

GENOME ANNOUNCEMENT

## Genome Sequences of *Brucella melitensis* 16M and Its Two Derivatives 16M1w and 16M13w, Which Evolved *In Vivo*

Yuehua Ke, Xitong Yuan, Yufei Wang, Yaoxia Bai, Jie Xu, Hongbin Song, Liuyu Huang, and Zeliang Chen Department of Infectious Disease Control, Beijing Institute of Disease Control and Prevention, Beijing, People's Republic of China

*Brucella melitensis* is an intracellular pathogen that induces chronic infection in humans. Here, we report the genome sequences of 16M and its two derivatives, 16M1w and 16M13w, which were allowed to adapt *in vivo* for 1 and 13 weeks, respectively. Our findings contribute to the investigation of adaptive mutations and mechanisms of chronic infection by *B. melitensis*.

*Trucella melitensis* is an important intracellular bacterium that D persists inside its host and results in chronic infection (8). The current view regarding chronic infection with B. melitensis presumes low replication and mutation rates when in the host. However, this cannot be easily tested in actual infections. Laboratory evolution studies of isolates of Mycobacterium tuberculosis showed accumulation of a large number of mutations during latency (4, 7). This implied that intracellular bacteria experience great adaptive mutations during survival in their hosts (2, 3). To investigate the mechanism of chronic infection for Brucella melitensis, we analyzed in vivo adaptive mutations in BALB/c mice. Here, we report the genome sequences of three Brucella melitensis strains: 16M, a laboratory strain, and two derivatives, 16M1w and 16M13w, which were isolated from BALB/c mice after infection for 1 (acute infection) and 13 (chronic infection) weeks, respectively.

Laboratory strain 16M was used to infect BALB/c mice, and 1 and 13 weeks postinfection, bacteria were isolated from the mice. Genomic DNA was directly extracted from the bacterial cultures. All the genomes were sequenced with an Illumina GA IIx sequencer with a paired-end protocol. Low-quality reads were removed, and the remaining reads were assembled with the Clcbio genomics workbench version 4.03. About 1.3 Gb of clear data was generated for each of the three strains, resulting in an average coverage of 450-fold. The clear data were assembled into 75, 83, and 83 contigs for 16M, 16M1w, and 16M13w, respectively. A total of 3.23, 3.23, and 3.21 Mb contig sequences were obtained for the three strains.

The genome sequences were then annotated with different tools. Open reading frames (ORFs) were identified by using the RAST (Rapid Annotation using Subsystem Technology) system (1). The rRNA and tRNA were predicted with RNAmmer (5) and tRNAscan-SE 1.21 (6), respectively. A total of 3,363, 3,363, and 3,335 coding sequences were predicted for 16M, 16M1w, and 16M13w. Each of the three genome sequences has one copy of 5S rRNA, two copies of large-subunit rRNA, and one copy of small-subunit rRNA.

Comparative genomic analysis was performed using the genome sequence of 16M as a reference. To our surprise, we found only 11 single-nucleotide polymorphisms (SNPs) for 16M1w, but we found 5,019 SNPs for 16M13w. Further analysis showed that, of the SNPs of 16M13w, 3,086 were nonsynonymous sites, 1,855 were synonymous sites, and 78 were nonsense mutations. A total of 3,184 SNPs are located on chromosome I, and 1,835 are located on chromosome II. The results showing that a large number of SNPs was observed in 16M13w implied that *Brucella melitensis* in chronic infection undergoes great adaptive mutations. This finding put forward great implications of adaptive mutations in the pathogenesis of intracellular bacteria. Further analysis of the adaptive mutations and their involvement in chronic infection will be provided in our future publications.

**Nucleotide sequence accession numbers.** The draft genome sequences of *B. melitensis* 16M, 16M1w, and 16M13w are available in GenBank under accession numbers AHWC00000000, AHWD00000000, and AHWE000000000, respectively. The version for strain 16M described in this paper, the first version, was assigned accession number AHWB01000000.

## ACKNOWLEDGMENTS

This work was supported by the National Basic Research Program of China (grant number 2009CB522602), the National Natural Science Foundation of China (31000548, 31000041, and 81071320), and the National Key Program for Infectious Diseases of China (2008ZX10004-015, 2009ZX10004-103, and 2008ZX10004-008).

## REFERENCES

- 1. Aziz RK, et al. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. doi:10.1186/1471-2164-9-75.
- Brockhurst MA, Colegrave N, Rozen DE. 2011. Next-generation sequencing as a tool to study microbial evolution. Mol. Ecol. 20:972–980.
- 3. Conrad TM, Lewis NE, Palsson BO. 2011. Microbial laboratory evolution in the era of genome-scale science. Mol. Syst. Biol. 7:509.
- Ford CB, et al. 2011. Use of whole genome sequencing to estimate the mutation rate of Mycobacterium tuberculosis during latent infection. Nat. Genet. 43:482–486.
- Lagesen K, et al. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964.
- Sherman DR, Gagneux S. 2011. Estimating the mutation rate of Mycobacterium tuberculosis during infection. Nat. Genet. 43:400–401.
- 8. von Bargen K, Gorvel JP, Salcedo SP. 2012. Internal affairs: investigating the Brucella intracellular lifestyle. FEMS Microbiol. Rev. 36:533–562.

Received 19 July 2012 Accepted 27 July 2012 Address correspondence to Liuyu Huang, Huangly@nic.bmi.ac.ncn, or Zeliang Chen, zeliangchen.amms@gmail.com. Y.K., X.Y., Y.W., and Y.B. contributed equally to this work. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/JB.01293-12