self-selected on	24.1 ± 1.1 kg/m ²	Conditions ≥ 4 weeks	hunger ratings	saturated fat intake in
day 5	2 weeks regular	apart		females (62%)
Free inpatient	bedtimes ^{c,d}			
movement				
Tasali <i>et al.</i> ¹²³	4 females, 5 males (<i>n</i> = 9)	1: 2 nights of	ivGTT; heart rate	↑ insulin resistance (25%)
Randomized	20–31	undisturbed sleep ^e	variability	~ cortisol; ~ daytime; ~
cross-over study	64.2 kg; 19–24 kg/m ²	2: 3 nights of	24 h, every 20 min	nocturnal
Controlled	NA	disturbed sleep by		↑ sympathovagal balance
Sedentary activities	1 week regular bedtimes ^d	acoustic stimuli		(14%)
		during SWS (~33		
		micro-arousals/h) ^e		
		Conditions ≥ 4 weeks		
		apart		
Thomas <i>et al.</i> ⁹¹	Men (<i>n</i> = 17)	1: 3 nights of 7-h	¹⁸ F-DG and PET,	↓ global cerebral glucose
Consecutive phases	24.7 ± 2.8	45-min bedtimes ^e	every 24 h	metabolic rate (8%),
study	NA	2: 85 h of TSD (only		especially the thalamus,
NA	≥ 7 days of 7-h 45 min	results from 24 h		prefrontal and posterior
NA	bedtimes ^d	shown)		parietal cortices
Van Cauter <i>et al.</i> ⁴⁵	Men (<i>n</i> = 8)	1: 2 nights of 8-h	Continuous glucose	↓ nocturnal GH pulses
Consecutive phases	22–27	bedtimes ^e	infusion during	(70%) ^a
study	22.7 ± 0.7 kg/m ²	2: 28 h of TSD	phase 2	~ cortisol, ↓ acrophase
Only breakfast on	1 week 23:00–07:00		24 h, every 30 min	(4%); ¹
study day	bedtimes ^c			1 hour earlier nadir
Bed rest				
VanHelder <i>et al.</i> ⁸⁹	Men (<i>n</i> = 10)	60 h of TSD followed	OGTT at 10 h, 60 h,	↑ insulin response (21% in
Consecutive phases	22 ± 3	by 7-h of recovery	and after recovery	group 1)
study Isocaloric	74.5 ± 11.8 kg	sleep ^e	sleep	
controlled	1 night of 7-h bedtimes ^e	1: sedentary activities		
Sedentary and exercise		2: daily exercise		
		Conditions ≥ 10 days		
		apart		
Van Leeuwen <i>et al.</i> ¹¹²	Men (<i>n</i> = 13)	1: 2 nights of 8-h	Flow cytometry, real	~ cortisol
Consecutive phases	23.1 ± 2.5	bedtimes ^e	time PCR (1	↑ IL-6 (63%)
study	NA	2: 5 nights of 4-h	morning sample),	↑ IL-1β (37%)
Isocaloric controlled	2 weeks regular	bedtimes ^e	salivary cortisol (10	~ TNF-α
Free outpatient	bedtimes ^c		times a day)	↑ hsCRP (45%)
movement				

Continued

Table 1. Continued

Study design	Gender/sample size			
Energy intake	Age \pm SEM (years)		Techniques used for	Main findings (sleep
Behavioral activity	Mean weight \pm SEM	Experimental sleep	blood sampling	deprived vs. longer
	Prestudy conditions	protocol	(time, frequency)	sleep)
Vgontas <i>et al.</i> ¹⁰⁶	13 females, 12 males	1: 4 nights of 8-h	24 h, every 30 min	\sim cortisol; \downarrow and 2 h earlier
Consecutive phases	($n = 25$)	bedtimes ^e	(ELISA, TNF- α ,	peak
study	25.2 \pm 3.8	2: 8 nights of 6-h	and IL-6)	\uparrow IL-6
Usual diet	23.8 \pm 2.3 kg/m ²	bedtimes ^e		\uparrow TNF- α in males; \sim in
Free outpatient	2 weeks regular			females
movement	bedtimes ^d			
(sedentary during				
sampling)				

NOTE: Sleep curtailment was verified by ^cquestionnaires/diaries, ^dby actigraphy, or ^eby polysomnography. The percentage of change in metabolic parameters was given in the article or calculated from ^aabsolute levels in the text/tables or from ^bgraph values of the original article. Metabolic measurements are in plasma/serum, unless otherwise specified. Bed rest was always required during the sleep-deprived state. \sim similar; TSD: total sleep deprivation; VAS: visual analogue scale; RMR: resting metabolic rate; IL-6: interleukin 6; IL-1 β : interleukin 1 β ; TNF- α : tumor necrosis factor- α ; EE: energy expenditure; ivGTT: intravenous glucose tolerance test; RQ: respiratory quotient; OGTT: oral glucose tolerance test; REM: rapid-eye movement; GH: growth hormone; ACTH: adrenocorticotrophic hormone; FDG: fludeoxyglucose; PET: position emission tomography.

fragmented sleep,¹²³ or total sleep deprivation⁹⁰ (Table 1). However, increased insulin secretion in response to ivGTTs was found after 60 hours of sleep deprivation⁸⁹ and two nights of sleep fragmentation.¹²⁴ Thus, almost all studies find decreased glucose tolerance and a dampened compensatory insulin response after sleep curtailment.

Effects on EE

Two groups found that one night of total sleep deprivation increased EE during the night, whereas the following day, resting and postprandial EE were reduced by 5% and 20%, respectively, overall leading to a positive energy balance^{43,100} (Fig. 3 and Table 1). Similarly, Schmid *et al.* reported that volunteers had decreased spontaneous physical activity after a night of short sleep compared to a night with restful sleep¹¹⁰ (Table 1). Fragmenting sleep for two consecutive nights lead to an increase in the respiratory quotient (RQ), indicating a shift from fat toward carbohydrate oxidation, without affecting total EE¹²⁵ (Table 1). High RQ predicts weight gain and insulin resistance.¹²⁶ Two weeks of partial sleep deprivation (5.5 hours) decreased resting metabolic rate and increased RQ in a randomized, cross-over study¹⁰⁹ (Table 1). In contrast, subjects with chronic insomnia had increased metabolic rate.¹²⁷ We have recently shown that poor sleep quality was associ-

ated with higher REE, a higher RQ, and an activation of the stress system in obese subjects with short sleep duration.¹²⁸ In summary, EE may be differentially affected by chronic versus acute sleep curtailment and may also depend on differences in reasons of sleep curtailment (experimentally induced short sleep vs. chronic insomnia).

Additional endocrine mechanisms linking sleep, glucose metabolism, and energy expenditure

As mentioned above, thyroid hormones, GH, cortisol, adipokines (e.g., adiponectin), and sympathovagal balance modulate glucose metabolism and energy homeostasis (Fig. 3). Acute total sleep deprivation increased TSH and free T₄ levels,⁸⁸ whereas partial sleep deprivation of 5.5-h nights for two weeks or six 4-h nights decreased TSH and T₄ levels, suggesting a dose-related effect^{104,116} (Table 1). Decreased peripheral thyroid hormone levels are likely secondary to suppression of the nocturnal TSH peak, possibly secondary to decreased drive from TRH neurons in the PVN.¹¹⁶ Sleep-deprived rodents show reduced levels of hypothalamic TRH mRNA in the PVN of the hypothalamus.¹²⁹ Of note, a different stressor such as acute immobilization also decreases TRH mRNA hypothalamic levels and plasma TSH levels in rodents.¹³⁰

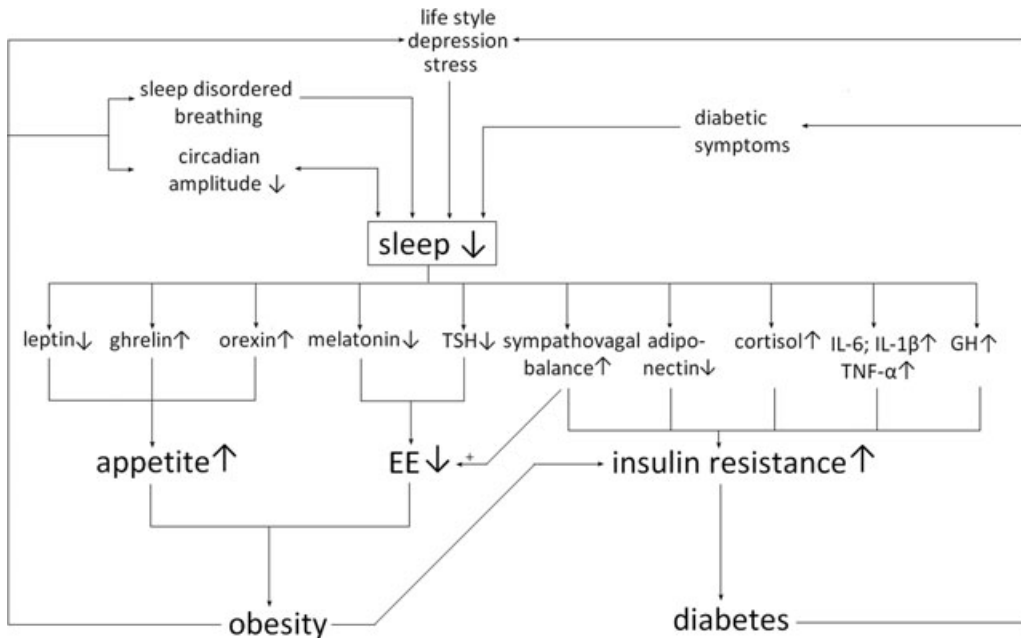


Figure 3. A simplified schematic representation of putative pathways of sleep curtailment leading to obesity and diabetes, via endocrine mechanisms that stimulate appetite, decrease energy expenditure, and increase insulin resistance. The direction of change (an increase or decrease) of each mechanism due to sleep loss is displayed; increased sympathovagal balance could stimulate EE, depicted by +, while a decreased EE would contribute to obesity. For readability, the relationships between decreases in leptin levels associated with IL-6 and TNF- α production are not displayed. Cortisol and proinflammatory cytokines display a positive bidirectional relationship. Increased insulin levels stimulate orexin secretion and orexic arcuate neurons, while decreasing the activity of anorexigenic arcuate neurons, leading to a further increase in appetite. Adiponectin influences energy homeostasis, possibly via modulating appetite through arcuate neurons and modulating EE.

Insulin resistance can result from increases in insulin counter-regulatory hormones, including GH and cortisol (Fig. 3). One night of total sleep deprivation markedly decreased the GH peak that usually occurs around sleep onset^{45,95} (Table 1). Partial sleep deprivation did not affect 24-h levels of GH in lean or obese individuals,^{101,109,118} but six 4-h nights changed GH nocturnal secretory patterns: sleep deprivation was associated with a biphasic pattern compared to the usual single large GH peak at sleep onset.¹⁰² This extended exposure to GH may decrease glucose uptake in muscles, thus contributing to insulin resistance. However, the threshold of GH levels contributing to insulin resistance is unknown. In contrast to the previous study, decreased GH levels during the first four hours of sleep were reported after a two-week partial sleep-deprivation period, in association with more SWS.^{109,118} GH is usually secreted during SWS, thus this unexpected finding may be due to a dissociation of GH secretion from SWS. In summary, the

effects of sleep deprivation on GH plasma levels are variable.

Partial and total sleep deprivation resulted in elevated salivary and plasma cortisol the following afternoon and evening^{100,101,109,114,117} (Table 1), while morning cortisol levels may be unchanged or even decreased.^{90,97,117} As awakening is usually associated with a cortisol surge, this finding probably reflects the lack of the awakening response. In addition, a delay and/or a reduction in the acrophase of cortisol have been reported after partial and total sleep deprivation.^{95,102,109} In a sleep-deprived state, cortisol levels decreased more slowly after reaching their acrophase, possibly indicating decreased sensitivity to the negative feedback effects on the hypothalamic–pituitary–adrenal axis. Mean 24-h levels of plasma cortisol often remain unchanged by sleep deprivation,^{43,95,96,99,106,109,123} but the circadian cortisol-pattern changes, as described above.

Adiponectin levels were similar after partial and total sleep deprivation and sleep

fragmentation^{99,121,124} (Table 1). However, subgroup analysis revealed that effects of sleep restriction on adiponectin levels may be gender and race dependent: adiponectin decreased in Caucasian women and increased in African American women, while adiponectin levels in males did not change after short sleep¹²¹ (Table 1). Besides influencing energy homeostasis, decreased adiponectin levels worsen insulin resistance.

Sympathovagal balance, defined as the ratio between the activity of the sympathetic and the parasympathetic nervous system, can be indirectly assessed with heart rate variability analysis. Since the activity of this system is very diverse, on the basis of the different anatomic branches, it is unclear to which extent perturbations in heart-rate variability may be predictive of changes in basal metabolic rate and insulin sensitivity. Furthermore, heart rate, EPI and NE levels are markers of sympathetic activity. Acute sleep deprivation is associated with increased sympathetic activity, decreased parasympathetic tone and therefore, with increased sympathovagal balance^{101,109,123,124} (Table 1). Total EE is usually decreased in acute sleep deprivation; thus, other mechanisms, such as decreased thyroid hormones or adiponectin levels, must prevail over increased sympathovagal tone (Fig. 3). Increased sympathovagal balance also adds to insulin resistance¹³¹ (Fig. 3). Furthermore, activation of the sympathetic nervous system and inhibition of the parasympathetic nerves decreases insulin secretion; thus, changes in the autonomic nervous system could explain the absence of a compensatory insulin response after sleep curtailment.¹³²

In summary, sleep loss impairs glucose metabolism and may decrease EE. This is mediated by multiple factors, such as decreased thyroid hormones and, possibly, adiponectin levels; increased sympathovagal balance; and altered patterns of cortisol secretion, while the role of GH is still unclear (Fig. 3).

Effects on appetite

It is not clear to what extent obesity in subjects with short sleep is caused by increased energy intake versus decreased EE¹³³ (Fig. 3 and Table 1). If food is provided *ad libitum*, partial or total sleep deprivation may not change hunger ratings, while likely increasing food intake.^{99,117,120} On the other hand, when food intake is controlled, sleep deprivation

usually increases appetite. Self-reported sleep quality was inversely related to appetite in 53 first-degree, normal weight relatives of subjects with diabetes.¹³⁴ After two 4-h nights, there was a 24% increase in daytime appetite ratings;¹⁰⁵ in addition, sleep deprivation specifically increased appetite for sweet and salty foods. The effects of sleep deprivation on appetite may not be immediate: one 4-h night of sleep deprivation was associated with increases in food intake 36 h later, while this parameter was unaffected immediately following sleep deprivation.¹¹³ In addition, the amount of sleep deprivation needed to increase hunger ratings may be variable. One night of total, but not partial, sleep deprivation, significantly increased hunger ratings in normal weight males.^{98,113} No differences in total food intake were observed after two 4-h, 15 min- versus 8-h, 15 min- nights, but subjects ate relatively more fat when sleeping less.¹¹⁹ In a randomized, cross-over, five-day study of 4 h versus 9 h of sleep, with a controlled diet on days 1–4 versus an *ad libitum* diet on day five, women, but not men, exhibited increased total caloric intake and a preference for fatty foods, especially if saturated, suggesting that women may be more susceptible to the orexic effects of sleep deprivation than men.¹²²

Chronotype, the individual attribute determining morning or evening preference, modulates metabolism. Obese subjects carrying the CLOCK 3111TC/CC polymorphism, a variant of one of the core clock genes associated with being an “owl” or an evening person, sleep 20 minutes less.¹³⁵ In addition, these subjects had a higher intake of trans-fatty acids and proteins, and consumed relatively later in the day, while having the same total energy intake of noncarriers. Subjects carrying the CLOCK 3111TC/CC genotype are also more resistant to weight loss: they lost about 2 kg less while on a hypocaloric diet for 30 weeks.¹³⁵

Leptin and ghrelin inhibit and stimulate appetite, respectively (Fig. 3). Following partial sleep deprivation, mean daytime values of leptin were approximately 18% lower compared to the rested state.^{104,105} In addition, the circadian amplitude of leptin was dampened and the acrophase was decreased and phase advanced by two hours. A decreased circadian amplitude was also observed after three nights of total sleep deprivation.⁹⁴ In a large cohort study of 1,024 subjects, shorter habitual sleep duration as assessed by sleep diaries related

to lower morning values of leptin.⁵ Some studies found no association between short sleep and leptin levels.^{94,98,100,119,136} On the contrary, morning leptin levels were found to be increased after five nights of four hours,¹²⁰ after one night of three hours¹¹⁷ and after one night of total sleep deprivation.⁹⁹ Short sleep duration, as assessed by one night of polysomnography in 561 subjects, also related to higher morning leptin levels.¹³⁷ It is difficult to explain the inconsistent results of these different studies. Of note, studies that found decreased leptin after short sleep controlled food intake, whereas when sleep-deprived subjects were allowed to eat *ad libitum*, leptin levels were unchanged or even increased.

In obese subjects, leptin levels are elevated but several of its actions are impaired, a concept referred to as *leptin resistance*. In a cross-over study design, no association among habitual sleep duration, measured actigraphically, and morning leptin levels was observed in obese subjects.¹³⁸ Mean 24-h leptin levels in obese subjects on a controlled diet were also unchanged after 14 nights of 5.5 h versus 8.5 hours.¹¹⁸

The physiological increase in total ghrelin levels in the early night was blunted when individuals were kept awake.⁹⁵ A similar phenomenon is reported in chronic insomniacs.¹³⁵ However, morning total ghrelin levels were increased after one night of sleep deprivation^{95,98} and in individuals with shorter habitual sleep durations.⁵ Mean daytime total ghrelin levels were higher after two nights of 4-h sleep.¹⁰⁵ CLOCK 3111TC/CC carriers, who display an evening preference and sleep 20 min shorter, also had increased morning ghrelin levels.¹³⁴ Mean 24-h levels of the metabolically active acetylated ghrelin were higher in obese subjects after two weeks of 5.5-h nights.¹¹⁸ Overall, sleep curtailment is consistently associated with higher daytime ghrelin.

The effect of sleep deprivation on the orexin system in normal subjects is not well studied. However, it is known that patients with narcolepsy have abnormally low orexin-A CSF levels, which correlate both with body weight and sleep abnormality.¹³⁹ Total sleep deprivation increased orexin levels in the CSF of squirrel monkeys, and increased cFos expression in orexin neurons in rats.^{81,140} It is challenging to assess the activity of the orexin system in humans. Central determination requires collection of CSF,

an invasive technique. Determination of orexin in plasma is hampered by the lack of adequate assays, in terms of sensitivity and specificity. Hyperactivity of the orexin system could contribute to increased food intake during sleep deprivation. In summary, appetite and food intake are often increased following sleep curtailment, possibly via decreased leptin, increased ghrelin and a hyperactive orexin system (Fig. 3).

Effects on the immune system

Enhanced levels of circulating leucocytes, pro-inflammatory cytokines (e.g., IL-6, TNF- α , IL-1 β) and CRP have been associated with an increased risk of cardiovascular disease and type 2 diabetes in long-term human and experimental studies.^{141–144} Increased plasma levels of IL-6 were reported after partial and total sleep deprivation in most cases,^{93,106,107,108,112} although two studies only reported a loss of circadian rhythms^{82,96} (Table 1). IL-1 β was elevated after partial,^{106,112} and total sleep deprivation,⁹⁶ but one study did not find any changes after total sleep deprivation⁸² (Table 1). TNF- α levels are most often,^{82,93,112} but not always,¹⁰⁷ unchanged after partial sleep restriction. One study only found increases in men, but not in women, after partial sleep deprivation¹⁰⁶ (Table 1). Another study reported higher levels of soluble TNF- α receptor I, through which TNF- α exerts its sleep promoting function.⁹³ Effects of sleep curtailment on CRP levels are inconsistent: they have been reported lower,⁹⁶ unchanged,¹²⁴ or higher.^{108,112} Prolonged total sleep deprivation has been associated with leucocytosis⁸² (Table 1). Thus, differences in study outcomes may be influenced by individual or gender differences. In addition, sensitivity and specificity of commercial cytokine assays are highly variable. Highly sensitive analytical methods, such as mass spectrometry, may be needed for accurate cytokine measurements.¹⁴⁵ We conclude that short sleep promotes a proinflammatory state, which, in turn, exerts its negative consequences on insulin resistance.

The other side of the coin: metabolic dysfunction influencing sleep

Most of the studies of sleep deprivation have been performed in healthy, lean subjects and therefore could not address the effect of sleep deprivation on subjects with obesity and diabetes. Obesity and

diabetes are often associated with increased stress or differences in life style that could negatively influence sleep¹⁴⁶ (Fig. 3). Depression and obesity may occur together as well: obesity may negatively affect mood and both pharmacologic antidepressant therapy and major depression itself are associated with weight gain.¹⁴⁷ We recently reported that obese subjects often suffer from sleep-disordered breathing, which induces frequent microawakenings and loss of SWS.¹⁴⁸ Similar effects of BMI on sleep have also been observed in youth. Decreased total sleep time and less sleep efficiency were observed in severely obese adolescents, of which 74% displayed sleep apnea (16.5 years; BMI 60.3 ± 2.1 kg/m²).¹⁴⁹ Changes in sleep architecture in obesity have been reported: severely obese adolescents (12–15 years; BMI 50.9; 39.8–63.0 kg/m², apnea index 14 episodes/h) have a decrease in REM sleep.¹⁵⁰

Symptoms associated with diabetes, such as thirst, nocturia, extreme glucose excursions, and mood alterations, may also interfere with sleep, independent of obesity. To this point, adults with type 1 diabetes, who are typically not obese, report a poorer sleep quality than their matched controls without diabetes.¹⁵¹ These awakenings were associated with a rapid decline of glucose levels, but not with the absolute values of hypoglycemia.¹⁵² Furthermore, studies in animals suggest that diet composition may directly influence sleep: mice fed a high-fat diet display an attenuated rhythm in the circadian expression of core clock genes.⁴⁰ Of note, augmented amplitude of the peripheral and central circadian rhythms favors wakefulness during the day and sleep at night, whereas an attenuated circadian amplitude disrupts the sleep–wake cycle.¹⁵³ Likewise, studies in rodents demonstrate that physical activity at the congruent time (i.e., nocturnal for mice) increases the central circadian amplitude, suggesting that active behavior during the day and sleep at night in humans may be beneficial for high-amplitude circadian rhythms.¹⁵⁴ Thus, sleep and circadian rhythm are bidirectionally connected (Fig. 3). Leptin influences sleep architecture in normal fed rodents, increasing SWS by 13% and decreasing REM by 30%.¹⁵⁵ Furthermore, *ob/ob* mice that lack leptin and *db/db* mice that lack the leptin receptor, both display sleep disturbances that are reverted by leptin replacement. In summary, metabolic dysfunction, altered hormone levels, and sleep abnormalities probably all influence each other.^{156,157}

In summary, sleep deprivation and obesity may potentiate each other in a vicious reverberating circuit; short sleep may induce weight gain, via the mechanisms outlined in this paper, whereas obesity, in turn, via sleep apnea and other symptoms, may disrupt sleep. Observational studies, especially if cross-sectional, may not be well suited to disentangle these aggregate, causative factors. This underlies the need for behavioral, life-style intervention studies.

Possible therapeutic role of melatonin

In nonequatorial areas, the duration, intensity, and spectral quality of natural light vary across seasons.¹⁵⁸ For example, during a summer in England, there was relatively more blue light versus green and red light exposure, in addition to increased light intensity and duration.¹⁵⁸ These are important signals for seasonal rhythms, transmitted to the body via the duration of nocturnal melatonin secretion. As a consequence of artificial lighting and other environmental modifications, seasonality is greatly attenuated in modern humans. Furthermore, because sleep is usually the only time during which humans do not experience light exposure, shorter sleep duration will result in longer light exposure. Light suppresses the pineal production of melatonin in a dose-related fashion. White artificial light greater than 200 lux completely suppressed nocturnal plasma melatonin production, dim room light (106 lux) reduced melatonin by 88% and light intensities lower than 80 lux did not affect melatonin levels.¹⁵⁹ As the night progresses, the intensity of continuous artificial light needed to suppress circulating melatonin increases.¹⁶⁰ Of note, light of shorter wavelength, such as the light that is typically emitted by electronic devices such as televisions and computers, induces relatively more melatonin suppression and likely leads to greater sleep disruption.¹⁶⁰

Urinary 24-h melatonin concentrations decreased by more than 50% when subjects slept five hours versus eight hours, while being continuously exposed to white light (700 lux) during wake hours.¹⁶¹ Diminished levels of melatonin were likely due to three additional hours of light exposure. Likewise, mean 24-h plasma melatonin concentrations were decreased after 6 nights of 4-h versus 12-h bedtimes.¹⁰³ The onset of melatonin secretion was delayed, due to four hours longer light

exposure before bedtime (300 lux) and the acrophase was reduced in sleep-deprived subjects.

Recent studies suggest that melatonin may have a physiological role in modulating metabolism. For example, removal of the pineal gland decreased the responsiveness of several insulin-dependent hepatic kinases during the dark phase in rats.¹⁶² This effect was reversed by nocturnal supplementation of melatonin in the drinking water. Chronic melatonin supplementation in drinking water also reduced body weight in rodents.^{163,164} Patients with metabolic syndrome have disturbances in melatonin production, characterized by an alteration of the night melatonin-insulin ratio of unclear clinical significance.¹⁶⁵ In a large genome-wide association studies conducted in individuals of European origin, certain genetic variants of the melatonin receptor 1B were associated with an increased risk of type 2 diabetes.^{166–168} However, the most common receptor variant did not influence the relationship between sleep abnormalities and type 2 diabetes.¹⁶⁶ Interestingly, the melatonin receptor 1B is present on pancreatic β -cells, and melatonin has an inhibitory effect on insulin secretion. Thus, it was feared that treatment with melatonin in subjects with diabetes would increase blood glucose levels. On the other hand, since type 2 diabetes is associated with reduced melatonin levels toward the end of the night;¹⁶⁹ thus, there may be a physiological rationale for melatonin supplementation at bedtime. Few studies have addressed these questions. Sleep efficiency improved after five months of treatment with 2 mg prolonged-release melatonin administered two hours before bedtime, while total sleep time was unaffected in 26 subjects with type 2 diabetes.¹⁷⁰ HbA1c levels also improved (8.47% vs. 9.13% at baseline). Lack of a control group prevented understanding whether improved HbA1c levels were in part due to other factors than melatonin administration, such as better pharmacological and diet compliance due to study participation. In lean subjects, melatonin administration at night for 2 months was associated with decreased LDL levels and improved oxidative status.¹⁷¹ Another study found a positive correlation between plasma melatonin levels at 2 am and HDL levels in 36 women treated with 1 mg melatonin for one month.¹⁷² As melatonin promotes sleep, the decrease in obesity and improvement in insulin resistance could be due to longer and better sleep per se, but it could also

relate to other melatonin-dependent pathways yet to be identified (Fig. 3).

Discussion and future directions

Epidemiological and laboratory data prove the existence of a relationship between short or disturbed sleep and adverse metabolic outcomes. In general, these effects are small in size, but certain specific populations may be more susceptible than others. For example, age plays a clear role as the relationship between short sleep and obesity is stronger in children. Sleep duration and sleep architecture change with age in healthy individuals, with SWS becoming less common with age.¹⁷³ It is also well known that sex and reproductive hormones affects sleep regulatory mechanisms¹⁷⁴ and that sleep disorder affects women differently from men.¹⁷⁵ It is unfortunate that experimental studies, in both humans and in rodents, are mostly conducted in the male gender, thus unduly discriminating the female gender.

Similarly, sleep deprivation is more common in minorities but the effects of socioeconomic status per se versus ethnicity remain to be determined. Individual susceptibility within these categories varies greatly: identification of contributing factors will greatly help prevention and treatment and be one of the tenets of individualized medicine.¹⁷⁶ Few negative effects are expected to be observed in individuals with voluntary sleep curtailment (natural short sleepers), in contrast to individuals with short sleep due to social or work-related pressure or chronic insomnia. Current epidemiologic studies often lack an objective measurement of sleep duration, such as actigraphy or repeated sleep diaries. Thus, prospective studies with objective sleep duration measurements are warranted in different subgroups to further explore this relationship.

In addition, the causality of the relationship between sleep and metabolic disorders remains unclear, even in prospective studies. Obese/diabetic individuals may be sleeping less than lean subjects because of differences in life style or stress; their sleep could be interrupted more often by symptoms of disturbed glucose levels or sleep apnea; evidence arises that possibly circulating hormones affect the brain inducing worsened sleep (Fig. 3). As already mentioned, causality can be addressed best in long-term studies where “prescribed” sleep duration is randomized, but this may present with practical difficulties and significant challenges,

including the fact that the biological need for optimal sleep duration varies and cannot be estimated at an individual level. Sleep is regulated by circadian and homeostatic mechanisms. As far as homeostatic mechanisms are concerned, it has been the matter of a long scientific debate whether during wakefulness, single or rather multiple metabolites accumulate, progressively increasing the pressure to sleep during the day. Classic experiments of parabiosis seem to support the notion of endogenous “sleep-promoting” substances. Recently, the importance of adenosinergic neurotransmission in the regulation of sleep, especially non-REM sleep, has been underlined.¹⁷⁷ Nevertheless, it is certain that there is a large individual variability in sleep needs. Sensitive and specific biomarkers of sleep deprivation are needed for various reasons, including clinical and medico-legal reasons.¹⁷⁸

The effect of sleep extension in subsets of short sleepers on metabolism can be addressed in randomized controlled trials. Currently, we are conducting such a study in short-sleeping obese individuals.¹⁷⁹ Investigating the effects of sleep extension in other subgroups of varying age, BMI, and ethnicity will also be of interest. The effect of metabolic disturbances on sleep, such as hyperinsulinemia or increased circulating levels of proinflammatory cytokines, may be empirically addressed in polysomnographical studies. As it is difficult to artificially mimic the internal milieu of obese/diabetic individuals, effects of metabolism may be modeled in animal studies using parabiosis, connecting circulations of lean and obese animals and comparing sleep characteristics.

As indicated by studies of sleep deprivation, the mechanisms connecting short sleep and obesity/insulin resistance are probably mediated by three pathways: increases in appetite, decreases in EE, and influences on glucose metabolism (Fig. 3). Endocrine mechanisms influencing these pathways are complex and interconnected. The existing studies of sleep deprivation have generated variable results; small sample size, differences in the control of food-intake and physical activity levels, and inherent individual differences may have all contributed to this variability. In addition, these studies only addressed the effects of acute sleep deprivation and were not performed in real-life situations. We suggest that future studies should focus on the effects of sleep deprivation in specific patient popula-

tions, including obese/insulin-resistant individuals and chronic insomniacs versus natural short sleepers, and should mimic real-life conditions.

Most studies find worsened glucose tolerance after sleep curtailment, which may be beneficial from a teleological perspective, at least in the short term. When our ancestors remained awake during the night, they were probably experiencing a threat (e.g., absence of food, presence of predators), so inducing a state in which glucose availability was advantageous. Likewise, increases in appetite result in more food intake. Together with reduced EE, this will further increase blood glucose availability. In modern societies, however, prolonged exposure to such a state may result in obesity and type 2 diabetes.

At this point, there is a general consensus that short sleep has negative effects on health, but the amount of sleep curtailment causing metabolic disturbances is unclear, and specific groups at greater risk are yet to be identified. Whether melatonin would benefit obese/diabetic subjects, or a subset of these individuals, remains to be proven in rigorous randomized clinical trials. Supplementation of melatonin has been shown to improve sleep efficiency, and HbA1c levels in patients with type 2 diabetes.¹⁷⁰ Lean subjects may also profit from melatonin administration as well, as it improved lipid profiles.^{171,172} Thus, melatonin appears to be a promising supplement in battling the obesity/diabetes epidemics, but controlled studies need to confirm its positive effect on metabolism. Furthermore, the long-term effects and side effects of chronic melatonin supplementation need to be examined in different subgroups.

Overall, improving sleep duration and quality is a potential tool to counteract the epidemics of obesity and diabetes. However, before sleep extension advice is translated to the clinic, more research is needed.

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Conflicts of interest

The authors declare no conflicts of interest.

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