

NIH Public Access

Author Manuscript

Int J Radiat Biol. Author manuscript; available in PMC 2012 September 27.

Published in final edited form as:

Int J Radiat Biol. 2008 December ; 84(12): 959–975. doi:10.1080/09553000802395527.

Auger processes in the 21st century

Roger W. Howell

Division of Radiation Research, Department of Radiology, New Jersey Medical School, University of Medicine and Dentistry of New Jersey, Newark, NJ, USA

Abstract

Purpose—The extreme radiotoxicity of Auger electrons and their exquisite capacity to irradiate specific molecular sites has prompted scientists to extensively investigate their radiobiological effects. Their efforts have been punctuated by quadrennial international symposia that have focused on biophysical aspects of Auger processes. The latest meeting, the 6th International Symposium on Physical, Molecular, Cellular, and Medical Aspects of Auger Processes, was held 5–6 July 2007 at Harvard Medical School in Boston, Massachusetts, USA. This article provides a review of the research in this field that was published during the years 2004–2007, the period that has elapsed since the previous meeting.

Conclusion—The field has advanced considerably. A glimpse of the potential of this unique form of ionizing radiation to contribute to future progress in a variety of fields of study is proffered.

Keywords

Auger emitters; radiation chemistry; radiation physics; DNA damage; radiotherapy; radionuclides

Introduction

When an atom is ionized by removing an electron from an inner atomic shell, the residual atom is in an excited state. Relaxation back to the ground state occurs rapidly via radiative and non-radiative processes. Radiative processes are those involving the emission of characteristic X-rays. Non-radiative processes, often referred to as Auger processes, result in the emission of Auger, Coster-Kronig (CK), and super-CK electrons. These are distinguished by the shells involved with the transition and are often collectively referred to as Auger electrons. These are competitive processes, with radiative processes being more probable for K-shell vacancies and non-radiative processes being more probable for vacancies in the L-shell and above. Thus, creation of an initial inner atomic shell vacancy

Publisher's Disclaimer: Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf> This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

^{© 2008} Informa Healthcare USA, Inc.

Correspondence: Roger W. Howell, PhD, Department of Radiology, MSB F-451, UMDNJ – New Jersey Medical School, 185 S. Orange Avenue, Newark, NJ 07103, USA. Tel: +1 973 972 5067. Fax: +1 973 972 6474. rhowell@umdnj.edu.

Declaration of interest: The author reports no conflicts of interest. The author alone is responsible for the content and writing of the paper.

leads to a series of atomic transitions involving the emission of characteristic X-rays and Auger electrons. This phenomenon has been historically referred to as the Auger effect.

Auger processes are induced by a number of mechanisms. The photoelectric effect can be used to create an inner atomic shell vacancy that leads to a subsequent shower of characterisitic X-rays and Auger electrons. This phenomenon was first observed by Pierre Auger when he exposed a cloud chamber to X-rays (Auger 1925). Inner atomic shell vacancies are also created when radionuclides decay by either electron capture or internal conversion. The ensuing shower of emitted Auger electrons was observed by Meitner while studying radioactive decay (Meitner 1923). The emitted electrons have discrete energies; however, due to the stochastic nature of the relaxation process, the numbers and energies of the emitted electrons vary for each initial vacancy created in a given subshell (i.e., K , L_1 , L_2 , etc.). Most of these Auger electrons have very low energies (\sim 20–500 eV) with ranges (\sim 1– 10 nm) in biological matter. As a consequence, biomolecules in the vicinity of the decay are subject to the direct effects of electron irradiation and indirect effects caused by radical species that arise principally from the radiolysis of water (Wright et al. 1990). In addition, the molecule containing the excited atom is also subject to damage caused by charge neutralization (Pomplun and Sutmann 2004).

Biological damage caused by Auger processes that arise from the photoelectric effect and radioactive decay has been a topic of considerable interest in basic radiobiology, radiation safety, diagnostic radiology, and radiation therapy. Radionuclides that decay by electron capture and internal conversion are commonly used in basic research laboratories and in diagnostic nuclear medicine. Examples of these Auger electron emitters include $99mTc$, $111In$, $123I$, $125I$, and $201Tl$. The keen interest in these radionuclides began when it was observed that Auger electron emitters can be highly radiotoxic when they decay in the vicinity of DNA in the cell nucleus (Ertl et al. 1970, Hofer and Hughes 1971, Bradley et al. 1975, Feinendegen 1975). In fact, some Auger electron emitters can be as radiotoxic as 210Po which emits 5.3 MeV alpha particles (Howell et al. 1990, Rao et al. 1990). Several comprehensive reviews have been published on the biological effects of Auger electron emitters (Sastry and Rao 1984, Sastry 1992, Kassis 2004, Buchegger et al. 2006, Nikjoo et al. 2006). These are an excellent resource for a complete background and analysis of the field.

The extreme radiotoxicity of Auger electron emitters prompted scientists to extensively investigate the radiobiological effects of Auger electron emitters as well as Auger electrons released as a consequence of the photoelectric effect. Their efforts have been punctuated by a series of international meetings that focused on biological aspects of Auger processes. These began with the founding meeting in 1975 that was organized by Ludwig Feinendegen in Jülich, Germany. This meeting was followed by the first in 1987 in Charney Basset, UK (Baverstock and Charton 1988), the second in 1991 in Amherst, USA (Howell et al. 1992), the third in 1995 in Lund, Sweden (Acta Oncologica 1996;35[7]), the fourth in 1999 in Lund, Sweden (*Acta Oncologica* 2000;39[6]), and the fifth in 2003 in Melbourne, Australia (International Journal of Radiation Biology 2004; 80[11–12]). The proceedings of each of these meetings have been published and are referenced above. The 2nd–5th proceedings contain a review of published work since the prior meeting (Adelstein 1992, Hofer 1996, Hofer 2000, Kassis 2004). The present manuscript continues in this tradition. It provides a review of articles related to biophysical aspects of Auger processes that were published from 2004–2007, excluding articles published in the previous proceedings.

Review of the literature 2004–2007

DNA damage by Auger processes induced by external beams

Perhaps one of the most extensively studied topics concerning biological damage caused by Auger processes has been DNA damage. These studies have been carried out with Auger processes induced by external beams of ionizing radiation as well as with radionuclides that emit Auger electrons. The last four years of research has continued this trend. Fujii et al. (2004) have used 0.538 keV synchrotron ultrasoft X-rays to create inner shell vacancies in oxygen atoms within molecular constituents of DNA including 2-deoxy-D-ribose, thymine, thymidine, and thymidine 5′-monophosphate. Upon irradiation of dry films of the sugar 2 deoxy-D-ribose, a variety of molecular fragments were observed including H^+ , CH^{-+}_{x} , $C_2H_x^+$, CO⁺, CH_xO⁺, C₃H_x⁺, C₂H₂O⁺ and C₃H_xO⁺ (x = 1, 2 and 3). When compared with the fragment yields from thymine, thymidine, and thymidine-5′-monophosphate, it was concluded that the sugar is more sensitive to the Auger effect than the thymine base.

Photon activation therapy has been under development since it was proposed by Fairchild et al. (1982). This involves using synchrotron radiation to create inner atomic vacancies in elements such as iodine and platinum, thereby utilizing the induced Auger cascades as a means to increase therapeutic effectiveness (Fairchild and Bond 1984, Sech et al. 2000). This concept was recently extended to He^{2+} beams (Usami et al. 2005). Usami et al. (2005) labeled plasmid DNA with terpyridine platinum (PtTC) and irradiated the samples with $He²⁺$ ions. They found strong correlations of single-strand break (SSB) and double-strand break (DSB) induction with concentration of PtTC, thereby suggesting that Auger cascades in ionized Pt were the key factor in this enhancement of DNA damage. Addition of dimethyl sulfoxide (DMSO) significantly reduced the enhancement. DMSO completely mitigated the enhancement for SSB induction but not for DSB.

DNA damage from Auger electron emitters in plasmid DNA

A topic of considerable importance to the field is to quantify the number of DSB that are produced per decay of radionuclides that emit Auger electrons. DNA damage caused by the Auger electron emitters 123 I and 125 I were compared by Lobachevsky and Martin (2005). They labeled plasmid DNA with ¹²³I- or ¹²⁵I-iodoHoechst 33258 and measured the SSB and DSB yields in the absence and presence of the radical scavenger DMSO. In the absence of DMSO, they found 0.62 and 0.82 DSB/decay for ^{123}I and ^{125}I , respectively. In the presence of DMSO, the values were 0.54 and 0.65, respectively. The authors indicate that their similarly high toxicity in terms of DSB/decay suggests that ¹²³I is well suited for therapeutic applications.

Balagurumoorthy et al. (2006) studied the dependence of DSB production by 125I-labeled m-iodo-p-ethoxy Hoechst 33342 (¹²⁵IEH) on the number of ¹²⁵I decays per DNA molecule. Early studies wherein the ¹²⁵IEH:DNA ratio was 42:1 indicated that ¹²⁵IEH causes ~1 DSB/ decay (Kassis et al. 1999). However, their recent studies with an ¹²⁵IEH:DNA ratio of 3:1 indicate that the previous experiments overestimated the DSB yield because additional decays in nicked DNA (the high probability of which arises as a consequence of the large number of decays per plasmid) could convert this form into the linearized form. Their analysis of their recent experiments with fewer decays per plasmid has led them to revise their DSB estimate for 125 IEH to ~ 0.5 DSB/decay.

Elmroth and Stenerlow (2005) have used pulsed field gel electrophoresis and DNA fragment analysis to measure DSB produced by 125 ₁-labeled 5-iodo-2[']-deoxyuridine (125 IdU) that was incorporated into the DNA of V79 cells. Cells were loaded with 125IdU and frozen in 10% DMSO at −70°C. They found that on average, each ¹²⁵I decay produces one local DSB (within 5 nm). In addition to this 'primary' DSB, there is on average around 0.6 DSB

produced, presumably within 1 Mbp from the primary DSB. Thus, a total of about 1.6 DSB are produced per decay. In contrast, when the conventional fraction of activity released (FAR) assay was used, only about 1 DSB per decay was measured. Even higher DSB are anticipated under physiological conditions.

Investigation continues on finding compounds that are able to direct Auger electron emitters to the DNA for therapeutic use. It has recently been shown that the aminoacridine compound N-(3-(acridine-9-ylamino)-propyl)-3-iodobenzamide (A3), labeled with 125I, binds to DNA and causes about 1–1.4 DSB/decay (Kullberg et al. 2005). Accordingly, this radiochemical has therapeutic promise provided that a pharmaceutical can be identified to deliver and internalize this agent specifically within tumor cells.

Theoretical models to study plasmid DNA damage caused by Auger electrons continue to evolve to levels of increasing sophistication. Lobachevsky et al. (2004) have created a semiempirical model to analyze plasmids following the decay of $125I$ on plasmid DNA. Their model accounts for various possibilities of creating linearized, relaxed, and supercoiled plasmids in the absence and presence of the radical scavenger DMSO. Among the possibilities are a single SSB, single DSB, or two SSB created by a single decay on a plasmid, and single SSB and DSB created by two decays on the same plasmid. Their model provided an excellent fit to the experimental data (i.e., SSB and DSB per decay) thereby affording a more comprehensive understanding of the DNA damage caused by Auger electron emitters. Kümmerle and Pomplun (2005) have taken a different approach and designed a new computer generated model of pUC19 plasmid DNA. This model can simulate the various conformations that the plasmid can undergo depending on the experimental conditions. To facilitate simulation of DNA-damage caused by Auger electrons, it has been designed to interface with Monte Carlo radiation track structure codes. With this model, Edel et al. (2006) have carried out a computer simulation of damage caused by 125I in plasmid DNA in the absence and presence of scavenging with DMSO. Their model shows that radiotoxicity of Auger electrons from 125I decay in plasmid DNA is caused mainly by direct mechanisms (92% of linearization events (LE) arises from direct hits). In contrast, indirect mechanisms were predominant when ^{125}I is freely distributed (> 87% of LE are scavengable). Good agreement was observed between the theoretical data and recent experimental findings. This indicates that their modifications including new target, new sampling algorithms, and new electron cross sections have substantially improved their capacity to simulate experimental observations.

The nature of the DNA damage caused by $125I$ decays has recently been studied by producing site-specific radiation-induced DSB in plasmid DNA by triplex forming oligonucleotide (TFO) targeted 125I (Datta et al. 2005a, 2005b, 2006). This approach enables precise delivery of 125I to a specific predetermined location on the DNA. In these studies, base damage around the site of decay was assessed on the fragments containing the 3′ and 5′ ends of the upper and lower strands. This novel approach precisely elucidated base damage clustering within eight bases of the targeted site. This damage was mitigated to some extent by the presence of DMSO. Interestingly, base damage also occurred >24 bp from the decay site. The authors hypothesize that this may be caused by a charge migration mechanism. Baverstock's soliton theory also comes to mind where it was proposed that the energy deposited in DNA by ionizing radiation can be transmitted over long distances along the molecule (Baverstock and Cundall 1988).

Interpretation of damage caused by multiple 125 I decays in TFO requires theoretical models of increasing complexity. Nikjoo et al. (2006) have developed a theoretical TFO model consisting of linked 10mers and three 125I decay sites. A total of 18,000 decays were simulated and the frequency distribution of strand breaks was recorded. The SSB

distributions around each decay site were symmetric and similar in magnitude. A highly sophisticated model has also been developed by Li and colleagues (Li et al. 2004, Li 2006) to simulate ¹²⁵I induced DNA fragmentation in DNA-protein and 41-mer synthetic oligodeoxynucleotide (oligoDNA). Radiation transport was simulated with PARTRAC and damage to DNA caused by direct (not scavengable) and indirect (scavengable) effects was studied. Direct effects included those due to direct interaction of the emitted radiations with the molecules, as well as those due to the charge neutralization processes associated with the Auger cascade process. With their inherent assumptions, the neutralization process was found to be primarily responsible for the damage caused within 5–7 base pairs of the decay site. This approach yielded good agreement between experimental data in the literature and their calculations.

Auger emitters as probes of molecular structure

Recently Auger electron emitters have been used as probes to ascertain important molecular structures. He et al. (2004b) have used ¹²⁵I to label oligonucleotides that contained four human telomeric repeats at a single internal position. The labeled oligos were then placed in different ionic conditions, allowed to fold into quadruplexes, and then frozen for decay accumulation. The probabilities of breaks at individual nucleotides were determined by sequencing gel electrophoresis. They then analyzed the quadruplex fold based on the comparison of the probability of breaks with the distance from ^{125}I to the corresponding nucleotides derived from the available nuclear magnetic resonance (NMR) and X-ray quadruplex structures. Interestingly, their data suggest that an antiparallel molecular conformation was present when both K^+ and Na^+ were present. However, Na^+ -containing solution favored the basket-type antiparallel quadruplex and K^+ favored the chair-type antiparallel quadruplex. Based on these data, it appears that antiparallel and parallel conformations coexist in solution and ion concentration determines which configuration is favored. This study attests to the unusual capacity of Auger electron emitters to elucidate important biological structures at the molecular level. This is likely to be an important use of Auger emitters as we continue with our work in this 21st century (Laughton et al. 2004, Nikjoo et al. 2006).

The experimental DNA damage data that arise from Auger processes are complex indeed. Theoretical modeling plays a key role in interpreting these data and outstanding progress has been made. The present generation of theoretical modeling of DNA damage caused by Auger processes is the result of an evolution that has occurred over more than 20 years. New capabilities in the realm of biological assays, such as those of Datta et al. (2005a, 2005b, 2006), will require that the models continue to evolve. Among the many aspects that are considered by the modelers are the stochastics of the number of decays per plasmid, electron cross sections for low energy electrons in water and in DNA, stochastics of the Auger electron spectra, direct effects of the electrons, indirect effects of radical species produced via radiolysis of water, charge transfer, charge neutralization, and the effect of molecular structure on all of the above. This list is by no means exhaustive. These models rely on the accuracy and completeness of the data that are put into them. Accordingly, it is essential that the latest discoveries in radiation physics and chemistry be incorporated into the models. The last four years have been productive and many new discoveries and technologies have emerged that are of importance to modeling Auger processes. Further evolution of our DNA damage models will likely take advantage of these new findings. These will be reviewed in the next subsection of this article.

Physics of Auger cascades

To facilitate calculation of the radiation absorbed dose, experimental investigations with radioactive materials necessarily require accurate determination of activity. Counting

efficiencies are necessary for quantifying activity of radionuclides that decay by electron capture or internal conversion. When using liquid scintillation techniques, LMM Auger electron energies are an important aspect of this determination (Malonda et al. 2006). Malonda et al. (2006) have calculated mean LMM Auger electron energies and counting efficiencies for the Auger electron emitters ${}^{55}Fe$ and ${}^{125}I$.

Emfietzglou and Nikjoo (2005) have studied the impact of various model approximations on inelastic low-energy electron cross sections for liquid water which are critical for Monte Carlo simulation of Auger electron track structure and radiation absorbed dose estimates in small volumes. They examined the influence of the optical data set, the dispersion algorithm, and the perturbation and exchange Born corrections. They found that the shape and position of the energy loss spectrum remains essentially unchanged, however, peak height is influenced up to a factor of 1.5. Larger discrepancies were observed for electron energies below 100 eV. These findings prompted them to calculate new inelastic cross sections for electrons in the energy range $0.1-10 \text{ keV}$ with accuracies $\sim 5\%$, comparable to higher energy electrons (Emfietzoglou and Nikjoo 2007). New ionization cross sections for the interaction of low energy electrons with DNA bases and sugars, and amino acids, have been developed by Peudon et al. (2006). Rather than relying on common methods that use extrapolations based on cross-sections for liquid water, their approach implements the binary-encounter dipole (BED) model. Their electron cross-sections for DNA are in good agreement with published values, and the amino acid results have not been reported on previously. Overall, these developments in cross sections for low energy electrons will play an important role in furthering our understanding of the biological effects of Auger electron emitters.

As interest in Auger electron emitters moves forward, there is continued need for knowledge of the spectrum of Auger electrons emitted by radionuclides. Given the biophysical importance of the very low energy Auger electrons, and the inherent difficulty in measuring their yields, theoretical calculation of Auger electron spectra has traditionally been the source of such data (Kassis et al. 1980, Charlton and Booz 1981, Rao et al. 1983, Pomplun et al. 1987, Howell 1992). Nikjoo et al. (2006) have recently estimated the Auger electron yields for 124I. Bé et al. (2006) have calculated highly detailed K- and L- Auger electron spectra for several radionuclides and found that their averaged values are in good agreement with values published in the literature. Both deterministic and Monte Carlo approaches have been used to calculate Auger electron spectra. The advantage of the Monte Carlo approach is that it provides information regarding the stochastics of the decay process, namely the spectrum for each individual decay. Recently, Carles and Malonda (2006) have devised a new deterministic approach that is capable of providing some detail regarding Auger spectra for individual decays. Yakushev et al. (2005) have measured the K and L Auger electron spectra of 140 Nd and 111 In. To compare their relative potential for therapeutic applications of Auger electron emitters, they determined ratios of Auger electron yields per decay. They found that 111In emitted more K- and L-shell Auger electrons than 140Nd; however, 140Nd emitted substantially more low energy L-shell Auger electrons (2.8–7 keV). Their findings were in good agreement with theoretical calculations.

Theoretical simulations of Auger electron spectra, and the ensuing interaction of Auger electrons with water, DNA, and other molecules of importance to biological processes, rely on the input data used to create the simulations. A number of recent developments in atomic physics are of importance to these simulations. The studies described in the remainder of this subsection review these important developments.

Takahashi et al. (2006) have developed a new approach for calculating Auger decay transition rates that include the effects of core-hole excited state dynamics. Their approach

Howell **Reserves and the Community Community** Page 7

was tested for normal and first resonant Auger processes in gas-phase water. Interestingly, the Auger decay of OH and O fragments that arise as a consequence of ultrafast dissociation of H2O, contributes to the total intensity. This component is of increasing importance with increasing excitation energy. With the exception of the Franck-Condon vibrational structure, their calculated Auger spectra are a close match to experimental data. Ohrwäll et al. (2005) have studied the Auger electron spectrum that arises following core ionization of liquid water clusters. The resulting Auger electron spectrum falls between those observed in the gaseous and solid phases and is characterized by broadened energy bands and a shoulder at the high energy region of the spectrum. This shoulder appears to be due to delocalized final states in which the two valence holes are mostly located at different water molecules. These studies prompt thought on the potential importance of Auger electrons emitted by ionized water molecules in the first layer of water around the DNA.

Feyer et al. (2005) have studied Auger spectra following ionization of CO and CO2. It has been shown that that proper consideration of the vibrational broadening and shift of the bands due to the dynamics of the nuclei is needed to accurately calculate the resulting Auger spectra. These findings confirm that typical calculation of Auger spectra based on vertical electronic transition energies and neglect of vibrational effects can lead to inaccuracies. This becomes more and more important as dicationic states of higher binding energies are considered. There are also many other studies that are of importance to modeling the Auger cascade process (Kato et al. 2004, Kou et al. 2004, Kugeler et al. 2004, Tanis et al. 2004, Viefhaus et al. 2004, Villani and Tarantelli 2004, Kawai 2005, Kawai et al. 2005, Hikosaka et al. 2006, Melero Garcia et al. 2006, Hikosaka et al. 2007).

Cederbaum et al. (1997) identified a new atomic de-excitation process called interatomic coulombic decay (ICD). This is distinct from Auger processes and is a competing process. When an inner atomic shell (*core*) vacancy is created as a consequence of photoionization, the resulting cation is normally found in a highly excited state above the double ionization threshold. A valence electron can fill the core hole liberating enough energy for the emission of a second electron, an Auger electron. The kinetic energy of the Auger electron is largely independent of the energy of the incident radiation and depends primarily on the intraatomic electronic structure. The Auger electron spectrum is not significantly dependent on the chemical environment. In contrast, an ionization from the inner-valence region $(\sim 20-80$ eV) leads to cationic final states below the double ionization threshold and an autoionization process on the same site is often inhibited. If the charge of a possible dication is delocalized, as it could be in a cluster, then the yielded excess energy obtained by filling the inner valence hole now is sufficient to ionize a valence electron on a *neighboring* site resulting in a two-site doubly-ionized system. This process is known as interatomic or intermolecular Coulombic decay. ICD has been shown to be a common decay mechanism in hydrogen bonded clusters as well as in van der Waals clusters (Cederbaum et al. 1997). Pernpointer et al. recently showed that inclusion of ICD has a substantial effect on Auger spectra that arise following the creation of vacancies in iodine atoms located within molecular structures (Pernpointner and Knecht 2005, Pernpointner et al. 2006). A further process called resonant ICD, analogous to resonant Auger decay, has also been demonstrated by Barth et al. (2005). Thus, it is safe to say that in the molecular environment, ICD processes compete with Auger decay. The current models used to calculate the yields and energies of Auger electrons do not consider ICD. However, it is not clear at present what significance the role that ICD would have on the radiobiological effects that result as a consequence of inner atomic shell vacancies created by photoionization, electron capture, or internal conversion. Further work is necessary to assess this.

Recent advances in experimental atomic physics have made it possible to observe atomic rearrangements in the attosecond time realm (Kienberger et al. 2004, Uiberacker et al.

2007). Laser technology was used to observe subfemtosecond ionization steps following the creation of atomic vacancies. This approach enables them to probe the transient population of short-lived valence electronic states in excited atoms or molecules. Most importantly, this provides a window into multi-electron dynamics such as Auger processes that occur on an attosecond to femtosecond timescale. Ultimately, these techniques are likely to provide valuable information of importance to biological aspects of Auger processes. These include more accurate transition rates for N- and O-shell transitions, effect of molecular interactions on Auger transition rates, and perhaps the possibility of observing Couloumb explosion in real time.

Cellular responses to Auger processes

In view of the extreme radiotoxicity of Auger electron emitters, there has been concern regarding the risk of using these radionuclides in diagnostic radiopharmaceuticals. While this debate has been quite active in the past as evidenced by numerous publications on this topic, relatively few articles were published on this topic over the last four years. The Auger electron emitter ^{99m}Tc is the most widely used Auger electron emitter in nuclear medicine. Kriehuber et al. (2004a) have exposed cultured SCLII cells to ^{99m}Tc-pertechnetate and assayed cell killing (colony forming assay), DNA damage (micronuclei), and apoptosis. Cellullar uptake was low and results for the cell killing and micronucleus end points indicate that $99mTc$ -pertechnetate is substantially less radiotoxic than acute $60C$ γ-rays (relative biological effectiveness (RBE) <1). This finding is in general agreement with those of Narra et al. (1994). Accordingly, the authors argue that the current risk estimates for $\frac{99 \text{m}}{2}$ pertechnetate are adequate and do not require reassessment. The computational modeling of Pomplun et al. arrived at a radiation weighting factor of 1.2 for $\frac{99 \text{m}}{\text{C}}$, a value in good agreement with the above data (Pomplun et al. 2006). Kriehuber et al. (2004b) did, however, find that 99mTc-pertechnetate appears to be more effective in causing apoptosis. This finding will be further investigated. In contrast to the results for ^{99m}Tc, Kriehuber et al. have used the same experimental system to show that ⁶⁵Zn is considerably more radiotoxic than acute ⁶⁰Co γ-rays. RBE values of 4, 2, and 5–8 have been found for cell killing, micronucleus formation, and apopotosis. The high radiotoxicity of this Auger electron emitter may be attributed to enhanced amounts of Zn^{2+} in the perinuclear region of the cells suggesting a major energy deposition close to the nuclear envelope.

Health risks associated with testicular exposure to diagnostic radiopharmaceuticals have historically been the subject of intense study (Hosain et al. 1978, Gupta et al. 1981, Rao et al. 1995). The high RBE of these radiochemicals in testis was demonstrated in rodents (Rao et al. 1983, 1988). Nettleton et al. (2004) recently administered 111 InCl or 201 TlCl to volunteers that were undergoing scheduled orchidectomies and, upon their surgical removal, quantified the testicular uptake of these radiopharmaceuticals. Measurements indicate that ICRP dose estimates for 111InCl and 201TlCl are under- and overestimated by a factor of about 4, respectively. As in the mouse studies (Rao et al. 1983, 1988), both radiochemicals gain access to the human seminiferous tubules (Nettleton et al. 2004). However, cellular uptake was not measured specifically. Nevertheless, their study provides valuable data for estimating the absorbed dose to human testes from 111 In and 201 Tl that should be taken into account in risk estimation.

Radioiodinated Hoechst-33342 has been used extensively to localize $125I$ in the DNA of the cell nucleus, a location generally considered to be required to realize the maximum toxicity imparted by a given Auger electron emitter. Yasui et al. have shown that cells labeled with ¹²⁵I-H33342 are killed in a dose-dependent manner consistent with a monoexponential function (Yasui et al. 2007). The mean lethal number of 125 I decays per cell was 122 and a linear increase in DNA DSB induction was observed that was equivalent to 15γ -H2AX foci/cell (Yasui et al. 2007).

Howell Page 9

Flow cytometric analysis has shown that induction of $γ$ -H2AX correlates with targeting of the nuclear genome with 125I-labeled triplex forming oligodeoxynucleotides (TFO) (Panyutin et al. 2005). Thus, γ-H2AX can be used as a sensitive indicator of targeting in order to rapidly screen a large number of 125 I-TFOs for potential application in gene therapy. Screening is essential to find optimal combinations of Auger electron emitters and TFO that will increase damage to the target region of the DNA and decrease DNA damage from non-specific irradiation (Panyutin and Neumann 2005). It stands to reason that the same approach could be adopted for other Auger electron emitting radiochemicals as well.

Urashima et al. (2006) have studied radiation-induced apoptosis caused by DNAincorporated 125IdU in several human tumor cell lines. They found that apoptosis initiated by 125IdU depends on dose, correlates with cell radiosensitivity and takes place through a caspase-3 (CASP-3)-mediated pathway. In contrast, γ-irradiation induced apoptosis does not correlate with cell radiosensitivity and appears to occur through a CASP-3-independent pathway and/or a CASP-3-mediated pathway. Thus, 125I-induced apoptosis appears to behave differently than for conventional radiations. Differences in cellular responses following irradiation with γ -rays and 125 I have also been observed in gene expression studies. Sokolov et al. (2006) compared genome-wide responses in IMR-90 human lung fibroblasts following irradiation with high- and low-dose rate γ-rays and 125IdU. They found that greater than 2000 genes were up- or down-regulated in the case of high- and lowdose rate γ-rays. In contrast, only 206 genes were altered in the case of ¹²⁵IdU. Furthermore, there was considerable overlap between changes in gene expression caused by 125 IdU and γ -rays.

An intriguing area of research is the capacity of Auger electron emitters to impart radiationinduced bystander effects. These studies were in their infancy at the time of the last Auger Symposium. Since then, Kishikawa et al. (2006) have studied the capacity of ¹²⁵I and ¹²³I to impart bystander effects in an in vivo tumor model. Specifically, they labeled LS174T cells with either ¹²⁵IdU or ¹²³IdU, mixed them with either unlabeled cells or lethally irradiated cells, and injected them subcutaneously into nude mice. Tumor size was monitored and it was found that ¹²⁵IdU caused inhibition of tumor growth whereas ¹²³IdU promoted tumor growth. The mechanisms underlying this observation were studied using microarray analysis which showed that the cell growth inhibitors TIMP1 (tissue inhibitor of metalloproteinase) and TIMP2 are overexpressed by dying 125I-labeled cells and the tumor-cell mitogen angiogenin is overexpressed by lethally irradiated 123I-labeled cells. Bystander effects caused by incorporated radionuclides have also been examined by Boyd et al. (2006). They transferred cell culture medium from flasks containing cells labeled with 125I-meta iodobenzylguanidine (MIBG), ¹³¹I-MIBG, or ²¹¹At-meta astatatobenzylguanidine (MABG) to flasks containing unirradiated recipient cells. This culture medium, which contained no significant radioactivity, was found to be toxic to the recipient cells. The cell survival response of the recipient bystander cells was dose dependent in the case of the low linear energy transfer (LET) 131 I-MIBG, but was U-shaped for the high-LET type irradiation afforded by $125I$ and $211At$. These studies again point out that Auger emitters can cause biological responses that are quite different in nature than those observed for low-LET radiations.

Finally, the effects of the Auger electron emitter 125 I on mitochondrial DNA has been a subject of interest for some time. Using a technique to inhibit mitochondrial uptake of ¹²⁵IdU, Yasui and Hofer (1986) were able to show that the radiotoxicity of ¹²⁵IdU was primarily caused by nuclear DNA damage and not mitochondrial DNA damage. This radiochemical localizes predominantly in the nuclear DNA. Given the substantial attention that mitochondrial DNA damage has received of late (Van Houten et al. 2006), it may be of interest to specifically target mitochondria with an Auger electron emitter such as $125I$. The

recent studies of VanBrocklin et al. (2007) have shown that one can target them with 125Iiodorotenol or ¹²⁵I-iodorotenone. These findings suggest that Auger electron emitters could be used as precision radioprobes to study mechanisms related to amplified oxidative stress caused by damage to mitochondrial DNA and its relationship to a variety of diseases.

Therapeutic aspects of Auger processes

Stereotactic radiosurgery plays an important role in radiation oncology. Exploration of the potential to combine stereotactic radiosurgery and photon activation therapy of cancer has recently been studied. Corde et al. (2004) have carried out studies to identify the optimal energy for IdU and contrast agent enhancement of the radiotoxicity of X-rays. They have identified 50 keV as the most effective energy which is considerably above the iodine Kedge of 33.169 keV. Monoenergetic X-rays just above the K-edge were less effective. Accordingly, Adam et al. (2006) stereotactically delivered 50 keV monochromatic X-rays to activate iodine upon its infusion into the carotid artery of glioma tumor bearing rats. They found that the combination of iodine and X-rays substantially improved survival compared to either iodine or X-rays alone. Recent studies indicate that even further enhancement can be achieved using novel approaches to enhance iodine uptake in the tumor (Rousseau et al. 2007). Studies have also carried out using the same approach with a combination of 78 keV monochromatic X-rays and cis-platinum (Biston et al. 2004).

Interest also continues in developing chemo-Auger combination therapy of cancer. This approach implements a chemotherapy agent that is labeled with an Auger electron emitter (Hou et al. 1985, Howell et al. 1986, Azure et al. 1992). Garnuszek (2004) has produced a $Pt(II)Cl₂⁻¹²⁵I-histamine complex and tested its capacity to inhibit the growth of$ transplantable colon carcinoma (C38) in mice. It was found that a combination of nonradioactive Pt(II)Cl₂-histamine and Pt(II)Cl₂-125_I-histamine was more effective than either alone in inhibiting tumor growth. It is interesting to speculate that radiations emitted by 125I, or ICD processes arising from vacancies in the residual tellurium atom, may be creating L-shell vacancies in Pt which in turn results in a large Auger cascade. Bleomycin, and the DNA-intercalators daunorubicin and dauxorubicin have also been explored for use in chemo-Auger combination therapy of cancer (Ghirmai et al. 2005, Jaaskela-Saari et al. 2005, Ickenstein et al. 2006). Recent studies with 125I-labeled dauxorubicin indicate that the radiopharmaceutical localizes in the cell nucleus and is dramatically more toxic than its unlabeled counterpart (Ickenstein et al. 2006). Finally, Bapat et al. (2005) showed that ¹²⁵Ilabeled bakuchiol, a meroterpine phenol isolated from plants, was cytotoxic when taken up by murine tumor cells and its cytotoxicity is significantly greater than unlabeled bakuchiol. Notably, mouse splenocytes were not adversely affected by 125I-bakuchiol.

There has been considerable interest in using ¹³¹I-MIBG for cancer therapy. However, adverse hematological toxicity has limited its use. He et al. have investigated the capacity of 123I-MIBG to kill cultured neuroblastoma cells with its short-range Auger and conversion electrons while sparing cells of haematopoietic lineage (He et al. 2004a). Their findings suggest that ¹²³I-MIBG may be useful for residual small volume and micrometastatic neuroblastoma.

Radiolabeled 5-iodo-2[']-deoxyuridine $(^{125}$ IdU) has been repeatedly shown to be highly cytotoxic when localized on the DNA in the cell nucleus. One of the well-known drawbacks of this radiochemical for therapeutic applications is its rapid dehalogenation in vivo. Over the past several years, efforts have been taken to explore the alternate thymidine analog 5 iodo-4′-thio-2′-deoxyuridine (ITdU) as a means to deliver 123I or 125I to the DNA of tumor cells (Reske et al. 2007). ITdU was selected for a number of reasons, among them are that it is incorporated into nuclear DNA through the thymidine salvage pathway, it is not incorporated into mitochondrial DNA, and most importantly, its in vivo stability is

considerably higher than other 5-halogenosubstituted deoxyuridines. Reske et al. (2007) have recently shown that ¹²³ITdU causes DNA damage and induction of apoptosis in leukemia cell lines that are both sensitive and resistant to β- or γ-irradiation and doxorubicin. Furthermore, like IdU, they were able to metabolically stabilize 123 ITdU by inhibiting thymidylate synthase (TS) activity with 5-fluoro-2′-deoxyuridine (FdU) and increase cellular uptake, DNA incorporation, and 123ITc–induced apoptosis. Perillo-Adamer et al. (2006) have also revisited the concept of pretreating cells with fluorodeoxyuridine (FdU) to increase the cytotoxicity of 125IdU. Early studies indicated that while FdU increases mean cellular uptake by about 3-fold, little improvement was observed in the overall killing of the population (Kassis et al. 1991). Perillo-Adamer et al. carried out additional studies with three cell lines and found that a 1-h incubation with 1 µM FdU resulted in ≥7.5-fold increase in 125IdU uptake at 16–24 h later. Cell synchronization in S phase was observed with a peak of 69.5% in the same timeframe. This corresponded to a 2.5- to 4.1-fold increase over baseline. These findings suggest that manipulation of the cellcycle with FdU may increase the therapeutic potency of ¹²⁵IdU. This strategy has been adopted for increasing tumor uptake of ¹²⁵IdU following its intratumoral administration (Buchegger et al. 2004). This approach involves pretreating glioblastoma xenograft bearing mice with FdU to block de novo synthesis of deoxythymidine to increase the uptake of 125IdU. Nearly a five-fold increase in retention of the 125IdU was observed in the FdUtreated mice compared to untreated.

Kit formulations are available for the preparation of radiolabeled IdU (Foulon et al. 1995, Schaffland et al. 2004). Semnani et al. (2005) recently showed that the formulation of a SnUdR based kit can be manipulated to maximize uptake of IdU by the tumor. Tumor to non-tumor uptake of 123 IdU/ 125 IdU was up to 6- to 7-fold higher than controls depending on the quantity of SnUdR used in the formulation. While clarification of the mechanism(s) involved in this enhancement is needed, it is possible that the SnUdR may be inhibiting/ modulating the de novo thymidylate synthetic pathways.

The radionuclide ⁶⁴Cu decays to either stable ⁶⁴Ni (β ⁺ decay, yield = 0.18; or electron capture, yield = 0.45) or to ⁶⁴Zn (β ⁻ decay, yield = 0.37). Accordingly, a number of Auger electrons are emitted by this radionuclide which has been of considerable interest for targeted radionuclide therapy. Recently, Obata et al. (2005) synthesized 64Cu-diacetyl-bis $(N⁴$ -methylthiosemicarbazone) (⁶⁴Cu-ATSM), a prospective radiotherapy agent for the treatment of hypoxic tumors, and carried out cytotoxicity studies in Lewis lung carcinoma cells. Like $124I$, this radionuclide irradiates the cells with Auger electrons, beta particles, and positrons. The radioactivity was distributed in both the cytoplasm and nucleus. About 4500 decays per cell were required to achieve 37% survival (Obata et al. 2005). This is considerably higher than the typical 100 decays required for DNA-incorporated 125I. However, assuming that the diameters of the nucleus and cell are 8 and 10 µm, respectively, and 80% of the intracellular activity is in the cytoplasm, the D_{37} dose to the cell nucleus can be estimated using the cellular S values of Goddu et al. (1997). With these assumptions one obtains D₃₇ = 4500 decays (0.8 * 3.17 × 10⁻⁴ Gy/decay + 0.2 * 1.48 × 10⁻³ Gy/decay) = 2.5 Gy.

Radioimmunotherapy of cancer with antibodies radiolabeled with Auger electron emitters is an evolving approach. In an effort to identify the optimal Auger electron emitter for this purpose, Michel et al. have recently carried out a number of studies (Michel et al. 2004, 2005a, 2005b, 2005c). Cell culture studies were undertaken with monolayers of human carcinoma cells and radiolabeled anti-epidermal growth factor receptor and anti-epithelial glycoprotein-1, conjugated to¹¹¹In or ¹²⁵I. Interestingly, their results indicate that the highest degree of cell killing can be obtained with the ¹¹¹In-labeled antibodies and this is due to the higher specific activity that can be attained in this case. Further studies were

carried out with 111 In- or 125 I-labeled anti-human epidermal growth factor receptor 2 (HER2) antibodies against breast carcinoma SK-BR-3 and ovarian carcinoma SK-OV-3.ip1 cells. Both showed good cytotoxicity. In vivo studies with anti-cluster of differentiation antigen 74 (CD74), anti-CD20, and anti-human leukocyte antigen (HLA-DR), labeled with the same Auger electron emitters, indicate that these radiopharmaceuticals can successfully treat subcutaneous Raji, Daudi, and RL-B lymphoma human tumors in severe combined immunodeficient mice. However, only disease with size <2 mm in diameter were sterilized indicating that radioimmunotherapy with Auger electron emitters will be useful for micrometastatic disease. Further progress in radioimmunotherapy has also been made in terms of using antibodies to deliver Auger electron emitters to the nucleus of tumor cells. An anti-CD33 monoclonal antibody HuM195 was modified with peptides that contain the nuclear localizing sequence of simian virus 40 large T antigen and labeled with 111 In (Chen et al. 2006b). Cell culture and animal studies indicate that the 111In was delivered to the nucleus of the tumor cells thereby indicating that this is a promising approach for treating acute myeloid leukemia (AML). Attainment of nuclear localization is a major step forward in radioimmunotherapy with Auger electron emitters.

A number of somatostatin analogs have been identified as having potential for targeted therapy of cancer. Capello et al. (2005) studied the effect of different administered activities on the therapeutic capacity of 111In-diethylenetriamine pentaacetic acid (DTPA)-octreotide against somatostatin receptor-positive rat pancreatic CA20948 tumours expressing the somatostatin receptor subtype 2 (sst2). They found a strong dependence of therapeutic effect on tumor size with therapy being most effective in animals with tumor volumes $\langle 1 \text{ cm}^3 \rangle$. Interestingly, it was also noted that the treatment upregulated receptor expression on the tumor cells thereby suggesting increased efficacy for repeat treatments. Development of new variants of ¹¹¹In labeled octreotide for diagnostic and therapeutic applications are also underway. Ginj et al. have designed and characterized new 1,4,7,10 tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA)-based peptides ¹¹¹In-DOTA-Nal³, Thr⁸-octreotide and ¹¹¹In-DOTA-BzThi³, Thr⁸-octreotide (Ginj et al. 2005a, 2005b). The peptides were compared with our clinical gold standard 111 In/ 90 Y-DOTATyr³-octreotide. They show superior pharmacologic properties when compared with with their clinical gold standard ¹¹¹In^{/90}Y-DOTATyr³-octreotide. Also studied recently is radiolabeled DOTA-Tyr³-octreotate (DOTA-TATE). Korde et al. (2007) have iodinated this analog to obtain 125I-DOTA-TATE and determined its biodistribution following intravenous administration in melanoma-bearing C57BL/6 mice. At 24 h post-injection, kidney and tumor activities were 4.7 and 2% injected dose per gram, respectively. This study suggests a role for 125 I in targeted therapies with somatostatin analogs. Clinical trials with 111 In-DTPA-Phe-Octreotide have been undertaken by Limouris et al. (2005). They selected patients with neuroendocrine liver metastases and administered 4070 to 7030 MBq at each of up to 12 administrations via selective hepatic catheterization. Follow-up was done with computed tomography (CT), magnetic resonance imaging (MRI), and ultrasound (US) with US being identified as the best of the three. Very good tumor response was observed. A pilot clinical trial that used 111In-DTPA-octreotide to treat differentiated thyroid carcinoma was carried out by Stokkel et al. (2004). They found that high doses (7,400 MBq at 2–3 week intervals) of 111In-DTPA-octreotide resulted in stabilization of the disease in a subgroup of patients. This therapeutic approach appears to be promising for patients that have a low pretreatment thyroglobulin value (i.e., small tumour load).

Kidney toxicity is often the dose-limiting factor in therapeutic applications of radiolabeled peptides, particularly for energetic beta emitters. De Jong et al. (2004) have shown that the radiolabeled peptide 111In-octreotide is distributed in a highly nonuniform fashion in human kidneys. The glomeruli, which form radiation-sensitive functional units for late radiation damage, are largely in the outer cortical regions of the kidney. However, they found most of

the radioactivity in the inner cortical zone. This accounts for the generally low radiotoxicity of radiolabeled peptides that emit particles with short ranges, such as Auger electron emitters, α-emitters, and low-energy β-emitters. A subsequent detailed radiation dosimetry analysis indicates the importance of considering these kidney radioactivity distributions when considering the biological effects of low energy electron emitters (Konijnenberg et al. 2007). Using a rat model, Rolleman et al. have recently demonstrated that colchecine can reduce kidney uptake of 111 In-DTPA-octreotide by 63%. D-lysine reduces uptake by 54%. When colchecine and D-lysine are given in combination, a 76% reduction was observed. This suggests that such pharmacologic approaches may also be used to reduce normal tissue toxicity associated with the therapeutic use of radiolabeled somatostatin analogs.

The capacity to achieve intranuclear targeting of tumor cells with 111 In labeled peptides has also been significantly advanced. Ginj et al. showed that a trifunctional N-terminal derivative of ((1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid)0,-Tyr³)octreotide (DOTA-TOC) bearing the nuclear localization signal (NLS) (of simian virus 40 large-T antigen) PKKKRKV was capable of delivering substantial amounts of radioactivity to tumor cell nuclei and with a high degree of retention (Ginj et al. 2005a, 2005b). Compared to the clinically used 111In-DOTA-TOC, one N-terminal derivative exhibited a 45-fold higher accumulation in the cell nuclei and 6-fold increase in cellular retention. Another approach involves the use of 111In and 123I labeled immunoconjugates composed of 16-mer peptides derived from HIV-1 transactivator of transcription (tat) protein and anti-mouse IgG (mIgG) (Cornelissen et al. 2007, Hu et al. 2007). These constructs were prepared and used to label cultured BT-474 breast cancer (BC) cells. Data showed that the tat peptides promoted cellular penetration and nuclear uptake of radioactivity. As pointed out by (Boswell and Brechbiel 2005), these approaches offer significant advances in using Auger electron emitters for therapy.

Watanabe et al. (2006) have targeted human neuroblastoma cells which overexpress N-myc with ¹¹¹In-labeled phosphorothioate antisense oligonucleotides. Cell culture studies showed that when 111In-labeled antisense hybridized with N-myc mRNA, N-myc expression was reduced and cell proliferation was inhibited. In vivo studies showed that when the treated neuroblastoma cells were inoculated intraperitoneally in nude mice, tumor growth was inhibited compared to controls. Based on these findings, they concluded that targeting of mRNA in tumor cells may be a valuable approach to treating cancer. Other 111 In labeled peptides for potential therapeutic applications have been created by He et al. (2004c). They have continued their progress toward the development of antisense/antigene radiotherapy approaches by chelation of peptide nucleic acid (PNA)-DTPA conjugates with 111In. The PNA-DTPA-¹¹¹In was used to target single stranded DNA and breaks were measured. The PNA directed delivery yielded about ~0.1 SSB per decay, which was three times more than for 111 In labeled DNA-oligonucleotides. Based on these findings, they concluded that PNA oligomers may be very effective for antisense/antigene radiotherapy.

Reilly et al. (2004, 2006) have continued their development of 111 In-labeled human epidermal growth factor (hEGF) to target breast cancer cells that are hEGF positive. Pharmacokinetic studies in mice and rabbits indicate that ¹¹¹In-DTPA-hEGF can be administered at multiples of the maximum dose planned for phase I human trials without causing untoward toxicity to normal tissues.

Recently, the possibility of using the ubiquitous Auger electron emitter ^{99m}Tc for cancer therapy has been explored. Häfliger et al. (2005) have synthesized a trifunctional bioconjugate consisting of the SV40 nuclear localization signal (NLS) peptide, an aliphatic triamine ligand, and the DNA intercalating pyrene and labeled it with $\frac{99 \text{m}}{\text{C}}$. This radiopharmaceutical localizes in the cell nucleus of B16F1 mouse melanoma cells and

causes a much stronger radiotoxic effect than non-nuclear-localizing ^{99m}Tc complexes. Additional complexes have also been studied (Häfliger et al. 2005). According to the authors, these data suggest that with suitable targeting, ^{99m}Tc may be useful for therapeutic applications in addition to its widespread use for diagnostic imaging. Schipper et al. (2007) have recently studied the therapeutic potential of ^{99m}Tc-pertechnetate in nude mice carrying sodium/iodide symporter (NIS) expressing neuroendocrine tumor xenografts. They found that 99mTc provided substantial tumor control; however, considerable normal tissue toxicity was present that necessitated premature sacrifice of the animals.

Antiangiogenesis approaches to cancer therapy are well established. McQuade et al. (2004) have considered the possibility of using $125I$ as an antiangiogenic agent. They used $125I$ labeled biostatin, a member of the disintegrin family of polypeptides, to target $ανβ3$ integrins in EMT-6 mammary carcinoma tumors. A maximum tumor uptake of 11.7% ID/g occurred 2 h post-injection. These data suggest that radiolabeled ligands of this type may be suitable for antiangiogenesis tumor ablation.

Chen et al. (2006a) have designed a new approach to targeting tumor cells that involves delivering a water-soluble 125I-labeled quinazolinone prodrug to the tumor cells whereupon it is enzymatically hydrolyzed to a water-insoluble form that precipitates. Intratumoral injection of this prodrug resulted in \sim 70% retention of the initially injected activity. Popisil et al. (2007) have identified prostatic acid phosphatase (PAP) as an enzyme overexpressed in prostate cancer and secreted in the extracellular space. Theoretical simulations showed that the prodrug ammonium 2-(2-phosphoryloxyphenyl)-6-iodo-4- $({}^{3}H)$ -quinazolinone (IQ₂-P) could be favorably docked into PAP. Autoradiographic studies with human prostate tumor cells showed that the radioiodinated prodrug accumulated around the cells. These and other data indicate potential for this enzyme mediated approach to cancer diagnosis and therapy (Wang et al. 2007).

The Auger emitter $125I$ was tested for its capacity to eliminate measles virus infection (Dingli et al. 2005). Cells were infected with a recombinant measles virus expressing the sodium iodide symporter (MV-NICE) and then exposed to Na¹²⁵I. Effective control of MV-NICE was obtained in cultured cells. However, preliminary in vivo studies were not similarly effective. Nevertheless, the use of Auger electron emitters to fight viral infection is intriguing.

Among the many considerations for therapy is choosing the optimal radionuclide, a topic that has been discussed extensively. Recently, Uusijärvi et al. (2006) evaluated a variety of Auger electron emitting radionuclides for their capacity to irradiate tumor tissue while minimizing the absorbed dose to normal tissues. Their analysis, which takes into account the subcellular distribution of radioactivity in tumor tissues and absorbed dose to healthy tissues, suggests that the Auger emitters with the most promise for cancer therapy from a purely dosimetric standpoint are ^{58m}Co , ^{189m}Os , ^{103m}Rh , ^{193m}Pt , and ^{195m}Pt . The latter two radionuclides have received substantial attention in the past because they emit in excess of ³⁰ Auger electrons per decay on average and they have been shown to be highly radiotoxic (Howell et al. 1986, 1994, Azure et al. 1992). However, issues related to specific activity have precluded their further advance. Avenues of producing carrier-free ^{193m}Pt have been suggested, specifically $^{192}Os(\alpha,3n)^{193m}Pt$ which has a cross section of 0.5 b with 40 MeV α-particles (Howell et al. 1986, 1994, Azure et al. 1992). This approach may be feasible in light of the recent work by Hilgers et al. (2005) who carried out $^{192}Os(p,n)^{192}Ir$. Strategies have also been devised to increase the specific activity of ^{195m}Pt by about 100-fold, a major advance in the use of this potent Auger electron emitter (Knapp et al. 2005). Based on this development, one may anticipate that further progress will made in the therapeutic use of these platinum radionuclides.

The future of Auger processes in biology and medicine

Through the efforts of a relatively few number of scientists, many substantial advances have been made in terms of our understanding of the biological effects of Auger processes and their application in medicine. A puzzling dichotomy of opinions remains with respect to the significance of Auger electrons in terms of their biological effects. They are often largely ignored by international organizations that assess radiation risks, while, at the same time, their extreme radiotoxicity is touted as the perfect precision strike against disease. One can only conclude that there is much to learn about these curious electrons which have seemingly impotent energies when considered alone, but deposit an unparalleled density of energy when emitted as a shower of low-energy electrons.

An area of intense research on Auger processes has been DNA damage. As described above, recent advances in site-specific targeting *and* damage assessment will likely facilitate further discoveries regarding DNA damage and repair. No other source of radiation has this targeting capability. At present, these studies have been largely limited to plasmid DNA, however, their implementation in cultured cells and in vivo can readily be envisioned. Also brought to the fore is the potential for Auger processes to be used for elucidating molecular structure. This approach was introduced at the last Auger Symposium in Melbourne. While an extremely potent cascade of Auger electrons may be desired for treating cancer, more delicate radiation patterns may be useful for these and other applications of these radionuclides. For example, Panyutin et al. proposed the use of Auger emitters for gene therapy (Panyutin and Neumann 1994, Panyutin et al. 2000). In these instances, there may be scope for using 'softer' Auger electron emitters that impart the least amount of collateral damage. Radionuclides that decay by electron capture to the ground state of a stable daughter atom may be useful for such applications. Such radionuclides emit only Auger electrons and X-rays. There are few such radionuclides with suitable physical half-lives, perhaps 71Ge may be ideal with a half-life of 11.4 d.

The most active area of research over the past four years has clearly been therapeutic applications of Auger electron emitters. Each meeting brings further evolution of existing strategies as well as new and novel therapeutic strategies. One important challenge to overcome in the therapeutic use of these radionuclides is differential uptake of radiopharmaceuticals by tumor cells even under ideal conditions where all the cells are exposed to the same drug concentration. Recent studies have suggested that log normal distributions of radioactivity among cell populations are likely to be the norm and that the breadth of this distribution can have a profound impact on a given radiopharmaceutical's capacity to sterilize the tumor tissue (Neti and Howell 2006). This is particularly significant for Auger electron emitters whose toxic effects are largely a result of the high self-dose to the labeled cells. With this in mind, Auger electron emitting radiopharmaceuticals that target cell populations more uniformly may ultimately be the most effective therapeutic agents. Compounds such as 125I-labeled Hoechst-33342 and -33258 enjoy highly uniform uptake which is why their unlabeled form is often used as a reagent for cell cycle analysis. While these examples may not have the targeting specificity that is desired, strategies that increase the uniformity of their uptake is something to strive for in the design of new Auger electron emitting radiopharmaceuticals and agents for photon activation therapy.

It is likely that Auger electron emitters can be used to make important discoveries in other fields of study as well. For example, Hofer (1992) suggested that the study of the aging process may benefit greatly from their use. Other topics include memory and cognitive function. This is an area of intense interest to the National Aeronautics and Space Administration (NASA) as they prepare for a manned space mission to Mars, a venture during which astronauts will receive sizeable absorbed doses from high-LET heavy ions.

Finally, one can envision the use of their precise targeting capability to gain insight into human fertility.

In conclusion, it was stated in a recent book by a well-known author that we have reached the end of the scientific age and no more major breakthrough discoveries will be made (Horgan 1996). This bleak outlook might daunt many scientists; however, similar prognostications have been made for centuries. The unique capacity of Auger electrons to deliver highly localized doses of radiation that are largely confined to nanometer dimensions make them potential tools for making breakthrough discoveries in the biological sciences. Only a small cadre of close-knit scientists with limited means have ventured into the arena of Auger processes. Much remains to be discovered by new scientists entering the field.

References

- Adam JF, Joubert A, Biston MC, Charvet AM, Peoc'h M, Le Bas JF, Balosso J, Esteve F, Elleaume H. Prolonged survival of Fischer rats bearing F98 glioma after iodine-enhanced synchrotron stereotactic radiotherapy. International Journal of Radiation Oncology Biology Physics. 2006; 64:603–611.
- Adelstein, SJ. Biophysical apsects of Auger processes: A review of the literature, 1987–1991. Howell, RW.; Narra, VR.; Sastry, KSR.; Rao, DV., editors. Woodbury, NY: American Institute of Physics; 1992. p. 1-13.[\(http://www.aapm.org/pubs/books/PROC_8.pdf](http://www.aapm.org/pubs/books/PROC_8.pdf))
- Auger P. Sur les rayons β secondaires produit dans un gaz par des rayons X. Comptes Rendues Hehdomadaires des Seances de L'Academie des Sciences. 1925; 180:65–68.
- Azure, MT.; Sastry, KSR.; Archer, RD.; Howell, RW.; Rao, DV. Microscale synthesis of carboplatin labeled with the Auger emitter Pt-193m: Radiotoxicity versus chemotoxicity of the antitumor drug in mammalian cells. Howell, RW.; Narra, VR.; Sastry, KSR.; Rao, DV., editors. Woodbury, NY: American Institute of Physics; 1992. p. 336-351.[\(http://www.aapm.org/pubs/books/default.asp](http://www.aapm.org/pubs/books/default.asp))
- Balagurumoorthy P, Chen K, Bash RC, Adelstein SJ, Kassis AI. Mechanisms underlying production of double-strand breaks in plasmid DNA after decay of 125I-Hoechst. Radiation Research. 2006; 166:333–344. [PubMed: 16881734]
- Bapat K, Chintalwar GJ, Pandey U, Thakur VS, Sarma HD, Samuel G, Pillai MR, Chattopadhyay S, Venkatesh M. Preparation and in vitro evaluation of radioiodinated bakuchiol as an anti tumor agent. Applied Radiation and Isotopes. 2005; 62:389–393. [PubMed: 15607914]
- Barth S, Joshi S, Marburger S, Ulrich V, Lindblad A, Ohrwall G, Bjorneholm O, Hergenhahn U. Observation of resonant interatomic coulombic decay in Ne clusters. Journal of Chemical Physics. 2005; 122 241102.
- Baverstock, KF.; Charton, DE., editors. DNA damage by Auger emitters. London: Taylor & Francis; 1988. 1988.
- Baverstock KF, Cundall RB. Solitons and energy transfer in DNA. Nature. 1988; 332:312–313. [PubMed: 3352732]
- Bé MM, Chiste V, Dulieu C. Detailed calculation of K- and L-Auger electron emission intensities following radioactive disintegration. Applied Radiation and Isotopes. 2006; 64:1435–1439. [PubMed: 16569503]
- Biston MC, Joubert A, Adam JF, Elleaume H, Bohic S, Charvet AM, Esteve F, Foray N, Balosso J. Cure of Fisher rats bearing radioresistant F98 glioma treated with cis-platinum and irradiated with monochromatic synchrotron X-rays. Cancer Research. 2004; 64:2317–2323. [PubMed: 15059878]
- Boswell CA, Brechbiel MW. Auger electrons: Lethal, low energy, and coming soon to a tumor cell nucleus near you. Journal of Nuclear Medicine. 2005; 46:1946–1947. [PubMed: 16330556]
- Boyd M, Ross SC, Dorrens J, Fullerton NE, Tan KW, Zalutsky MR, Mairs RJ. Radiation-induced biologic bystander effect elicited in vitro by targeted radiopharmaceuticals labeled with alpha-, beta-, and auger electron-emitting radionuclides. Journal of Nuclear Medicine. 2006; 47:1007– 1015. [PubMed: 16741311]

- Bradley EW, Chan PC, Adelstein SJ. The radiotoxicity of iodin-125 in mammalian cells. I. Effects on the survival curve of radioiodine incorporated into DNA. Radiation Research. 1975; 64:555–563. [PubMed: 1197658]
- Buchegger F, Adamer F, Schaffland AO, Kosinski M, Grannavel C, Dupertuis YM, de Tribolet N, Mach JP, Delaloye AB. Highly efficient DNA incorporation of intratumourally injected [¹²⁵I]iododeoxyuridine under thymidine synthesis blocking in human glioblastoma xenografts. International Journal of Cancer. 2004; 110:145–149.
- Buchegger F, Perillo-Adamer F, Dupertuis YM, Delaloye AB. Auger radiation targeted into DNA: A therapy perspective. European Journal of Nuclear Medicine Molecular Imaging. 2006; 33:1352– 1363.
- Capello A, Krenning E, Bernard B, Reubi JC, Breeman W, de Jong M. ¹¹¹In-labelled somatostatin analogues in a rat tumour model: Somatostatin receptor status and effects of peptide receptor radionuclide therapy. European Journal of Nuclear Medicine and Molecular Imaging. 2005; 32:1288–1295. [PubMed: 16021448]
- Carles AG, Malonda AG. Combined electron emission effects on the biologically damaging efficiency of ¹²⁵I. International Journal of Radiation Biology. 2006; 82:211-220. [PubMed: 16638718]
- Cederbaum LS, Zobeley J, Tarantelli F. Giant intermolecular decay and fragmentation of clusters. Physical Review Letters. 1997; 79:4778.
- Charlton DE, Booz J. A Monte Carlo treatment of the decay of I-125. Radiation Research. 1981; 87:10–23. [PubMed: 7255664]
- Chen K, Wang K, Kirichian AM, Al Aowad AF, Iyer LK, Adelstein SJ, Kassis AI. In silico design, synthesis, and biological evaluation of radioiodinated quinazolinone derivatives for alkaline phosphatase-mediated cancer diagnosis and therapy. Molecular Cancer Therapeutics. 2006a; 5:3001–3013. [PubMed: 17172404]
- Chen P, Wang J, Hope K, Jin L, Dick J, Cameron R, Brandwein J, Minden M, Reilly RM. Nuclear localizing sequences promote nuclear translocation and enhance the radiotoxicity of the anti-CD33 monoclonal antibody HuM195 labeled with 111 In in human myeloid leukemia cells. Journal of Nuclear Medicine. 2006b; 47:827–836. [PubMed: 16644753]
- Corde S, Joubert A, Adam JF, Charvet AM, Le Bas JF, Esteve F, Elleaume H, Balosso J. Synchrotron radiation-based experimental determination of the optimal energy for cell radiotoxicity enhancement following photoelectric effect on stable iodinated compounds. British Journal of Cancer. 2004; 91:544–551. [PubMed: 15266326]
- Cornelissen B, Hu M, McLarty K, Costantini D, Reilly RM. Cellular penetration and nuclear importation properties of 111 In-labeled and 123 I-labeled HIV-1 tat peptide immunoconjugates in BT-474 human breast cancer cells. Nuclear Medicine and Biology. 2007; 34:37–46. [PubMed: 17210460]
- Datta K, Jaruga P, Dizdaroglu M, Neumann RD, Winters TA. Molecular analysis of base damage clustering associated with a site-specific radiation-induced DNA double-strand break. Radiation Research. 2006; 166:767–781. [PubMed: 17067210]
- Datta K, Neumann RD, Winters TA. Characterization of complex apurinic/apyrimidinic-site clustering associated with an authentic site-specific radiation-induced DNA double-strand break. Proceedings of the National Academy of Sciences of the USA. 2005a; 102 10569–10574.
- Datta K, Neumann RD, Winters TA. Characterization of a complex ¹²⁵I-induced DNA double-strand break: Implications for repair. International Journal of Radiation Biology. 2005b; 81:13–21. [PubMed: 15962759]
- De Jong M, Valkema R, Van Gameren A, Van Boven H, Bex A, Van De Weyer EP, Burggraaf JD, Korner M, Reubi JC, Krenning EP. Inhomogeneous localization of radioactivity in the human kidney after injection of $[(111)$ In-DTPA]octreotide. Journal of Nuclear Medicine. 2004; 45:1168– 1171. [PubMed: 15235063]
- Dingli D, Peng KW, Harvey ME, Vongpunsawad S, Bergert ER, Kyle RA, Cattaneo R, Morris JC, Russell SJ. Interaction of measles virus vectors with Auger electron emitting radioisotopes. Biochemical Biophysical Research Communications. 2005; 337:22–29.

- Edel S, Terrissol M, Peudon A, Kummerle E, Pomplun E. Computer simulation of strand break yields in plasmid pBR322: DNA damage following 125 I decay. Radiation Protection Dosimetry. 2006; 122:136–140. [PubMed: 17185311]
- Elmroth K, Stenerlow B. DNA-incorporated 125 I induces more than one double-strand break per decay in mammalian cells. Radiation Research. 2005; 163:369–373. [PubMed: 15799691]
- Emfietzoglou D, Nikjoo H. The effect of model approximations on single-collision distributions of low-energy electrons in liquid water. Radiation Research. 2005; 163:98–111. [PubMed: 15606313]
- Emfietzoglou D, Nikjoo H. Accurate electron inelastic cross sections and stopping powers for liquid water over the 0.1–10 keV range based on an improved dielectric description of the Bethe surface. Radiation Research. 2007; 167:110–120. [PubMed: 17214512]
- Ertl HH, Feinendegen LE, Heiniger HJ. Iodine-125, a tracer in cell biology: Physical properties and biological aspects. Physics of Medicine and Biology. 1970; 15:447–456.
- Fairchild RG, Bond VP. Photon activation therapy. Strahlentherapie. 1984; 160:758–763. [PubMed: 6515666]
- Fairchild RG, Brill AB, Ettinger KV. Radiation enhancement with iodinated deoxyuridine. Investigative Radiology. 1982; 17:407–416. [PubMed: 6813285]
- Feinendegen LE. Biological damage from the Auger effect, possible benefits. Radiation Environmental Biophysics. 1975; 12:85–99.
- Feyer V, Bolognesi P, Coreno M, Prince KC, Avaldi L, Storchi L, Tarantelli F. Effects of nuclear dynamics in the low-kinetic-energy Auger spectra of CO and CO2. Journal of Chemical Physics. 2005; 123 224306.
- Foulon CF, Zhang YZ, Adelstein SJ, Kassis AI. Instantaneous preparation of radiolabeled 5-iodo-2' deoxyuridine. Applied Radiation and Isotopes. 1995; 46:1039–1046. [PubMed: 7496371]
- Fujii K, Akamatsu K, Yokoya A. Ion desorption from DNA components irradiated with 0.5 keV ultrasoft X-ray photons. Radiation Research. 2004; 161:435–441. [PubMed: 15038768]
- Garnuszek P. Marked survival prolongation of mice bearing a transplantable colon adenocarcinoma by treatment with radioactive platinum-[125I]histamine complex. Preliminary report. Nuclear Medicine Review. Central and Eastern Europe. 2004; 7:113–116. [PubMed: 15968596]
- Ghirmai S, Mume E, Tolmachev V, Sjoberg S. Synthesis and radioiodination of some daunorubicin and doxorubicin derivatives. Carbohydrate Research. 2005; 340:15–24. [PubMed: 15620662]
- Ginj M, Chen J, Walter MA, Eltschinger V, Reubi JC, Maecke HR. Preclinical evaluation of new and highly potent analogues of octreotide for predictive imaging and targeted radiotherapy. Clinical Cancer Research. 2005a; 11:1136–1145. [PubMed: 15709181]
- Ginj M, Hinni K, Tschumi S, Schulz S, Maecke HR. Trifunctional somatostatin-based derivatives designed for targeted radiotherapy using Auger electron emitters. Journal of Nuclear Medicine. 2005b; 46:2097–2103. [PubMed: 16330576]
- Goddu, SM.; Howell, RW.; Bouchet, LG.; Bolch, WE.; Rao, DV. MIRD Cellular S values: Selfabsorbed dose per unit cumulated activity for selected radionuclides and monoenergetic electron and alpha particle emitters incorporated into different cell compartments. Reston, VA: Society of Nuclear Medicine; 1997.
- Gupta SM, Herrera N, Spencer RP, Hosain F, Crucitti TW. Testicular-scrotal content of ²⁰¹Tl and ⁶⁷Ga after intravenous administration. International Journal of Nuclear Medicine and Biology. 1981; 8:211–212. [PubMed: 7319717]
- Haefliger P, Agorastos N, Renard A, Giambonini-Brugnoli G, Marty C, Alberto R. Cell uptake and radiotoxicity studies of an nuclear localization signal peptide-intercalator conjugate labeled with $[{}^{99m}Tc(CO)_3]$ ⁺ Bioconjugate Chemistry. 2005; 16:582–587. [PubMed: 15898725]
- Häfliger P, Agorastos N, Spingler B, Georgiev O, Viola G, Alberto R. Induction of DNA-doublestrand breaks by auger electrons from ^{99m}Tc complexes with DNA-binding ligands. ChemBioChem. 2005; 6:414–421. [PubMed: 15651047]
- He Y, Das B, Baruchel S, Kumar P, Wiebe L, Reilly RM. Meta-[123I]iodobenzylguanidine is selectively radiotoxic to neuroblastoma cells at concentrations that spare cells of haematopoietic lineage. Nuclear Medicine Communications. 2004a; 25:1125–1130. [PubMed: 15577592]

- He Y, Neumann RD, Panyutin IG. Intramolecular quadruplex conformation of human telomeric DNA assessed with ¹²⁵I-radioprobing. Nucleic Acids Research. 2004b; 32:5359-5367. [PubMed: 15475390]
- He Y, Panyutin IG, Karavanov A, Demidov VV, Neumann RD. Sequence-specific DNA strand cleavage by 111In-labeled peptide nucleic acids. European Journal of Nuclear Medicine and Molecular Imaging. 2004c; 31:837–845. [PubMed: 14762696]
- Hikosaka Y, Aoto T, Lablanquie P, Penent F, Shigemasa E, Ito K. Experimental investigation of corevalence double photoionization. Physical Review Letters. 2006; 97 053003.
- Hikosaka Y, Lablanquie P, Penent F, Kaneyasu T, Shigemasa E, Eland JH, Aoto T, Ito K. Double photoionization into double core-hole states in Xe. Physical Review Letters. 2007; 98 183002.
- Hilgers K, Sudar S, Qaim SM. Experimental study and nuclear model calculations on the $^{192}Os(p,n)$ ¹⁹²Ir reaction: Comparison of reactor and cyclotron production of the therapeutic radionuclide ¹⁹²Ir. Applied Radiation and Isotopes. 2005; 63:93–98. [PubMed: 15866453]
- Hofer, KG. Closing address: Biophysical aspects of Auger processes. Howell, RW.; Narra, VR.; Sastry, KSR.; Rao, DV., editors. Woodbury, NY: American Institute of Physics; 1992. p. 403-412. ([http://www.aapm.org/pubs/books/PROC_8.pdf\)](http://www.aapm.org/pubs/books/PROC_8.pdf)
- Hofer KG. Biophysical aspects of Auger processes a review. Acta Oncologica. 1996; 35:789–796. [PubMed: 9004754]
- Hofer KG. Biophysical aspects of Auger processes. Acta Oncologica. 2000; 39:651–657. [PubMed: 11130000]
- Hofer KG, Hughes WL. Radiotoxicity of intranuclear tritium, iodine-125 and iodine-131. Radiation Research. 1971; 47:94–109. [PubMed: 5559387]
- Horgan, J. The end of science: Facing the limits of knowledge in the twilight of the scientific age. Reading, MA: Addison-Wesley; 1996.
- Hosain F, Hosain P, Spencer RP. Testicular imaging with thallium-201 and comparison with other radionuclides. Journal of Nuclear Medicine. 1978; 19:720–720.
- Hou DY, Hoch H, Johnston GS, Tsou KC, Jones AE, Farkas RJ, Miller EE, Larson SM. A new In-111 bleomycin complex for combined radiotherapy and chemotherapy. Journal of Surgical Oncology. 1985; 29:91–98. [PubMed: 2417055]
- Howell RW. Radiation spectra for Auger-electron emitting radionuclides: Report No. 2 of AAPM Nuclear Medicine Task Group No. 6. Medical Physics. 1992; 19:1371–1383. [PubMed: 1461199]
- Howell RW, Kassis AI, Adelstein SJ, Rao DV, Wright HA, Hamm RN, Turner JE, Sastry KSR. Radiotoxicity of ^{195m}Pt labeled *trans*-platinum(II) in mammalian cells. Radiation Research. 1994; 140:55–62. [PubMed: 7938455]
- Howell RW, Narra VR, Rao DV, Sastry KSR. Radiobiological effects of intracellular polonium-210 alpha emissions: A comparison with Auger-emitters. Radiation Protection Dosimetry. 1990; 31:325–328.
- Howell, RW.; Narra, VR.; Sastry, KSR.; Rao, DV., editors. Biophysical aspects of Auger processes. New York: American Institute of Physics; 1992. [\(http://www.aapm.org/pubs/books/PROC_8.pdf](http://www.aapm.org/pubs/books/PROC_8.pdf))
- Howell, RW.; Sastry, KSR.; Hill, HZ.; Rao, DV. Cis-platinum-193m: Its microdosimetry and potential for chemo-Auger combination therapy of cancer. Schlafke-Stelson, AT.; Watson, EE., editors. Springfield, VA: National Technical Information Service; 1986. p. 493-513.
- Hu M, Chen P, Wang J, Scollard DA, Vallis KA, Reilly RM. (¹²³)I-labeled HIV-1 tat peptide radioimmunoconjugates are imported into the nucleus of human breast cancer cells and functionally interact in vitro and in vivo with the cyclin-dependent kinase inhibitor, p21(WAF-1/ Cip-1). European Journal of Nuclear Medicine and Molecular Imaging. 2007; 34:368–377. [PubMed: 17021818]
- Ickenstein LM, Edwards K, Sjoberg S, Carlsson J, Gedda L. A novel 125I-labeled daunorubicin derivative for radionuclide-based cancer therapy. Nuclear Medicine and Biology. 2006; 33:773– 783. [PubMed: 16934696]
- Jaaskela-Saari HA, Grenman R, Ramsay HA, Tarkkanen J, Paavonen T, Kairemo KJ. Indium-111 bleomycin complex in squamous cell cancer xenograft tumors of nude mice. Cancer Biotherapy and Radiopharmaceuticals. 2005; 20:426–435. [PubMed: 16114991]

- Kassis AI. The amazing world of Auger electrons. International Journal of Radiation Biology. 2004; 80:789–803. [PubMed: 15764386]
- Kassis AI, Adelstein SJ, Haydock C, Sastry KSR. Radiotoxicity of Se-75 and S-35: Theory and application to a cellular model. Radiation Research. 1980; 84:407–425. [PubMed: 7454987]
- Kassis AI, Guptill WE, Taube RA, Adelstein SJ. Radiotoxicity of $5-[125]$ iodo-2'-deoxyuridine in mammalian cells following treatment with 5-fluoro-2'-deoxyuridine. Journal Nuclear Biology and Medicine. 1991; 35:167–173.
- Kassis AI, Harapanhalli RS, Adelstein SJ. Comparison of strand breaks in plasmid DNA after positional changes of Auger electron-emitting iodine-125. Radiation Research. 1999; 151:167– 176. [PubMed: 9952301]
- Kato HS, Furukawa M, Kawai M, Taniguchi M, Kawai T, Hatsui T, Kosugi N. Electronic structure of bases in DNA duplexes characterized by resonant photoemission spectroscopy near the Fermi level. Physical Review Letters. 2004; 93 086403.
- Kawai J. Radiative Auger effect and extended X-ray emission fine structure (EXEFS). Analytical Sciences. 2005; 21:733–735. [PubMed: 16038486]
- Kawai J, Ohta M, Konishi T. Chemical effects in high-resolution nickel Kα X-ray fluorescence spectra. Analytical Sciences. 2005; 21:865–868. [PubMed: 16038511]
- Kienberger R, Goulielmakis E, Uiberacker M, Baltuska A, Yakovlev V, Bammer F, Scrinzi A, Westerwalbesloh T, Kleineberg U, Heinzmann U, Drescher M, Krausz F. Atomic transient recorder. Nature. 2004; 427:817–821. [PubMed: 14985755]
- Kishikawa H, Wang K, Adelstein SJ, Kassis AI. Inhibitory and stimulatory bystander effects are differentially induced by Iodine-125 and Iodine-123. Radiation Research. 2006; 165:688–694. [PubMed: 16802869]
- Knapp FF, Mirzadeh H, Beets AL, Du M. Production of therapeutic isotopes in the ORNL High Flux Isotope Reactor (HIFR) for applications in nuclear medicine, oncology, and interventional cardiology. Journal of Radiochemical and Nuclear Chemistry. 2005; 263:503–509.
- Konijnenberg M, Melis M, Valkema R, Krenning E, de Jong M. Radiation dose distribution in human kidneys by octreotides in peptide receptor radionuclide therapy. Journal of Nuclear Medicine. 2007; 48:134–142. [PubMed: 17204710]
- Korde A, Pandey U, Banerjee S, Dev Sarma H, Mukherjee A, Venkatesh M. Preparation and biological studies of ¹²⁵IDOTA- TATE. Applied Radiation and Isotopes. 2007; 65:687–690. [PubMed: 17137786]
- Kou J, Mori T, Kumar SV, Haruyama Y, Kubozono Y, Mitsuke K. Double photoionization of C(60) and C(70) in the valence region. Journal of Chemical Physics. 2004; 120:6005–6009. [PubMed: 15267482]
- Kriehuber R, Kadenbach K, Schultz F, Weiss DG. Study on cell survival, induction of apoptosis and micronucleus formation in SCL-II cells after exposure to the Auger electron emitter ^(99m)Tc. International Journal of Radiation Biology. 2004a; 80:875–880. [PubMed: 15764396]
- Kriehuber R, Riedling M, Simko M, Weiss DG. Cytotoxicity, genotoxicity and intracellular distribution of the Auger electron emitter ⁶⁵Zn in two human cell lines. Radiation and Environmental Biophysics. 2004b; 43:15–22. [PubMed: 15106009]
- Kugeler O, Prumper G, Hentges R, Viefhaus J, Rolles D, Becker U, Marburger S, Hergenhahn U. Intramolecular electron scattering and electron transfer following autoionization in dissociating molecules. Physical Review Letters. 2004; 93 033002.
- Kullberg EB, Stenerlow B, Ghirmai S, Lundqvist H, Malmstrom PU, Orlova A, Tolmachev V, Gedda L. An aminoacridine derivative for radionuclide therapy: DNA binding properties studied in a novel cell-free in vitro assay. International Journal of Oncology. 2005; 27:1355–1360. [PubMed: 16211232]
- Kümmerle EA, Pomplun E. A computer-generated super-coiled model of the pUC19 plasmid. European Biophysics Journal. 2005; 34:13–18. [PubMed: 15300338]
- Laughton CA, Grindon C, Girard P, Nikjoo H. The mysteries of telomere structure and recognition: could radioprobing help? International Journal of Radiation Biology. 2004; 80:805–811. [PubMed: 15764387]

- Li WB. Calculation of DNA strand breakage by neutralisation effect after ^{125}I decays in a synthetic oligodeoxynucleotide using charge transfer theory. Radiation Protection Dosimetry. 2006; 122:89– 94. [PubMed: 17132665]
- Li WB, Friedland W, Jacob P, Panyutin IG, Paretzke HG. Simulation of 125 I decay in a synthetic oligodeoxynucleotide with normal and distorted geometry and the role of radiation and nonradiation actions. Radiation and Environmental Biophysics. 2004; 43:23–33. [PubMed: 15042380]
- Limouris GS, Dimitropoulos N, Kontogeorgakos D, Papanikolos G, Koutoulidis V, Hatzioannou A, Mourikis D, Lyra M, Dimitriou P, Stravaka A, Vlahos L. Evaluation of the therapeutic response to In-111-DTPA octreotide-based targeted therapy in liver metastatic neuroendocrine tumors according to CT/MRI/US findings. Cancer Biotherapy and Radiopharmaceuticals. 2005; 20:215– 217. [PubMed: 15869458]
- Lobachevsky PN, Karagiannis TC, Martin RF. Plasmid DNA breakage by decay of DNA-associated Auger electron emitters: Approaches to analysis of experimental data. Radiation Research. 2004; 162:84–95. [PubMed: 15222798]
- Lobachevsky PN, Martin RF. DNA breakage by decay of Auger electron emitters: experiments with ¹²³I-iodo-Hoechst 33258 and plasmid DNA. Radiation Research. 2005; 164:766–773. [PubMed: 16296882]
- Malonda AG, Carles AG, Garcia G. Mean values of the LMM Auger transition in a KLM model. Applied Radiation and Isotopes. 2006; 64:1485–1491. [PubMed: 16574421]
- McQuade P, Knight LC, Welch MJ. Evaluation of ⁶⁴Cu and ¹²⁵I-radiolabeled bitistatin as potential agents for targeting $\alpha \vee \beta_3$ integrins in tumor angiogenesis. Bioconjugate Chemistry. 2004; 15:988–996. [PubMed: 15366951]
- Meitner L. Das β-strahlenspektrum von UX₁ und seine deutung. Zeitschrift für Physik. 1923; 17:54– 66.
- Melero Garcia E, Kivimaki A, Pettersson LG, Alvarez Ruiz J, Coreno M, de Simone M, Richter R, Prince KC. Fluorescence emission of excited hydrogen atoms after core excitation of water vapor. Physical Review Letters. 2006; 96 063003.
- Michel RB, Andrews PM, Castillo ME, Mattes MJ. In vitro cytotoxicity of carcinoma cells with ¹¹¹Inlabeled antibodies to HER-2. Molecular Cancer Therapy. 2005a; 4:927–937.
- Michel RB, Andrews PM, Rosario AV, Goldenberg DM, Mattes MJ. ¹⁷⁷Lu-antibody conjugates for single-cell kill of B-lymphoma cells in vitro and for therapy of micrometastases in vivo. Nuclear Medicine and Biology. 2005b; 32:269–278. [PubMed: 15820762]
- Michel RB, Castillo ME, Andrews PM, Mattes MJ. In vitro toxicity of A-431 carcinoma cells with antibodies to epidermal growth factor receptor and epithelial glycoprotein-1 conjugated to radionuclides emitting low-energy electrons. Clinical Cancer Research. 2004; 10:5957–5966. [PubMed: 15355929]
- Michel RB, Rosario AV, Andrews PM, Goldenberg DM, Mattes MJ. Therapy of small subcutaneous B-lymphoma xenografts with antibodies conjugated to radionuclides emitting low-energy electrons. Clinical Cancer Research. 2005c; 11:777–786. [PubMed: 15701868]
- Narra VR, Sastry KSR, Goddu SM, Howell RW, Strand SE, Rao DV. Relative biological effectiveness of 99mTc radiopharmaceuticals. Medical Physics. 1994; 21:1921–1926. [PubMed: 7700199]
- Neti PVSV, Howell RW. Log normally distributed cellular uptake of radioactivity: Implications for biological responses to radiopharmaceuticals. Journal of Nuclear Medicine. 2006; 47:1049–1058. [PubMed: 16741316]
- Nettleton JS, Lawson RS, Prescott MC, Morris ID. Uptake, localization, and dosimetry of ¹¹¹In and ²⁰¹Tl in human testes. Journal of Nuclear Medicine. 2004; 45:138–146. [PubMed: 14734686]
- Nikjoo H, Girard P, Charlton DE, Hofer KG, Laughton CA. Auger electrons a nanoprobe for structural, molecular and cellular processes. Radiation Protection Dosimetry. 2006; 122:72–79. [PubMed: 17132671]
- Obata A, Kasamatsu S, Lewis JS, Furukawa T, Takamatsu S, Toyohara J, Asai T, Welch MJ, Adams SG, Saji H, Yonekura Y, Fujibayashi Y. Basic characterization of ⁶⁴Cu-ATSM as a radiotherapy agent. Nuclear Medicine and Biology. 2005; 32:21–28. [PubMed: 15691658]
- Ohrwall G, Fink RF, Tchaplyguine M, Ojamae L, Lundwall M, Marinho RR, Naves de Brito A, Sorensen SL, Gisselbrecht M, Feifel R, Rander T, Lindblad A, Schulz J, Saethre LJ, Martensson

N, Svensson S, Bjorneholm O. The electronic structure of free water clusters probed by Auger electron spectroscopy. Journal of Chemical Physics. 2005; 123 054310.

- Panyutin IG, Neumann RD. Sequence-specific DNA double-strand breaks induced by triplex forming ¹²⁵I labeled oligonucleotides. Nucleic Acids Research. 1994; 22:4979–4982. [PubMed: 7800489]
- Panyutin IG, Neumann RD. The potential for gene-targeted radiation therapy of cancers. Trends in Biotechnology. 2005; 23:492–496. [PubMed: 16125814]
- Panyutin IG, Winters TA, Feinendegen LE, Neumann RD. Development of DNA-based radiopharmaceuticals carrying Auger-electron emitters for anti-gene radiotherapy. Quarterly Journal of Nuclear Medicine. 2000; 44:256–267. [PubMed: 11105589]
- Panyutin IV, Sedelnikova OA, Bonner WM, Panyutin IG, Neumann RD. DNA damage produced by ¹²⁵I-triplex-forming oligonucleotides as a measure of their successful delivery into cell nuclei. Annals of the New York Academy of Sciences. 2005; 1058:140–150. [PubMed: 16394133]
- Perillo-Adamer F, Delaloye AB, Genton CS, Schaffland AO, Dupertuis YM, Buchegger F. Short fluorodeoxyuridine exposure of different human glioblastoma lines induces high-level accumulation of S-phase cells that avidly incorporate ¹²⁵I-iododeoxyuridine. European Journal of Nuclear Medicine and Molecular Imaging. 2006; 33:613–620. [PubMed: 16450135]
- Pernpointner M, Knecht S. The influence of relativistic effects on the ionization spectra of the alkali iodides. Chemical Physics Letters. 2005; 410:423–429.
- Pernpointner M, Knecht S, Cederbaum LS. Ionization spectra and electronic decay in small iodide clusters: fully relativistic results. Journal of Chemical Physics. 2006; 125 34309.
- Peudon A, Edel S, Terrissol M. Molecular basic data calculation for radiation transport in chromatin. Radiation Protection Dosimetry. 2006; 122:128–135. [PubMed: 17166876]
- Pomplun E, Booz J, Charlton DE. A Monte Carlo simulation of Auger cascades. Radiation Research. 1987; 111:533–552. [PubMed: 3659286]
- Pomplun E, Sutmann G. Is coulomb explosion a damaging mechanism for $125 \text{I} \text{U} \text{d} \text{R}$? International Journal of Radiation Biology. 2004; 80:855–860. [PubMed: 15764393]
- Pomplun E, Terrissol M, Kummerle E. Estimation of a radiation weighting factor for ^{99m}Tc. Radiation Protection Dosimetry. 2006; 122:80–81. [PubMed: 17145726]
- Pospisil P, Wang K, Al Aowad AF, Iyer LK, Adelstein SJ, Kassis AI. Computational modeling and experimental evaluation of a novel prodrug for targeting the extracellular space of prostate tumors. Cancer Research. 2007; 67:2197–2205. [PubMed: 17332350]
- Rao DV, Govelitz GF, Sastry KSR. Radiotoxicity of thallium-201 in mouse testes: Inadequacy of conventional dosimetry. Journal of Nuclear Medicine. 1983; 24:145–153. [PubMed: 6822877]
- Rao DV, Narra VR, Govelitz GF, Lanka VK, Howell RW, Sastry KSR. In vivo effects of 5.3 MeV alpha particles from Po-210 in mouse testes: Comparison with internal Auger emitters. Radiation Protection Dosimetry. 1990; 31:329–332.
- Rao DV, Sastry KSR, Grimmond HE, Howell RW, Govelitz GF, Lanka VK, Mylavarapu VB. Cytotoxicity of some indium radiopharmaceuticals in mouse testes. Journal of Nuclear Medicine. 1988; 29:375–384. [PubMed: 3126279]
- Rao DV, Shepstone B, Wilkins HG, Howell RW. Testicular kinetics and dosimetry of Tl-201 in humans. Journal of Nuclear Medicine. 1995; 36:607–609. [PubMed: 7699450]
- Reilly RM, Chen P, Wang J, Scollard D, Cameron R, Vallis KA. Preclinical pharmacokinetic, biodistribution, toxicology, and dosimetry studies of 111 In-DTPA-human epidermal growth factor: An Auger electron-emitting radiotherapeutic agent for epidermal growth factor receptorpositive breast cancer. Journal of Nuclear Medicine. 2006; 47:1023–1031. [PubMed: 16741313]
- Reilly RM, Scollard DA, Wang J, Mondal H, Chen P, Henderson LA, Bowen BM, Vallis KA. A kit formulated under good manufacturing practices for labeling human epidermal growth factor with ¹¹¹In for radiotherapeutic applications. Journal of Nuclear Medicine. 2004; 45:701–708. [PubMed: 15073268]
- Reske SN, Deisenhofer S, Glatting G, Zlatopolskiy BD, Morgenroth A, Vogg AT, Buck AK, Friesen C. 123IITdU- mediated nanoirradiation of DNA efficiently induces cell kill in HL60 leukemia cells and in Doxorubicin-, β-, or γ-radiation-resistant cell lines. Journal of Nuclear Medicine. 2007; 48:1000–1007. [PubMed: 17504875]

- Rousseau J, Boudou C, Esteve F, Elleaume H. Convection-enhanced delivery of an iodine tracer into rat brain for synchrotron stereotactic radiotherapy. International Journal of Radiation Oncology Biology Physics. 2007; 68:943–951.
- Sastry KSR. Biological effects of the Auger emitter ¹²⁵I: A review. Report No. 1 of AAPM Nuclear Medicine Task Group No. 6. Medical Physics. 1992; 19:1361–1370. [PubMed: 1461198]
- Sastry, KSR.; Rao, DV. Dosimetry of low energy electrons. Rao, DV.; Chandra, R.; Graham, M., editors. New York: American Institute of Physics; 1984. p. 169-208.
- Schaffland AO, Delaloye AB, Kosinski M, Dupertuis YM, Buchegger F. The preparation of clinical grade 5- $[$ ¹²³I]iodo-2'-deoxyuridine and 5- $[$ ¹²⁵I]iodo-2'-deoxyuridine with high *in vitro* stability and the potential for early proliferation scintigraphy. Nuclear Medicine Communications. 2004; 25:461–468. [PubMed: 15100504]
- Schipper ML, Riese CG, Seitz S, Weber A, Behe M, Schurrat T, Schramm N, Keil B, Alfke H, Behr TM. Efficacy of (99^{cm}) Tc pertechnetate and $(1³¹)$ I radioisotope therapy in sodium/iodide symporter (NIS)-expressing neuroendocrine tumors in vivo. European Journal of Nuclear Medicine Molecular Imaging. 2007; 34:638–650.
- Sech CL, Takakura K, Saint-Marc C, Frohlich H, Charlier M, Usami N, Kobayashi K. Strand break induction by photoabsorption in DNA-bound molecules. Radiation Research. 2000; 153:454– 458. [PubMed: 10761007]
- Semnani ES, Wang K, Adelstein SJ, Kassis AI. 5- $[1^{23}I/125]$]iodo-2'-deoxyuridine in metastatic lung cancer: Radiopharmaceutical formulation affects targeting. Journal of Nuclear Medicine. 2005; 46:800–806. [PubMed: 15872354]
- Sokolov MV, Smirnova NA, Camerini-Otero RD, Neumann RD, Panyutin IG. Microarray analysis of differentially expressed genes after exposure of normal human fibroblasts to ionizing radiation from an external source and from DNA-incorporated iodine-125 radionuclide. Gene. 2006; 382:47–56. [PubMed: 16876969]
- Stokkel MP, Verkooijen RB, Bouwsma H, Smit JW. Six month follow-up after ¹¹¹In-DTPAoctreotide therapy in patients with progressive radioiodine non-responsive thyroid cancer: A pilot study. Nuclear Medicine Communications. 2004; 25:683–690. [PubMed: 15208495]
- Takahashi O, Odelius M, Nordlund D, Nilsson A, Bluhm H, Pettersson LG. Auger decay calculations with core-hole excited-state molecular-dynamics simulations of water. Journal of Chemical Physics. 2006; 124 64307.
- Tanis JA, Landers AL, Pole DJ, Alnaser AS, Hossain S, Kirchner T. Evidence for Pauli exchange leading to excited-state enhancement in electron transfer. Physical Review Letters. 2004; 92 133201.
- Uiberacker M, Uphues T, Schultze M, Verhoef AJ, Yakovlev V, Kling MF, Rauschenberger J, Kabachnik NM, Schroder H, Lezius M, Kompa KL, Muller HG, Vrakking MJ, Hendel S, Kleineberg U, Heinzmann U, Drescher M, Krausz F. Attosecond real-time observation of electron tunnelling in atoms. Nature. 2007; 446:627–632. [PubMed: 17410167]
- Urashima T, Nagasawa H, Wang K, Adelstein SJ, Little JB, Kassis AI. Induction of apoptosis in human tumor cells after exposure to Auger electrons: Comparison with gamma-ray exposure. Nuclear Medicine and Biology. 2006; 33:1055–1063. [PubMed: 17127180]
- Usami N, Furusawa Y, Kobayashi K, Frohlich H, Lacombe S, Le Sech C. Fast He 2+ ion irradiation of DNA loaded with platinum-containing molecules. International Journal of Radiation Biology. 2005; 81:515–522. [PubMed: 16263655]
- Uusijärvi H, Bernhardt P, Ericsson T, Forssell-Aronsson E. Dosimetric characterization of radionuclides for systemic tumor therapy: Influence of particle range, photon emission, and subcellular distribution. Medical Physics. 2006; 33:3260–3269. [PubMed: 17022220]
- Van Houten B, Woshner V, Santos JH. Role of mitochondrial DNA in toxic responses to oxidative stress. DNA Repair (Amsterdam). 2006; 5:145–152.
- VanBrocklin HF, Hanrahan SM, Enas JD, Nandanan E, O'Neil JP. Mitochondrial avid radioprobes. Preparation and evaluation of 7' (Z)- $[125]$ Jiodorotenone and 7' (Z)- $[125]$ Jiodorotenol. Nuclear Medicine and Biology. 2007; 34:109–116. [PubMed: 17210467]

- Viefhaus J, Cvejanovic S, Langer B, Lischke T, Prumper G, Rolles D, Golovin AV, Grum-Grzhimailo AN, Kabachnik NM, Becker U. Energy and angular distributions of electrons emitted by direct double auger decay. Physical Review Letters. 2004; 92 083001.
- Villani C, Tarantelli F. Double ionization of fluorinated benzenes: Hole localization and delocalization effects. Journal of Chemical Physics. 2004; 120:1775–1791. [PubMed: 15268307]
- Wang K, Kirichian AM, Aowad AF, Adelstein SJ, Kassis AI. Evaluation of chemical, physical, and biologic properties of tumor-targeting radioiodinated quinazolinone derivative. Bioconjugate Chemistry. 2007; 18:754–764. [PubMed: 17385903]
- Watanabe N, Sawai H, Ogihara-Umeda I, Tanada S, Kim EE, Yonekura Y, Sasaki Y. Molecular therapy of human neuroblastoma cells using Auger electrons of ¹¹¹In-labeled N-myc antisense oligonucleotides. Journal of Nuclear Medicine. 2006; 47:1670–1677. [PubMed: 17015904]
- Wright HA, Hamm RN, Turner JE, Howell RW, Rao DV, Sastry KSR. Calculations of physical and chemical reactions with DNA in aqueous solution from Auger cascades. Radiation Protection Dosimetry. 1990; 31:59–62.
- Yakushev EA, Kovalik A, Filosofov DV, Korolev NA, Lebedev NA, Lubashevski AV, Rosch F, Novgorodov AF. An experimental comparison of the K- and L-Auger electron spectra generated in the decays of ¹⁴⁰Nd and ¹¹¹In. Applied Radiation and Isotopes. 2005; 62:451-456. [PubMed: 15607923]
- Yasui LS, Chen K, Wang K, Jones TP, Caldwell J, Guse D, Kassis AI. Using Hoechst 33342 to target radioactivity to the cell nucleus. Radiation Research. 2007; 167:167–175. [PubMed: 17390724]
- Yasui LS, Hofer KG. Role of mitochondrial DNA in cell death induced by I-125 decay. International Journal of Radiation Biology. 1986; 49:601–610.