

Reprogramming IgH isotype-switched B cells to functional-grade induced pluripotent stem cells

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Induced pluripotent stem cells (iPSCs) can be formed from somatic cells by a defined set of genetic factors; however, aberrant epigenetic silencing of the imprinted *Dlk1-Dio3* gene cluster often hinders their developmental potency and ability to contribute to high-grade chimerism in mice. Here, we describe an approach that allows splenic B cells activated to undergo Ig heavy-chain (*IgH*) class-switch recombination (CSR) to be reprogrammed into iPSCs that contribute to high-grade chimerism in mice. Treatment of naïve splenic B cells in culture with anti-CD40 plus IL-4 induces *IgH* CSR from IgM to IgG1 and IgE. CSR leads to irreversible *IgH* locus deletions wherein the IgM-producing C μ exons are permanently excised from the B-cell genome. We find that anti-CD40 plus IL-4-activated B cells produce iPSCs that are uniformly hypermethylated in the imprinted *Dlk1-Dio3* gene cluster and fail to produce chimerism in mice. However, treatment of activated B cells with the methyltransferase inhibitor 5-aza-2'-deoxycytidine before and at early stages of reprogramming attenuates hypermethylation of the *Dlk1-Dio3* locus in resultant iPSCs and enables them to form high-grade chimerism in mice. These conditions allowed us to produce chimeric mice in which all mature B cells were derived entirely from IgG1-expressing B-cell-derived iPSCs. We conclude that culture conditions of activated B cells before and at early stages of reprogramming influence the developmental potency of resultant iPSCs.

Enforced expression of *Oct4*, *Klf4*, *Sox2*, and *c-myc* (OKSM) in somatic cells can result in the formation of induced pluripotent stem cells (iPSCs), which possess characteristics of blastocyst-derived embryonic stem cells (ES cells) (1–4). Although the iPSC label is applied to somatic cell-derived self-renewing cells that display ES cell-like morphology and growth characteristics, further characterization must be done to define iPSC functional developmental potency (5–7). In this context, iPSCs can vary widely in level of functional potency, with only a minority of iPSCs able to support high-grade chimerism and germline contribution in mice (8). Recent reports suggest that iPSCs may harbor epigenetic differences compared with ES cells that retard developmental potency (8). For example, recent work showed that iPSCs with methylation-induced *Dlk1-Dio3* gene cluster silencing contribute poorly to chimerism in mice, whereas iPSCs with an active *Dlk1-Dio3* region produce high-grade chimeras (9). Expression of the *Meg3* gene (also known as *Gtl2*), located within the *Dlk1-Dio3* locus, inversely correlates with the CpG methylation status of this locus and therefore serves as an indirect marker for iPSC functional quality in terms of developmental potency (9). In this context, iPSC clones that express elevated *Meg3* levels (defined by >80% of ES cell levels) referred to as *Meg3*^{on} clones exhibit low CpG methylation and high functional quality, whereas *Meg3*^{off} iPSCs contribute poorly to mouse chimerism in general (9).

Another factor influencing reprogramming potential is the developmental stage of somatic cells, with less-differentiated cells in some cases being more amenable to epigenetic reprogramming (10). In this regard, enforced expression of the four standard OKSM factors can reprogram progenitor B cells more efficiently

than mature naïve (i.e., IgM⁺ IgD⁺) B cells (10). Additionally, iPSCs derived from progenitor B cells through standard OKSM-mediated reprogramming have been shown to more readily contribute to high-grade chimerism compared with iPSCs derived from mature naïve B cells (10, 11). However, reprogramming of B cells representing developmental stages beyond the mature naïve B-cell stage has not been reported previously.

Products of the *Rag1* and *Rag2* genes are required for assembly of component variable (V), diversity (D), and joining (J) gene segments (“VDJ recombination”) to form *IgH* and T-cell receptor (*TCR*) variable region exons (12, 13). Because productive assembly of *IgH* or *TCR* variable region exons is required for the development of B and T cells, respectively, mice deficient in either *Rag1* or *Rag2* have no mature B or T cells due to a block in lymphocyte development at the early progenitor stage owing to inability to undergo V(D)J recombination (14, 15). Upon activation by antigen in peripheral lymphoid organs, mature B cells may undergo *IgH* class-switch recombination (CSR). CSR is a process in which the *IgH* μ constant region exons (C μ) are deleted and replaced by one of several sets of downstream constant region exons (C_{Hs}; e.g., C γ , C ϵ , and C α), thereby allowing for the formation of other *Ig* classes (e.g., IgG, IgE, or IgA). CSR occurs within switch (S) regions, which are 1- to 10-kb sequences located 5' to each set of C_{Hs} (16). The activation-induced cytidine deaminase (AID) enzyme initiates both CSR and the related process of somatic hypermutation (SHM) of *Ig* variable-region exons via cytidine deamination activity. During CSR, AID activity leads to DNA double-strand breaks (DSBs) in a donor S region (S μ) upstream of C μ and in a downstream acceptor S region; these DSBs then are joined so that C μ is irreversibly deleted and replaced with one of the downstream C_{Hs} (17).

Each S region is preceded by a promoter and noncoding exon termed an “I” exon (18). Different forms of activation and/or cytokines provided by helper T cells or other cells can direct AID and, as a result, CSR to a particular target S region by specifically stimulating transcription from upstream I region promoters (16, 18). In this regard, stimulation of cultured splenic IgM⁺ B cells with anti-CD40 plus IL-4, mimicking in vivo activation by T helper type 2 (T_{H2}) T cells, induces B-cell plasmablast differentiation and class switching to IgG1 and IgE (19, 20). Though anti-CD40 plus IL-4 treatment theoretically can lead to direct CSR from C μ to either C γ 1 or C ϵ , direct CSR to C ϵ occurs less frequently than to C γ 1 in mature B cells (20–24). In this regard, various studies have shown that IgE switching largely occurs

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through a sequential CSR mechanism in which activated B cells first switch from IgM to IgG1 via direct CSR between S_{μ} and $S_{\gamma 1}$ followed by switching to IgE via a “second step” recombination via breaking and joining of the newly made hybrid $S_{\mu}/S_{\gamma 1}$ and S_{ϵ} (25–29). IgG1⁺ B-cell intermediates in sequential CSR have been proposed to be a required part of the development of high-affinity IgE responses *in vivo* (29, 30); however, the mechanisms regulating CSR from IgG1 to IgE remain poorly understood, partly because of the difficulty in isolating IgG1⁺ B-cell intermediates. In this context, the ability to reprogram IgG1⁺ B cells into iPSCs could allow the development of stable IgG1⁺ mouse models for studying second-step IgG1-to-IgE switching.

We now describe culture conditions that allow us to generate IgG1⁺ B-cell-derived iPSCs with sufficient developmental potency to produce chimeric animals harboring mature T and B cells derived entirely from IgG1-switched iPSCs and show that these B cells can be activated to switch from IgG1 to IgE.

Results

OKSM-Mediated Reprogramming of Activated B Cells Results in iPSCs Derived from *IgH* Class-Switched B Cells. To develop an approach to reprogram *IgH* class-switched B cells, we used mice harboring a transgenic reprogramming system in which the OKSM reprogramming factors are expressed in a doxycycline-inducible fashion (31). We activated splenic B cells with anti-CD40 and IL-4 for 4 d to induce *IgH* class switching before addition of doxycycline to increase the probability of obtaining iPSC clones from *IgH* class-switched B cells. Cells were then plated on irradiated mouse embryonic fibroblasts (MEFs) before colonies with ES cell-like morphology and growth characteristics became apparent after 10–12 d of doxycycline treatment. We picked 72 colonies with ES cell-like morphology for analysis, and of these, 59 clones grew to be stable doxycycline-independent iPSC clones.

To determine if iPSC clones were indeed derived from B cells that have been activated for CSR, we characterized them by Southern blotting EcoRI-digested DNA with probes that flank both sides of S_{μ} . AID activity in B cells stimulated to undergo CSR induces multiple DSBs within a given S region (32–35), which may be relegated, joined to another DSB in the same S region, or ligated to a downstream S region to affect CSR. Rejoining DSBs within an S region, accompanied by resection or joining two DSBs within the same S region, can cause internal switch deletions (ISDs), which often are large enough to be observed via Southern blotting (33, 36). Therefore, a probe recognizing the 5' end of S_{μ} (i.e., I_{μ} probe), which hybridizes to an EcoRI fragment that contains the entire S_{μ} region (Fig. 1A), will show different-sized restriction fragments compared with germline configuration if the B-cell had undergone either S_{μ} ISD or CSR to a downstream C_H region. A probe 3' to S_{μ} ($I_{\gamma 1}$ probe) hybridizing to a separate EcoRI fragment (Fig. 1A) will be absent in cells that have undergone CSR to a downstream C_H region, because intervening DNA in between S regions that have undergone CSR is excised in the process (18).

We observed that nearly every iPSC clone demonstrated different-sized I_{μ} probe-reactive bands compared with germline configuration, suggesting that these clones were derived from B cells that had expressed AID and had undergone either ISD or CSR events (Fig. 1B and C, Upper). We also observed that $I_{\gamma 1}$ probe-reactive restriction fragments were absent in 37 of 59 clones (Fig. 1B and C, Lower), indicating that at least 63% of these iPSC clones derived from B cells that had undergone CSR. The $I_{\gamma 1}$ probe-reactive bands of sizes differing from germline could signify other AID-mediated recombination events such as $I_{\gamma 1}$ ISD. Another probe hybridizing to C_{ϵ} revealed that at least 11 clones had two clear bands (representing two alleles) diverging from germline configuration, suggesting that some iPSCs derived from B cells had undergone CSR to IgE (Fig. S1).

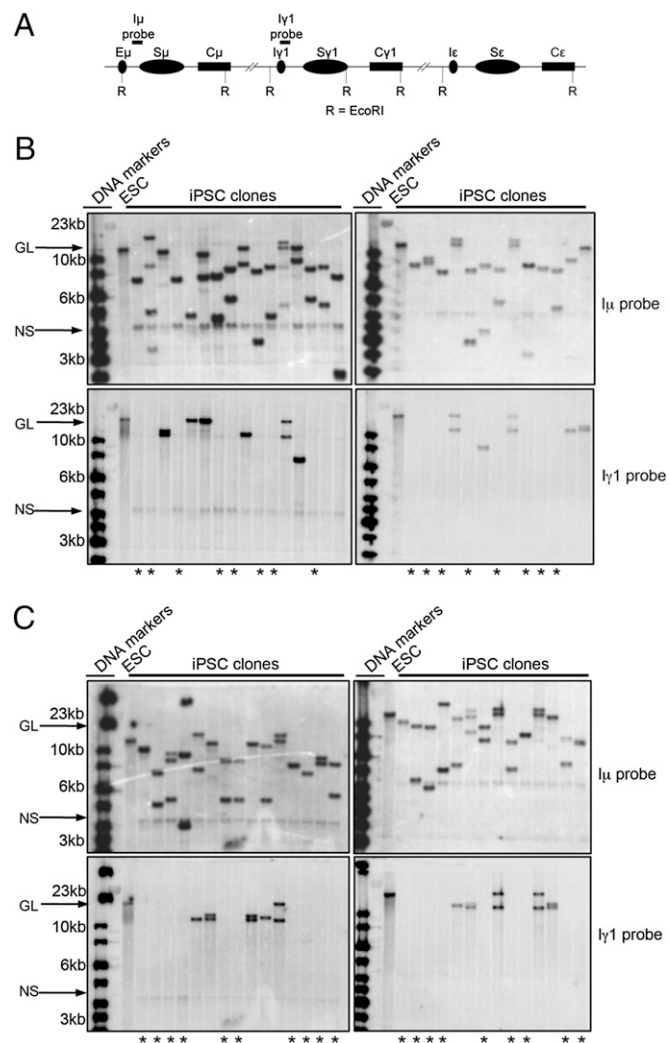


Fig. 1. *IgH* class-switched B cells can be reprogrammed to iPSCs. (A) A schematic of the *IgH* constant region. EcoRI (R) restriction sites and locations of the I_{μ} and $I_{\gamma 1}$ probes are indicated. (B and C) Southern blot analysis of EcoRI-digested genomic DNA extracted from ES cells and iPSC clones reprogrammed from day 4 anti-CD40/IL-4-stimulated B cells hybridized to an I_{μ} (Upper) or $I_{\gamma 1}$ (Lower) probe. Both I_{μ} - and $I_{\gamma 1}$ -hybridizing germline (GL) bands defined by EcoRI-digested ES cell DNA are indicated with an arrow and labeled. Signals likely representing nonspecific (NS) background bands are likewise indicated. The first two lanes contain DNA size markers, and DNA sizes are indicated in kilobases on the left. Lanes containing prominent I_{μ} -hybridizing bands different from GL indicate alleles that have undergone ISD or CSR. Lanes with asterisks (*) show no bands hybridizing to the $I_{\gamma 1}$ probe, indicating that these clones have likely undergone CSR to either $C_{\gamma 1}$ or to other downstream C_H .

5-Aza-2'-Deoxycytidine Treatment of Activated B Cells at Early Stages of Reprogramming Allows for the Development of *Meg3*^{on} iPSCs.

Given the predictive ability of iPSC *Meg3* expression to identify iPSCs of high developmental potency (9), we assessed *Meg3* levels by quantitative PCR. The TC1 and C37 ES cells, which showed nearly identical *Meg3* expression levels, were used to compare with iPSC clones. We observed that all iPSC colonies derived from activated B cells (B-iPSCs) had *Meg3* expression levels well below that of ES cell controls (*Meg3*^{off}; Fig. 2A), which suggests low developmental potency (9). Because *Meg3*^{off} status in general correlates with *Dlk-Dio3* locus hypermethylation and poor iPSC developmental potency (9), we hypothesized that treatment of B cells with the methyltransferase inhibitor 5-aza-2'-deoxycytidine

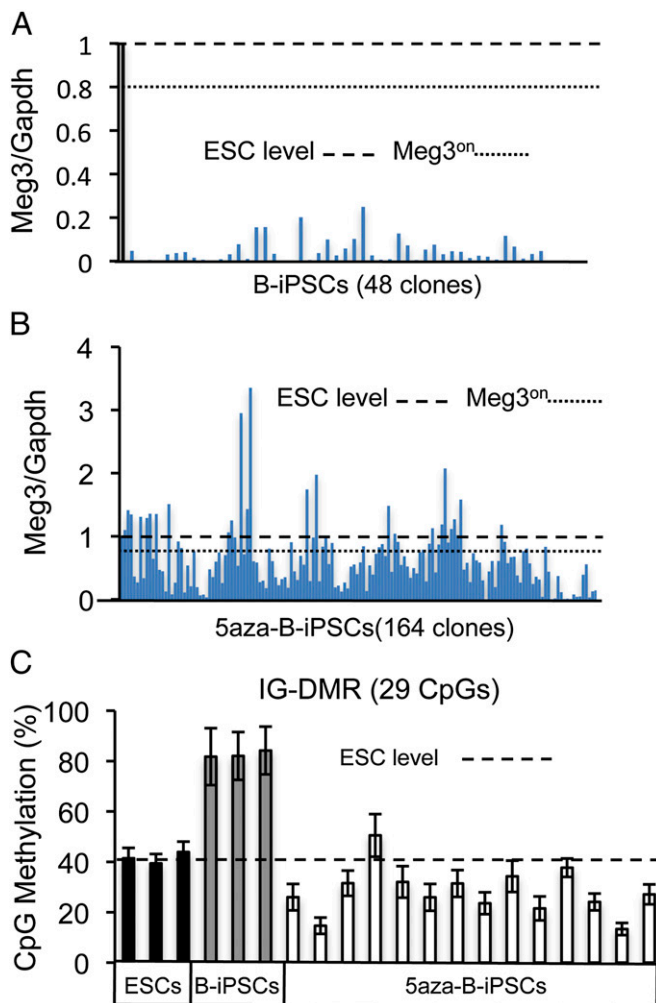


Fig. 2. Treatment of activated B cells before reprogramming with 5aza leads to *Meg3*^{on} B-cell-derived iPSCs with attenuated IG-DMR hypermethylation. (A and B) Quantitative PCR analysis of *Meg3* expression in ES cell and iPSC clones derived from day 4 anti-CD40/IL-4-stimulated B cells normalized to *Gapdh* expression. The iPSCs were derived without 5aza (B-iPSCs) (A) or with a 2-d 5aza treatment at the beginning of reprogramming (5aza-B-iPSCs) (B). Y axis indicates values relative to ES cell *Meg3* expression. The black bar indicates ES cell *Meg3* expression. ES cell expression is also indicated by the dashed horizontal line. The dotted horizontal line indicates 80% ES cell *Meg3* expression, which is the threshold we used to define *Meg3*^{on} status. (C) Degree of CpG DNA methylation at the IG-DMR in ES cell clones (black bars), untreated B-cell-derived iPSCs (gray bars), and 5aza-treated *Meg3*^{on} iPSC clones (white bars) as measured by bisulfite sequencing. Dashed line indicates the average ES cell level of IG-DMR CpG methylation. Error bars indicate SDs of methylation status of the 29 CpGs within the IG-DMR.

(5aza) may give rise to iPSCs with ES cell-like *Meg3* expression levels and *Dlk-Dio3* locus methylation status, perhaps resulting in improved iPSC quality. In this regard, previous attempts using 5aza during late stages of reprogramming have shown increased efficiency of iPSC generation, but these studies did not demonstrate contribution of 5aza-treated iPSCs to chimerism in mice (37, 38). We therefore used a similar 5aza treatment method; but in our approach, we provided a 2-d treatment course of 5aza beginning 1 d before induction of the OKSM reprogramming factors. We rationalized that the toxic effects of a global methyltransferase inhibition may be ameliorated during reprogramming by the reprogramming process itself and/or by allowing several days of selection against potential toxicities associated with genome-wide hypomethylation during the reprogramming process.

We observed that this early 5aza treatment approach given to day 4 anti-CD40 plus IL-4-activated B cells resulted in doxycycline-independent iPSCs (5aza-B-iPSCs) that were morphologically indistinguishable from B-iPSCs (Fig. S2). In terms of *Meg3* expression, *Meg3* qPCR assays showed that 51 of a total of 165 5aza-B-iPSC clones were *Meg3*^{on} (Fig. 2B). Because *Meg3* expression is used here as a surrogate for *Dlk-Dio3* DNA methylation status, we used bisulfite sequencing to measure the DNA methylation within this locus, specifically at the intergenic differentially methylated region (IG-DMR), which has been established as a useful test to assess iPSC potency (9, 39, 40). As expected, we find that our *Meg3*^{on} 5aza-B-iPSC clones showed decreased IG-DMR CpG methylation compared with B-iPSCs (Fig. 2C).

5aza iPSCs Contribute to Mouse Chimerism and Can Make “All-IgG1” Mice in the Context of *Rag2*^{-/-} Blastocyst Complementation. It has previously been shown that iPSCs derived from mature naïve B cells by standard OKSM reprogramming contribute poorly to mouse chimerism (10, 11). It has also been previously shown that *Meg3*^{off} iPSCs in general display reduced developmental potency and contribute poorly to mouse chimerism (9, 39, 40). Given these prior studies, we predicted that our B-iPSCs, which are uniformly *Meg3*^{off} and derived via standard OKSM reprogramming, would display low developmental potency. To test this notion, we evaluated developmental potency of three independent B-iPSC clones in the context of *Rag*-deficient blastocyst complementation (RDBC). RDBC involves injection of selected ES cells into blastocysts from *Rag2*-deficient mice, leading to the generation of somatic chimeras harboring mature T and B cells derived entirely from the injected ES cells (41). Thus, we used RDBC to test iPSC developmental potency by assaying for the presence of mature B and T cells by FACS analysis of surface markers for B cells (B220) and T cells (Thy1.2) in peripheral blood of resultant chimeras. We found that consistent with the *Meg3*^{off} status of B-iPSCs, neither B nor T cells could be detected in the peripheral blood of B-iPSC *Rag2*^{-/-} chimeras from all three independent B-iPSCs (Table 1 and Fig. S3A).

To test the effect of 5aza treatment on developmental potency of iPSCs reprogrammed from activated B cells, we assayed five different 5aza-B-iPSCs by RDBC. Notably, all five *Meg3*^{on} 5aza-B-iPSCs injected for RDBC gave rise to mouse chimerism, with three giving rise to T + B cells in peripheral blood, and two giving rise only to detectable T cells (Table 1 and Fig. S3B). We also found that injected 5aza-B-iPSCs into C57BL/6 blastocysts frequently give rise to high-grade (>80%) chimerism as determined by percentage of agouti (iPSC-derived) coat color (Fig. S4). Of eight 5aza-B-iPSCs tested, six had clear evidence of coat color chimerism with four displaying high-grade (>50% agouti) chimerism (Fig. S4B). Together, our findings suggest that 5aza treatment of activated B cells before reprogramming leads to *Meg3*^{on} clones with developmental potency sufficient for B and T cell differentiation and high-grade coat color chimerism.

To further examine RDBC chimeric mice derived from the two independent 5aza-B-iPSC clones that gave T + B cells, we assayed splenic B cells for B220, as well as IgM and IgG1 by surface stains and flow cytometry. We observed that B cells from both clones were IgM⁻ and IgG1⁺ (Fig. 3A and B). These data indicate that the 5aza-B-iPSCs used to generate these chimeras were derived from B cells that had undergone CSR to IgG1, and that all mature B cells in these RDBC chimeras were iPSC derived. Further Southern blot analysis revealed that four of the five 5aza-B-iPSC clones used for RDBC had evidence of V(D)J rearrangements on both alleles, and that three of the five clones had undergone CSR on the nonproductive allele (Fig. S5). To determine whether the IgG1⁺ B cells derived from 5aza-B-iPSCs could be activated in culture for CSR to IgE, IgG1⁺ 5aza-B-iPSC-derived B cells and control IgM⁺ B cells from age-matched OKSM transgenic mice were treated with anti-CD40 plus IL-4, and IgE CSR was measured using a FACS-based approach we

Table 1. Results of RDBC assay

Injected clone	Live-born	T cell chimerism	T + B cell chimerism
B-iPSC1	6	0	0
B-iPSC2	6	0	0
B-iPSC3	6	0	0
5aza-B-iPSC1	15	0	7
5aza-B-iPSC2	12	0	6
5aza-B-iPSC3	5	3	0
5aza-B-iPSC4	12	6	0
5aza-B-iPSC5	9	1	3

The total number of live-born mice and number of T and T + B-cell chimerism resulting from *Rag2*^{-/-} blastocyst complementation with the indicated iPSC clones are shown. The presence of T and T + B-cell chimerism was determined by FACS analysis of Thy1.2 (T-cell)- and B220 (B-cell)-expressing cells in peripheral blood as shown in Fig. S3.

recently developed that involves removal of nonspecific surface IgE before detection of cytoplasmic IgE (20). We observed that 5aza-B-iPSC-derived IgG1⁺ B cells can be activated for switching to IgE similarly to control IgM⁺ B cells (Fig. 3 C and D). Notably, the overall kinetics of CSR to IgE also was similar in the 5aza-B-iPSC-derived IgG1⁺ B cells and the IgM⁺ control cells (Fig. 3D). Thus, the 5aza-B-iPSC-derived IgG1⁺ B cells can develop into naïve-like IgG1⁺ B cells that are functional in terms of their ability to be activated in culture to undergo further CSR reactions.

Discussion

Our goal was to develop a method to produce *IgH*-switched B-cell-derived iPSCs that are able to support high-grade mouse chimeras to expand tools for the study of B-cell development and function. Given that tested iPSCs derived from activated B cells

were *Meg3*^{off} and did not support lymphocyte development in the setting of RDBC, we developed conditions using 5aza treatment of *IgH* class-switched B cells that allowed us to isolate *Meg3*^{on} iPSCs that were able to form high-grade coat color chimerism. In addition, we found that several tested 5aza-B-iPSC cells derived from IgG1⁺ B cells were capable of generating all-IgG1 mice following injection into *Rag2*-deficient host blastocysts. Finally, we found that iPSC-derived all-IgG1 B cells can be efficiently activated to switch to IgE.

We have found that iPSCs derived from day 4-activated B cells from OKSM are uniformly *Meg3*^{off}, and several tested failed to produce T or T + B-cell chimerism in *Rag2*^{-/-} mice. Correspondingly, a contemporaneous study found that iPSCs derived from OKSM B cells taken directly from mice were uniformly *Meg3*^{off} with reduced developmental potency (39). Similar to our findings, this study also found that attenuation of hypermethylation at the *Dlk-Dio3* locus by treatment of B cells with ascorbic acid (39) resulted in *Meg3*^{on} iPSCs with high developmental potential (39). In our studies, 5aza treatment of day 4-activated B cells to induce demethylation before iPSC generation enabled us to produce iPSC-derived B cells that had previously undergone CSR. Moreover, we found that these iPSC-derived IgG1⁺ B cells are functional in terms of the ability to undergo further activation and CSR to IgE in culture. Thus, our 5aza study, coupled with the ascorbic acid study (39), highlight the usefulness of developing reprogramming conditions aimed at attenuating *Dlk-Dio3* locus hypermethylation during iPSC derivation, and show that different approaches to achieve this goal may be used.

Because our approach involved treatment of the day 4-activated B cells with 5aza before the onset of reprogramming, it may offer insights into the potential utility of epigenetic modification before and at early stages of reprogramming to positively influence resultant iPSCs. However, the detailed molecular mechanisms underlying this effect remain to be determined. Given that both loss of DNA methylation and de novo DNA methylation likely occur

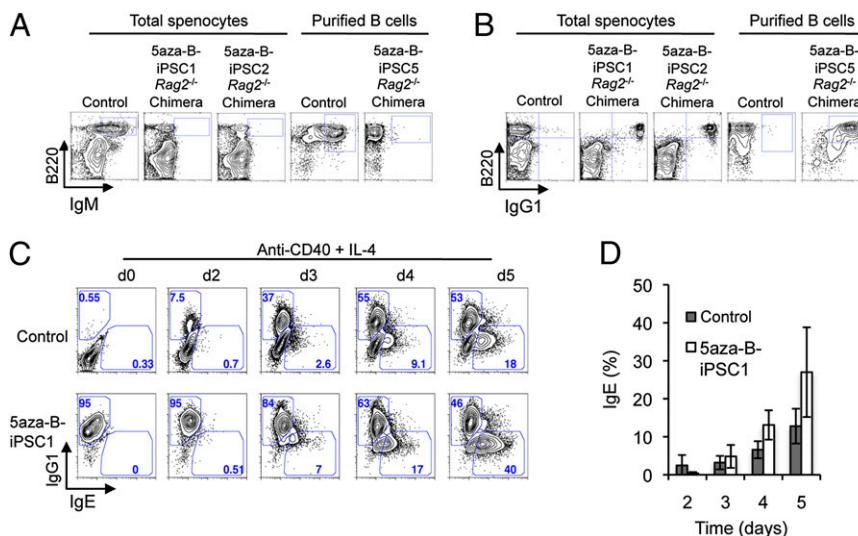


Fig. 3. The 5aza-B-iPSCs can form all-IgG1 chimeric mice in *Rag2*^{-/-} hosts, which can support further CSR to IgE. (A) FACS plots of splenocytes from 5aza-B-iPSC-complemented *Rag2*^{-/-} chimeras stained for surface expression of the pan-B-cell marker B220 and IgM. For comparison, splenocytes from reprogrammable mice indicated as control were analyzed in parallel. Cells expressing both B220 and IgM are demarcated within FACS plots by a box in the upper right-hand corner. (B) FACS plots of splenocytes from control mice and 5aza-B-iPSC-complemented *Rag2*^{-/-} chimeras stained for surface expression of B220 and IgG1. Cells expressing both B220 and IgG1 are shown by a box in the upper right-hand corner. (C) FACS analysis of day 0, 2, 3, 4, and 5 anti-CD40/IL-4-stimulated B cells from reprogrammable mice (control) or 5aza-B-iPSC-derived B cells stained for cytoplasmic IgG1 and IgE expression as described in the text. Gates within the plots demarcate IgG1⁺ and IgE⁺ B cells in the upper left and lower right portion of box, respectively. Numbers indicate percentage of cells within the gates. (D) Statistical analysis of three independent FACS analysis experiments for IgE switching of control and 5aza-B-iPSC-derived B cells activated with anti-CD40 plus IL-4 for the indicated time points. Shown are mean values \pm SD of three independent experiments involving *Rag2*^{-/-} iPSC chimeras derived from 5aza-B-iPSC#2.

during the reprogramming process (42), 5aza treatment early during reprogramming may allow the reprogramming process to ameliorate potential negative effects of global hypomethylation. In this context, whether the reprogramming process provides a window for specific epigenetic remodeling not available in stable cycling iPSCs remains to be determined.

The irreversible genetic deletions associated with B-cell development and CSR provide a means to assess the C_H configuration and clonality of activated B-cell–derived iPSCs. Furthermore, given that the process of CSR excises 100–200 kb of DNA between C_H s, the derivation of B-cell models from *IgH* class-switched B-cell–derived iPSCs provides an approach to assess the potential role of sequences internal to the C_H region in regulation of transcription, CSR and other processes during B-cell development and activation. Additionally, the ability to efficiently generate all-iPSC–switched B cells allows questions to be addressed regarding the functional consequences of expressing various *IgH* isotypes on B-cell development and function. In this regard, because $IgG1^+$ B cells are often intermediates in normal sequential B-cell switching to IgE (20, 26–29), the creation of all- $IgG1$ chimeric mice provides a valuable tool for studies related to mechanisms regulating second-step switching from $IgG1$ to IgE and the stability of $IgG1^+$ B cells.

An antigen-specific B-cell mouse model has been generated with somatic cell nuclear transfer technology (43). In that context, the approaches we describe here involving generation of iPSCs derived from activated, class-switched B cells followed by RDBC to generate mice in which all B cells derive from the clonally rearranged iPSC cells may provide a useful extension of that general method of generating monoclonal mice with selected antibody specificities. In particular, the RDBC approach could allow a more rapid method for initial screening for mice that make the desired antibody specificities and in the context of desired *IgH* isotypes. Though not yet demonstrated, we would expect that the high-level chimerism we have achieved with 5aza-B-iPSCs will eventually allow germline transmission of desired iPSC *Ig* genotypes after RDBC screening.

Materials and Methods

Mice, Cell Culture, and Reprogramming. Doxycycline-inducible reprogrammable mice used in these experiments have been described previously (31) and were a gift from Konrad Hochedlinger (Massachusetts General Hospital, Boston, MA). All experiments with mice followed the protocols approved by the Children's Hospital Boston Animal Care and Use Committee. Splenic cells from reprogrammable mice were isolated by B220⁺ selection via magnetic columns (Miltenyi Biotech) and cultured in RPMI (Invitrogen) supplemented with L-glutamine, penicillin, streptomycin, and 15% (vol/vol) FBS together

with anti-CD40 plus IL-4 to induce CSR to $IgG1$ and IgE as described (20). Fresh media was added to the cells on days 2, 3, and 4. After 4 d of culture, OKSM reprogramming factors were induced by the addition of 2 μ g/mL doxycycline, and cells were transferred to ES cell medium consisting of DMEM (Invitrogen) in 15% FBS on gelatinized plates seeded with a feeder layer of irradiated MEFs for 10–12 d with medium exchanges every 2 d. Colonies with ES cell morphology were then picked and expanded in the absence of doxycycline for further analysis. For 5aza (Sigma) treatment, a 2.5- μ M 5aza solution in ES cell medium was given to day 4-activated B cells 24 h before the addition of doxycycline, and another fresh 2.5 μ M 5aza solution was provided at the time of doxycycline addition. 5aza was removed 24 h after doxycycline addition for a total 5aza treatment period of 48 h. Following 5aza removal, cells were incubated on MEFs with doxycycline for 10–12 d before ES cell-like colonies were picked and expanded in ES cell media without doxycycline to isolate stable iPSC clones. Media was exchanged every 2 d during reprogramming. Functional analysis of iPSCs was performed by injection of iPSCs into *Rag2*^{-/-} or C57B/6 blastocysts after verifying normal karyotype.

Analysis of DNA Methylation. iPSC clones were lysed in PK lysis buffer [100 mM Tris-HCl (pH 8.0), 5 mM EDTA, 0.2% SDS and 200 mM NaCl, 200 μ g/mL proteinase K] followed by genomic DNA precipitation with isopropanol (50% vol/vol). After washing with 70% ethanol, DNA was resuspended in T low E buffer [10 mM Tris (pH 8.0), 0.1 mM EDTA]. DNA was bisulfite converted using the EpiTect Bisulfite Kit (Qiagen) and analyzed by EpigenDX using the ADS1452 (IG-DMR) assay.

RNA Isolation and Quantification. Total RNA was extracted using the TRIzol method (Invitrogen) and reverse transcribed into cDNA using qScript (Quanta Biosciences). *Meg3* and *Gapdh* transcripts were quantified using SYBR Green qPCR (Applied Biosystems) primers described elsewhere (9).

Southern Blot Analysis. Genomic DNA (20 μ g) was digested with EcoRI before separation on a 1% agarose gel and blotting to Hybond-XL membrane (Amersham Biosciences). Blots were hybridized to probes specific to DFL16, $I\mu$, $C\mu$, $I\gamma 1$, and $C\epsilon$ as described elsewhere (44), washed, and put on phosphorimaging cassettes for exposure.

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