

REVIEW ARTICLE

ROLE OF OXIDATIVE STRESS WHILE CONTROLLING IRON DEFICIENCY ANEMIA DURING PREGNANCY - INDIAN SCENARIO

Neeta Kumar, Nomita Chandhiok, Balwan S Dhillon and Pratik Kumar *

Division of Reproductive Health and Nutrition, Indian Council of Medical Research, *Department of Medical Physics, IRCH, All India Institute of Medical Sciences, Ansari Nagar, New Delhi-110029

ABSTRACT

Iron Deficiency anemia ranks 9th among 26 diseases with highest burden. Asia bears 71% of this global burden. Adverse maternal and birth outcome associated with hemoglobin status renders the issue worth attention. Indian scenario has worsened over the period despite continuous international and national efforts. This indicates some lacunae in the approach and strategies applied. Various reports state that even with maximum effort to increase outreach and monitoring for adherence to Iron schedule, consumer's compliance remains abysmally low. Recent studies has pointed out biological basis of side effects (gastrointestinal complains and systemic events) as raised oxidative stress for which iron is the key catalyst. Up till now the only target of research has been to raise hemoglobin of pregnant women above 11gm/dl. With the reports of pregnancy specific morbidities i.e. hemorrhage and septicemia with low hemoglobin, eclampsia, small for gestation age, gestational diabetes with higher ranges of hemoglobin, alarm is raised to define optimum range. Use of oxidative stress as biochemical marker with different doses and schedules has been defined because India lack information for its own population upon oxidative stress status when iron is supplemented as per current guidelines. Studies done in India and abroad have defined that too much and too less, both may raise oxidative stress and studies of this sort may provide biochemical scale for optimization. This review therefore has evaluated currently available Indian research and reports to understand the need of future research area. Important findings from other countries have been incorporated for comparison.

KEY WORDS

Oxidative stress, Iron supplementation, Anemia during pregnancy.

Iron balance is critical to maintain normal erythropoiesis (1 ,2). Optimum balance is highly needed for growing children and pregnant women. Daily requirement for iron is 6 times greater for women in the last trimester of pregnancy than for non pregnant women. Currently 100mg elemental iron for 100 days is given during pregnancy in India (3). However the mode / strategies of supplementation have come under scrutiny. Investigations world wide have recently begun to measure

Address for Correspondence :

Dr. Neeta Kumar,
Scientist B, Div. of RHN,
Indian Council of Medical Research
Ansari Nagar, New Delhi-110029
E-mail: neeta@icmr.org.in, delhineeta@yahoo.co.in
Phone: 09313195247

the effect of ferrous iron (4) supplementation on indices of Oxidative Stress (OS) in pregnant women. A fine line between deficiency and overdoses in case of iron (or any micronutrient supplementation for that matter) is evident in literature and an utmost care is required while deciding doses and schedules (5 , 20). However during pregnancy these effects are not well studied. Efforts have begun to combat this situation without compromising beneficial effects of iron. Considering the age old problem of anemia in India and massive efforts going in vain this review applies extensive literature search by using key words Iron, pregnancy, oxidative stress, and anemia. Cross references and personal communications were collected through local and National Library to put forward a comprehensive Indian scenario in global perspective upon what happen to oxidative reactions when iron is deficient and consumed.

Oxidative stress and its significance for maternal health:

Oxidative stress is defined as disturbance in the prooxidant-antioxidant balance in favour of the prooxidant, leading to potential damage producing oxidative stress. Free radical is any atom (e.g. oxygen, nitrogen) with at least one unpaired electron in the outermost shell, and is capable of independent existence. There are numerous types of free radicals that can be formed within the body, but here oxygen centered free radicals most commonly the superoxide anion (O_2^-), the hydroxyl radical (OH^\cdot), singlet oxygen, and hydrogen peroxide (H_2O_2) are studied (6).

The interaction of copper or iron and H_2O_2 also produce OH^\cdot as first observed by Fenton. Iron has the ability to gain and lose electrons i.e. ($Fe^{2+} \leftrightarrow Fe^{3+}$) very easily. This property makes iron and copper two common catalysts of oxidation reactions. The release of iron can be detrimental to cellular membranes because of the pro-oxidation effects it may have (7, 8). Raised OS have been implicated not only for chronic diseases like cardiovascular disease, cancer, diabetes, cirrhosis, atherosclerosis, Alzheimer's disease, and Parkinson's disease, it has been found altered with pregnancy, iron deficiency and overdoses (9, 26). Importance of attaining optimum range is desirable as low hemoglobin levels as well as high hemoglobin levels have been associated with adverse events (107). An elevated Hb level may be an indicator for possible pregnancy complications associated with poor plasma volume expansion, and should not be mistaken for good iron status (10). In addition to this, speculations have been made that an uncompensated OS experienced during pregnancy could predict or affect some pathological conditions of mother. Its effect on the developing fetus cannot be excluded either (11). Oxidative stress as biochemical indicator has been found proportionate to side effects in animal studies (64) and hence may be used to optimize iron status.

Burden due to low hemoglobin: An Indian review finds 19-22% of total maternal deaths due to anemia. Deaths due to septicemia and hemorrhage may indirectly be attributable to anemia (12). Indian Council of Medical Research (ICMR) reports prevalence of anemia among women 75% (31% moderate, 3% severe) (13). ICMR study on 1,66,996 live births across all over India found 11.5% direct maternal deaths due to anemia (14). Approximately 50% of the anemia is attributable to iron deficiency.

Average estimates for all anemia attributed mortality (both direct & indirect) are 7.26 for Asia which is high when compared

with other developing region like Africa (6.37%) and Latin America (3.0%). Globally 841,000 deaths and 35,057,000 disability-adjusted life years lost go in to account of anemia. Asia bears 71% of this global mortality burden and 65% of the disability-adjusted life years lost as compared with North America which bears only 1.4% (15). Recently completed National Health and Family welfare Survey (NFHS-III 2006) reported 57.9% anemia (Hb <11gm/dl) among pregnant women. It is higher than 49.7%, reported in 1999 NFHS-II (16). Several factors identified for increasing burden i.e. low bioavailability of iron in Indian diet, high phytates, low content (17), high worm load & malaria prevalence(18), poor outreach and quality of iron supplementation, non-compliance and logistics difficulties (19, 20).

Adverse impact of low hemoglobin (<11g/dl) on maternal status have been widely proved (21, 22). Gomber et al reported rise in baby's birth weight from 2509 to 2803g when Hb rises from 7 to 11 g/dl (23). Sharma et al reported pregnant women (n=447) with 4-6 fold higher risk of prolonged labor at Hb<8.9 g%, 4.8-fold higher risk of delivery complications with Hb≤7.5 g% and maximum birth weight at Hb 9.6-10.5 g% (24). RR 1.35 (0.92-2.00) at Hb 40-80 g/L and 3.51 (2.05-6.00) at Hb<47 g/L is reported for maternal mortality (25).

An USA based retrospective analysis upon 173,031 pregnant women showed odd ratio (OR) of 1.68 for preterm birth at Hb <7g/dl (Z score <-3.0) and 0.91 at Z score - 1.1-2.0 (26). Hematocrit (Hct) from 41% to 44% had lowest risk of premature birth and double at Hct<37% (27). Decreasing the Hct by single point had 24% rise in prematurity (OR 1.24), 5-point decrease tripled risk of prematurity (OR 2.98) (9). Scholl reported preterm delivery five fold high with iron deficiency anemia. Low birth weight and low pregnancy weight gain was three fold high in his famous Camden trial (28). Similar equations are yet to be derived for Indian population for all the ranges of Hb.

Burden due to high hemoglobin: Indian prevalence of high Hb is not known. Indirect evidence (ICMR study) finds 24% prevalence of pregnancy induced hypertension in a multicenter data. Approximately 10-24% maternal deaths attributed to high Hb, that comes equivalent to maternal deaths due to anemia (11.5%) and hemorrhage (23.6%) (14). Sirinivasan (Chandigarh, India) encountered 50 cases of severe cerebrovascular thrombosis due to hemoconcentration amongst 1000 deliveries performed (29). Iron overload, plasma volume nonexpanding due to protein nutritional deficiency are some factors identified behind high Hb at term (30). Pregnancy specific morbidities like PIH, eclampsia, septic

abortion and gestational diabetes are found to be associated with high Hb at term (31, 32, 33). Prevalence of such events in developing countries are 10 times more than in developed countries. Plasma volume study (n= 92 Indian, 191 Fijian women) showed lower red cell volumes, blood volumes & high incidence of toxemia, lower plasma volume for a given birth weight/ maternal size ratio in Indian women (34).

Study from USA found 0.43% women with very high Hb (>14gm/l) (0.25% with very low Hb). Koller found 62% women having small for date baby at Hb >2SD (35). Lao, China reports high iron levels in the group with Gestational Diabetes Mellitus. Hb > 13 gm/dl has been taken as hemoconcentration in Cochrane meta-analysis of eight trials. Women on daily iron were three times more likely to have hemoconcentration (RR 3.01; 95% CI 1.46-6.19) (36). Steer found Hb above 105 g/l associated to increased risk of low birth weight and preterm delivery (PTD) (37). This phenomenon is seen in all ethnic groups. Zhou et al reports RR 2.52 for PTD at Hb \geq 130 g/l. Scanlon et al find OR: 1.79 with high Hb in small for gestational age (SGA) (10). Other studies too report similar findings (38, 39, 40).

Non compliance and side effects with Iron consumption:

Various surveys from India found that even if logistics are taken care of, the compliance remains abysmally low. Its average is 22% and as low as 2% in some EAG states (Empowered Action Group states). This is despite increment of attendance

at antenatal clinics (ANC) from 44% (NFHS-II, 1999) to 51% in NFHS III 2006 (41, 42). Recently launched NRHM (National Rural Health Mission-2005) has concerns that in spite of improved attendance at ANC clinics situation of iron consumption has not improved. Daily oral iron supplement has shown improvement of hematological indices but with frequent gastrointestinal (GI) symptoms. ICMR study evaluating dose effect of 60-120 mg iron supplementation in 115 participants faced 32 lost to follow-up (43). Despite constant monitoring to ensure compliance and manage the side effects 47.2% of the subjects consumed less than 90 tablets of 60 mg daily iron dose, 14% reported side effects (44). Prevalence of side effects doubled with 200 mg daily dose (44). Another study from New Delhi reported 40% compliance with daily 100 mg iron, while 85% in weekly. Majority (75%) of the non-compliance in oral iron group was due to GI side effects (45).

From outside India, RR1.13 is reported for nausea, 1.21 for stomach pain and 1.12 for hard stool at 20 mg iron daily (4). Cochrane analysis (6 trials) on women receiving daily oral iron reported more side-effects of any kind than women taking placebo or not taking any iron supplementation at all (26% versus 11.9%): (RR 1.90; 95% CI 1.09 to 1.33) (107). Vomiting in 15.7% iron users, 8.94% in nonusers, RR 1.69 (CI 95% 1.15 to 2.47) has been reported. Upper tolerable dose of iron is 45mg according to CDC Atlanta.

Table 1 : Side effect profile at 60- 120 mg daily iron doses (currently in practice in Indian subcontinent)

Year	Journal/Country	Noncompliance / attrition (%)	Noncompliance due to side effects (%)	Causes for side effect and noncompliance -%
Studies from Indian subcontinent				
1975	Q J Med (58)	30	6	Mostly due to non cooperation and refusal to draw blood
1989	GOI-Report (59)	No mention	No mention	GI side effects-1%
1991	ACC/SCN (57)	58	No mention	Lack of supply
1992	ICMR technical report (44)	47.2 (at 120 mg iron)	noncompliance due to communication skills in pursuing study subjects	GI side effects-14.2% nausea-8.8% gastritis-6.5%, Diarrhea-0.5%, Constipation- 0.8%, abdominal distention- 0.3%
1999	Natl Med J India (43)	38%		Majority GI side effects
2000	Journal of Nutrition. (Pakistan) (60)	45%	Only one subject with minor side effect migration, change of center	No GI symptoms, withdrawal due to other personal reasons,
2002	J Health Popul Nutr.(Dhaka) (61)	18%	61.5	GI symptoms:(%): Heartburn 11.5, Nausea 14.4, Vomiting9.6, Diarrhoea 10.6, Constipation 59.6
2002	Indian Pediatr (23)	27%	No side effect reported	Not reported
2004	Am J Clin Nutr (21)	48%	21% had side effects	Dyspepsia10%,Constipation-5%, Diarrhea-3%, Vomiting-2%, Rash and itching-1%
2004	J Obstet Gynaecol Res (45)	60%	30% women with symptoms	75% of the non compliance had GI symptoms: nausea, vomiting, constipation
2007	Mescape general medicine (62)	Full compliance	78% subjects had side effects	GI intolerance 65.4%, Constipation 48%, Metallic taste 15.3%, Diarrhea 3.8%, Rashes 1.9%

Common drawback with iron studies is that criteria for lost to follow up are found to be the major bias in final analysis of many hospital based randomized case control trials. For example a widely cited study evaluated effect of iron on birth outcome started with 513 pregnant women and only 144 women's status in IIIrd trimester could be obtained (46). Most of the studies did not analyze morbidities during pregnancy in different groups. Analysis to see the effect of supplementation in different trimester is also required. Majority Indian women are approaching for health care and supervised delivery either at the time of delivery or in third trimester. They are given extensive iron therapies (if detected anemic) in third trimester or just before delivery. Benefit to maternal health and birth outcome obtained by such last moment supplementation is not documented in India (38, 47). Many reports count lost to follow up as noncompliance (48). 18-60% noncompliance is reported in an Indian review. It is varying from 9-87% from other countries. (Table 1). Significant withdrawal due to side effects that was associated with hemo-concentration has been reported from Mexico and other countries outside India (36, 49-57).

OXIDATIVE STRESS ESTIMATION DURING IRON SUPPLEMENTATION

Animal studies:

Effects at the site of absorption: Studies from National Institute of Nutrition-ICMR reports oxidative status after iron consumption. Generation of apoptosis in GI tract in addition to reduction of microvillus height, mucosal erosion in rats was seen with different schedules of iron orally as observed by transmission electron microscope. EPR spectroscopy identified production of hydroxyl and methoxyl radicals in both the luminal and mucosal contents in the GI tract of iron supplemented rats (63). Surprisingly these changes became more pronounced when iron was given to anemic rats (64, 65).

Systemic effect: Elevated free radical levels from 25 to 40%, reduction of scavenging enzymes-catalase, SOD, glutathione peroxidase and alpha tocopherol in rat liver and kidney homogenates at 20 hours reported after iron administration at dose equal the dose of human consumption (100mg daily) (14). Continuous absorption of a fraction of supplemental iron at a rate beyond normal, perhaps by passive diffusion mass effect resulted in excessive liver iron levels, particularly in the previously deficient animals (66). Genotoxic and reproductive changes have been extensively highlighted in animal studies (Table 2). Organs have been found responding differently to different dose of iron (84).

Human studies: *In India* there is no randomized controlled study of oxidative status with iron during pregnancy, except currently ongoing trial at ICMR started as task force study at two Obstetrics and Gynae centers: AIIMS, New Delhi and PGIMER, Chandigarh. Preliminary findings (n=20) show TBARS level is decreased in nonanemic pregnant women in comparison to anemic group. Oxidized glutathione peroxidase is found raised during pregnancy ($p < 0.05$) when compared with non pregnant age matched controls. Effect estimation of daily vs. weekly iron supplementation on oxidative stress is ongoing under this study.

An American pilot study (n=19) on healthy pregnant women with borderline anemia with oral ferrous iron in prophylactic doses (36 mg daily) found 2 fold elevation oxidized glutathione in Fe + group ($p=0.02$) (1). Lachili et al (2001) investigated the effect of a daily iron supplementation (100 mg/d as fumarate) and vitamin C (500 mg/d as ascorbate) for the third trimester of pregnancy on lipid peroxidation (plasma TBARS) and found them enhanced in the iron supplemented group ($p < 0.05$) (23). Lund et al found fecal iron raised from 60 mmol/l at base line to 300 mmol/l after iron supplementation, which returned to base line within 2 wks after stopping iron (67). This resulted in 40% increased production of free radicals at colon, which could cause mucosal cell damage or increased production of carcinogens.

Iron found enhancing free radical production by RNAs and hyperglycemia through glycation. Elevated non-transferrin-bound iron (NTBI) was detected shortly after the ingestion of iron supplements in plasma and umbilical cord blood (68). Transferrin binds the iron leaving the cell and entering the circulation, thus avoiding the entrance of free iron. However, this mechanism appears to be overwhelmed by amounts of passively diffused iron when large iron boluses are presented to the intestine, and NTBI may reach the liver, causing systemically raised OS. Not only dose and schedule, effect of timing of supplementation has been highlighted in some reports (69). Important work done in India to seek answers of the questions rose by John in the year 2000 (108) is listed in Table 2. All human studies from India and seeing its limited number, some foreign studies reporting salient changes of OS status are included. Indian human data show increased OS during anemia and reversal with iron supplementation while findings from other countries are contrary to that (2, 4, 8, 10, 18, 80, 81, 82). Indian animal studies are also evaluated as their findings are contrary to what is found with human studies in India. However animal studies from foreign resonate the similar findings (70-71). Majority of them found raised OS with anemia and further rising with iron doses.

CONCLUSION

Oxidative stress worsens with anemia in most of the animal and human studies (43, 26-28, 33, 63, 64, 66). Status improved with iron in some animal and human studies; however majority animal as well as studies involving pregnant women found it

increased (Table 2). Investigation to check whether OS increase is such that it can alter birth outcome are required. In Indian conditions where the ANC strategies are not much supervised, there is scarcity of information upon side effects and noncompliance too. In view of major studies and meta-analysis, it is concluded that there is a need of large, good

Table 2 : All Indian and prominent foreign studies done on human participants and animals to estimate oxidative stress with different iron doses at the site of absorption, systemic and at reproductive organs

Ref.	Iron dose	Sample/site	OS parameters				
			SOD	MDA/ TABRS	GSHPx	GSH Rd	Catalase
Human Studies : India							
Acharya et al (72)	Iron not supplemented	Human RBC cell lines, anemic subjects	↑		↓		↓
Ramachandran et al (73)	-Do-	Human RBCs in iron deficiency					
Sundaram et al (74)	60mg iron daily for one month	n = 20 anemic adults, 16 control, serum tested		↑ pre Fe, ↓ ed after Fe		↑	↑ ed fructosamine pre Fe, Post therapy ↓ ed
Human Studies : from outside India							
Lachilim et al (8)	100mg/day as fumerate + 500mg vit. C in third trimester pregnancy	n = 27 Fe + Fe, 27 No supplementation Maternal blood used for OS parameters		↑ ed 3.62±0.36 in Fe + arm 3.01±0.37 in control			α-tocopherol ↓ in Fe+ group
Rehema et al (2)	36 mg ferrous iron supplemented Fe+ Group	n=19 pregnant women with borderline anemia, 13 Fe+, 6 Fe-ve		2 fold ↓ ed in Fe+ group			Other parameters no significant change
Scholl et al (28)	N=360, 60mg daily iron Camedon's ongoing Research	n=350 Urinary excretion of 8-hydroxyguanosine (8-OH-dG)					8-OH-dG ↑ ed (2.7 fold) post iron, AOR-3.3 at Serum transferring saturation>21.7%
Isler (75)	n=8 anemic received oral iron, 18 healthy control	Whole blood sample	↑		NS		
Ferreira et al (76)	effect of iron at concentrations (0, 1, 5, 10, 50 and 100 microM Fe3+)	Human normal RBC's antioxidant estimated, RBC incubated with colloidal iron hydroxide		Minimum at 5 & max. at 100 mcm	↓ with time at 10-100 mcm		NS
Kurtoglu et al (9)	60 mg iron given	Adult RBCs studied for OS in 63 anemic before (b) and after (a)	↓ b ↑ a	↑ b ↓ a	↓ b ↑ a		↓ b ↑ a
Sevgi et al (77)	Supplemented iron	n=30 anemic & 25 normal children			↓ b ↑ a		
Olivares et al (78)	100 mg/d Fe, as ferric polymaltose	n=12 anemic, 12 control women RBC studied					↓ CuZn-SOD activity
Binkoski et al (79)	50 mg iron gr. A, 100 mg iron gr. B.	26 non anemic women		NS			
Kumerova et al (80)	No iron	n=56 anemic, 50 healthy control adults			↓	↓	↓ G6PD in anemic

Ref.	Iron dose	Sample/site	OS parameters					Other
			SOD	MDA/ TABRS	GSHPx	GSH Rd	Catalase	
Rehman et al (81)	14 mg/day Gr A, 60mg/day-Gr B with 260mg vit C iron	n=38 healthy adults, DNA extracted from white blood cells, tested before (b) & after (a) 6 weeks of Fe consumption	↓ b ↑ a	↑ b ↓ a	↓ b ↑ a		↓ b ↑ a ↓ a	↑ b Oxidative DNA damage
Yang et al (82)	14mg (Gr A), 60mg (Gr B) iron with Vit. C RDA	n=20 adults each their own control, platelet function & low density lipoprotein oxidation tested			NS with added Vit. C			↑ mean lag phase oxidation of low density lipoprotein (LDL) after iron
Troost et al (83)	Single 100mg dose	N=6, GI Lumen wash collected from double lumen perfusion tube		↑ (0.07 – to 3.35) on iron perfusion			↑ ed NPAC	Expression of 89 genes & six biological processes altered with iron
Devrim et al (84)	N=15 on 100 mg iron, 13-no iron	N=27 pregnant women maternal plasma & placenta tested		↑				
Yip et al (85)	No iron given	RBCs of iron deficient adults		↑				
Petukhov et al (86)	-do-	RBC of polycythemia subjects		↑				↓ ed selenium dependent GSHPx
Vives et al (87)	-do-	Microrcytic RBC of patients	↓	↑	↓			
Avissar et al (88)	-do-	RBCs of iron deficient adults	↓	↑	↓			
Riazantsev et al (89)	-do-	RBCs of anemic pregnant women			↑		↓	
Panchenko et al (90)	-do-	RBCs of anemic children	↓					
Cellerino et al (91)	-do-	n=9 normal, 15 anemic adults	↓	↑	↓			
Mehmet et al (92)	-do-	22 anemic and 22 healthy females	↓	↑	↓		↓	↑ lymphocyte DNA damage
Golovin and Konvai (93)	-do-	N=55 anemic adults	NS	NS	NS		NS	NS
Krause et al (94)	-do-	n=29, Blood of anemic adults			↑			
Animal Studies : India								
Jain et al (95)	rats fed Fe-deficient (2 ppm Fe) Fe diets	RBC membrane lipids and proteins tested		↑ ed in anemic			↑ ed NPAC in anemic	↓ ed RBC T½ in iron deficient group
Rao and Jagdeesan (96)	Fischer rats iron sufficient (C) or Fe deficient (d) diets	Hepatic enzymes	↓ by 28% Gr. D.	↑ by 50% Gr. D	by 50% Gr. D			
Srigiridhar and Nair (97)	effects of excess free iron, was tested in rats fed with iron	site of iron absorption-duodenum	↑ a		↓ a		↑ a	Ratios of SOD/Gpx, Cat/Gpx ↑ ed in the Fe fed group
Srigiridhar and Nair (64)	WKY female rate 8 mg iron for 15 days	Effect after supplement	NS		NS			protein carbonyl formation
Srigiridhar and Nair (63)	Fe fed rats Given Vit E and C	Site-rat mucosa Before vit E & C (b), after (a)	↓ b ↑ a	↑ b ↑ a	↓ b ↑ a		↓ b ↑ a	Vit E & C acted antioxidant
Kaur and Mehmood (98)	Wistar rats fed iron 29 mg/kg body weight (or 6.58 mg/kg Fe) daily	Site-GI brush border						↓ Alp, sucrose maltase, lactase rehalase, NS- leucine amino peptidase & γGT

quality trials assessing clinical outcomes of various iron supplementation strategies with respect to beneficial / adverse effects and OS status (99). Major thrust of public health policy is to increase demand and compliance to control anemia, but without addressing compliance, optimizing oxidative stress and minimizing side effects it may again be a series of failed strategies. Other factors responsible for altered OS i.e. pregnancy itself, age, parity, chronic diseases, diet, environment, season, emotional quotient/ stress, protein intake, intestinal parasites and malaria etc are unique in India. Therefore study should be a multi-center, on large sample to provide concrete answers pertaining to situations in India.

Questions asked by John, years ago still remain unanswered. He put queries (100) i.e. what approach will provide the most efficacious outcome? If iron supplementation is provided, what dosage and schedule are the most beneficial? Is there a real health risk of too much iron? Can iron supplementation prevent as well as induce oxidative stress? What changes in oxidative stress patterns occur, if normal as well as anemic women are supplemented? Upcoming information from Camedon study and ICMR task force study would be providing some of these answers.

Abbreviation: Malondialdehyde: MDA, Oxidized low density lipoprotein: OX-LDL, Superoxide dismutase: SOD, Glutathione peroxidase: GSH-Px, Glutathione reductase: GSHRd, OS: Oxidative stress, Thiobarbituric acid reacting substances (TBARs), ↑ raised, ↓ decreased, NS- No significant change, Iron = elemental iron doses, Non-protein antioxidant capacity-NPAC, Alp-alkaline phosphatase, b= before, a=after

REFERENCES

- Sloan NL, Jordan E, Winikoff B. Effects of Iron Supplementation on Maternal Hematologic Status in Pregnancy. *Am J Public Health* 2002; 92 (2): 288-93.
- Rehema A, Zilmer K, Klaar U, Karro H, Kullisaar T, Zilmer M. Ferrous iron administration during pregnancy and adaptational oxidative stress (Pilot study). *Medicina (kaunas)* 2004; 40(6): 547-52.
- Kumar A. National nutritional anaemia control programme in India. *Ind J Public Health* 1999; 43(1): 3-5.
- Makrides M, Crowther CA, Gibson RA, Gibson RS, Skeaff CM. Efficacy and tolerability of low dose iron supplements during pregnancy: a randomized controlled trial. *Am J Clin Nutr* 2003; 78(1): 145-53.
- Galleano M, Puntarulo S. Effect of mild iron overload on liver and kidney lipid peroxidation. *Braz J Med Biol Res* 1994; 27(10): 2349-58.
- Zwart LL, Meerman JN, Commandeur JNM, Vermeulen NPE. Biomarkers of free radical damage. Applications in experimental animals and humans. *Free Radic Biol Med* 1999; 26: 202-26.
- Walter PB, Knutson MD, Martinez AP, Lee S, Xu Y, Viteri FE, Ames BN. Iron deficiency and iron excess damage mitochondria and mitochondrial DNA in rats. *Proc Natl Acad Sci* 2002; 99: 2264-69.
- Lachili B, Hininger I, Faure H, Arnaud J, Richard MJ, Favier A, et al. Increased lipid peroxidation in pregnant women after iron and vitamin C supplementation. *Biol Trace Elem Res* 2001; 83(2): 103-10.
- Kurtoglu E, Ugur A, Baltaci AK, Undar L. Effect of iron supplementation on oxidative stress and antioxidant status in iron deficiency anemia. *Biol Trace Elem Res* 2003; 96(1-3): 117-23.
- Scanlon KS, Yip R, Schieve LA, Cogswell ME. High and low hemoglobin levels during pregnancy: differential risks for preterm birth and small for gestational age. *Obstet Gynecol* 2000; 96(5 Pt 1): 741-8.
- Wisdom SJ, Wilson R, McKillop JH, Walker JJ. Antioxidant systems in normal pregnancy and in pregnancy-induced hypertension *Am J Obstet Gynecol* 1991; 165: 170-74.
- Anand A. Anemia- a major cause of maternal death. *Ind Med Trib* 1995; 3(1): 5-8.
- NNMB Technical report No. 24. National nutrition Monitoring Bureau-Diet and Nutrition status of population and prevalence of hypertension among adults in rural areas. NIN, ICMR, 2006.
- Bedi N, Kambo I, Dhillon BS, Saxena BN, Singh P. Maternal deaths in India - Preventable tragedies (An ICMR Task force Study). *J Obst Gyn of Ind* 2001; 51(2): 86-92.
- Stoltzfus RJ. Iron deficiency: global prevalence and consequences. *Food Nutr Bull* 2003; 24(4 Suppl): S99-103.
- International Institute of Population Sciences and ORC Macro. National Family Health Survey - 3. International Institute of Population Sciences, Mumbai. (<http://www.iipsindia.org/nfhs3.html>) (Accessed on October 2007)
- NNMB Technical Report No. 22. National Nutrition Monitoring Bureau. Prevalence of Micronutrient Deficiencies. National Institute of Nutrition. Indian Council of Medical Research. Hyderabad - 500 007, 2003.
- Awasthi S, Pande VK. Prevalence of malnutrition and intestinal parasites in preschool slum children in lucknow. *Ind Pead* 1997; 34.
- Community-Level Interventions to Prevent and Treat Anemia: A Review of Evidence from India March 2008, Evidence review series-3, USAIDS- Vistaar Project link: www.intrahealth.org
- Vijayaraghavan K, Brahmam GNV, Nair KM, Akbar D, Rao NP. Evaluation of national nutritional anemia prophylaxis programme. *Ind J Pead* 1990; 7(2): 183-90.

21. Sharma JB, Jain S, Mallika V, Singh T, Kumar A, Arora R, et al. A prospective, partially randomized study of pregnancy outcomes and hematologic responses to oral and intramuscular iron treatment in moderately anemic pregnant women. *Am J Clin Nutr* 2004; 79(1): 116-22.
22. Nair KM, Bhaskaram N, Balakrishna P, Sesikeran RB. Response of hemoglobin, serum ferritin, and serum transferrin receptor during iron supplementation in pregnancy: a prospective study. *Nutrition* 2004; 20(10): 896-9.
23. Gomber S, Agarwal KN, Mahajan C, Agarwal N. Impact of daily versus weekly hematinic supplementation on anemia in pregnant women. *Ind Pediatr* 2002; 39(4): 339-46.
24. Malhotra M, Sharma JB, Batra S, Sharma S, Murthy NS, Arora R. Maternal and perinatal outcome in varying degrees of anemia. *Int J Gynaecol Obs* 2002; 79(2): 93-100.
25. Brabin BJ, Hakimi M, Pelletier D. An analysis of anemia and pregnancy-related maternal mortality. *J Nutr* 2001; 131(2S-2): 604S-614S.
26. Scanlon KS, Yip R, Schieve LA, Cogswell ME. High and Low Hemoglobin Levels During Pregnancy: Differential Risks for Preterm Birth and Small for Gestational Age. *Obs & Gynecol* 2000; 96: 741-8.
27. Lieberman E, Ryan KJ, Monson RR, Schoenbaum SC. Association of maternal hematocrit with premature labor. *Am J Obs Gynecol* 1988; 159(1): 107-14.
28. Scholl TO, Hediger ML, Fischer RL, Shearer JW. Anemia versus iron deficiency: increased risk of preterm delivery. *Am J Clin Nutr* 1992; 55: 985-8.
29. Chopra JS, Prabhakar S. Clinical features and risk factors in stroke in young. *Acta Neurologica Scandinavia* 1979; 43: 289-300.
30. Newman V, Judith TF. Role of nutrition in the prevention of preeclampsia. Review of the literature. *J Nurse Midwifery* 1990; 35(5): 282-91.
31. Raman L, Pawashe AB, Yasodhara P. Hyperferritinemia in pregnancy induced hypertension and eclampsia. *J Postgrad Med* 1992; 38(2): 65-7.
32. Afkhami-Ardekani M, Rashidi M. Iron status in women with and without gestational diabetes mellitus. *J Diabetes Complications* 2008; 4 (Epub ahead of print).
33. Lao TT, Chan LY, Tam KF, Ho LF. Maternal hemoglobin and risk of gestational diabetes mellitus in Chinese women. *Obs Gynaecol* 2002 May; 99(5 Pt 1): 807-12.
34. Coleman RJ. Comparison of plasma volume levels in normal pregnancy between two ethnic groups in Fiji. *The Australian and New Zealand J Obs Gynaecol* 1978; 18(2): 127-32.
35. Koller O, Sandvei R, Sagen N. High hemoglobin levels during pregnancy and fetal risk. *Int J Gynaecol Obs* 1980; 18(1): 53-6.
36. Pena-Rosas JP, Viteri FE. Effects of routine oral iron supplementation with or without folic acid for women during pregnancy. *Cochrane Database of Systematic Reviews* 2006; Issue 3. Art. No.: CD004736. DOI: 10.1002/14651858.CD004736.
37. Steer P, Alam MA, Wadsworth J, Welch A. Relation between maternal haemoglobin concentration and birth weight in different ethnic groups. *BMJ* 1995; 310(6978): 489-91.
38. Zhou LM, Yang WW, Hua JZ, Deng CQ, Tao X, Stoltzfus RJ. Relation of hemoglobin measured at different times in pregnancy to preterm birth and low birth weight in Shanghai. *Am J Epidemiol* 1998; 148(10): 998-1006.
39. Stephansson O, Dickman P, Johansson A, Cnattingius S. Maternal hemoglobin concentration during pregnancy and risk of stillbirth. *JAMA* 2000; 284(20): 2611-7.
40. Scholl TO. High third-trimester ferritin concentration: associations with very preterm delivery, infection, and maternal nutritional status. *Obs Gynaecol* 1998; 92(2): 162-66.
41. National Nutrition Monitoring bureau, Technical report No. 22, Prevalence of micronutrient deficiencies, National Institute of Nutrition- ICMR, 2003.
42. Toteja GS, Singh P, Dhillon BS, Saxena BN. Micronutrient deficiency disorders in 16 Distt. of India – ICMR Task Force Study in 13 States. 1997-99.
43. Shatrugna V, Raman L, Kailash U, Balakrishna N, Rao KV. Effect of dose and formulation on iron tolerance in pregnancy. *Natl Med J India* 1999; 12(1): 18-20.
44. Fields supplement trial in pregnant women with 60 mg, 120 mg and 180 mg of iron with 500mcg of folic acid-An ICMR task force study. Published 1992.
45. Mukhopadhyay A, Bhatla N, Kriplani A, Pandey RM, Saxena R. Daily versus intermittent iron supplementation in pregnant women: Hematological and pregnancy outcome. *J Obs Gynaecol Res* 2004; 30(6): 409-13.
46. Cogswell ME, Parvanta I, Ickes L, Yip R, Brittenham GM. Iron supplementation during pregnancy anemia, and birth weight: a randomized controlled trial. *Am J Clin Nutr* 2003; 78(4): 773-81.
47. Gambling L, Andersen HS, Czopek A, Wojciak R, Krejpcio Z, McArdle HJ. Effect of timing of iron supplementation on maternal and neonatal growth and iron status of iron-deficient pregnant rats. *J Physiol* 2004; 561(1): 195-203.
48. Galloway R, Judith MG. Determinants of compliance with iron supplementation: supplies, side effects, or psychology? *Soc Sci Med* 1994; 39(3): 381-90.
49. Chisholm M. A controlled clinical trial of prophylactic folic acid and iron in pregnancy. *Br J Obs Gynae* 1966; 73: 191 -6.

50. Svanberg B, Arvidsson B, Norrby A, Rybo G, Sölvell L. Svanberg B, Arvidsson B, et al. Absorption of supplemental iron during pregnancy—a longitudinal study with repeated bone marrow studies and absorption measurements. *Acta Obstet Gynecol Scand* 1975; 48: 87-108.
51. Griffiths M. Concept testing nutrition communication and behaviour change components. Manoff International, Indonesia Nutrition Development program 1980; Volume 1: p 38.
52. Blot I, Papiernik E, Kaltwasser JP, Werner E, Tchernia G. Influence of routine administration of folic acid and iron during pregnancy. *Gynecol Obs Invest* 1981; 12(6): 294-304.
53. Romslo I, Haram K, Sagen N, Augensen K. Iron requirement in normal pregnancy as assessed by serum ferritin, serum transferrin saturation and erythrocyte protoporphyrin determinations. *Br J Obs Gynaecol* 1983; 90: 101-07.
54. Charoenlarp P, Dhanamitta S, Kaewvichit R, Silprasert A, Suwanaradd C, Na-Nakorn S, et al. A WHO collaborative study on iron supplementation in Burma and Thailand. *Am J Clin Nutr* 1988; 47: 280-97.
55. de Souza AI, Batista FM, Cardoso FL, Natal FJ. The effectiveness of three regimens using ferrous sulfate to treat anemia in pregnant women. *Rev Panam Salud Publica* 2004; 15(5): 313-19.
56. Ekström EC, Kavishe FP, Habicht JP, Frongillo EA, Rasmussen KM, Hemed L, et al. Adherence to iron supplementation during pregnancy in Tanzania: determinants and hematologic consequences. *Am J Clin Nutr* 1996; 64: 368 - 74.
57. ACC/SCN. Controlling iron deficiency. A report based on an ACC/SCN workshop, State-of-the-art series. Nutrition policy discussion paper no. 9 1991; p. 4.
58. Sood SK, Ramachandran K, Rani K, Ramalingaswami V, Mathan VI, Ponniah J, et al. W.H.O. sponsored collaborative studies on nutritional anaemia in India. 1. The effects of supplemental oral iron administration to pregnant women. *Q J Med* 1975; 44(174): 241-58.
59. GOI- Government of India. Ministry of health and Family welfare, UNICEF/New Delhi, report of the meeting on the prevention and control of nutritional anemia 1989; pg34.
60. Mumtaz Z, Shahab S, Butt N, Rab MA, DeMuyneck A. Daily iron supplementation is more effective than twice weekly iron supplementation in pregnant women in Pakistan in a randomized double-blind clinical trial. *J Nutr* 2000; 130: 2697-702.
61. Hyder SMZ, Persson LA, Chowdhury AMR, Ekström EC. Do side-effects reduce compliance to iron supplementation? A study of daily- and weekly-dose regimens in pregnancy. *J Health Popul Nutr* 2002; 20(2): 175-9.
62. Saha L, Pandhi P, Gopalan S, Malhotra S, Saha PK. Comparison of efficacy, tolerability, and cost of iron polymaltose complex with ferrous sulphate in the treatment of iron deficiency anemia in pregnant women. *Med Gen Med* 2007; 9(1): 1.
63. Srigiridhar K, Nair KM. Supplementation with alpha-tocopherol or a combination of alpha-tocopherol and ascorbic acid protects the gastrointestinal tract of iron-deficient rats against iron-induced oxidative damage during iron repletion. *Br J Nutr* 2000; 84: 165-73.
64. Srigiridhar K, Nair KM. Iron-deficient intestine is more susceptible to peroxidative damage during iron supplementation in rats. *Free Radic Biol Med* 1998; 25: 660-65.
65. Srigiridhar K, Nair KM, Subramanian R, Singotamu L. Oral repletion of iron induces free radical mediated alterations in the gastrointestinal tract of rat. *Molecular and Cellular Biochemistry* 2001; 219 (1-2): 91-8.
66. Viteri FE. Iron supplementation for the control of iron deficiency in populations at risk. *Nutr Rev* 1997; 55: 195-209.
67. Lund EK, Wharf SG, Fairweather-Tait SJ, Johnson IT. Oral ferrous sulfate supplements increase the free radical-generating capacity of feces from healthy volunteers. *Am J Clin Nutr* 2003; 78(3): 498.
68. William B, Aharon R, Itzhak NS, Ayala A, Chaim HZ, Ioav C. The assessment of serum nontransferrin-bound iron in chelation therapy and iron supplementation. *Blood* 2000; 95: 2975-82.
69. Gambling L, Andersen HS, Czopek A, Wojciak R, Krejpcio Z, McArdle HJ. Effect of timing of iron supplementation on maternal and neonatal pregnant rats. *J Physiol* 2004; 561 (15): 195-203.
70. Jansson LT, Perkiö MV, Willis WT, Refino CJ, Dallman PR. Red cell superoxide dismutase is increased in iron deficiency anemia. *Acta Haematol* 1985; 74(4): 218-21.
71. Chen Q, Le GW, Shi YH, Zhang SM, Jin X. Effect of iron supplementation on intestinal function and oxidative stress in piglets with induced colitis. *J Animal Feed Sciences* 2007; 16: 205-13.
72. Acharya J, Punchard NA, Taylor JA, Thompson RP, Pearson TC. Red cell lipid peroxidation and antioxidant enzymes in iron deficiency. *Eur J Haematol* 1991; 47(4): 287-91.
73. Ramachandran M, Iyer GY. Ramachandran M, Iyer GY. Erythrocyte glutathione reductase in iron deficiency anaemia. *Clin Chim Acta* 1974; 52(2): 225-29.
74. Sundaram RC, Selvaraj N, Vijayan G, Bobby Z, Hamide A, Dasse NR. Increased plasma malondialdehyde and fructosamine in iron deficiency anemia: Effect of treatment. *Biomedicine & Pharmacotherapy* 2007; 61(10): 682-5.

75. Isler M, Delibas N, Guclu M, Gultekin F, Sutcu R, Bahceci M, et al. Superoxide dismutase and glutathione peroxidase in erythrocytes of patients with iron deficiency anemia: effects of different treatment modalities. *Croat Med J* 2002; 43(1): 16-9.
76. Ferreira ALA, Machado PEA, Matsubara LS. Lipid peroxidation, antioxidant enzymes and glutathione levels in human erythrocytes exposed to colloidal iron hydroxide in vitro. *Braz J Med Biol Res* 1999; 32(6): 689-94.
77. Sevgi Y, Gönenç C, Ciğdem A. Neutrophil glutathione peroxidase activity in iron deficiency anaemia. *Scand J Haematol* 1986; 36(1): 58-60.
78. Olivares M, Araya M, Pizarro F, Letelier A. Erythrocyte Cu-Zn superoxide dismutase activity is decreased in iron-deficiency anemia. *Biological Trace Element Research* 2006; 112(3): 193-203.
79. Binkoski AE, Kris-Etherton PM, Beard JL. Iron Supplementation Does Not Affect the Susceptibility of LDL to Oxidative Modification in Women with Low Iron Status. *J Nutr* 2004; 134: 99-103.
80. Kumerova A, Lece A, Skesters A, Silova A, Petuhovs V. Anaemia and antioxidant defense of the red blood cells. *Mater Med Pol* 1998; 30(1-2): 12-5.
81. Rehman A, Collis CS, Yang M, Kelly M, Diplock AT, Halliwell B, et al. The Effects of iron and vitamin C co-supplementation on oxidative damage to DNA in healthy volunteers. *Biochemical and Biophysical Research Communications* 1998; 246(1): 293-8.
82. Yang M, Collis CS, Kelly M, Diplock AT, Evans CR. Do iron and vitamin C co-supplementation influence platelet function or LDL oxidizability in healthy volunteers? *Eur J Clin Nutr* 1999; 53(5): 367-74.
83. Troost FJ, Saris WH, Haenen GR, Bast A, Brummer RJ. New method to study oxidative damage and antioxidants in the human small bowel: effects of iron application. *Am J Physiol Gastrointest Liver Physiol* 2003; 285: G354-G359.
84. Devrim E, Tarhan I, Ergüder IB, Durak I. Oxidant/Antioxidant Status of Placenta, Blood, and Cord Blood Samples From Pregnant Women Supplemented With Iron. *Journal of the Society for Gynecologic Investigation* 2006; 13(7): 502-05.
85. Yip R, Mohandas N, Clark MR, Jain S, Shohet SB, Dallman PR. Red cell membrane stiffness in iron deficiency. *Blood* 1983; 62(1): 99-106.
86. Petukhov VI, Kumerova AO, Letse AG, Silova AA, Shkesters AP, Krishchuna MA, et al. Erythremia: the activity of erythrocyte antioxidant enzymes and the association with iron deficiency. *Ter Arkh* 1997; 69(4): 57-61.
87. Vives Corrons JL, Miguel-García A, Pujades MA, Miguel-Sosa A, Cambiazzo S, Linares M, et al. Increased susceptibility of microcytic red blood cells to in vitro oxidative stress. *Eur J Haematol* 1995; 55(5): 327-31.
88. Avissar N, Farkash Y, Shaklai M. Erythrocyte enzymes in polycythemia vera: a comparison to erythrocyte enzyme activities of patients with iron deficiency anemia. *Acta Haematol* 1986; 76(1): 37-43.
89. Riazantsev VV, Grishchenko OV, Pereira AA, Belous AM. Intensity of lipid peroxidation processes and activity of antioxidant enzymes in erythrocytes during anemia in pregnancy. *Ukr Biokhim Zh* 1996; 68(3): 116-20.
90. Panchenko LF, Lamchingiin T, Gerasimov AM, Sukhanov IS, Konoplina LA. Superoxide dismutase activity in the blood of children with iron deficiency anemia. *Vopr Med Khim* 1979; 25(2): 181-5.
91. Cellerino R, Guidi G, Perona G. Plasma iron and erythrocytic glutathione peroxidase activity. A possible mechanism for oxidative hemolysis in iron deficiency anemia. *Scand J Haematol* 1976; 17(2): 111-6.
92. Mehmet A, Mehmet H, Abdurrahim K, Saadet O, Hakim C, Metin C. Lymphocyte DNA damage and oxidative stress in patients with iron deficiency anemia. *Mutation research. Fundamental and molecular mechanisms of mutagenesis*. 2006; 601(2): 144-49.
93. Golovin AA, Konvai VD. Lipid peroxidation in patients with iron deficiency anemia complicated by frequent acute respiratory diseases. *Klin Med (Mosk)* 1991; 69(11): 73-5.
94. Krause A, Baerwald C, Goebel KM. Red cell metabolism and ferritin levels in iron deficiency anaemia. *Biomed Biochim Acta* 1987; 46(2-3): S218-22.
95. Jain SK, Yip R, Hoesch RM, Pramanik AK, Dallman PR, Shohet SB. Evidence of peroxidative damage to the erythrocyte membrane in iron deficiency. *Am J Clin Nutr* 1983; 37(1): 26-30.
96. Rao J, Jagadeesan V. Lipid peroxidation and activities of antioxidant enzymes in iron deficiency and effect of carcinogen feeding. *Free Radic Biol Med* 1996; 21(1): 103-8.
97. Srigiridhar K, Nair KM. Protective effects of antioxidant enzymes and GSH in vivo on iron mediated lipid peroxidation in gastrointestinal tract of rat. *Ind J Biochem Biophys* 1997; 34(4): 402-5.
98. Vir P, Kaur J, Mahmood A. Effect of Chronic Iron Ingestion on the Development of Brush Border Enzymes in Rat Intestine. *Toxicology Mechanisms and Methods* 2007; 17(7): 393 – 9.
99. Reveiz L, Gyte GM, Cuervo LG. Treatments for iron-deficiency anemia in pregnancy. *Cochrane Database Syst Rev* 2007; 18(2): CD003094.
100. Beard JL. Effectiveness and strategies of iron supplementation during pregnancy. *Am J Clin Nutr* 2000; 71(5): 1288S-1294s.