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Learning and the motivation to eat: Forebrain circuitry

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Abstract

Appetite and eating are not only controlled by energy needs, but also by extrinsic factors that are not directly related to energy balance. Environmental signals that acquire motivational properties through associative learning-learned cues-can override homeostatic signals and stimulate eating in sated states, or inhibit eating in states of hunger. Such influences are important, as environmental factors are believed to contribute to the increased susceptibility to overeating and the rise in obesity in the developed world. Similarly, environmental and social factors contribute to the onset and maintenance of anorexia nervosa and other eating disorders through interactions with the individual genetic background. Nevertheless, how learning enables environmental signals to control feeding, and the underlying brain mechanisms are poorly understood. We developed two rodent models to study how learned cues are integrated with homeostatic signals within functional forebrain networks, and how these networks are modulated by experience. In one model, a cue previously paired with food when an animal was hungry induces eating in sated rats. In the other model, food-deprived rats inhibit feeding when presented with a cue that signals danger, a tone previously paired with footshocks. Here evidence will be reviewed that the forebrain network formed by the amygdala, lateral hypothalamus and medial prefrontal cortex mediates cue-driven feeding, while a parallel amygdalar circuitry mediates suppression of eating by the fear cue. Findings from the animal models may be relevant for understanding aspects of human appetite and eating, and maladaptive mechanisms that could lead to overeating and anorexia.

Keywords

Animal Models; Amygdala; Anorexia; Anxiety; Conditioning; Eating Disorders; Fear; Feeding; Hypothalamus; Learning; Memory; Motivation; Obesity; Prefrontal Cortex

1. Introduction

The motivation to eat is not only controlled by the physiological signals that convey energy and nutrient needs. Appetite and eating are also driven by environmental and social factors unrelated to homeostasis (for reviews see [1-11]). Notably, cues from the environment that acquire motivational properties through learning exert powerful control over food consumption. Learned cues can override homeostatic regulatory signals to stimulate eating

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in sated states, or to inhibit eating in states of hunger [12–15]. Such influences are important, and if persistent could lead to dysregulation of eating.

Indeed, environmental rather than metabolic changes are believed to underlie the increased susceptibility to overeating and the rise in obesity in the developed world [10, 16–20]. But, the changes that have been reshaping our environment are multifaceted, and could influence eating behavior by exceedingly complex means. Thus, the current obesity models also span across diverse functional systems that contribute to the regulation of feeding beyond the critical homeostatic and metabolic control [21]. Dysfunction in reward processing and the underlying brain systems and similarities with drug addiction have been proposed [22–27]. Other models have focused on the role of stress, or on a relationship between Western diet and cognitive impairments (for reviews see [28–31]). Still, an important facet of our environment is the prevalence of food cues.

Food-associated cues powerfully promote eating in laboratory animals and in humans [12, 13, 32]. Thus, it is easy to envision how in our environment, which is abundant in easily accessible palatable foods, the ubiquitous images and messages with cues for food that stimulate appetite could aid overeating.

In parallel with obesity, anorexia nervosa and other eating disorders are also more prevalent in Western societies, and have been on the rise [33–36]. Likewise, environmental and social factors are believed to impinge on the genetic background of the vulnerable to increase recruitment to eating disorders. Nevertheless, how environmental cues gain the ability to control feeding, and the underlying brain mechanisms remain largely unknown.

Recently, we developed two behavioral preparations to study how learned cues are integrated with homeostatic signals within functional forebrain networks. We use associative learning, Pavlovian conditioning, to enable initially neutral environmental signals to modulate food intake based on prior associations with either rewarding or aversive events. Thus, in one setting, a cue that signals food based on prior associations with food consumption when an animal was hungry, stimulates feeding in sated rats, *food cue induced feeding*. In the other setting, a cue that signals danger based on its prior pairings with an aversive event inhibits feeding in food-deprived rats, *fear cue induced cessation of feeding*.

Here we provide an overview of our recent findings and other evidence that learned cues modulate food consumption, and that the critical forebrain network includes the amygdala, lateral hypothalamus and medial prefrontal cortex. The findings in animals might also be informative for understanding the control of appetite in humans including maladaptive environmental influences that could lead to eating disorders.

2. Learning and the motivation to eat I: Cue-induced feeding

Others and we have shown that a cue that signals food can stimulate eating in stated states, and this ability is acquired through associative learning. We use a preparation that is based on the protocol by Weingarten [13], and work of Zamble [37], and behavioral aspects were described in our recent reviews [14, 38–40].

In brief, an initially neutral signal from the environment, such as a tone (conditioned stimulus, CS) is paired with food (unconditioned stimulus, US). That is the tone is repeatedly presented just prior to food delivery to food-deprived rats. The tone (CS) becomes a signal for food and it brings animals to the site of food delivery, the food cup. The amount of time spent at the food cup (conditioned response, CR) is a well-defined measure of associative learning, and during training rats learn to approach the food cup when the cue that predicts food is presented. A control stimulus, typically another auditory

cue, such as a noise, that is not followed by food delivery is also presented during training, and that cue does not bring rats to the food cup.

After training, sated rats are tested for food consumption during tests with cues presentations. The cue-induced feeding is evident in such tests; rats consume more food in the presence of the cue that signals food compared to tests with the presentation of the control cue. Importantly, cue-induced eating is not simply a byproduct of the CRs that bring the rats to the food cup. Enhanced eating also occurs in tests when food is presented in a receptacle that is different in appearance and location from the food cup used in training [41–43]. Thus, the cue's ability to stimulate eating is a motivational property acquired through conditioning.

We have typically used discrete cues as conditioned stimuli in our preparations [39]. Recently, we showed that the environment in which food is consumed during training also serve as a conditioned stimulus to promote eating [42]. In that protocol, we trained rats to consume food pellets in a distinct environment (context), while another control group of food-deprived rats were exposed to the same context, but received food pellets in their home cages. Then we tested sated rats for food pellet consumption in the conditioning context. Rats that were previously fed in the conditioning context when hungry consumed more food pellets in the conditioning context during tests compared to the rats in the control group that were never fed in that context. These results showed that contextual conditioned stimuli, similar to discrete cues, could promote food consumption, in agreement with a study in mice [44] and a recent study that used female rats [45].

The rodent cue-induced feeding model is relevant to human eating. Classical conditioning supports cue-driven eating in preschool children. When presented with a distinct song and a flashing light that were previously paired with snacks, sated children begin to eat faster and consume lager amounts compared to their consumption in the presence of another song and light that were not paired with snacks [32]. Additionally, cues associated with the sensory properties of the food itself such as a brief sight, smell, or taste of a food prior to a meal can stimulate sated individuals to eat [12]. This is exaggerated in restrained eaters (dieters) [46], and in obese children [47], suggesting heightened vulnerability in these populations to cue-triggered overeating. In that regard, obese children show bias for food-associated cues (words) [48], and obese women show attentional bias for food images regardless of hunger state [49] and exaggerated brain response (fMRI) to pictures of high-calorie foods compared to controls [50].

Similarities between the rodent and human data underscore that a common, fundamental mechanism supports the ability of learned cues to modulate feeding (also see section 2.3.). This, in turn, underscores the importance of animal models, which allow examination of the brain mechanisms at a level currently impossible with imaging methods in humans. Likewise, findings from human studies provide a valuable guide to future rodent experiments.

There are also some possible translational implications for treatment of overeating. Learning and associative cues serve an adaptive function (see section 2.3.), but are becoming maladaptive, for at least some humans in the developed world. Through these mechanisms, plentiful palatable foods and cues in our surroundings provide constant appetite stimulant. Thus, an obvious, and yet hard to achieve, strategy would be to limit the exposure to the cues for highly palatable, high-calorie foods. Another strategy might be to use associative learning to form new preferences and reminders for "healthy" foods.

2.1. Motivation underlying cue-induced feeding: Appetite for the training food

The motivational basis for feeding under the learned cue is acquired through associative learning, however its exact nature is not known. Recently we showed that it appears to involve specific drive for the training food, rather than a general drive to eat [42, 43]. We showed that sated rats enhanced consumption of the training pellets, but not other familiar, or novel foods in the conditioning context [42, 43]. These findings suggest that through conditioning the CS becomes a signal for the training food (US) specifically, rather than a signal for feeding.

Other recent studies corroborate our findings [51, 52]. In these studies rats were trained with two different CSs (tone or noise) that were each paired with a distinct food, US (sucrose or maltodextrine). Then rats were tested for food consumption during tests with presentations of either the CS that was previously paired with the test food, presentations of the CS for different food, or no CS. The cue-driven feeding occurred only in the presence of the CS for the test food, but not when the CS signaled the other food, in accordance with our findings [42, 43].

Collectively, these findings suggest that through associative learning the cue (CS) gains the ability to evoke a sensory-specific representation of the food (US). In turn selective consumption of the signaled food suggests induction of a motivational state similar to appetite, or craving rather than induction of huger. In that regard, studies in humans that primed subjects with brief food presentations induced specific desires for the food the subject was primed with, and the appetite was correlated with the amounts consumed [12, 53].

Additional features suggest parallels with food cravings, although such comparisons should be taken with caution because food cravings are difficult to define in animal models [54]. Nevertheless, both are food selective and can be elicited by exposure to cues associated with food [54, 55]. Furthermore, binge eating in humans is associated with cue-elicited cravings, and animals can consume a large amount in a very short time in the context associated with food [42, 43, 55, 56].

2.2. Forebrain circuitry for cue-induced feeding

The cue-induced feeding model provides a framework for analysis of the brain circuitry and plasticity that underlies integration between environmental and homeostatic signals. Our focus has been on the forebrain contribution and specifically the telencephalon communication with the lateral hypothalamus (LHA). Our studies build on the hypothesis that the LHA is an integrative site for signals underlying the motivation to eat that include the physiological signals from the body ("intrinsic"), and "extrinsic", such as environmental, emotional, and cognitive signals.

The LHA receives the inputs that could relay the physiological signals relevant for food intake regulation via the arcuate nucleus of the hypothalamus and other hypothalamic and brainstem areas, and the inputs from the telencephalic areas related to motivation, emotion, and cognition [2, 4, 57, 58]. In turn, the LHA sends widespread outputs to the brainstem and forebrain. Thus, the LHA is well positioned to contribute to its historically assigned functions, the initiation of feeding, reward, and motivation (for reviews see [2, 59] [4].

Within the telencephalon, areas well known for their roles in associative learning and decision-making, the amygdala and medial prefrontal cortex, respectively, send substantial input to the LHA. We examined each of these areas and showed that cue-driven food consumption critically depends on an intact basolateral area of the amygdala (BLA, includes basolateral, basomedial, and lateral nuclei), the ventromedial prefrontal cortex (vmPFC,

includes infralimbic, prelimbic and ventral medial orbitofrontal areas), and the BLA-LHA communications [41, 43, 60, 61].

The BLA communication with the LHA includes direct, and indirect pathways, and we used a preparation that disconnected both types of relays to show that the BLA-LHA system is necessary for cue-induced eating [61]. The vmPFC received input from the BLA and in turn sends output to the LHA and is thus, well positioned to relay the information between the BLA and LHA [57, 62–64]. Therefore, we examined whether the vmPFC and BLA neurons that send direct pathways to the LHA are functionally activated (immediate early gene induction) during the cue-induced feeding tests. To accomplish this we applied a combination of immediate early gene induction imaging method with detection of a retrograde tracer within the vmPFC and BLA after injections into the LHA. We showed that subpopulations of neurons within the vmPFC and BLA that send input to the LHA (retrogradely labeled) were selectively activated during the cue-induced feeding tests [65].

Then we targeted the vmPFC with bilateral, neurotoxic lesions, and found that the lesions abolished conditioned context-driven food consumption [43]. The vmPFC lesions in that study also produced changes in food consumption in novel settings. The vmPFC-lesioned rats consumed less than controls when fed in a novel environment, or when a novel food was first presented in a familiar environment. Importantly, the vmPFC-lesions deficits were specific to food consumption during tests and did not impact food intake in the home cage or body weight. Similarly, a recent study found no changes in body weight after lesions of the vmPFC region somewhat overlapping with the lesioned area in our study [66]. Collectively, these results suggest that the vmPFC might not be critical for the homeostatic control of food intake and body weight. Instead, it might be critically recruited when evaluation of environmental signals based on prior experience is required to guide goal-directed behaviors, such as eating under the learned cue or in a novel setting.

The specific roles of the basolateral amygdala, and prefrontal cortex in cue driven feeding remain unknown. Our lesions were made before behavioral training, and thus could have interfered with the learning acquisition, memory recall, or behavioral expression phase of the cue-induced feeding. The basolateral amygdala and prefrontal cortex are engaged differently in other tasks that are similar to our paradigm in that they also rely on CS properties acquired through associative learning to modulate behavior (devaluation task, and second order conditioning; [39]). The basolateral amygdala is needed during the acquisition phase, but not when the flexible use of the CS acquired value is used to modulate behavior in the devaluation task, and in the second order conditioning [67, 68]. In contrast, a region of the prefrontal cortex (the lateral orbitofrontal cortical area) is needed during both the acquisition, and expression phase in the devaluation task [68]. In that regard, the lateral orbitofrontal cortex encoding, and updating of the acquired associative value of the CS depends on communications with the BLA [69, 70].

Similar dissociable roles for the BLA and vmPFC have been shown in aversive conditioning. The vmPFC's responses to aversive CSs [71], and cannabinoid potentiation of learning plasticity within the vmPFC requires BLA input during the acquisition, but not once the association has been formed [72]. Thus, the basolateral amygdala plays a role in guiding prefrontal cortical responses during the learning acquisition based on the incentive value acquired through the association between environmental signals and rewards or punishments. This general framework might apply to the BLA-vmPFC system in cue-induced feeding.

The anatomical connections between the BLA, vmPFC, and LHA support this possibility. The BLA and vmPFC share extensive bidirectional connections and each send direct

pathways to the LHA [62–64, 73–76]. Thus, the BLA and vmPFC could modulate the LHA processing via direct pathways, as well as through the communications with each other.

Additional complexity in this system is provided by heterogeneity within the vmPFC. The distinct regions within the vmPFC (the infralimbic, ventral prelimbic, and ventro-medial orbitofrontal areas) might serve different functions in the appetitive processing, and associated motivation for food and drugs [77, 78]. For example, the initial choice to explore novel food, and responding to conflicting information guided by context is attributed to the prelimbic area [79, 80]. On the other hand the infralimbic area, which is critical for extinction of aversive CSs, also supports functions associated with extinction of appetitive CSs [81–83]. Notably, cue-induced cravings for food and drugs in humans are attributed to the medial orbitofrontal area [84] [85–87], a region that appears somewhat homologous to the ventro-medial orbitofrontal area in rats [43]. Nevertheless, whether the ventral medial orbitofrontal area might be necessary for the cue memory recall and subsequent induction of the drive to eat remains to be determined.

2.3. Learning and the homeostatic control

In our preparations we pit learned cues against homeostatic signals to stimulate eating in sated states or inhibit feeding in states of hunger. However, associative learning endows an organism with an adaptive function to facilitate regulation of feeding and other goal-directed behaviors.

As such associative learning supports acquisition of preferences and aversions to foods and associated cues and environments based on taste and post-ingestive consequences (for reviews see [38, 88–90]). Associative learning and memory are also critical in reward processing mechanisms that contribute to regulation of feeding, and much has been elucidated about the underlying brain systems (for reviews see [23, 91–94]).

Importantly, learning and associated anticipatory motivation function in concert with the homeostatic regulation [95–97]. The ability to predict a meal, and the anticipatory motivation prepare the body for the incoming nutrients, and as such assist homeostatic regulation [95–97]. The well-known example is control of insulin, a hormone released by the pancreas that peripherally regulates glucose metabolism, and acts as an adiposity signal in the brain [98].

Insulin is regulated by both the physiological and anticipatory signals. Its release is triggered by a meal-induced increase in blood glucose, as well as by the cues that predict a meal [95], and gain control through conditioning [99]. The anticipatory insulin release ("cephalic insulin") is regulated by the brain via the vagus nerve, and is an adaptive response that helps prevent hyperglycemia that would otherwise occur because of the delay in meal-induced insulin cascade [95, 100].

Other physiological signals and associated brain regulatory substrates are also under anticipatory control. A recent study examined the role of ghrelin in food anticipatory functions with ghrelin-receptor deficient mice [101]. Ghrelin is an orexigenic peptide released by the stomach before meals that acts through the vagus nerve to reach the brain substrates including ghrelin-producing neurons [102, 103]. Thus, ghrelin receptors in the body and the brain are necessary for ghrelin homeostatic signaling and function. Interestingly, Davis and colleagues found that ghrelin-receptor signaling is also necessary for adaptive, food anticipatory activity, and modulation of spatial memory [101].

Finally, the brain feeding regulatory systems can be conditioned to function independent from the physiological signals that normally regulate them. This was shown in a recent study

[104] with exogenous manipulation of a potent feeding stimulant, the neuropeptide Y (NPY) [105]. In the study of Drazen and colleagues, NPY was injected into the brain repeatedly at the same time of the day, and then at testing the injection was omitted to examine the consequences of anticipation of the NPY burst based on the time cue [104]. The anticipation of the NPY burst induced eating that was comparable to that produced by the actual NPY injection. Thus, the learned cue (injection time) was able to recruit the NPY input-dependent substrates to initiate feeding in the absence of the physiological signals.

In turn, these findings might provide insights into the mechanisms underlying food-cue driven feeding that also occurs in the absence of physiological hunger. In particular the plasticity within the NPY input system is of interest. In that regard, the NPY is important for the motivation to eat [106], and can initiate feeding under sated conditions. Importantly, its ability to stimulate feeding in satiated rats is mediated via the LHA [107].

2.3.1. Cue integration with the hypothalamic homeostatic system—As follows from the above discussion, the NPY input system within the lateral hypothalamus might be important for convergence between the telencephalic influences and homeostatic regulatory control. The anatomical evidence supports this possibility. The NPY neurons from the ARH send input to the LHA, where they end on the neurons that express orexigenic neuropeptides, melanin concentrating hormone (MCH) and orexin/hypocretin (ORX) [3, 108]. The BLA, and vmPFC also send direct pathways to the LHA [58, 62–64, 109], and the vmPFC innervates the area with ORX- and MCH-neurons [2, 58, 108–110].

The two orexigenic peptides appear to sub serve different functions in the control of food intake. The MCH role in the homeostatic regulation is well established. The MCH injections induce feeding and its mRNA is increased by fasting, and is over expressed in genetically obese (ob/ob) mice [111]. Genetic manipulations that eliminated [112] or enhanced MCH production [113] confirmed its importance in food intake and body weight regulation.

On the other hand, the role of ORX system in feeding [114] might be related to adaptive functions that complement homeostatic control. The ORX system, which is critical for control of wakefulness, mediates arousal in response to fasting, and contributes to other functions that depend on behavioral states, including reward [115–117]. Accordingly, neurons that produce ORX, which are located exclusively in the LHA, have the capability to influence diverse brain systems via widespread pathways, and similarly broadly distributed receptors [110, 118, 119].

Importantly, the ORX system is involved in processing of food and drug rewards via circuitry that engages the LHA, nucleus accumbens, ventral tegmental area, and the prefrontal cortex (e.g., [120–124]). Notably, ORX neurons have been shown to respond to contextual cues for food and drugs in the conditioned place preference task [122, 125]. Similarly, a recent study showed that the ORX neurons located in the perifornical area responded to the contextual cues that were previously paired with food [126]. Interestingly, the ORX neurons responded regardless of the type of food the context signaled, while the prefrontal cortex was selectively activated when the context signaled palatable food (chocolate), but not when the context signaled an ordinary meal (lab chow) [126]. These findings highlight the role of ORX neurons in food anticipatory mechanisms, and suggest that communication with the prefrontal cortex, and other forebrain and brainstem targets, might be important for dissociation between general and specific food motivations.

In accordance, our preliminary findings suggest that the ORX neurons are also important for cue-induced feeding (unpublished observations). Thus, we have begun work that examines

whether the ORX and MCH neuron are engaged (immediate early gene induction) by the learned cue that stimulates feeding.

Other hypothalamic regions that are under telencephalic influences via indirect pathways might also be important for cue-driven feeding. A polysynaptic pathway could allow the BLA influence on the arcuate nucleus of the hypothalamus [127], which is considered a primary sensory area for the physiological signals related to energy needs [3]. Similarly, the amygdala and prefrontal cortex could reach another critical node of the hypothalamic feeding circuitry, the paraventricular nucleus of the hypothalamus [2–4] via indirect pathways through the bed nuclei of the stria terminalis [57, 58, 64, 128] [57].

3. Learning and the motivation to eat II: Fear cue induced cessation of feeding

In parallel with cue-induced feeding we have been developing a preparation that uses learned cues to inhibit feeding. In that protocol a cue that predicts danger (fear cue) inhibits food intake in food-deprived rats [15, 40]. Similar to food cue's ability to stimulate feeding in sated states, fear cue's ability to inhibit feeding in hungry states is acquired through associative learning. As such both behavioral preparations provide models for integration between the environmental and homeostatic signals, and for mapping of the underlying brain networks.

Our fear-induced feeding cessation model builds on well-established aversive, fear conditioning paradigms (e.g., [129–132]). In these preparations an initially neutral, environmental signal such as a tone (conditioned stimulus, CS) is paired with an aversive event such as a mild, electric, foot-shock (unconditioned stimulus, US). Through associative learning the tone becomes a predictor for the shock, and this is manifested in its acquired ability to produce fear-related behavioral responses (conditioned responses, CRs).

We use aversive conditioning to modulate feeding. In our preparation, rats receive toneshock pairings during aversive training sessions, and are given food pellets that they consume during appetitive training sessions. The aversive and appetitive training sessions occur in alternating order, and are conducted in distinct environments (contexts). After training, rats are tested for food consumption during tests with tone (CS; fear cue) presentations in the appetitive context. Prior to testing rats are food deprived, and accordingly rats in the control condition (that did not receive tone-shock pairings during training) consume substantial amounts of food during the tests. Rats that previously received tone-shock pairings consume much less food than rats in the control condition. Thus, the fear cue effectively competes with the homeostatic signals induced by food deprivation to inhibit feeding.

We also measure CS-induced freezing behavior during food consumption tests. Freezing is a species-typical defense response that has been extensively studied in rodent fear conditioning paradigms. It is an easily recognized behavior, characterized by the absence of all movement except that required for breathing [133, 134], During our food consumption tests with tone (CS) presentations, rats show both behavioral responses, the freezing behavior and the inhibition of feeding. Importantly, the CS-induced feeding cessation is not merely a consequence of immobilization due to CS-induced conditioned freezing behavior. We showed previously that the CS-induced feeding cessation and CS-induced freezing are dissociable behavioral responses induced by the same CS. Brain lesions that abolished conditioned freezing left inhibition of eating intact—lesions of the ventrolateral region of the periaqueductal gray [135], an area critical for conditioned freezing [136], or lesions of the basolateral amygdala [15]. Thus, the CS's influence on feeding is not directly depended

on CS-induced freezing, and engages somewhat dissociable amygdalar and brainstem substrates.

Nevertheless, cessation of eating and freezing behavior are both components of a preparatory motivational system critical for defensive behavior triggered by the CS. Cessation of eating in anticipation of danger is an adaptive response that prepares an organism for an imminent threat, but could become maladaptive when persistent. Sustained fear is associated with anxiety, and its effects on food intake might relate to aspect of disordered eating in humans.

Anorexia nervosa is an eating disorder characterized by relentless maintenance of extremely low body weight through restricted eating, which is often combined with excessive exercise and even purging [34–36, 137]. The ability to maintain restricted eating in emaciated states is paradoxical, and occurs in conflict with physiological hunger signals [36, 138]. We hypothesize that another core symptom of anorexia nervosa might be informative, the obsessive fear of weight gain despite being underweight [137].

Fear inhibits food intake [139, 140]. Thus, the sustained fear might not only be the key symptom, but also an important contributor that facilitates the maintenance of restricted eating in anorexia. In that regard, anorexia nervosa shows high co-morbidity with anxiety disorders (reviewed in [36]), and support for fear network dysfunction has been found in recent imaging studies with anorexia nervosa patients.

Within the amygdala, abnormal functioning, and a decrease in its volume were found in anorexia patients [141, 142]. Enhanced medial prefrontal cortex activity was associated with responses to food images that the patients with eating disorders (anorexia and bulimia) found threatening and disgusting [143]. Similarly, greater amygdala activation was found among anorexia patients when confronted with a threatening, symptom-provoking cue—their distorted body image [144]. Interestingly, amygdala recruitment was also correlated with increased anxiety in non-eating disordered young women when viewing pictures of slim, idealized female bodies [145].

Thus, our fear cue-induced feeding cessation rodent model is relevant to human eating, and provides a behavioral framework for defining the critical brain substrates. We have begun the analysis of the brain systems that allow fear cues access to the feeding circuitry, and the findings are discussed next.

3.1. The forebrain circuitry for fear cue feeding cessation

We began to delineate the forebrain circuitry underlying fear cue-induced feeding cessation in a study that examined the involvement of amygdalar subregions. The central nucleus (CEA) and the BLA are necessary components of the conditioned fear circuit [129–132, 146], and have been linked to a range of functions that rely on associative learning to control goal-directed behaviors (for reviews see [92, 129–132, 146–132, 148]. Furthermore, relevant to fear cue-mediated regulation of feeding the BLA and CEA could both influence hypothalamic and brainstem function via distributed network of direct and indirect connections [58, 64]. Thus we examined whether one or both of these structures are critical.

We found that bilateral, neurotoxic lesions of the central amygdala, but not bilateral, neurotoxic lesions of the basolateral amygdala, abolished CS-induced feeding cessation [15]. On the other hand, lesions of each of the structures abolished CS-induced freezing behavior. Thus, both the BLA- and CEA-lesioned rats greatly reduced freezing compared to the control (sham-lesioned) rats, but only the CEA-lesioned rats failed to inhibit feeding

during the same test. Thus, the data showed that the CS's influence on feeding is independent of CS-induced freezing, and engages dissociable amygdalar subsystems.

The CEA influence on food consumption could be accomplished via complex output network that reaches multiple components of the feeding system via direct and indirect pathways. The CEA sends direct projections to the brainstem, lateral hypothalamus, and bed nuclei of the stria terminalis [64, 128, 149], as well as indirect pathways to the paraventricular nucleus of the hypothalamus [128, 150]. Thus, future studies will examine where within the distributed feeding network the CEA exerts its action, and the plasticity that underlies learned cue inhibition of feeding [3–6, 10, 64, 81, 151].

Similarly, future studies are needed to address whether repeated exposure to aversive events, and associated fear and anxiety, might contribute to long-term suppression of eating and changes in body weight. The fear cue model presented here, therefore, provides a framework for future brain and behavioral analyses.

4. Conclusion

In summary, we have developed two behavioral preparations that rely on associative learning to acquire motivational powers to modulate feeding independent of existing hungersatiety state. In one preparation a cue becomes a signal for food and gains the ability to stimulate consumption of that food in sated states. In the other preparation a cue becomes a signal for an imminent danger and gains the ability to powerfully inhibit feeding in hungry states. As such these models are helpful for analysis of the integration between the systems that process associative learning (the amygdala, prefrontal cortex and associated network) and the hypothalamic and brainstem feeding regulatory systems.

Thus far, our work has identified several critical components of the forebrain circuitry for cue-induced feeding. Nevertheless, the entire network, which is undoubtedly complex, and highly interconnected, remains unknown. Even less is known about the circuitry through which fear and anxiety inhibit feeding. Thus, determining the critical brain circuitries remains an important step in illuminating the mechanisms underlying environmental contribution to the regulation and dysregulation of food intake and body weight.

On a higher level, these models should provide a framework for understanding the role of the telencephalon in control of feeding and other goal-directed behaviors in the context of learning and anticipatory motivation. Ultimately they should help elucidate how the amygdala and its associated network processes biologically relevant sensory stimuli and modulates behavior accordingly.

Research Highlights

- Food cues stimulate feeding in sated states (cue-induced feeding model).
- Cue feeding circuit: basolateral amygdala, prefrontal cortex, lateral hypothalamus.
- Fear cues inhibit feeding in hungry states (fear-induced anorexia model).
- Fear cue-induced feeding cessation depends on the central nucleus of the amygdala.

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References

- Booth DA. Mood- and nutrient-conditioned appetites. Cultural and physiological bases for eating disorders. Ann. N.Y. Acad. Sci. 1989; 575:122–135. [PubMed: 2699183]
- 2. Elmquist JK, Elias CF, Saper CB. From lesions to leptin: Hypothalamic control of food intake and body weight. Neuron. 1999; 22:221–232. [PubMed: 10069329]
- Schwartz MW, Woods SC, Porte DJ, Seeley RJ, Baskin DG. Central nervous system control of food intake. Nature. 2000; 404:661–671. [PubMed: 10766253]
- Swanson LW. Cerebral hemisphere regulation of motivated behavior. Brain Research. 2000; 886:113–164. [PubMed: 11119693]
- Grill HJ, Kaplan JM. The neuroanatomical axis for control of energy balance. Front Neuroendocrinol. 2002; 23:2–40. [PubMed: 11906202]
- Saper CB, Chou TC, Elmquist JK. The need to feed: homeostatic and hedonic control of eating. Neuron. 2002; 36:199–211. [PubMed: 12383777]
- Moran TH. Gut peptide signaling in the controls of food intake. Obesity. 2006; 14(Suppl 5):250S– 253S. [PubMed: 17021376]
- Morton GJ, Cummings DE, Baskin DG, Barsh GS, Schwartz MW. Central nervous system control of food intake and body weight. Nature. 2006; 443:289–295. [PubMed: 16988703]
- Stroebele N, de Castro JM. Effect of ambience on food intake and food choice. Nutrition. 2004; 20:821–838. [PubMed: 15325695]
- Berthoud HR. Interactions between the "cognitive" and "metabolic" brain in the control of food intake. Physiol Behav. 2007; 91:486–498. [PubMed: 17307205]
- Herman CP, Polivy J. External cues in the control of food intake in humans: The sensorynormative distinction. Physiol Behav. 2008; 94:722–728. [PubMed: 18499202]
- Cornell CE, Rodin J, Weingarten H. Stimulus-induced eating when satiated. Physiol Behav. 1989; 45:695–704. [PubMed: 2780836]
- Weingarten HP. Conditioned cues elicit feeding in sated rats: A role for learning in meal initiation. Science. 1983; 220:431–433. [PubMed: 6836286]
- 14. Petrovich GD, Gallagher M. Control of food consumption by learned cues: A forebrainhypothalamic network. Physiol Behav. 2007; 91:397–403. [PubMed: 17498758]
- Petrovich GD, Ross CA, Mody P, Holland PC, Gallagher M. Central but not basolateral amygdala is critical for control of feeding by aversive conditioned cues. J Neurosci. 2009; 29:15205–15212. [PubMed: 19955373]
- Schachter S. Obesity and eating. Internal and external cues differentially affect the eating behavior of obese and normal subjects. Science. 1968; 161:751–756. [PubMed: 5663800]
- Hill JO, Wyatt HR, Reed GW, Peters JC. Obesity and the environment: where do we go from here? Science. 2003; 161:751–756.
- Levitsky DA. The non-regulation of food intake in humans: hope for reversing the epidemic of obesity. Physiol Behav. 2005; 86:623–632. [PubMed: 16263145]
- 19. Popkin BM, Duffey K, Gordon-Larsen P. Environmental influences on food choice, physical activity and energy balance. Physiol Behav. 2005; 86:603–613. [PubMed: 16246381]
- 20. Berthoud HR, Morrison C. The brain, appetite, and obesity. Annu Rev Psychol. 2008; 59:55–92. [PubMed: 18154499]
- 21. Friedman JM. Obesity in the new millennium. Nature. 2000; 404:632–634. [PubMed: 10766249]
- 22. Volkow ND, Wise RA. How can drug addiction help us understand obesity? Nature Neurosci. 2005; 8:555–560. [PubMed: 15856062]
- Cota D, Tschöp MH, Horvath TL, Levine AS. Cannabinoids, opioids and eating behavior: the molecular face of hedonism? Brain Research Reviews. 2006; 51:85–107. [PubMed: 16364446]

- Trinko R, Sears RM, Guarnieri DJ, DiLeone RJ. Neural mechanisms underlying obesity and drug addiction. Physiol Behav. 2007; 91:499–505. [PubMed: 17292426]
- Small DM. Individual differences in the neurophysiology of reward and the obesity epidemic. International Journal of Obesity. 2009; 33:S44–S48. [PubMed: 19528979]
- Berridge KC, Ho CY, Richard JM, DiFeliceantonio AG. The tempted brain eats: pleasure and desire circuits in obesity and eating disorders. Brain Research. 2010; 1350:43–64. [PubMed: 20388498]
- 27. Cason AM, Smith RJ, Tahsili-Fahadan P, Moorman DE, Sartor GC, Aston-Jones G. Role of orexin/hypocretin in reward-seeking and addiction: implications for obesity. 2010
- Hagan MM, Wauford PK, Chandler PC, Jarrett LA, Rybak RJ, Blackburn K. A new animal model of binge eating: Key synergistic role of pars caloric restriction and stress. physiology & Behavior. 2002; 77:45–54. [PubMed: 12213501]
- Tamashiro KL, Hegeman MA, Nguyen MM, Melhorn SJ, Ma LY, Woods SC, et al. Dynamic body weight and body composition changes in response to subordination stress. Physiol Behav. 2007; 91:440–448. [PubMed: 17512562]
- Dallman MF. Stress-induced obesity and the emotional nervous system. Trends Endocrinol Metab. 2009; 21:159–165. [PubMed: 19926299]
- 31. Kanoski SE, Davidson TL. Western diet consumption and cognitive impairment: Links to hippocampal dysfunction and obesity. Physiol Behav. 2010 Epub ahead of print.
- Birch LL, McPhee L, Sullivan S, Johnson S. Conditioned meal initiation in young children. Appetite. 1989; 13:105–113. [PubMed: 2802592]
- Bulik CM. Exploring the gene-environment nexus in eating disorders. J Psychiatry Neurosci. 2005; 30:335–339. [PubMed: 16151538]
- Treasure J, Claudino AM, Zucker N. Eating disorders. Lancet. 2010; 375:583–593. [PubMed: 19931176]
- Klein DA, Walsh BT. Eating disorders: clinical features and pathophysiology. Physiol Behav. 2004; 81:359–374. [PubMed: 15159176]
- Kaye W. Neurobiology of anorexia and bulimia nervosa. Physiol Behav. 2008; 94:121–135. [PubMed: 18164737]
- 37. Zamble E. Augmentation of eating following a signal for feeding in rats. Learn Motiv. 1973; 4:138–142.
- Petrovich GD, Gallagher M. Amygdala sybsystems and control of feeding behavior by learned cues. Ann NY Acad Sci. 2003; 985:251–262. [PubMed: 12724163]
- Holland PC, Petrovich GD. A neural systems analysis of the potentiation of feeding by conditioned stimuli. Physiol Behav. 2005; 86:747–761. [PubMed: 16256152]
- Petrovich GD. Forebrain circuits and control of feeding by learned cues. Neurobiol Learn Mem. 2010 Epub.
- 41. Holland PC, Petrovich GD, Gallagher M. The effects of amygdala lesions on conditioned stimuluspotentiated eating in rats. Physiology & Behavior. 2002; 76:117–129. [PubMed: 12175595]
- 42. Petrovich GD, Ross CA, Gallagher M, Holland PC. Learned contextual cue potentiates eating in rats. Physiol Behav. 2007; 90:362–367. [PubMed: 17078980]
- Petrovich GD, Ross CA, Holland PC, Gallagher M. Medial prefrontal cortex is necessary for an appetitive contextual conditioned stimulus to promote eating in sated rats. J Neurosci. 2007; 27:6436–6441. [PubMed: 17567804]
- 44. Le Merrer J, Stephens DN. Food-induced behavioral sensitization, its cross-sensitization to cocaine and morphine, pharmacological blockade, and effect on food intake. J Neurosci. 2006; 26:7163– 7171. [PubMed: 16822973]
- Boggiano MM, Dorsey JR, Thomas JM, Murdaugh DL. The Pavlovian power of palatable food: lessons for weight-loss adherence from a new rodent model of cue-induced overeating. Int J Obes. 2009; 33:693–701.
- Fedoroff IC, Polivy J, Herman CP. The effect of pre-exposure to food cues on the eating behavior of restrained and unrestrained eaters. Appetite. 1997; 28:33–47. [PubMed: 9134093]

- 47. Jansen A, Theunissen N, Slechten K, Nederkoorn C, Boom B, Mulkens S, et al. Overweight children overeat after exposure to food cues. Eating Behaviors. 2003; 4:197–209. [PubMed: 15000982]
- Braet C, Crombez G. Cognitive interference due to food cues in childhood obesity. Journal of clinical child and adolescent psychology. 2003; 32:32–39. [PubMed: 12573930]
- 49. Castellanos EH, Charboneau E, Dietrich MS, Park S, Bradley BP, Mogg K, et al. Obese adults have visual attention bias for food cue images: evidence for altered reward system function. Int J Obes. 2009; 33:1063–1073.
- Stoeckel LE, Weller RE, Cook EWr, Twieg DB, Knowlton RC, Cox JE. Widespread rewardsystem activation in obese women in response to pictures of high-calorie foods. Neuroimage. 2008; 41:636–647. [PubMed: 18413289]
- Galarce EM, Crombarg HS, Holland PC. Reinforcer-specificity of appetitive and consummatory behavior of rats after Pavlovian conditioning with food reinforcers. Physiol Behav. 2007; 91:95– 105. [PubMed: 17346758]
- Delamater AR, Holland PC. The influence of CS-US interval on several different indices of learning in appetitive conditioning. Journal of Experimental Psychology: Animal Behavior Processes. 2008; 34:202–222. [PubMed: 18426304]
- Fedoroff I, Polivy J, Herman CP. The specificity of restrained versus unrestrained eaters' responses to food cues: general desire to eat, or craving for the cued food? Appetite. 2003; 41:7–13. [PubMed: 12880616]
- 54. Weingarten HP, Elston D. The phenomenology of food craving. Appetite. 1990; 15:231–246. [PubMed: 2281953]
- 55. Jansen A. A learning model of binge eating: cue reactivity and cue exposure. Behav Res Ther. 1998; 36:257–272. [PubMed: 9642846]
- Sobik L, Hutchison K, Craighead L. Cue-elicited craving for food: a fresh approach to the study of binge eating. Appetite. 2005; 44:253–261. [PubMed: 15876472]
- Risold PY, Thompson RH, Swanson LW. The structural organization of connections between hypothalamus and cerebral cortex. Brain Research Reviews. 1997; 24:197–254. [PubMed: 9385455]
- Petrovich GD, Canteras NS, Swanson LW. Combinatorial amygdalar inputs to hippocampal domains and hypothalamic behavior systems. Brain Research Reviews. 2001; 38:247–289. [PubMed: 11750934]
- Wise RA. Lateral hypothalamic electrical stimulation: does it make animals "hungry"? Brain Research. 1974; 67:187–209. [PubMed: 4620218]
- 60. Holland PC, Hatfield T, Gallagher M. Rats with basolateral amygdala lesions show normal increases in conditioned stimulus processing but reduced conditioned potentiation of eating. Behavioral Neuroscience. 2001; 115:945–950. [PubMed: 11508734]
- Petrovich GD, Setlow B, Holland PC, Gallagher M. Amygdalo-hypothalamic circuit allows learned cues to override satiety and promote eating. Journal of Neuroscience. 2002; 22:8748– 8753. [PubMed: 12351750]
- 62. Sesack SR, Deutch AY, Roth RH, Bunney BS. Topographical Organization of the Efferent Projections of the Medial Prefrontal Cortex in the Rat: An Anterograde Tract-Tracing Study With *Phaseolus vulgaris* Leucoagglutinin. Journal of Comparative Neurology. 1989; 290:213–242. [PubMed: 2592611]
- 63. Hurley KM, Herbert H, Moga MM, Saper CB. Efferent Projections of the Infralimbic Cortex of the Rat. Journal of Comparative Neurology. 1991; 308:249–276. [PubMed: 1716270]
- 64. Swanson LW, Petrovich GD. What is the amygdala? Trends Neurosci. 1998; 21:323–331. [PubMed: 9720596]
- Petrovich GD, Holland PC, Gallagher M. Amygdalar and prefrontal pathways to the lateral hypothalamus are activated by a learned cue that stimulates eating. J Neurosci. 2005; 25:8295– 8302. [PubMed: 16148237]
- 66. Davidson TL, Chan K, Jarrard LE, Kanoski SE, Clegg DJ, Benoit SC. Contributions of the hippocampus and medial prefrontal cortex to energy and body weight regulation. Hippocampus. 2009; 19:235–252. [PubMed: 18831000]

- 67. Setlow B, Gallagher M, Holland PC. The basolateral complex of the amygdala is necessary for acquisition but not expression of CS motivational value in appetitive Pavlovian second-order conditioning. European Journal of Neuroscience. 2002; 15:1841–1853. [PubMed: 12081664]
- Pickens CL, Saddoris MP, Setlow B, Gallagher M, Holland PC, Schoenbaum G. Different roles for orbitofrontal cortex and basolateral amygdala in a reinforcer devaluation task. J Neurosci. 2003; 23:11078–11084. [PubMed: 14657165]
- 69. Schoenbaum G, Setlow B, Saddoris MP, Gallagher M. Encoding predicted outcome and acquired value in orbitofrontal cortex during cue sampling depends upon input from basolateral amygdala. Neuron. 2003; 39:855–867. [PubMed: 12948451]
- 70. Stalnaker TA, Franz TM, Dingh T, Schoenbaum G. Basolateral amygdala lesions abolish obrbitofrontal-dependent reversal impairments. Neuron. 2007; 54:51–58. [PubMed: 17408577]
- 71. Laviolette SR, Lipski WJ, Grace AA. A subpopulation of neurons in the medial prefrontal cortex encodes emotional learning with burst and frequency codes through a dopamine D4 receptordependent basolateral amygdala input. J Neurosci. 2005; 25:6066–6075. [PubMed: 15987936]
- Laviolette SR, Grace AA. Cannabinoids Potentiate Emotional Learning Plasticity in Neurons of the Medial Prefrontal Cortex through Basolateral Amygdala Input. J Neurosci. 2006; 26:6458–6468. [PubMed: 16775133]
- 73. Krettek JE, Price JL. Projections from the amygdaloid complex to the cerebral cortex and thalamus in the rat and cat. J. Comp. Neurol. 1977; 172:687–722. [PubMed: 838895]
- 74. Kita H, Kitai ST. Amygdaloid Projections to the Frontal Cortex and the Striatum in the Rat. Journal of Comparative Neurology. 1990; 298:40–49. [PubMed: 1698828]
- Gabbott PL, Warner TA, Jays PR, Salway P, Busby SJ. Prefrontal cortex in the rat: projections to subcortical autonomic, motor, and limbic centers. J Comp Neurol. 2005; 492:145–177. [PubMed: 16196030]
- 76. Hoover WB, Vertes RP. Anatomical analysis of afferent projections to the medial prefrontal cortex in the rat. Brain structure & function. 2007; 212:149–179. [PubMed: 17717690]
- Dalley JW, Cardinal RN, Robbins TW. Prefrontal executive and cognitive functions in rodents: neural and neurochemical substrates. Neuroscience and Biobehavioral Reviews. 2004; 28:771– 784. [PubMed: 15555683]
- Schroeder BE, Binzak JM, Kelley AE. A common profile of prefrontal cortical activation following exposure to nicotine- or chocolate-associated contextual cues. Neuroscience. 2001; 105:535–545. [PubMed: 11516821]
- Burns LH, Annett L, Kelley AE, Everitt BJ, Robbins TW. Effects of lesions to amygdala, ventral subiculum, medial prefrontal cortex, and nucleus accumbens on the reaction to novelty: implication for limbic-striatal interactions. Behavioral Neuroscience. 1996; 110:60–73. [PubMed: 8652073]
- Marquis J-P, Killcross AS, Haddon JE. Inactivation of the prelimbic, but not infralimbic prefrontal cortex impairs the contextual control of response conflict in rats. European Journal of Neuroscience. 2007; 25:559–566. [PubMed: 17284198]
- Quirk GJ, Garcia R, Gonzalez-Lima F. Prefrontal mechanisms in extinction of conditioned fear. Biol Psychiatry. 2006; 60:337–343. [PubMed: 16712801]
- 82. Rhodes SEV, Killcross S. Lesions of rat infralimbic cortex enhance recovery and reinstatement of an appetitive Pavlovian response. Learn Mem. 2004; 11:611–616. [PubMed: 15466316]
- 83. Peters J, LaLumiere RT, Kalivas PW. Infralimbic prefrontal cortex is responsible for inhibiting cocaine seeking in extinguished rats. J Neurosci. 2008; 28:6046–6053. [PubMed: 18524910]
- Hinton EC, Parkinson JA, Holland AJ, Arana FS, Roberts AC, Owen AM. Neural contributions to the motivational control of appetite in humans. European Journal of Neuroscience. 2004; 20:1411– 1418. [PubMed: 15341613]
- Arana FS, Parkinson JA, Hinton E, Holland AJ, Owen AM, Roberts AC. Dissociable contributions of the human amygdala and orbitofrontal cortex to incentive motivation and goal selection. J Neurosci. 2003; 23:9632–9638. [PubMed: 14573543]
- 86. Small DM, Zatorre RJ, Dagher A, Evans AC, Jones-Gotman M. Changes in brain activity related to eating chocolate: from pleasure to aversion. Brain. 2001; 124:1720–1733. [PubMed: 11522575]

- Brody AL, Mandelkern MA, London ED, Childress AR, Lee GS, Bota RG, et al. Brain metabolic changes during cigarette craving. Arch Gen Psychiatry. 2002; 59:1162–1172. [PubMed: 12470133]
- Yamamoto T, Shimura T, Sako N, Yasoshima Y, Sakai N. Neural substrates for conditioned taste aversion in the rat. Behav. Brain Res. 1994; 65:123–137. [PubMed: 7718144]
- Bernstein IL. Taste aversion learning: a contemporary perspective. Nutrition. 1999; 15:229–234. [PubMed: 10198919]
- Sclafani A. Oral and postoral determinants of food reward. Physiology & Behavior. 2004; 81:773– 779. [PubMed: 15234183]
- 91. Baxter MG, Murray EA. The amygdala and reward. Nature Rev. Neurosci. 2002; 3:563–573. [PubMed: 12094212]
- 92. Cardinal RN, Parkinson JA, Hall J, Everitt BJ. Emotion and motivation: the role of the amygdala, ventral striatum, and prefrontal cortex. Neurosci. Biobehav. Rev. 2002; 26:321–352. [PubMed: 12034134]
- Kelley AE, Baldo BA, Pratt WE, Will MJ. Corticostriatal-hypothalamic circuitry and food motivation: integration of energy, action and reward. Physiol Behav. 2005; 86:773–795. [PubMed: 16289609]
- Berridge KC, Robinson TE, Aldridge JW. Dissecting components of reward: 'liking', 'wanting', and learning. Curr Opin Pharmacol. 2009; 9:65–73. [PubMed: 19162544]
- 95. Woods SC. The eating paradox: How we tolerate food. Psychol Rev. 1991; 98:488–505. [PubMed: 1961770]
- Woods SC, Ramsay DS. Pavlovian influence over food and drug intake. Behavioural Brain Research. 2000; 110:175–182. [PubMed: 10802313]
- 97. Berridge KC. Motivation concepts in behavioral neuroscience. Physiol Behav. 2004; 81:179–209. [PubMed: 15159167]
- 98. Woods SC. Signals that influence food intake and body weight. Physiol Behav. 2005; 86:709–716. [PubMed: 16260007]
- Woods SC, Vasselli JR, Kaestner E, Szakmary GA, Milburn P, Vitiello MV. Conditioned insulin secretion and meal feeding in rats. J Comp Physiol Psychol. 1977; 91:128–133. [PubMed: 838910]
- Berthoud HR, Bereiter DA, Trimble ER, Siegel EG, Jeanrenaud B. Cephalic phase, reflex insulin secretion. Neuroanatomical and physiological characterization. Diabetologia. 1981; (Supppl): 393–401. [PubMed: 7014335]
- 101. Davis JF, Choi DL, Clegg DJ, Benoit SC. Signaling through the ghrelin receptor modulates hippocampal function and meal anticipation in mice. Physiol Behav. 2010 Epub.
- 102. Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. Diabetes. 2001; 50:1714– 1719. [PubMed: 11473029]
- 103. Castañeda TR, Tong J, Datta R, Culler M, Tschöp MH. Ghrelin in the regulation of body weight and metabolism. Front Neuroendocrinol. 2010; 31:44–60. [PubMed: 19896496]
- 104. Drazen DL, Wortman MD, Seeley RJ, Woods SC. Neuropeptide Y prepares rats for scheduled feeding. Am J Physiol Regul Integr Comp Physiol. 2005; 288:R1606–R1611. [PubMed: 15695319]
- 105. Leibowitz SF. Specificity of hypothalamic peptides in the control of behavioral and physiological processesses. Ann. N. Y. Acad. Sci. 1994; 739:12–35. [PubMed: 7530429]
- 106. Flood JF, Morley JE. Increased food intake by neuropeptide Y is due to an increased motivation to eat. Peptides. 1991; 12:1329–1332. [PubMed: 1815219]
- 107. Stanley BG, Chin AS, Leibowitz SF. Feeding and drinking elicited by central injection of neuropeptide Y: evidence for a hypothalamic site(s) of action. Brain Research Bulletin. 1985; 14:521–524. [PubMed: 3839709]
- 108. Elias CF, Saper CB, Maratos-Flier E, Tritos NA, Lee C, Kelly J, et al. Chemically defined projections linking the mediobasal hypothalamus and the lateral hypothalamic area. Journal of comparative neurology. 1998; 402:442–459. [PubMed: 9862320]

- 109. Floyd NS, Price JL, Ferry AT, Keay KA, Bandler R. Orbitomedial prefrontal cortical projections to hypothalamus in the rat. Journal of Comparative Neurology. 2001; 432:307–328. [PubMed: 11246210]
- 110. Swanson LW, Sanchez-Watts G, Watts AG. Comparison of melanin-concentrating hormone and hypocretin/orexin mRNA expression patterns in a new parceling scheme of the lateral hypothalamic zone. Neurosci Lett. 2005; 387:80–84. [PubMed: 16084021]
- 111. Qu D, Ludwig DS, Gammeltoft S, Piper M, Pelleymounter MA, Cullen MJ, et al. A role for melanin-concentrating hormone in the central regulation of feeding behaviour. Nature. 1996; 380:243–247. [PubMed: 8637571]
- 112. Shimada M, Tritos NA, Lowell BB, Flier JS, Maratos-Flier E. Mice lacking melaninconcentrating hormone are hypophagic and lean. Nature. 1998; 396:670–674. [PubMed: 9872314]
- 113. Ludwig DS, Tritos NA, Mastaistis JW, Kulkarni R, Kokkotoy E, Elmquist J, et al. Melaninconcentrating hormone overexpression in transgenic mice leads to obesity and insulin resistance. J Clin Invest. 2001; 107:379–386. [PubMed: 11160162]
- 114. Sakurai T, Amemiya A, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, et al. Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. Cell. 1998; 92:573–585. [PubMed: 9491897]
- 115. Mieda M, Willie JT, Hara J, Sinton CM, Sakurai T, Yanagisawa M. Orexin peptides prevent cataplexy and improve wakefulness in an orexin neuron-ablated model of narcolepsy in mice. Proc Natl Acad Sci USA. 2004; 101:4649–4654. [PubMed: 15070772]
- 116. Yamanaka A, Beuckmann CT, Willie JT, Hara J, Tsujino N, Mieda M, et al. Hypothalamic orexin neurons regulate arousal according to energy balance in mice. Neuron. 2003; 38:701–713. [PubMed: 12797956]
- 117. Boutrel B, Cannella N, de Lecea L. The role of hypocretin in driving arousal and goal-oriented behaviors. Brain Research. 2010; 1314:103–111. [PubMed: 19948148]
- 118. Mieda M, Yanagisawa M. Sleep, feeding, and neuropeptides: roles of orexins and orexin receptors. Curr Opin Neurobiol. 2002; 12:339–345. [PubMed: 12049942]
- 119. Baldo BA, Daniel RA, Berridge CW, Kelley AE. Overlaping distribution of orexin/hypocretinand dopamine-β-hydroxylase immunoreactive fibers in rat brain regions mediating arousal, motivation, and stress. Journal of Comparative Neurology. 2003; 464:220–237. [PubMed: 12898614]
- Georgescu D, Zachariou V, Barrot M, Mieda M, Willie JT, Eisch AJ, et al. Involvement of the lateral hypothalamic peptide orexin in morphine dependence and withdrawal. J Neurosci. 2003; 23:3106–3111. [PubMed: 12716916]
- 121. Zheng H, Corkern M, Stoyanova I, Patterson LM, Tian R, Berthoud HR. Peptides that regulate food intake: appetite-inducing accumbens manipulation activates hypothalamic orexin neurons and inhibits POMC neurons. Am J Physiol. 2003; 284:R1436–R1444.
- Harris GC, Wimmer M, Aston-Jones G. A role for lateral hypothalamic orexin neurons in reward seeking. Nature. 2005; 437:556–559. [PubMed: 16100511]
- 123. Zheng H, Patterson LM, Berthoud HR. Orexin signaling in the ventral tegmental area is required for high-fat appetite induced by opioid stimulation of the nucleus accumbens. J Neurosci. 2007; 27:11075–11082. [PubMed: 17928449]
- 124. Morshedi MM, Meredith GE. Repeated amphetamine administration induces Fos in prefrontal cortical neurons that project to the lateral hypothalamus but not the nucleus accumbens or basolateral amygdala. Psychopharmacology. 2008; 197:179–189. [PubMed: 18080115]
- 125. Harris GC, Aston-Jones G. Arousal and reward: a dichotomy in orexin function. Trends in Neurosciences. 2006; 29:571–577. [PubMed: 16904760]
- 126. Choi DL, Davis JF, Fitzerald ME, Benoit SC. The role of orexin-A in food motivation, rewardbased feeding behavior and food-induced neuronal activation in rats. Neuroscience. 2010; 167:11–20. [PubMed: 20149847]
- 127. DeFalco J, Tomishima M, Liu H, Zhao C, Cai XL, Marth JD, et al. Virus-assisted mapping of neural inputs to a feeding center in the hypothalamus. Science. 2001; 291:2608–2613. [PubMed: 11283374]

- 128. Dong HW, Petrovich GD, Swanson LW. Topography of projections from amygdala to bed nuclei of the stria terminalis. Brain Res Rev. 2001; 38:192–246. [PubMed: 11750933]
- 129. Davis M. The role of the amygdala in fear and anxiety. Annu. Rev. Neurosci. 1992; 15:353–375. [PubMed: 1575447]
- Fendt M, Fanselow MS. The neuroanatomical and neurochemical basis of conditioned fear. Biobehav. Rev. 1999; 23:743–760.
- LeDoux JE. Emotion circuits in the brain. Annu. Rev. Neurosci. 2000; 23:155–184. [PubMed: 10845062]
- Maren S. Neurobiology of Pavlovian fear conditioning. Annu. Rev. Neurosci. 2001; 24:897–931. [PubMed: 11520922]
- 133. Blanchard RJ, Blanchard DC. Crouching as an index of fear. Journal of Comparative and Physiological Psychology. 1969; 67:370–375. [PubMed: 5787388]
- 134. Fanselow MS. What is conditioned fear? Trends in Neuroscience. 1984; 7:400-402.
- 135. Petrovich GD, Ross CA, Holland PC, Gallagher M. Central but not basolateral amygdala is critical for control of feeding by aversive, conditioned cues. Soc Neurosci. 2006; 32 (Abst.).
- 136. Amorapanth P, Nader K, LeDoux JE. Lesions of periaqueductal gray dissociate-conditioned freezing from conditioned suppression behavior in rats. Learn. Mem. 1999; 6:491–499. [PubMed: 10541469]
- 137. Association, A. P.. Diagnostic and Statistical Manual of Mental Disorders. 4 ed.. Washington, DC: American Psychiatric Press; 1994.
- 138. Jimerson, DC.; Wolfe, BE. Psychobiology of eating disorders. In: Wonderlich, A.; Mitchell, JE.; de Zwaan, M.; Steiger, H., editors. Annual Review of Eating Disorders part 2–2006. Oxford: Radcliffe Publishing; 2006. p. 1-15.
- 139. Cannon, WB. Bodily changes in pain, hunger, fear and rage. 2nd ed.. New York: Appleton; 1915.
- 140. Schachter S, Goldman R, Gordon R. Effects of fear, food deprivation, and obesity on eating. Journal of Personality amd Social Psychology. 1968; 10:91–97.
- 141. Giordano GD, Renzetti P, Parodi RC, Foppiani L, Zandrino F, Giordano G, et al. Volume measurement with magnetic resonance imaging of hippocampus-amygdala formation in patients with anorexia nervosa. J Endocrinol Invest. 2001; 24:510–514. [PubMed: 11508785]
- 142. Takano A, Shiga T, Kitagawa N, Koyama T, Katoh C, Tsukamoto E, et al. Abnormal neuronal network in anorexia nervosa studied with I-123-IMP SPECT. Psychiatry Res. 2001; 107:45–50. [PubMed: 11472863]
- 143. Uher R, Murphy T, Brammer MJ, Dalgleish T, Phillips ML, Ng VW, et al. Medial prefrontal cortex activity associated with symptom provocation in eating disorders. Am J Psychiatry. 2004; 161:1238–1246. [PubMed: 15229057]
- 144. Seeger G, Braus DF, Ruf M, Goldberger U, Schmidt MH. Body image distortion reveals amygdala activation in patients with anorexia nervosa –a functional magnetic resonance imaging study. Neurosci Lett. 2002; 326:25–28. [PubMed: 12052530]
- 145. Friederich HC, Uher R, Brooks S, Giampietro V, Brammer M, Williams SC, et al. I'm not as slim as that girl: neural bases of body shape self-comparison to media images. Neuroimage. 2007; 37:674–681. [PubMed: 17604649]
- 146. Kapp, BS.; Pascoe, JP.; Bixler, MA. The amygdala: A neuroanatomical systems approach to its contribution to aversive conditioning. In: Squire, L.; Butters, N., editors. The Neuropsychology of Memory. New York: The Guilford Press; 1984. p. 473-488.
- 147. Holland P, Gallagher M. Amygdala circuitry in attentional and representational processes. Trends Cogn. Sci. 1999; 3:65–73. [PubMed: 10234229]
- 148. Gallagher, M. The amygdala and associative learning. In: Aggleton, JP., editor. The Amygdala: A functional analysis. New York: Oxford University Press; 2000. p. 311-329.
- 149. Hahn JD, Swanson LW. Distinct patterns of neuronal inputs and outputs of the juxtaparaventricular and suprafornical regions of the lateral hypothalamic area in the male rat. Brain Research Reviews. 2010; 64:14–103. [PubMed: 20170674]

- 150. Prewitt CM, Herman JP. Anatomical interactions between the central amygdaloid nucleus and the hypothalamic paraventricular nucleus of the rat: a dual tract-tracing analysis. J. Chem. Neuroanat. 1998; 15:173–185. [PubMed: 9797074]
- 151. Walker DL, Toufexis DJ, Davis M. Role of the bed nucleus of the stria terminalis versus the amygdala in fear, stress, and anxiety. Eur J Pharmacol. 2003; 463:199–216. [PubMed: 12600711]