

Cyclin D1 predicts the prognosis of advanced serous ovarian cancer

TOMOKO HASHIMOTO¹, NOZOMU YANAIHARA¹, AIKOU OKAMOTO¹, TAKASHI NIKAIIDO², MISATO SAITO¹, SATOSHI TAKAKURA¹, MAKOTO YASUDA³, HIROSHI SASAKI³, KAZUNORI OCHIAI¹ and TADAO TANAKA¹

¹Department of Obstetrics and Gynecology, The Jikei University School of Medicine; ²Department of Pathology, Kosei General Hospital, Tokyo; ³Department of Obstetrics and Gynecology, Jikei Kashiwa Hospital, Chiba, Japan

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Abstract. We previously reported that cyclin E (*CCNE1*) amplification is strongly associated with resistance to treatment in serous ovarian cancer by high-resolution oligonucleotide copy number analysis. Dysregulation of cell cycle control has been implicated as the key event in human oncogenesis, and aberrant expression of G1-S phase-related genes in particular has been reported in epithelial ovarian cancer (EOC). Nevertheless, there are conflicting results concerning the prognostic values of these abnormalities in EOC. This study focused on advanced serous EOC cases and investigated the association between the expression of G1-S phase-regulatory proteins and clinicopathological parameters. The utility of these proteins as prognostic factors was assessed, and whether these targets reflect chemoresistance of advanced serous EOC was investigated. A total of 66 patients treated by primary surgery were evaluated in this study. Immunohistochemical analysis for cyclin D1, pRb, p16, p53, p27^{Kip1}, p21^{Waf1/Cip1} and cyclin E was performed on formalin-fixed tissue sections collected from primary surgical specimens. The correlations between the expression of these proteins and the clinicopathological parameters, including progression-free survival (PFS), overall survival (OS) and chemosensitivity, were examined. Upon univariate analysis, overexpression of cyclin D1 was positively correlated with reduced PFS ($p=0.00062$) and OS ($p=0.00037$). Reduced expression of p27^{Kip1} was associated with shorter OS ($p=0.064$). Upon multivariate analysis, overexpression of cyclin D1 ($p=0.0019$), reduced expression of p27^{Kip1} ($p=0.042$) and residual tumor volume ($p=0.0092$) were identified as independent predictors of OS. Overexpression of cyclin D1 ($p=0.011$) as well as residual tumor volume ($p=0.006$) were significantly associated with first-line chemosensitivity. In advanced serous EOC, overexpression of cyclin D1 contributed largely to poor prognosis, and this may

have been in part mediated by chemoresistance. Cyclin D1 is a possible target for overcoming the refractory nature of advanced serous EOC.

Introduction

Ovarian cancer is the most lethal gynecological malignancy in developed countries and is the 9th most common cancer in Japanese females. An estimated 8,304 new cases and 4,467 mortalities occurred in 2005 (Center for Cancer Control and Information Services, National Cancer Center, Japan). Although ovarian cancer patients respond to cytoreductive surgery and combination chemotherapy satisfactorily, advanced cases exhibit a high level of recurrence, and the overall survival (OS) rate has not significantly changed for decades. However, clinical trials have been undertaken to improve prognosis. Since there are different clinical behavior patterns for certain histopathological subgroups, separate trials have been developed for clear cell (1) and mucinous carcinomas (2). The alteration of dose/schedule and the use of intraperitoneal therapy have been shown to be superior in at least one trial (3,4).

A number of clinicopathological factors, including the volume of residual tumor after primary surgery, FIGO stage and tumor grade, are reported to be the key prognostic factors (5,6). Cytoreduction to a non-macroscopic residual tumor is the ultimate goal, and it improves prognosis (7-9). However, in order to improve the prognosis of advanced epithelial ovarian cancer (EOC) cases, other predictive biomarkers should also be elucidated.

We previously described that cyclin E (*CCNE1*) amplification was strongly associated with resistance to treatment in serous ovarian cancer by high-resolution oligonucleotide copy number analysis (10). Therefore, the amplification status of cyclin E has potential for therapeutic exploitation, whereby patients exhibiting cyclin E amplification may benefit from novel, cyclin-related targeted treatments. Dysregulation of cell cycle control has been implicated as the key event in human oncogenesis, and aberrant expression of G1-S phase-related genes in particular has been reported in a number of human cancers, including EOC (5,11). Aberrant expression of the p16-cyclin D1-CDK4/6-pRb and p21-p27-cyclin E-CDK2 pathways have been reported to correlate with prognosis (5,11-14).

Barbieri *et al* reported in their series of 70 EOC cases that overexpression of cyclin D1 was associated with a shorter

Correspondence to: Dr Aikou Okamoto, Department of Obstetrics and Gynecology, The Jikei University School of Medicine, 3-25-8, Nishi-Shinbashi, Minato-ku, Tokyo 105-8461, Japan
E-mail: aikou@jikei.ac.jp

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OS. In particular, among patients with stage III/IV tumors and residual disease greater than 2 cm, cyclin D1 expression significantly influenced clinical outcome (5). Bali *et al* reported on 134 serous ovarian cancers for which molecular markers predicted reduced OS in univariate analysis, which included overexpression of cyclin D1 and p53, and reduced expression of p27^{Kip1} and p21^{Waf1/Cip1} (11). In contrast, it was reported that low nuclear p27 expression was associated with improved 3-year OS and progression-free survival (PFS) in 150 advanced stage (FIGO stages II, III and IV) EOC patients (12). Kommoss *et al* carried out immunohistochemical analysis of p16^{Ink4a} and pRb expression levels and found that they correlated with survival in a series of 300 patients with FIGO stage II-IV ovarian carcinoma. They reported that p16^{Ink4a}-negative tumors had a significantly worse prognosis in both univariate and multivariate analyses. High expression levels of pRb were associated with an incremental deterioration of prognosis, which was also the case in the subgroup of optimally debulked patients (15). Meanwhile, Khouja *et al*, using immunohistochemistry, evaluated 171 primary stage III ovarian carcinoma tumors for expression of Ki-67, p16, p14 and p57. High expression of p16 was correlated with poor differentiation and survival in univariate analysis. However, in multivariate analysis, p16 expression was not significantly associated with shorter survival (13). Some of the results are contradictory, probably due to the variety of histotypes and stages of EOC as well as disease heterogeneity, different research methodologies or the sample sizes of the studies. As serous ovarian cancer is the most common histological type of EOC and the prognosis of advanced cases remains poor, we limited our analysis to the serous histotype and advanced cases to eliminate such bias.

We focused on advanced serous EOC (stage III/IV) cases in particular and investigated the association between the expression of G1-S phase-regulatory proteins and the clinicopathological parameters. We aimed to identify the utility of these proteins as prognostic factors and to evaluate whether these targets reflect chemoresistance of advanced serous EOC.

Patients and methods

Patients and tumor specimens. The Jikei University School of Medicine Ethics Review Committee approved the study protocol, and informed consent was obtained from the patients. The tumor specimens were surgically obtained from a group of 66 patients with advanced primary ovarian serous adenocarcinoma who were treated at the Department of Obstetrics and Gynecology, The Jikei University School of Medicine, and Jikei University Kashiwa Hospital. The tumors were staged in accordance with the International Federation of Gynecology and Obstetrics (FIGO) system (1988). The clinical and pathological characteristics of the patient cohort are shown in Table I. The age at diagnosis, volume of postoperative residual disease, FIGO stage, presence of intraoperative ascites and patient outcome were obtained retrospectively from patient records as shown. The median follow-up time for the cohort was 15.5 months (range 3-72). The 66 patients received first-line platinum-based chemotherapy. Among them, 62 cases received taxane simultaneously as T-C chemotherapy following primary surgery (93.9%).

Table I. Clinical and pathological characteristics of the serous epithelial ovarian cancer patient cohort (n=66).

Clinicopathological parameters	No. of patients (%)
Age	
≤65 years	55 (83.3)
>65 years	11 (16.7)
FIGO stage	
III	52 (78.8)
IV	14 (21.2)
Residual disease	
≤2 cm	28 (42.4)
>2 cm	38 (57.6)
Ascites	
≤500 ml	25 (37.9)
>500 ml	41 (62.1)
Disease progression	
No	9 (13.6)
Yes	57 (86.4)

Immunohistochemistry. Immunostaining was performed on buffered formalin-fixed, paraffin-embedded tissue sections. The sections were deparaffinized, and standard immunohistochemical techniques were performed using Ventana XT system (BenchMark[®] XT; Ventana Medical Systems, Inc., Tuscon, AZ, USA) in accordance with the manufacturer's instructions. Antigen epitopes were retrieved using Ventana Benchmark CC1 standard program. The primary antibodies used in this study were: anti-cyclin D1 (rabbit monoclonal clone SP4; Ventana Medical Systems, Inc.), anti-pRb (mouse monoclonal clone 13A10; Novocastra Laboratories Ltd., UK; 1:100), anti-p16 (mouse monoclonal clone 16P04; Ventana Medical Systems, Inc.), anti-p53 (mouse monoclonal clone DO-7; Ventana Medical Systems, Inc.), anti-p21^{Waf1} (mouse monoclonal clone EA10; Calbiochem, Darmstadt, Germany; 1:50), anti-p27^{Kip1} (mouse monoclonal clone SX53G8; Dako, Glostrup, Denmark; 1:20) and anti-cyclin E (mouse monoclonal clone HE12; Medical and Biological Laboratories Co., Ltd., Japan; 1:500). Antibodies from Ventana Medical Systems, Inc. were pre-diluted. The antibodies were incubated at 37°C for 32 min (60 min for p21, Rb and cyclin E). The slides were counterstained with hematoxylin and mounted for microscopic examination. Positive and negative controls were tested in parallel for each staining.

Immunostaining evaluation. At least 500 tumor cells were evaluated for immunostaining, and the percentage of stained cells was calculated. The evaluation of immunostaining was conducted in a blinded manner by two independent screeners, without knowledge of the clinical and pathological characteristics of the cases. Standardization of scoring was achieved by comparison of scores between screeners, and discrepancies were resolved by consensus. The percentage of positive nuclear immunostaining in cells of the tumor sections was calculated. Scores are expressed as a percentage of positive nuclear

Table II. Immunohistochemical analysis of cell cycle gene expression in advanced serous epithelial ovarian cancer.

	Cyclin D1	pRb	p16 ^{Ink4a}	p53	p27 ^{Kip1}	p21 ^{Waf1/Cip1}	Cyclin E
Range of staining (% positive)	0-50	0-90	0-90	0-90	0-80	0-70	0-90
Median staining (%)	10	50	80	60	50	5	50
Cutoff value (%) (positive staining)	>20	>50	>50	>40	>40	>5	>70
Positive tumors (%)	11 (16.7)	26 (39.4)	42 (63.6)	37 (56.1)	37 (56.1)	20 (30.3)	11 (16.7)

Range of staining indicates the proportion of positive nuclear staining within representative areas of the tumor samples. Cut-off value is based on published reports. Number of specimens showing positive staining is provided in the bottom row.

staining within representative areas of the tumor sample. The percentage score above, whose staining is considered representative of overexpression, is based on published reports (11). The range and the median percentage of immunostaining, percentage values regarded as overexpression (cutoff value) and the numbers of specimens displaying positive staining are shown in Table II. Representative photomicrographs of tumor tissue showing positive and negative staining for the specific antigens are presented in Fig. 1.

Statistical analysis. The associations between clinicopathological parameters and the immunostaining scores were analyzed. The correlations between the expression of each gene and the clinicopathological parameters were analyzed using the Chi-square test. $p \leq 0.05$ was considered to be statistically significant. For survival analysis, event time distributions were evaluated using the Kaplan-Meier method, and differences in survival rates were compared using the log-rank test for univariate analysis and Cox proportional hazards regression model for multivariate analysis. PFS was calculated from the date of primary surgery to the date of disease progression. The duration of OS was defined from the date of primary surgery to the date the patient succumbed to the disease or to the date of last follow-up. The treatment-free interval (TFI) was defined as being from the last date of first-line chemotherapy to the date of recurrence or last follow-up without recurrence.

Results

Expression of G1-S phase-regulatory proteins and the association with clinicopathological parameters. The expression of G1-S phase-regulatory proteins was analyzed by immunohistochemistry in advanced serous EOC. Overexpression of cyclin D1, pRb, p16, p53, p27^{Kip1}, p21^{Waf1/Cip1} and cyclin E was detected with incidences of 16.7, 39.4, 63.6, 56.1, 56.1, 30.3 and 16.7%, respectively. Associations of the expression of each protein and clinicopathological parameters are shown in Table III. The volume of postoperative residual disease and the presence of ascites were not correlated with the expression pattern of any of the studied proteins. The expression of p53 appeared to be positively correlated with that of p16^{Ink4a} (Table III). No other significant association among the gene expressions was observed.

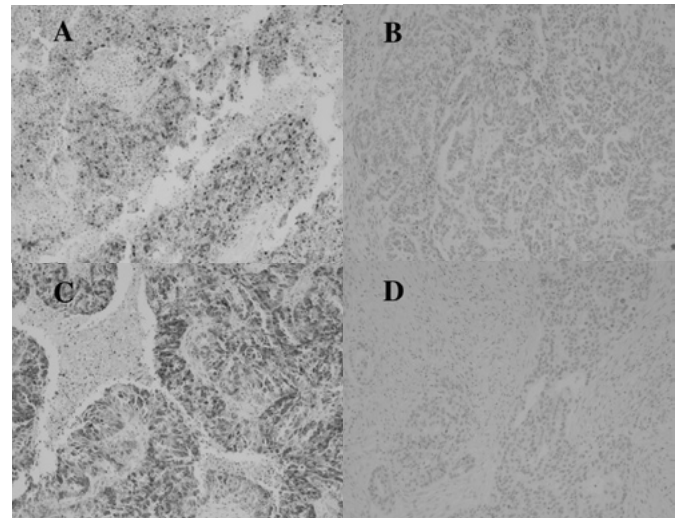


Figure 1. Immunohistochemistry on primary tumor tissue. Representative stainings for cyclin D1 and p27^{Kip1} expression are shown. Magnification, x100. (A) Cyclin D1-positive staining. (B) Cyclin D1-negative staining. (C) p27^{Kip1}-positive staining. (D) p27^{Kip1}-negative staining.

Correlation between G1-S phase-regulatory protein expression and patient outcome in advanced serous epithelial ovarian cancer. The relationship between gene expression and patient prognosis was assessed. Upon univariate analysis, the clinicopathological determinants of reduced OS included age and volume of residual disease >2 cm (Table IV). A molecular marker predictive of reduced OS upon univariate analysis was overexpression of cyclin D1 ($p=0.00037$, RR=0.28, 95% CI 0.044-0.40). Reduced expression of p27^{Kip1} had a trend of association with a shorter OS ($p=0.064$, RR=1.88, 95% CI 0.97-4.21). Regarding PFS, overexpression of cyclin D1 ($p=0.00063$, RR=0.34, 95% CI 0.054-0.43) was significantly correlated with reduced PFS, but reduced expression of p27^{Kip1} had no statistically significant correlation with PFS. The CA125 level, volume of intraoperative ascites, pRb, p16, p53, p21^{Waf1/Cip1} and cyclin E expression exhibited no statistically significant correlation with either OS or PFS. Kaplan-Meier curves and log-rank p-values according to cyclin D1 expression, p27^{Kip1} expression and residual tumor volume are shown in Fig. 2.

In the multivariate analysis using the Cox proportional hazards model, overexpression of cyclin D1 was identified

Table III. Association of gene expression and clinicopathological parameters in serous epithelial ovarian cancer.

	Cyclin D1	pRb	p16 ^{Ink4a}	p53	p27 ^{Kip1}	p21 ^{Waf1/Cip1}	Cyclin E
Residual disease	0.15	0.99	0.14	0.7300	0.097	0.17	0.91
Ascites	0.82	0.10	0.12	0.3000	0.310	0.81	0.82
Cyclin D1		0.43	0.30	0.6600	0.820	0.19	0.77
pRb			0.45	0.0820	0.420	0.95	0.14
p16 ^{Ink4a}				0.0049	0.190	0.34	0.73
p53					0.380	0.51	0.37
p27 ^{Kip1}						0.67	0.82
p21 ^{Waf1/Cip1}							0.90

Significant p-values are indicated in boldface type.

Table IV. Univariate analysis for the association of clinicopathological parameters and gene expression with clinical outcome in serous epithelial ovarian cancer.

Parameters	PFS			OS		
	RR	95% CI	p-value	RR	95% CI	p-value
Age (≤ 65 vs. >65 years)	0.71	0.300-1.46	0.32000	0.42	0.100-0.87	0.02900
Residual disease (≤ 2 vs. >2 cm)	0.62	0.360-1.04	0.07800	0.27	0.150-0.60	0.00087
CA125 (≤ 500 vs. >500)	0.83	0.440-1.54	0.56000	1.10	0.470-2.65	0.81000
Cyclin D1 (≤ 20 vs. $>20\%$)	0.34	0.054-0.43	0.00063	0.28	0.044-0.40	0.00037
pRb (≤ 50 vs. $>50\%$)	1.00	0.580-1.73	0.99000	0.90	0.440-1.81	0.76000
p16 ^{Ink4a} (≤ 50 vs. $>50\%$)	1.13	0.650-2.00	0.65000	0.97	0.480-1.96	0.93000
p53 (≤ 40 vs. $>40\%$)	1.43	0.850-2.59	0.17000	1.38	0.690-2.83	0.35000
p27 ^{Kip1} (≤ 40 vs. $>40\%$)	1.08	0.630-1.87	0.78000	1.88	0.970-4.21	0.06400
p21 ^{Waf1/Cip1} (≤ 5 vs. $>5\%$)	0.82	0.440-1.45	0.48000	1.48	0.710-3.00	0.31000
Cyclin E (≤ 70 vs. $>70\%$)	0.97	0.460-2.04	0.94000	0.86	0.330-2.19	0.75000
Ascites (≤ 500 vs. >500 ml)	0.81	0.470-1.37	0.43000	0.63	0.320-1.28	0.21000

Data census was at 75 months. Significant p-values are indicated in boldface type. PFS, progression-free survival; OS, overall survival; RR, relative risk; CI, confidence interval.

as the key determinant of OS ($p=0.0019$, RR=3.61, 95% CI 1.61-8.12) and PFS ($p=0.0052$, RR=2.70, 95% CI 1.35-5.41) (Table V). The volume of residual disease and reduced expression of p27^{Kip1} were found to be independent predictors of OS ($p=0.0092$ and $p=0.042$, respectively), but not of PFS when incorporated into a multivariate model (Table V).

Association between chemosensitivity and G1-S phase-regulatory protein expression. In order to assess whether the clinicopathological parameters reflect the chemosensitivity, the cohort was divided into two groups: patients who relapsed within 6 months after the last date of first-line chemotherapy; and patients who had no disease progression within 6 months after the last date of first-line chemotherapy. Using the Chi-square test, overexpression of cyclin D1 ($p=0.011$) as well as residual tumor volume >2 cm ($p=0.006$) were found to be significantly associated with TFI, suggesting that these parameters are correlated with first-line chemosensitivity (Table VI).

In contrast, expression of pRb, p16, p53, p27^{Kip1}, p21^{Waf1/Cip1} and cyclin E had no statistical correlation with chemosensitivity.

Discussion

Various studies exist concerning the association between G1-S phase-related genes and EOC prognosis, however, the results are conflicting. Amplification of cyclin E in high-resolution oligonucleotide microarrays was previously found to be associated with poor response to primary treatment in serous ovarian cancer (10), but in the present study, cyclin E expression in immunohistochemical analysis revealed no significant correlation with patient outcome of advanced serous EOC. It is considered that the variety of histological types of EOC, different tumor stages, tumor heterogeneity, racial backgrounds of patients, research methodologies and sample sizes may contribute to inconsistent results. In this study, we focused on advanced serous cases (limited to stage

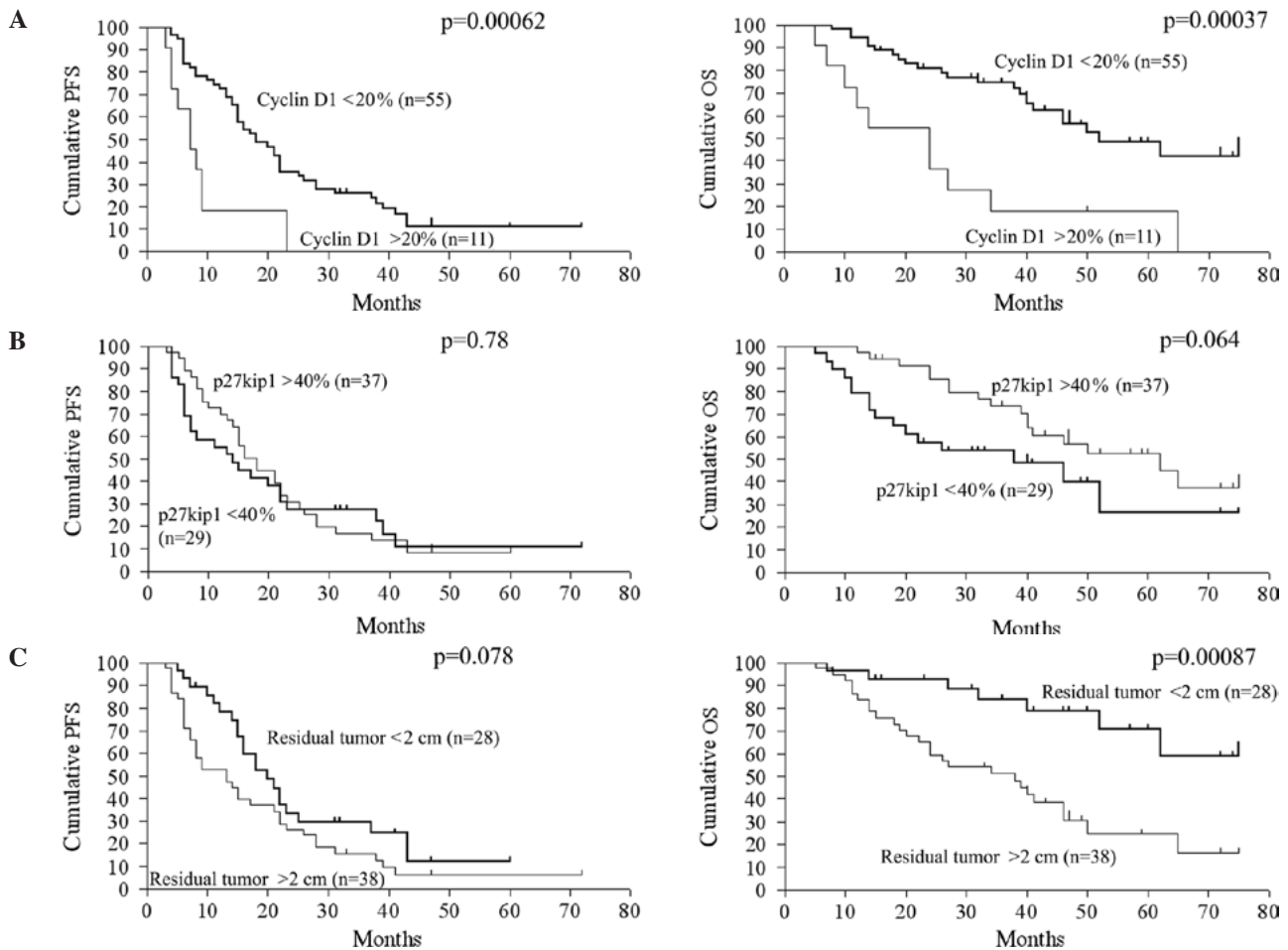


Figure 2. Kaplan-Meier curve. Kaplan-Meier curves and log-rank p-values for 5-year PFS and OS in the context of cyclin D1 expression (A), p27^{Kip1} expression (B) and residual tumor volume (C) are shown. PFS, progression-free survival; OS, overall survival.

Table V. Multivariate Cox regression analysis of PFS and OS of patients with serous EOC.

Clinical parameter	PFS			OS		
	RR	95% CI	p-value	RR	95% CI	p-value
Residual disease (≤ 2 vs. > 2 cm)	1.53	0.89-2.64	0.1200	3.06	1.32-7.12	0.0093
Cyclin D1 (≤ 20 vs. $> 20\%$)	2.70	1.35-5.41	0.0052	3.61	1.61-8.12	0.0019
p27 ^{Kip1} (≤ 40 vs. $> 40\%$)	1.01	0.59-1.72	0.9700	2.15	1.03-4.51	0.0420

Significant p-values are indicated in boldface type. EOC, epithelial ovarian cancer; PFS, progression-free survival; OS, overall survival; RR, relative risk; CI, confidence interval.

III/IV cases) at a single institution with similar surgical and chemotherapeutic procedures administered in order to eliminate such bias.

Cyclin D1, a regulatory kinase subunit that is selectively associated with cyclin-dependent kinase 4 (CDK4), is a crucial modulator of G1 progression in the cell cycle (16). In our analysis, overexpression of cyclin D1 was detected in 11% of the cases (Table II). The overexpression of cyclin D1 was previously observed in 14-89% of EOC cases (11,17-19), but the underlying mechanism has yet to be elucidated. Amplification of cyclin D1 in ovarian tumors occurs infrequently (20).

Furthermore, the mostly small cyclin D1 copy gains are not associated with an increase in detectable cyclin D1 protein by immunohistochemistry (21). These findings suggest that the post-transcriptional regulation of cyclin D1 protein production is complex. Recently, Jiang *et al* performed a systematic validation of the predicted microRNAs for cyclin D1 and revealed that microRNAs suppressed the endogenous cyclin D1 protein and mRNA levels *in vitro* (22). microRNAs may aid in determining the mechanism of cyclin D1 expression.

Barbieri *et al* reported that cyclin D1 overexpression significantly influenced the clinical outcome in advanced EOC

Table VI. Association of the TFI and clinicopathological parameters in serous EOC.

	TFI <6 months, n (%)	TFI ≥6 months, n (%)	p-value
Residual disease			
≤2 cm (n=28)	4 (6.0)	24 (36.4)	0.006
>2 cm (n=38)	19 (28.8)	19 (28.8)	
Cyclin D1			
≤20% (n=55)	15 (22.7)	40 (60.6)	0.011
>20% (n=11)	8 (12.1)	3 (4.6)	
pRB			
≤50% (n=40)	12 (18.2)	28 (42.4)	0.305
>50% (n=26)	11 (16.7)	15 (22.7)	
p16 ^{Ink4a}			
≤50% (n=24)	7 (10.6)	17 (25.8)	0.464
>50% (n=42)	16 (24.3)	26 (39.4)	
p53			
≤40% (n=29)	11 (16.7)	18 (27.3)	0.642
>40% (n=37)	12 (18.2)	25 (37.9)	
p27 ^{Kip1}			
≤40% (n=29)	13 (19.7)	16 (24.3)	0.132
>40% (n=37)	10 (15.2)	27 (40.9)	
p21 ^{Waf/Cip1}			
≤5% (n=46)	19 (28.8)	27 (40.9)	0.165
>5% (n=20)	4 (6.0)	16 (24.3)	
Cyclin E			
≤70% (n=55)	18 (27.3)	37 (56.1)	0.644
>70% (n=11)	5 (7.6)	6 (9.1)	

p-values are for TFI <6 months vs. TFI ≥6 months (Chi-square test, Yates correlation). Significant p-values are indicated in boldface type. EOC, epithelial ovarian cancer; TFI, treatment-free interval.

cases with residual disease greater than 2 cm. They identified cyclin D1 overexpression as an independent prognostic factor in multivariate analysis (5). Similarly, Bali *et al* identified cyclin D1 overexpression as an independent prognostic factor in the multivariate analysis of 134 serous EOC cases (11). In our study, overexpression of cyclin D1 was significantly correlated with reduced OS and PFS in both univariate and multivariate analyses, suggesting that overexpression of cyclin D1 actually contributes to the prognosis of advanced serous EOCs; therefore, its application to clinical practice is expected.

We found that both overexpression of cyclin D1 and residual tumor volume were significantly associated with TFI, suggesting that these parameters are correlated with first-line chemosensitivity (Table VI). Zhou *et al* showed that inhibition of cyclin D1 expression by siRNA in oral squamous cell carcinoma cells resulted in a decrease in cisplatin IC₅₀ level. *In vivo* transplantation models also confirmed a cisplatin-sensitizing effect of cyclin D1 knockdown in these cell lines (23). In addition, it was reported that overexpression of cyclin D1 was associated with reduced chemosensitivity and a higher survival rate upon cisplatin administration in a pancreatic cancer model (24). Moreover, inhibition of cyclin D1 expression rendered cells more susceptible to cisplatin-mediated apoptosis in the same model (24). Taken together, these findings indicate that

cyclin D1 expression may contribute to chemoresistance in a number of cancers, although further investigation is required. Therefore, we speculate that overexpression of cyclin D1 contributes to poor prognosis, which may in part be mediated by chemoresistance in ovarian cancer.

p27^{Kip1} is a cyclin-dependent kinase (cdk) inhibitor that regulates cell cycle progression from the G1 to S-phase. In non-cycling cells, p27^{Kip1} binds to cyclin E-cdk2 complexes and inhibits their activation. In contrast, p27^{Kip1} binding to catalytically active cyclin D-cdk4/6 complexes results in p27^{Kip1} degradation and the subsequent release of cdk2 from inhibition in proliferating cells (12,25). Therefore, p27^{Kip1} helps to coordinate a balance between proliferation and arrest (12,25). In the present study, reduced expression of p27^{Kip1} was detected in 43.9% of the cases. In previous immunohistochemical studies of ovarian tumors, 36.2-100% exhibited low expression of p27^{Kip1} (26). We found that reduced expression of p27^{Kip1} was associated with shorter OS (p=0.064), but had no statistically significant correlation with PFS (p=0.78). When incorporated into a multivariate model, reduced expression of p27^{Kip1} was found to be an independent predictor of OS (p=0.042) (Table V). The relationship between p27^{Kip1} expression levels and prognosis is controversial. Conflicting data regarding the possible prognostic role of p27^{Kip1} status also exist for ovarian cancer. Psyrrri *et al* evaluated subcellular

localization and protein levels of p27^{Kip1} in 150 advanced EOCs and found that low nuclear p27^{Kip1} expression was associated with improved prognosis, suggesting its potential as a strong predictor of outcome in advanced EOCs (12). On the other hand, Shigemasa *et al* reported that negative p27^{Kip1} expression was significantly correlated with poor survival in serous EOC patients, suggesting that the underexpression of p27^{Kip1} caused by a post-translational mechanism may contribute to development and progression and may result in poor prognosis of serous EOCs (27). It is hypothesized that different methodologies of immunohistochemical grading may account for these discrepancies. Our result suggests that p27^{Kip1} is associated with the prognosis of this disease, as previously reported.

In conclusion, overexpression of cyclin D1 contributed markedly to poor prognosis in advanced serous EOC; this may in part be mediated by chemoresistance. Cyclin D1 may be a target for overcoming the refractory nature of advanced serous EOC.

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References

- Takakura S, Takano M, Takahashi F, Saito T, Aoki D, Inaba N, Noda K, Sugiyama T, Ochiai K; Japanese Gynecologic Oncology Group: Randomized phase II trial of paclitaxel plus carboplatin therapy versus irinotecan plus cisplatin therapy as first-line chemotherapy for clear cell adenocarcinoma of the ovary: a JGOG study. *Int J Gynecol Cancer* 20: 240-247, 2010.
- Shimada M, Kigawa J, Ohishi Y, Yasuda M, Suzuki M, Hiura M, Nishimura R, Tabata T, Sugiyama T and Kaku T: Clinicopathological characteristics of mucinous adenocarcinoma of the ovary. *Gynecol Oncol* 113: 331-334, 2009.
- Katsumata N, Yasuda M, Takahashi F, Isonishi S, Jobo T, Aoki D, Tsuda H, Sugiyama T, Kodama S, Kimura E, Ochiai K, Noda K; Japanese Gynecologic Oncology Group: Dose-dense paclitaxel once a week in combination with carboplatin every 3 weeks for advanced ovarian cancer: a phase 3, open-label, randomised controlled trial. *Lancet* 374: 1331-1338, 2009.
- Armstrong DK, Bundy B, Wenzel L, Huang HQ, Baergen R, Lele S, Copeland LJ, Walker JL and Burger RA: Intraperitoneal cisplatin and paclitaxel in ovarian cancer. *N Engl J Med* 354: 34-43, 2006.
- Barbieri F, Lorenzi P, Ragni N, Schettini G, Bruzzo C, Pedullà F and Alama A: Overexpression of cyclin D1 is associated with poor survival in epithelial ovarian cancer. *Oncology* 66: 310-315, 2004.
- NIH Consensus Conference: Ovarian cancer. Screening, treatment, and follow-up. *JAMA* 273: 491-497, 1995.
- Terauchi F, Nishi H, Moritake T, *et al*: Prognostic factor on optimal debulking surgery by maximum effort for stage IIIC epithelial ovarian cancer. *J Obstet Gynaecol Res* 35: 315-319, 2009.
- Du Bois A, Reuss A, Pujade-Lauraine E, Harter P, Ray-Coquard I and Pfisterer J: Role of surgical outcome as prognostic factor in advanced epithelial ovarian cancer: a combined exploratory analysis of 3 prospectively randomized phase 3 multicenter trials: by the Arbeitsgemeinschaft Gynaekologische Onkologie Studiengruppe Ovarialkarzinom (AGO-OVAR) and the Groupe d'Investigateurs Nationaux Pour les Etudes des Cancers de l'Ovaire (GINECO). *Cancer* 115: 1234-1244, 2009.
- Chi DS, Eisenhauer EL, Zivanovic O, *et al*: Improved progression-free and overall survival in advanced ovarian cancer as a result of a change in surgical paradigm. *Gynecol Oncol* 114: 26-31, 2009.
- Etamadmoghadam D, deFazio A, Beroukhim R, Mermel C, George J, Getz G, Tothill R, Okamoto A, Raeder MB, Harnett P, Lade S, Akslen LA, Tinker AV, Locandro B, Alsop K, Chiew YE, Traficante N, Fereday S, Johnson D, Fox S, Sellers W, Urashima M, Salvesen HB, Meyerson M, Bowtell D; AOCs Study Group: Integrated copy number and expression analysis of chemoresistant ovarian carcinomas. *Clin Cancer Res* 15: 1417-1427, 2009.
- Bali A, O'Brien PM, Edwards LS, Sutherland RL, Hacker NF and Henshall SM: Cyclin D1, p53, and p21^{Waf1/Cip1} expression is predictive of poor clinical outcome in serous epithelial ovarian cancer. *Clin Cancer Res* 10: 5168-5177, 2004.
- Psyrrri A, Bamias A, Yu Z, *et al*: Subcellular localization and protein levels of cyclin-dependent kinase inhibitor p27 independently predict for survival in epithelial ovarian cancer. *Clin Cancer Res* 11: 8384-8390, 2005.
- Khouja MH, Baekelandt M, Nesland JM and Holm R: The clinical importance of Ki-67, p16, p14, and p57 expression in patients with advanced ovarian carcinoma. *Int J Gynecol Pathol* 26: 418-425, 2007.
- Milde-Langosch K and Riethdorf S: Role of cell-cycle regulatory proteins in gynecological cancer. *J Cell Physiol* 196: 224-244, 2003.
- Kommoss S, du Bois A, Ridder R, Trunk MJ, Schmidt D, Pfisterer J, Kommoss F; AGO-OVAR: Independent prognostic significance of cell cycle regulator proteins p16(INK4a) and pRb in advanced-stage ovarian carcinoma including optimally debulked patients: a translational research subprotocol of a randomised study of the Arbeitsgemeinschaft Gynaekologische Onkologie Ovarian Cancer Study Group. *Br J Cancer* 96: 306-313, 2007.
- Sherr CJ: G1 phase progression: cycling on cue. *Cell* 79: 551-555, 1994.
- Dhar KK, Branigan K, Parkes J, *et al*: Expression and subcellular localization of cyclin D1 protein in epithelial ovarian tumour cells. *Br J Cancer* 81: 1174-1181, 1999.
- Worsley SD, Ponder BA and Davies BR: Overexpression of cyclin D1 in epithelial ovarian cancers. *Gynecol Oncol* 64: 189-195, 1997.
- Barbieri F, Cagnoli M, Ragni N, Foglia G, Bruzzo C, Pedullà F and Alama A: Increased cyclin D1 expression is associated with features of malignancy and disease recurrence in ovarian tumors. *Clin Cancer Res* 5: 1837-1842, 1999.
- Masciullo V, Scambia G, Marone M, *et al*: Altered expression of cyclin D1 and CDK4 genes in ovarian carcinomas. *Int J Cancer* 74: 390-395, 1997.
- Diebold J, Mösinger K, Peiro G, *et al*: 20q13 and cyclin D1 in ovarian carcinomas. Analysis by fluorescence in situ hybridization. *J Pathol* 190: 564-571, 2000.
- Jiang Q, Feng MG and Mo YY: Systematic validation of predicted microRNAs for cyclin D1. *BMC Cancer* 9: 194, 2009.
- Zhou X, Zhang Z, Yang X, Chen W and Zhang P: Inhibition of cyclin D1 expression by cyclin D1 shRNAs in human oral squamous cell carcinoma cells is associated with increased cisplatin chemosensitivity. *Int J Cancer* 124: 483-489, 2009.
- Biliran H Jr, Wang Y, Banerjee S, *et al*: Overexpression of cyclin D1 promotes tumor cell growth and confers resistance to cisplatin-mediated apoptosis in an elastase-myc transgene-expressing pancreatic tumor cell line. *Clin Cancer Res* 11: 6075-6086, 2005.
- Blain SW, Scher HI, Cordon-Cardo C and Koff A: p27 as a target for cancer therapeutics. *Cancer Cell* 3: 111-115, 2003.
- Nam EJ and Kim YT: Alteration of cell-cycle regulation in epithelial ovarian cancer. *Int J Gynecol Cancer* 18: 1169-1182, 2008.
- Shigemasa K, Shiroyama Y, Sawasaki T, *et al*: Underexpression of cyclin-dependent kinase inhibitor p27 is associated with poor prognosis in serous ovarian carcinomas. *Int J Oncol* 18: 953-958, 2001.