

β -Endorphin: Replacement of tyrosine in position 27 by tryptophan increases analgesic potency—preparation and properties of the 2-nitrophenylsulfenyl derivative

(peptide synthesis/mouse tail-flick method/opiate receptor-binding assay)

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ABSTRACT An analog of human β -endorphin with tryptophan in position 27 has been synthesized by the solid-phase method. Bioassay of the analog showed that it had almost 4 times the analgesic potency of the parent hormone but only 68% of the opiate receptor-binding activity. This peptide is the most potent analgesic among the known synthetic analogs of β -endorphin. The 2-nitrophenylsulfenyl derivative of the analog has been prepared and shown to have a lower analgesic potency and a higher opiate receptor-binding activity than the parent compound.

All known β -endorphins, (β -EPs) isolated from pituitary glands of various species have 31 amino acids and similar primary structure (1-3). In residue position 27, the amino acid is either tyrosine or histidine (Fig. 1). Since there is no tryptophan in the β -EP molecule, it appeared of interest to synthesize an analog that has tryptophan in position 27. Moreover, photoreactive 2-nitro-4(5)-azidophenylsulfenyl derivatives of tryptophan-containing peptides have recently been described (4). If the analog that has tryptophan in position 27 is biologically active, it will be useful to study the characteristics of the β -EB receptor by using these photoaffinity labeling reagents. In this communication, we describe the synthesis and properties of the analog and its 2-nitrophenylsulfenyl derivative.

EXPERIMENTAL PROCEDURES

Aminomethyl-resin was prepared as described by Mitchell *et al.* (5) to give a load of 0.286 mmol/g by picric acid titration (6).

tert-Butoxycarbonyl(benzyl)glutamyl-4-(oxymethyl)phenylacetic acid. This compound (Boc, *tert*-butoxycarbonyl; Bzl, benzyl) was prepared as described for derivatives of other amino acids (5). The cesium salt of Boc-Glu(Bzl)-OH (3.2 mmol) (7) was allowed to react with 4-(bromomethyl)-phenylacetic acid phenacyl ester (3.2 mmol) in dimethylformamide (15 ml) for 2 hr at 24°C to give 1.607 g (83%) of a crystalline product: mp 38-40°C. After removal of the phenacyl group, the resulting product (0.985 g) was recrystallized twice from dichloromethane/petroleum ether to give a final yield of 0.74 g (59% based on phenacyl ester intermediate): mp 119-120°C; TLC (chloroform/acetic acid, 15:1) R_F 0.77 (single spot with Cl_2 /toluidine detection); $[\alpha]_D^{24}$ -17.6° (*c* 2, methanol). Analysis: Calculated for $C_{26}H_{31}NO_8$ (485.53): C, 64.32; H, 6.44; N, 2.88. Found: C, 64.28; H, 6.33; N, 2.84.

Boc-Glu(Bzl)-4-(oxymethyl)-phenylacetamidomethyl (Pam)-resin. Boc-Glu(Bzl)-4-(oxymethyl)phenylacetic acid (0.60 g, 1.24 mmol) was dissolved in CH_2Cl_2 (5 ml) and mixed at 0°C with 1.0 ml of a solution of dicyclohexylcarbodiimide (0.60

mmol) in CH_2Cl_2 . After 15 min at 0°C and 10 min with warming to 24°C, the mixture was filtered and the filtrate was added to aminomethyl-resin (2.1 g holding 11 ml of CH_2Cl_2). The mixture was stirred for 1 hr at 24°C and then treated with an additional 0.30 mmol of dicyclohexylcarbodiimide for 2 hr. The mixture was filtered and the resin was washed with CH_2Cl_2 (three 25-ml portions) and absolute ethanol (three 25-ml portions). The yield of dried product was 2.23 g; its amine content was 0.4 nmol/g by picric acid titration (6).

Protected Human β -[Trp²⁷]Endorphinyl-4-(oxymethyl)-Pam-resin. A sample (673 mg, 0.18 mmole) of Boc-Glu(Bzl)-4-(oxymethyl)-Pam-resin in a Beckman 990 peptide synthesizer was carried through the schedules described for the synthesis of human β -EP (β_h -EP) (8) with the following exceptions: (i) the use of trifluoroethanol for coupling was omitted and (ii) Boc-Trp(formyl)-OH (1.5 mmol) (9) was dissolved in a mixture of dimethylformamide (0.5 ml) and CH_2Cl_2 (2.0 ml) before carrying out coupling by the symmetrical anhydride procedure. After removal of the last Boc group, the yield of protected peptide resin was 1.494 g.

β_h -[Trp²⁷]EP. Protected peptide resin (710 mg, corresponding to 86 μ mol of starting load) was treated with liquid HF and processed through Sephadex G-10 as described (8) to give 268 mg of product. For deformylation (10), this material was dissolved in water (30 ml) and taken to pH 11.3 with 1 M NaOH. After 1.5 min, the solution was adjusted to pH 5.1 with glacial acetic acid and subjected to chromatography on CM-cellulose at 24°C in a 2.0 \times 48 cm column initially equilibrated with 0.01 M NH_4OAc , pH 4.5. A gradient was effected through a 500-ml constant volume mixing chamber containing the initial buffer by introduction of 0.4 M NH_4OAc . Collection of eighty 10-ml fractions gave two major peaks (tubes 38 and 46) and two minor ones (tubes 33 and 58), a pattern that is more complex than those consistently observed for analogs of β -EP. The inference that this result was due to side reactions in the deformylation step (9) was confirmed by the simple pattern given by chromatography on CM-cellulose of material that had not been deformylated (one major peak followed by one minor peak). In this manner, the peak centered in tube 46 was tentatively identified as corresponding to the desired product (yield, 80.5 mg). Final purification was achieved by partition chromatography on Sephadex G-50 in a 2.56 \times 45 cm column eluting with 1-butanol/pyridine/acetic acid/water, 500:300:1:1000 (vol/vol) and collecting 4.4-ml fractions. The product eluted with R_F 0.4 and isolation by lyophilization gave 25 mg of highly purified β_h -[Trp²⁷]EP (ca. 8% yield based on starting aminomethyl-resin).

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Abbreviations: β -EP, β -endorphin; β_h -EP, human β -EP; Boc, *tert*-butoxycarbonyl; Bzl, benzyl; Pam-resin, phenylacetamidomethyl-resin; NPS-Cl, 2-nitrophenylsulfenyl chloride.

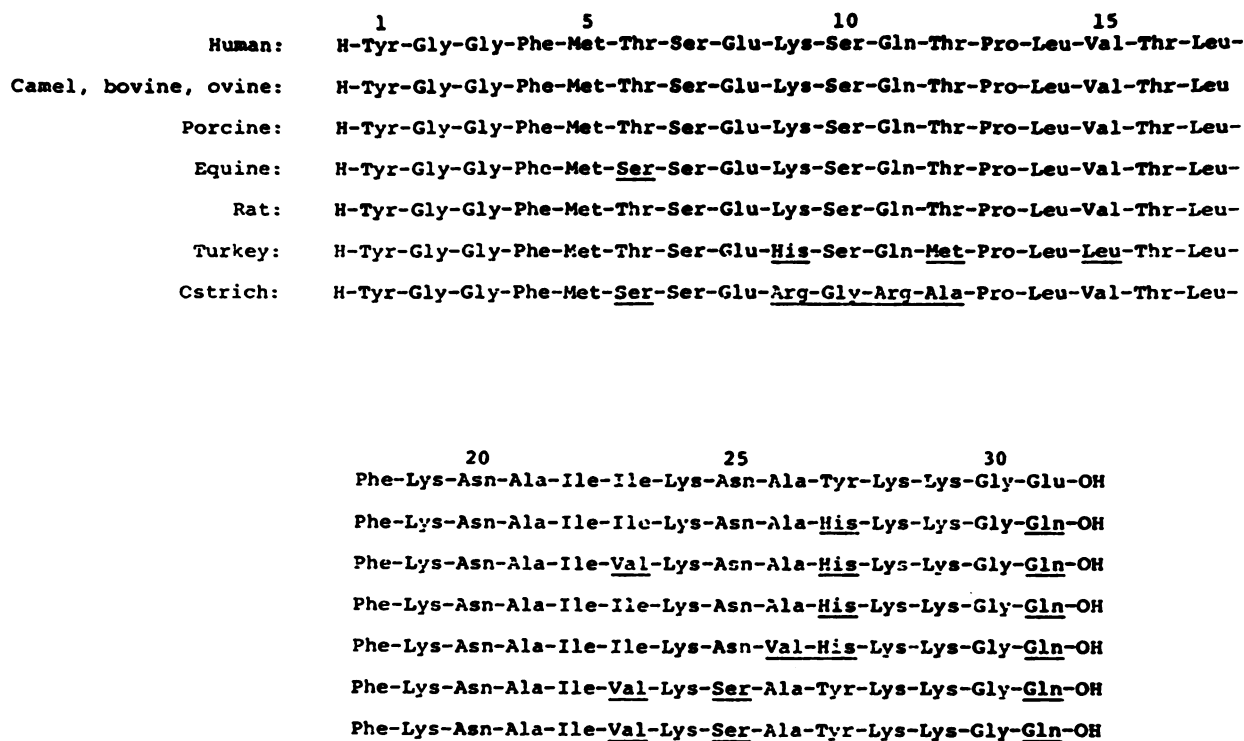


FIG. 1. Amino acid sequences of β -EP from various species.

It was homogeneous in TLC (silica gel) in 1-butanol/pyridine/ acetic acid/H₂O (5:5:1:4) (*R_F* 0.65) and on paper electrophoresis on Whatman 3 MM (400 V, 5 hr) at pH 3.7 and 6.7 (*R_F* 0.55 and 0.40, respectively, relative to lysine). In all cases 50 μ g of peptide was spotted and detection was by ninhydrin and Cl₂/toluidine.

Amino acid analysis of a 24-hr HCl hydrolysate gave (theoretical values in parentheses) aspartic acid, 2.10 (two); threonine 2.70 (three); serine, 1.67 (two); glutamic acid, 3.04 (three); proline, 0.98 (one); glycine, 2.95 (three); alanine, 2.17 (two); valine, 1.07 (one); methionine, 0.82 (one); isoleucine, 1.50 (two); leucine, 2.15 (two); tyrosine, 0.89 (one); phenylalanine, 1.91 (two); lysine, 5.08 (five); tryptophan, destroyed (the low value for isoleucine is accounted for by the acid-resistant Ile-Ile moiety). Amino acid analysis of an enzymic digest (trypsin/chymotrypsin followed by leucine aminopeptidase) gave aspartic acid, 0.22 (zero); threonine/serine/asparagine/glutamine, 7.35 (eight); glutamic acid, 2.17 (two); proline, 0.62 (one); glycine, 2.84 (three); alanine, 2.33 (two); valine, 1.16 (one); methionine, 0.99 (one); isoleucine, 1.86 (two); leucine, 1.90 (two); tyrosine, 1.06 (one); phenylalanine, 2.06 (two); lysine, 5.05 (five); tryptophane, 1.00 (one). Our current commercial batch of leucine aminopeptidase releases proline poorly.

Reaction of β_h -[Trp²⁷]-EP with 2-Nitrophenylsulfenyl Chloride. 2-Nitrophenylsulfenyl chloride (NPS-Cl) from Eastman was recrystallized as described (11). In a typical experiment, the synthetic analog (8 mg) was dissolved in 0.5 ml of 0.1 M HOAc and 22 mg of NPS-Cl in 0.5 ml of glacial HOAc was added with continuous stirring. After 1 hr at 22°C, the product was separated from excess reagent by gel filtration on a Sephadex G-10 column (1.2 \times 26 cm) previously equilibrated with 0.2 M HOAc. The peptide was recovered by lyophilization (yield, 6 mg).

To estimate the extent of modification, the NPS derivative was dissolved in 80% HOAc (1.0 mg/ml) and the amount of NPS

chromophore was determined spectrophotometrically (ϵ , 4000 at 365 m μ) according to Scoffone *et al.* (11).

Bioassay. The opiate receptor-binding assay was carried out as described (12) using rat brain membrane preparations with tritiated β_h -EP (13) as primary ligand and synthetic β_h -EP as standard competing ligand (8). Analgesic potency was estimated by the tail-flick method (14) using groups of 8–10 mice per dose as described (15).

RESULTS AND DISCUSSION

Solid-phase synthesis (16) of β_h -[Trp²⁷]EP was carried out by using the Pam-resin described by Mitchell *et al.* (5) with tryptophan protected by the formyl group (9, 17). In all other respects, synthesis and purification were carried out essentially as described for the preparation of β_h -EP (8). The analog was

Table 1. Amino acid composition of β_h -[NPS-Trp²⁷]EP

Amino acid	β_h -[NPS-Trp ²⁷]EP	β_h -[Trp ²⁷]EP	Theoretical
Lysine	4.8	5.4	5
Phenylalanine	2.0	2.0	2
Tyrosine	0.9	1.0	1
Leucine	2.1	2.1	2
Isoleucine*	1.4	1.3	2
Methionine	1.0	1.0	1
Valine	1.0	1.0	1
Alanine	1.9	1.9	2
Glycine	2.9	2.8	3
Proline	1.0	1.0	1
Glutamic acid	2.9	2.9	3
Serine†	1.6	1.6	2
Threonine†	2.8	2.7	3
Aspartic acid	2.0	2.0	2

* The Ile-Ile bond is known for resistance to 24-hr acid hydrolysis.

† Values not corrected for destruction.

Table 2. Analgesic potency and opiate receptor-binding activity of synthetic β_h -[Trp²⁷]EP and its NPS derivative

Peptide	Analgesic potency (A)		Opiate receptor-binding activity (B)		B/A
	AD ₅₀ , μ g per mouse	Relative*	IC ₅₀ , nM	Relative	
β_h -EP	0.104	100	0.615	100	1.00
β_h -[Trp ²⁷]EP	0.028	371	0.905	68	0.18
β_h -[NPS-Trp ²⁷]EP	0.168	62	0.200	260	4.20

AD₅₀, Mean analgesic dose.

* On a mol/mol basis.

obtained in highly purified form, as studies on the partition properties of β -EP and its analogs have shown (18). The relatively low yield (ca. 8%) as compared with that of the parent hormone (ca. 32%) in an earlier synthesis (8) can probably be accounted for in part by side reactions occurring during the removal of the formyl group (9). Such side reactions, which involve transfer of a formyl group to an amino group, would be expected to be more serious in a peptide containing a large number of lysine residues as β_h -EP.

The reaction of NPS-Cl with β_h -[Trp²⁷]EP in 50% HOAc was found to alkylate quantitatively the tryptophan residue as determined spectrophotometrically at 365 m μ (1.0 mol of NPS/mol of derivative). To ascertain that only the tryptophan residue was modified by NPS-Cl, the derivative was submitted to amino acid analysis after acid analysis. The results indicate that the amino acid contents of the derivative and β_h -[Trp²⁷]EP are identical (Table 1).

The biological activities of the two analogs are presented in Table 2. β_h -[Trp²⁷]EP is the most potent analgesic peptide

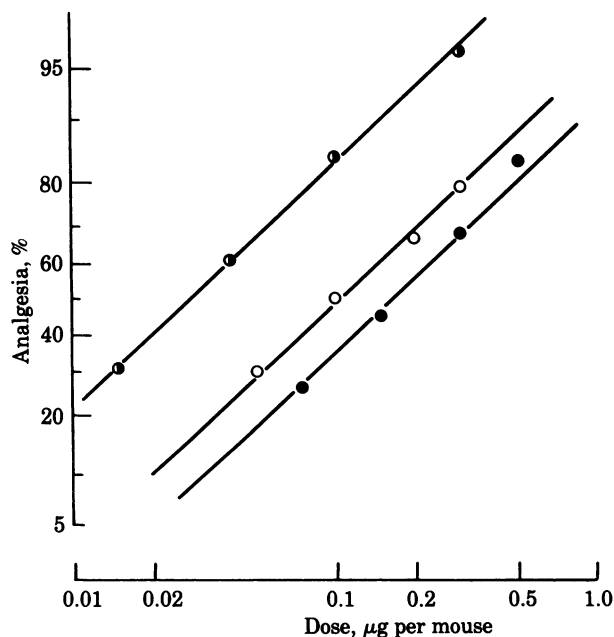


FIG. 2. Logarithmic probit dose-response relationship of antinociceptive effect produced by intracerebroventricular injection of β_h -EP and analogs. After control latency was obtained, groups of eight mice were injected and the tail-flick response was determined at 0.25, 0.50, and 1 hr. \circ , β_h -EP; \circ , β_h -[Trp²⁷]EP; \bullet , β_h -[NPS-Trp²⁷]EP.

among the various synthetic analogs of β -EP tested. It exhibits nearly 4 times the analgesic potency of the parent molecule (Fig. 2). However, it possesses only 68% of the opiate receptor-binding activity in comparison with that of β_h -EP. Thus, the ratio of binding activity to analgesic potency is 0.18, the lowest value ever obtained.

Modification of β_h -[Trp²⁷]EP with NPS-Cl decreased the analgesic potency to 1/6 that of the parent compound and increased the binding activity 4-fold. In comparison with β_h -EP, β_h -[NPS-Trp²⁷]EP possesses 62% analgesic potency. Addition of the NPS group in the tryptophan residue may cause steric hindrance, which could reduce analgesic potency, but increases the hydrophobicity, which enhances the opiate receptor-binding activity.

It has recently been reported (19) that replacement of glutamic acid-8 by glutamine increases the analgesic potency of β_h -EP 2.7-fold. The data presented here show that replacement of tyrosine-27 by tryptophan elevates the potency even more (Table 2). Earlier studies indicate that residue position 27 in β_h -EP can be replaced by histidine (20) or phenylalanine (21, 22) with moderate increase of analgesic potency. It may be possible to obtain a very potent analgesic analog by modification of residue positions 8 and 27 in the β_h -EP molecule.

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- Li, C. H. (1977) *Arch. Biochem. Biophys.* **183**, 592-604.
- Naudé, R. J., Chung, D., Li, C. H. & Oelofsen, W. (1981) *Biochem. Biophys. Res. Commun.* **98**, 108-114.
- Li, C. H., Ng, T. B., Yamashiro, D., Chung, D. & Hammonds, R. G., Jr. & Tseng, L.-F. (1981) *Int. J. Pept. Protein Res.* **18**, 242-248.
- Muramoto, K. & Ramachandran, J. (1980) *Biochemistry* **19**, 3280-3286.
- Mitchell, A. R., Kent, S. B. H., Engelhard, M. & Merrifield, R. B. (1978) *J. Org. Chem.* **43**, 2845-2852.
- Gisin, B. F. (1972) *Anal. Chim. Acta* **58**, 248-249.
- Gisin, B. F. (1973) *Helv. Chim. Acta* **56**, 1476-1482.
- Li, C. H., Yamashiro, D., Tseng, L.-F. & Loh, H. H. (1977) *J. Med. Chem.* **20**, 325-328.
- Yamashiro, D. & Li, C. H. (1973) *J. Org. Chem.* **38**, 2594-2597.
- Lemaire, S., Yamashiro, D. & Li, C. H. (1976) *J. Med. Chem.* **19**, 373-376.
- Scoffone, E., Fontana, A. & Rocchi, R. (1968) *Biochemistry* **7**, 971-979.
- Ferrara, P. & Li, C. H. (1980) *Int. J. Pept. Protein Res.* **16**, 66-69.
- Houghten, R. A., Chang, W.-C. & Li, C. H. (1980) *Int. J. Pept. Protein Res.* **16**, 311-320.
- D'Amour, F. E. & Smith, D. L. (1941) *J. Pharmacol. Exp. Ther.* **72**, 74-79.
- Loh, H. H., Tseng, L.-F., Wei, E. & Li, C. H. (1976) *Proc. Natl. Acad. Sci. USA* **73**, 2895-2898.
- Merrifield, R. B. (1963) *J. Am. Chem. Soc.* **85**, 2149-2154.
- Ohno, M., Tsukamoto, S., Makisumi, S. & Izumiya, N. (1972) *Bull. Chem. Soc. Jpn.* **45**, 2852-2855.
- Yamashiro, D. (1980) in *Hormonal Proteins and Peptides IX*, ed. Li, C. H. (Academic, New York), pp. 25-107.
- Li, C. H., Yamashiro, D., Hammonds, R. G., Jr., Nicolas, P. & Tseng, L.-F. (1981) *Biochem. Biophys. Res. Commun.* **101**, 118-123.
- Hammonds, R. G., Jr., Nicolas, P. & Li, C. H. (1982) *Int. J. Pept. Protein Res.*, in press.
- Blake, J., Tseng, L.-F., Chang, W.-C. & Li, C. H. (1978) *Int. J. Pept. Protein Res.* **11**, 323-328.
- Yamashiro, D., Ferrara, P. & Li, C. H. (1980) *Int. J. Pept. Protein Res.* **16**, 70-74.