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Computational identification and experimental validation of microRNAs binding to the Alzheimer-related gene *ADAM10*

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Abstract

Background: MicroRNAs (miRNAs) are post-transcriptional regulators involved in numerous biological processes including the pathogenesis of Alzheimer's disease (AD). A key gene of AD, *ADAM10*, controls the proteolytic processing of *APP* and the formation of the amyloid plaques and is known to be regulated by miRNA in hepatic cancer cell lines. To predict miRNAs regulating *ADAM10* expression concerning AD, we developed a computational approach.

Methods: MiRNA binding sites in the human *ADAM10* 3' untranslated region were predicted using the RNA22, RNAhybrid and miRanda programs and ranked by specific selection criteria with respect to AD such as differential regulation in AD patients and tissue-specific expression. Furthermore, target genes of *miR-103*, *miR-107* and *miR-1306* were derived from six publicly available miRNA target site prediction databases. Only target genes predicted in at least four out of six databases in the case of *miR-103* and *miR-107* were compared to genes listed in the AlzGene database including genes possibly involved in AD. In addition, the target genes were used for Gene Ontology analysis and literature mining. Finally, we used a luciferase assay to verify the potential effect of these three miRNAs on *ADAM10* 3'UTR in SH-SY5Y cells.

Results: Eleven miRNAs were selected, which have evolutionary conserved binding sites. Three of them (*miR-103*, *miR-107*, *miR-1306*) were further analysed as they are linked to AD and most strictly conserved between different species. Predicted target genes of *miR-103* (p -value = 0.0065) and *miR-107* (p -value = 0.0009) showed significant overlap with the AlzGene database except for *miR-1306*. Interactions between *miR-103* and *miR-107* to genes were revealed playing a role in processes leading to AD. *ADAM10* expression in the reporter assay was reduced by *miR-1306* (28%), *miR-103* (45%) and *miR-107* (52%).

Conclusions: Our approach shows the requirement of incorporating specific, disease-associated selection criteria into the prediction process to reduce the amount of false positive predictions. In summary, our method identified three miRNAs strongly suggested to be involved in AD, which possibly regulate *ADAM10* expression and hence offer possibilities for the development of therapeutic treatments of AD.

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Background

MicroRNAs (miRNAs) are on average 22 nucleotides long and play a pivotal role in gene regulation. These small RNAs regulate the gene expression post-transcriptionally by suppression of mRNA translation, stimulation of mRNA deadenylation and degradation or induction of target mRNA cleavage, but have also the potential to activate translation [1,2]. Over half of the mammalian protein coding-genes are regulated by miRNAs and most human mRNAs have binding sites for miRNAs [3]. The interaction of miRNA and target mRNA requires base pairing between the seed sequence (positions 2–8) of the miRNA at the 5' end and a sequence most frequently found in the 3' untranslated region (UTR) of the target mRNA [4]. MiRNAs are involved in neuronal functions like neurite outgrowth and brain development. They were recently described to play a role in human neurodegenerative diseases. Changes in miRNA expression profiles or miRNA target sequences could contribute to the development of Parkinson's disease and Alzheimer's disease (AD) [5,6].

Characteristics of AD are insoluble plaques of amyloid β (A β) peptides emerging from the cleavage of the amyloid beta precursor protein (*APP*) and neuro-fibrillary tangles in the brains of AD patients [7,8]. The alpha-secretase "a disintegrin and metalloproteinase 10" (*ADAM10*) [9-11] generates soluble secreted amyloid precursor protein-alpha (sAPP α) and avoids formation of plaques, because it cleaves *APP* inside the A β sequence [12].

Numerous available computational methods predict a large number of genes targeted by miRNAs regulating gene expression, but only few have been validated experimentally. Many computational predictions are false positives and therefore have to be filtered out [13]. The requirement of target-site conservation in different species including far related species would be a potential way to reduce the false positive rate [14].

In this study we established an approach to identify miRNAs regulating *ADAM10* expression which therefore might influence the progression of AD. The three programs RNA22, RNAhybrid and miRanda predicted potential miRNA binding sites to *ADAM10*. We sought to identify the most interesting miRNAs possibly binding to *ADAM10* with additional selection criteria in particular whether they play a role in AD. Additionally, the most interesting miRNAs were experimentally verified by a luciferase assay. Our results show that *miR-103*, *miR-107* and *miR-1306* influence the expression of *ADAM10* at least in the reporter assay system. These miRNAs could play a role in AD and therefore are interesting candidates to be further analysed concerning their biological function and relation to AD.

Methods

miRNA target site prediction databases

MiRNA binding sites to target genes were downloaded from seven different databases: miRBase, 5-Nov-2007, <http://www.mirbase.org/> [15]; microRNA, September 2008 Release, <http://www.microrna.org/microrna/home.do> [16]; PicTar via UCSC Table Browser, assembly = May 2004 (NCBI35/hg17), group = Regulation, track = PicTar miRNA, <http://genome.ucsc.edu/> [17]; PITA, version 6 (31-Aug-2008), <http://genie.weizmann.ac.il/pubs/mir07/index.html> [18]; RNA22, March 2007, <http://cbsrv.watson.ibm.com/rna22.html> [19]; TarBase, June 2008, <http://diana.cslab.ece.ntua.gr/tarbase/> [20]; TargetScan, Release 5, <http://www.targetscan.org/> [3]. We established a workflow considering all miRNA target site predictions downloaded.

miRNA target prediction

We used three prediction programs RNA22, RNAhybrid, miRanda and predicted all binding sites of the miRNA sequences to the 3'UTR sequence of human *ADAM10*.

RNA22 is a pattern-based method for the identification of miRNA-target sites. The method has high sensitivity, is resilient to noise, can be applied to the analysis of any genome without requiring genome-specific retraining and does not rely upon cross-species conservation. Focusing on novel features of miRNA-mRNA interaction RNA22 first finds putative miRNA binding sites in the sequence of interest then identifies the targeting miRNA and hence allows to identify sites targeted by yet-undiscovered miRNAs. An implementation of RNA22 (19-May-2008) is available online at <http://cbsrv.watson.ibm.com/rna22.html> [19,21].

The second program RNAhybrid is an extension of the classical RNA secondary structure prediction algorithm from Zuker and Stiegler [22]. It finds the energetically most favorable hybridization sites of a small RNA in a large RNA incorporating 'seed-match speed-up', which first searches for seed matches in the candidate targets and only upon finding such matches the complete hybridization around the seed-match is calculated. The user can define the position and length of the seed region with the option to allow for G:U wobble base pairs in the seed pairing. Intramolecular base pairings and branching structures are forbidden and statistical significance of predicted targets is assessed with an extreme value statistics of length normalized minimum free energies, a Poisson approximation of multiple binding sites, and the calculation of effective numbers of orthologous targets in comparative studies of multiple organisms. RNAhybrid, Version 2.1, is available online at <http://bibiserv.techfak.uni-bielefeld.de/rnahybrid/> [23,24].

The miRanda algorithm is similar to the Smith-Waterman algorithm, but scores based on the complementarity of nucleotides ($A = U$ or $G \equiv C$) and one G:U wobble pair is allowed in the seed region but has to be compensated by matches in the 3' end of miRNA. In order to estimate the thermodynamic properties of a predicted pairing between miRNA and 3'UTR sequence, the algorithm uses folding routines from the Vienna 1.3 RNA secondary structure programming library (RNALib) [25]. A conservation filter is used and optionally some rudimentary statistics about each target site can be generated. MiRanda, September 2008 Release, is available online at <http://www.microrna.org/microrna/home.do> [21,26].

The parameter setting for RNA22 is: maximum number of "UN-paired" bases within the extent of the seed = 0, extent of seed in nucleotides = 6, minimum number of paired-up bases that you want to see in any reported heteroduplex = 14, maximum value for the folding energy in any reported heteroduplex = -25 kcal/mol. The parameter setting for RNAhybrid is: "-s 3utr_human" ("-s" tells RNAhybrid to quickly estimate statistical parameters from "minimal duplex energies" under the assumption that the target sequences are human 3'UTR sequences). The parameter setting for miRanda is the default parameter setting: gap open penalty = -8, gap extend = -2, score threshold = 50, energy threshold = -20 kcal/mol, scaling parameter = 4.

We retrieved the 3'UTR sequence of *ADAM10* (human *ADAM10* 3'UTR based on transcript NM_001110 (chr15:58888510-58889745)) from NCBI <http://www.ncbi.nlm.nih.gov/>. We downloaded 703 mature miRNA sequences for *Homo sapiens* from miRBase, version 13.0 <http://www.mirbase.org/> [15].

Extraction of best miRNA predictions

The extraction of miRNAs was applied according to the following selection criteria. We checked for each miRNA how many programs predicted the miRNA to bind to human *ADAM10* 3'UTR. The regulation of miRNAs in AD was verified by the publication of Cogswell et al. [27], which provides a list of miRNAs expressed in the tissues hippocampus, cerebellum and medial frontal gyrus. Another possibility to check the expression of miRNAs in the brain is the Mouse Genome Informatics (MGI) database (Mouse Genome Database, The Jackson Laboratory, Bar Harbor, Maine; <http://www.informatics.jax.org/>) [28]. Literature search by PubMed was done as an additional approval, to search for already described target genes of the miRNAs, especially for target genes involved in AD. Mouse *ADAM10* 3'UTR based on transcript NM_007399 (chr9:70625902-70628036) from NCBI <http://www.ncbi.nlm.nih.gov/> was used for binding site search of mouse miRNAs from miRBase, version

13.0 <http://www.mirbase.org/> [15]. The parameter setting for RNA22 and miRanda is the same as for human miRNA binding site prediction at the human *ADAM10* 3'UTR. The parameter setting for RNAhybrid is "-d 1.9,0.28" (1.9 is the location parameter and 0.28 the shape parameter of the assumed extreme value distribution). Additionally, we searched by TargetScan database <http://www.targetscan.org/> [3] and microRNA database <http://www.microrna.org/microrna/home.do> [16] for miRNAs binding to human *ADAM10* 3'UTR and compared the TargetScan and microRNA predictions to our list of miRNAs for equal miRNAs. We identified the number of binding sites of a miRNA in the human *ADAM10* 3'UTR predicted by each program. *ADAM10* 3'UTR sequences from ten different species were analysed for conserved regions. The following sequences were taken: human *ADAM10* 3'UTR from transcript NM_001110 (chr15:58888510-58889745), mouse *ADAM10* 3'UTR from transcript NM_007399 (chr9:70625902-70628036), horse *ADAM10* 3'UTR from transcript XM_001498169.1 (chr1:132875124-132876868), dog *ADAM10* 3'UTR from transcript XM_858910 (chr30:26596273-26598436), chimp *ADAM10* 3'UTR from transcript XM_001172393.1 (chr15:55942343-55944774), chicken *ADAM10* 3'UTR from transcript ENSGALT00000034458 (chr10:7949768-7951846), rhesus monkey *ADAM10* 3'UTR from transcript XM_001096908 (chr7:36929437-36932008), zebra fish *ADAM10* 3'UTR from transcript NM_001159314 (chr7:31745579-31747655), opossum *ADAM10* 3'UTR from transcript ENSMODT00000011088 (chr1:162230000-162230183), zebra finch *ADAM10* 3'UTR from transcript XR_054746 (chr10:6638729-6639273). For multiple sequence alignment of the ten *ADAM10* 3'UTR sequences we applied ClustalW Version 2.1 from the European Bioinformatics Institute (EBI) <http://www.ebi.ac.uk/> [29,30]. We used default parameters except: DNA Weight Matrix = 'ClustalW', Clustering = 'UPGMA'. After extraction of the conserved regions between at least seven species we looked for miRNA binding sites localized in these conserved regions. Additionally, we determined the conservation (given in percentage) of the miRNA binding site sequence from human to each species.

Statistical analysis

Statistical analysis was performed with R statistical software (R 2.8.0, <http://www.r-project.org/>). The *p*-value was computed by the R function `fisher.test` with default settings. The Fisher's exact test is used to examine the significance of the association (contingency) between the two kinds of classification. Significantly regulated genes were considered, if the *p*-value is equal or below 0.05. We generated Venn diagrams to see the overlap between target genes of *miR-103* and *miR-107* common in 4 out

of 6 databases as well as genes in the AlzGene database (<http://www.alzgene.org/>; Version: 20.06.2011) [31]. Each set of target genes of *miR-103* and *miR-107* common in 4 out of 6 databases as well as the set of target genes of *miR-1306* in the database PITA was explored for enrichment in Gene Ontology [32] by the software Pathway Studio 8.0 (Ariadne Genomics) based on database ResNet 8.0.

Literature mining and pathway analysis

Literature search by PubMed was done to extract information about the target genes of the miRNAs resulting from Pathway Studio analysis and their relation to AD. To verify the miRNAs searches were performed for miRNA interactions in all PubMed abstracts with the help of the text mining program Pathway Studio 8.0 (Ariadne Genomics) based on the Natural Language Processing (NLP) Technology. Pathway analysis was done with the software Ingenuity Systems IPA 9.0 (<http://www.ingenuity.com/>) especially with the Path Designer.

Material

Mature miRNAs and the inactive negative control were from Invitrogen (No. PM11012, PM13206, PM10632, PM10056). All RNA species were dissolved to 5 pmol/ μ l in nuclease-free water upon arrival, aliquoted and stored at -20°C .

Cloning of the ADAM10 3'UTR luciferase reporter construct

The 3'UTR of human *ADAM10* was amplified from THP-1 chromosomal DNA using the FailSafe PCR kit (Epicentre) and the following primers:

```
AD10_3UTR_for 5'GCGGCCGCGCCCATTCAGCA  
ACCCAG 3'  
AD10_3UTR_rev 5'GCGGCCGCCACTTGTGCCCC  
TAGCAGCC 3'
```

The obtained DNA fragment was verified by restriction digestion and sequencing. The 3'UTR was subsequently cloned into the *NotI* site of the pCMV-GLuc vector (NEB), which allows to monitor regulated Gaussia luciferase expression in the cell supernatant.

Cell culture

SH-SY5Y cells were cultivated in phenol red-free DMEM/F12, supplemented with 10% FCS and 1% glutamine at 37°C , 95% air moisture, 5% CO_2 and passaged twice a week with a splitting rate of $\frac{1}{2}$ to $\frac{1}{4}$.

3'UTR luciferase reporter assay

Retro-transfection was performed using 0.005 μ l Lipofectamine 2000 (Invitrogen) per μ l OptiMEM-medium and 0.1 pmol/ μ l miRNA (Invitrogen) or negative control. For combination of miRNA 1306 together with miRNA 103 or 107 a concentration of 0.05 pmol/ μ l each was used. 2 ng/ μ l endotoxin-free plasmid DNA of the 3'UTR-reporter vector were added to 45.000 cells per well in 96 well format. Control cells were mock-treated with nuclease-free water instead of RNA molecules.

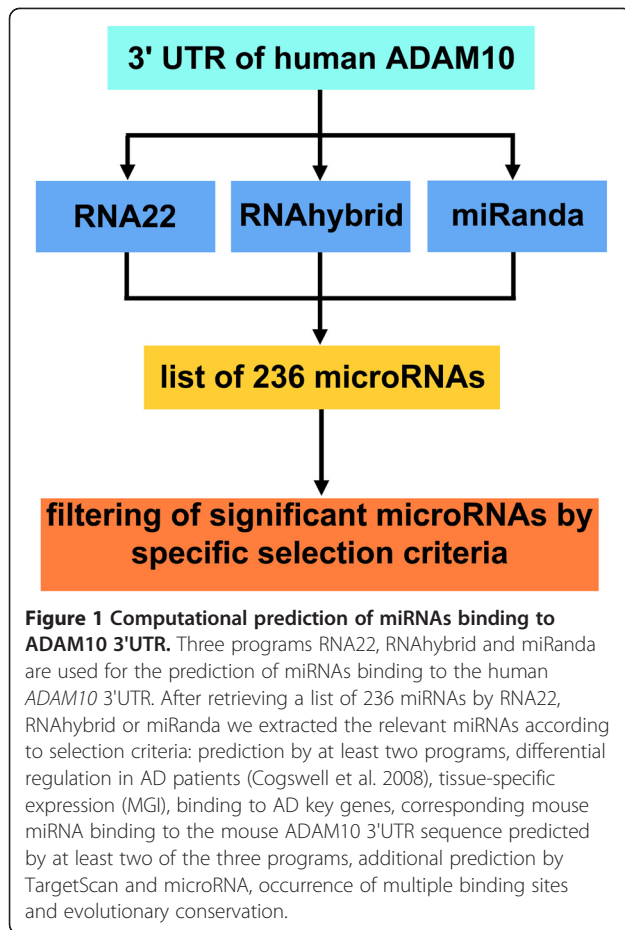
5 hrs after transfection, the cell supernatant was exchanged to 200 μ l culture medium per well. In a preliminary experiment, 10 μ l cell supernatant were collected at various time points over a 72 hour period; 48 hours were determined to be the optimal incubation time (data not shown). Therefore, 10 μ l cell supernatant were aspirated 48 hours after transfection and stored at -20°C until samples were measured. Secreted Gaussia luciferase was quantitatively analyzed (Renilla-Luciferase assay, Promega) using the FluostarOptima luminometer (BMG). Cell densities were checked by quantitation of protein content in the cell lysate by NanoQuant assay (Roth).

Results and discussion

MiRNA prediction

We established a workflow (Figure 1) to obtain miRNAs possibly binding to the human *ADAM10* 3'UTR. Instead of looking up miRNA target sites of *ADAM10* in the miRNA target site prediction databases, we predicted the binding sites of human miRNAs to human *ADAM10* 3'UTR by the three programs, RNA22, RNAhybrid and miRanda, with the aim to yield a more accurate miRNA target site prediction. Due to the fact that the three prediction programs focus on different aspects for miRNA target site prediction (pattern-based search, seed matching, conservation, energy or structure) various properties of the target sequence are covered (see Methods: "miRNA target prediction").

122 miRNAs are predicted by at least two programs to bind to human *ADAM10* 3'UTR sequence and 52 of them are significant according to expression and selection criteria described in the following. To consider different aspects of the distinct prediction algorithms at least two programs should predict a miRNA binding site. Important is also the expression of the miRNA in brain provided by the MGI database or the regulation of the miRNA in AD as described by Cogswell and colleagues (2008) in the tissues hippocampus, cerebellum and medial frontal gyrus [27]. An additional confirmation of miRNA being involved in AD is a binding site to a target gene, which is involved in AD, described in the literature and thus the miRNA might regulate also other AD key genes such as ADAM10. Furthermore, the miRNA



prediction is strengthened by the corresponding mouse miRNA binding to the mouse ADAM10 3'UTR sequence predicted by at least two of the three miRNA prediction programs RNA22, RNAhybrid and miRanda. Prediction of the miRNA by other webtools such as TargetScan and microRNA is also a confirmation of the miRNA. Multiple binding sites of a single miRNA in the 3'UTR verify the prediction [33].

The list of 52 miRNAs incorporates not conserved as well as conserved miRNAs or rather miRNA binding sites. Filtering according to conservation helps to reduce the number of miRNAs and improves the selection of candidates for further experimental validations. Consideration of conservation across species, including those not developing AD, was chosen as a filter criterion because a correlation between miRNA conservation and disease susceptibility has in general been suggested by Lu et al. [34]. Our selection procedure therefore excludes non-conserved miRNA binding sites, which also might be relevant for development of the disease but are not lost by being included in the list of 52 miRNAs. In regard to AD being a human disease, future analysis of non-conserved miRNAs might also represent a valuable approach which we did not follow in the

context of this manuscript. In our analysis eleven miRNA binding sites are conserved across at least seven species, at which four of these miRNAs are also conserved in the far related species zebra fish. The only miRNA binding site *miR-1306* in the human ADAM10 3'UTR predicted by all three programs is also conserved in the far related zebra fish as well as in mouse, horse, dog, chimp, chicken, rhesus monkey and zebra finch (Figure 2A). This binding site is located on chromosome 15 positions 58889309–58889324 and the programs RNA22, RNAhybrid and miRanda predicted the binding energy -32.29 , -25.7 and -22.55 kcal/mol, respectively. The conservation of the miRNA binding site sequence between human and the species mouse, horse, dog, chimp as well as rhesus monkey is 100%. The conservation of chicken, zebra finch and zebra fish to human in this binding region is 94%, 88% and 75%, respectively. The second most interesting miRNAs possibly binding to human ADAM10 3'UTR are *miR-103* as well as *miR-107* both having the same binding site located on chromosome 15 positions 58889443–58889468. This site is predicted by the two programs RNAhybrid and miRanda with binding energy -27.9 and -23.66 kcal/mol for *miR-103*, respectively, as well as -26.2 and -22.28 kcal/mol for *miR-107*, respectively. The conservation of the miRNA binding site sequence between human and the species mouse, horse, dog and chimp is 96%, while the conservation of chicken, rhesus monkey and finch to human is 65%, 100% and 73%, respectively (Figure 2B–C). Additionally, *miR-202*, *miR-423-5p*, *miR-503*, *miR-184* and *miR-922* bind also to the conserved binding region chromosome 15 positions 58889443–58889473 and *miR-330-5p* (chr15:58889149–58889178), *miR-671-5p* (chr15:58889720–58889745) and *miR-432* (chr15:58889688–58889718) bind to a region with good conservation also to the far related species zebra fish, but these eight miRNAs have no indication to be involved in AD (Table 1). Table 1 shows a ranked list of the best miRNA binding site predictions according to the specific selection criteria. We chose the three most interesting miRNAs 1306, 107 and 103 and performed analyses with the AlzGene database, further miRNA target site prediction databases, Gene Ontology, literature mining and validation experiments to identify the involvement in AD. *MiR-107* and *miR-103* are downregulated with age [35] as well as in AD gray matter [36] and repress the translation of cofilin. In brains of a transgenic mouse model of AD the level of *miR-103* and *miR-107* is decreased while the cofilin protein level is increased which results in the formation of rod-like structures [37]. Furthermore, *miR-107* expression is decreased even in the earliest stages of AD. As *miR-107* regulates beta-site APP-cleaving enzyme 1 (*BACE1*) it might be involved in accelerated disease progression

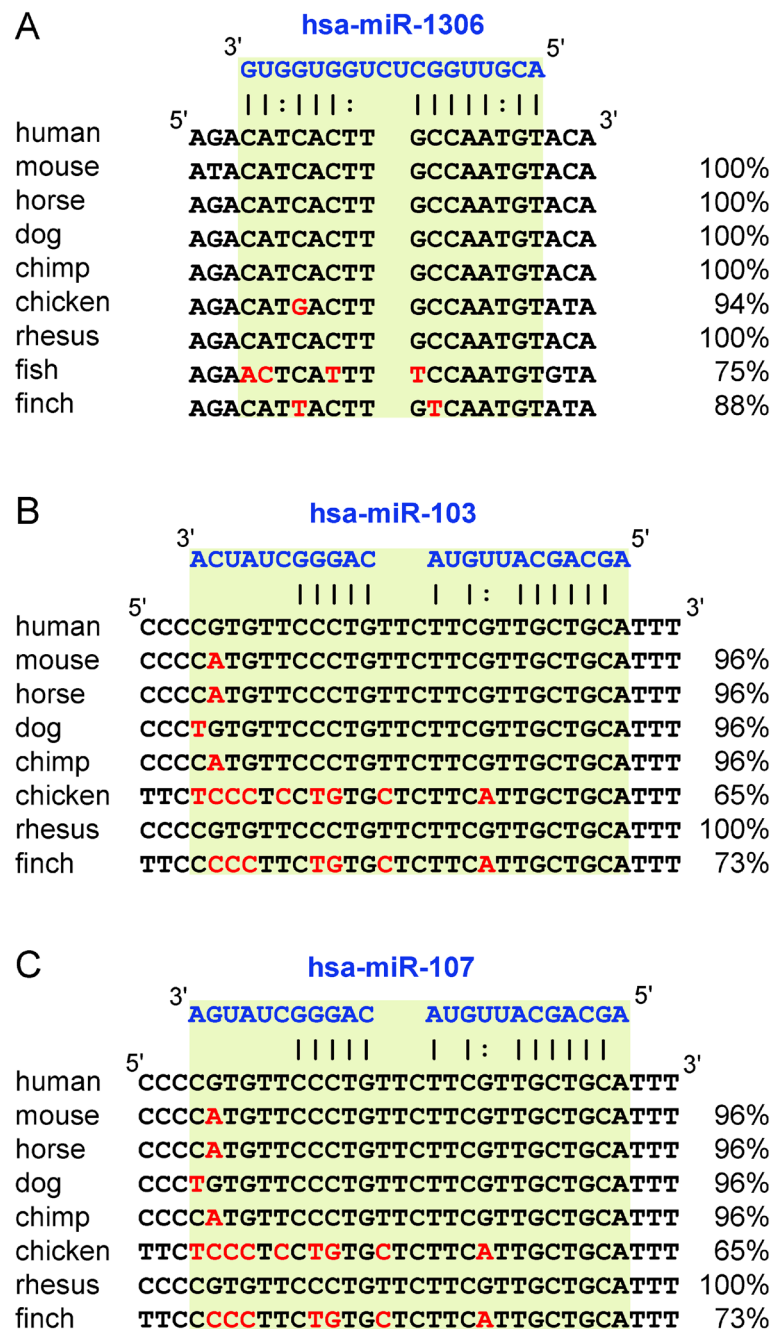


Figure 2 Conservation of the three miRNA binding sites within the ADAM10 3'UTR. The figure shows the conservation of the *miR-1306* (A), *miR-103* (B) and *miR-107* (C) binding region (light green) between different species. The blue sequence represents the miRNA binding to the DNA of different species as listed on the left side. On the right side the conservation of the miRNA binding region (light green) from different species to human sequence is given. Nucleotide mismatches in the binding region to human binding region are marked in red. The lines and colons below the miRNA sequence show perfect nucleotide matches and G:U/T wobble pairs, respectively.

[38]. The downregulation of miR-103 and miR-107 with age could concern a protective effect against plaque formation because reduced levels of these miRNAs would lead to an increased level of the predicted target ADAM10 and its neuroprotective product sAPP α in brains of AD patients. Our observations of strong

inhibition (> 40%) of ADAM10 expression in the reporter assay upon application of miR-103 and miR-107 would coincide with such a possible protective influence on amyloid pathology (see "Experimental validation of bioinformatically predicted miRNAs"). According to the publication from Cogswell et al. [27] *miR-103* is

Table 1 List of predicted miRNAs binding to a conserved region of human ADAM10 3'UTR

miRNA	∅ kcal/mol	Confirmations	Differential regulated in AD	Conservation zebra fish
1306	-26.85	predicted by 3 programs, mouse ADAM10		+
107	-24.24	targets BACE1, predicted by TargetScan, literature for AD, mouse ADAM10		
103	-25.78	predicted by TargetScan, literature for AD, mouse ADAM10	hippocampus, cerebellum	
330-5p	-27.20	predicted by microRNA, mouse ADAM10	hippocampus	+
432	-22.81	predicted by microRNA	cerebellum	+
423-5p	-22.1	mouse ADAM10	hippocampus, medial frontal gyrus	
671-5p	-27.61	mouse ADAM10		+
922	-27.99	predicted by microRNA		
503	-25.41	predicted by microRNA, mouse ADAM10		
202	-25.33	predicted by microRNA, mouse ADAM10		
184	-23.33	mouse ADAM10		

The best eleven predicted miRNAs, which bind to a conserved region of human *ADAM10* 3'UTR, are shown with additional information like the average predicted binding energy in kcal/mol (column 2), verifications of the miRNAs from literature, by additional mouse ADAM10 prediction or other prediction programs (column 3), the regulation of miRNAs in AD according to Cogswell et al. (column 4) and the conservation of the binding region in zebra fish marked with + in the last column.

differentially expressed in hippocampus and cerebellum in AD. In addition, the program TargetScan verifies the same binding site of *miR-103* and *miR-107* to human *ADAM10*. *MiR-1306* is further analysed due to its good conservation to the far related species zebra fish with only one mismatch in the seed region. It is the only miRNA whose binding site to human *ADAM10* is predicted by all three programs RNA22, RNAhybrid and miRanda, which strengthens the assumption that this binding site is functionally active. Additionally as shown in Figure 3 the hypothesis is verified that *miR-1306* is associated to AD: Twelve predicted target genes of *miR-1306* are involved in processes and functions playing a role in AD, the nervous system and other

neurodegenerative diseases. The predictions rely on TargetScan while the dedicated functions are Ingenuity Expert Findings or from Gene Ontology. *MiR-1306* possibly regulates genes like the cholinergic receptor, nicotinic, alpha 4 (*CHRNA4*), tumor necrosis factor receptor superfamily member 1B (*TNFRSF1B*) and mitogen-activated protein kinase kinase 4 (*MAP2K4*), which are associated to AD by the functions frontotemporal dementia, demyelination of neurons and Huntington's disease, respectively. *MAP2K4* has been found to be involved in AD and is putatively regulated by modules of transcription factor binding sites [39]. Furthermore, *miR-1306* is located on chromosome 22 within the second exon of DiGeorge syndrome critical region gene

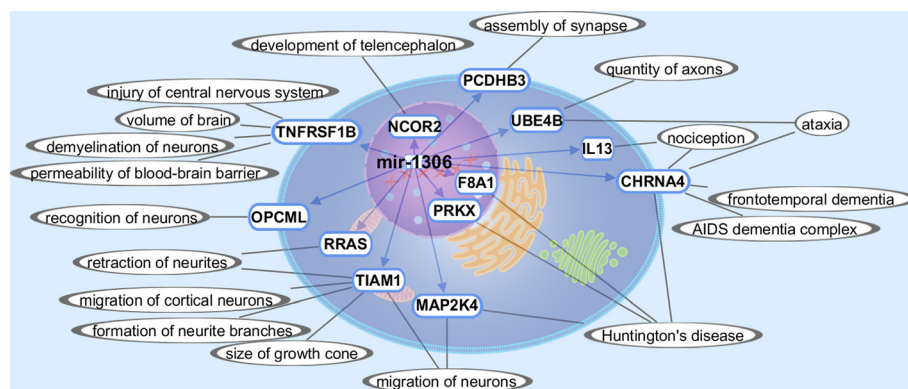


Figure 3 Target gene predictions for miR-1306. Twelve target genes of *miR-1306* were predicted by TargetScan concerning their function in AD, brain, nervous system or other neurodegenerative diseases and graphically interrelated by Path Designer (Ingenuity). The miRNA is located in the nucleus and the predicted target genes indicated by blue arrows and blue framed ellipses are located according to their Gene Ontology either in the nucleus, cytoplasm or membrane. The functions of the target genes are denoted by grey lines and grey framed ellipses and derived from Gene Ontology or Ingenuity Expert Findings, which are substantiated by literature.

8 (DGCR8), which is essential for miRNA biogenesis by being a subunit of the microprocessor complex [40]. Evers et al. presents a case of a DiGeorge syndrome patient with the typical deletion in chromosome band 22q11.2, which contains DGCR8, suffering from dementia [41].

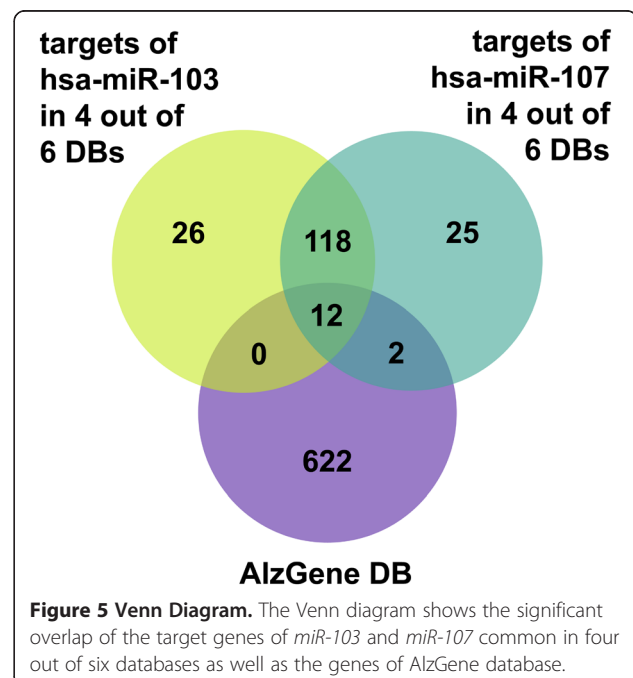
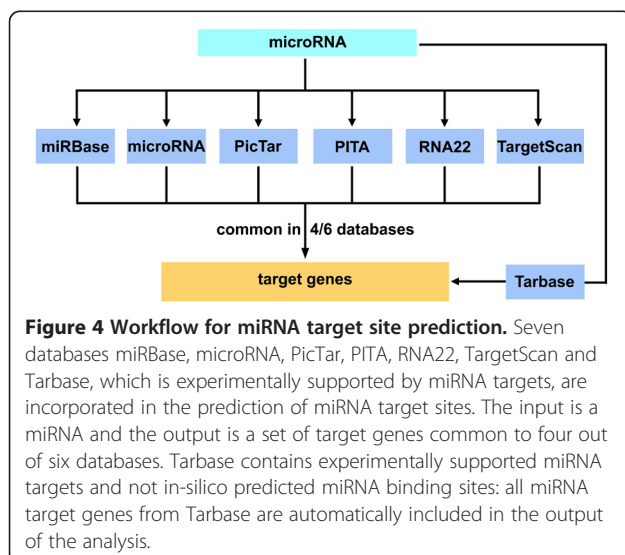
Prediction of miRNA target genes and their relation to AD

To confirm the relationship of the three miRNAs to AD we searched for target gene predictions of these miRNAs in the six databases miRBase, microRNA, PicTar, PITA, RNA22, as well as TargetScan (Figure 4). The combination of different miRNA target site prediction databases and the restriction of the output to four out of six databases in the case of *miR-103* and *miR-107* are important to reduce the amount of false positive miRNA target sites in the end. In the case of *miR-1306* there is no restriction of the output, because only the database PITA incorporates predictions for this miRNA. Additionally, Tarbase can be included in the analysis. The database Tarbase has a special position, because in contrast to the other six databases Tarbase contains experimentally supported miRNA targets and not in-silico predicted miRNA binding sites. All miRNA target sites from Tarbase are automatically included in the output of the analysis. In contrast to the other six databases Tarbase doesn't contain false positive miRNA target sites. In our case Tarbase doesn't contain target genes for the miRNAs: *miR-103*, *miR-107* and *miR-1306*. The six miRNA target site prediction databases miRBase, microRNA, PicTar, PITA, RNA22, and TargetScan contain altogether 18915 different (according to EntrezGene) human genes at which PITA alone contains 16819 genes. After the analysis we got 156 and 157 target genes for *miR-103* and *miR-107*, respectively, common in four out of six databases, and 890 target genes for *miR-1306* of database

PITA. As we focus on miRNAs playing a role in AD, we used the AlzGene database to see which target genes of the miRNAs have a genetic association with AD. AlzGene database is a regularly updated aggregation of all published genetic association studies including GWAS (genome-wide association studies) performed on AD phenotypes. 636 and 591 genes of AlzGene database overlap with 18915 genes of the six miRNA target site prediction databases and 16819 genes of database PITA, respectively. The overlap between AlzGene database genes and the target genes of *miR-103*, *miR-107* as well as *miR-1306* are 12, 14 (Figure 5) and 24 genes, respectively (Additional file 1). *MiR-103* and *miR-107* have 130 target genes in common (Figure 5, Additional file 1). Applying a Fisher's exact test we got a *p*-value of 0.0065, 0.0009 and 0.1904 for the overlap of *miR-103*, *miR-107* and *miR-1306*, respectively, with the AlzGene database, which shows that 12 and 14 are significant high numbers of overlapping genes between the target genes and the AlzGene database. This result suggests that *miR-103* and *miR-107* might play a role in AD. It is not remarkable that the *p*-value for the overlap of *miR-1306* with the AlzGene database is not significant as the restriction to four out of six databases was not possible, which consequentially leads to the inclusion of a lot of false positive target genes in the 890 target genes of *miR-1306*.

Gene ontology

Additionally, we did a Gene Ontology analysis with the predicted target genes of the three miRNAs to validate the functionality of the miRNAs and their involvement



to AD. With the help of the literature mining tool Pathway Studio we searched for molecular functions and biological processes common to the target genes of the three miRNAs. The molecular function calcium ion binding is significant in all three miRNA analyses (*miR-103*: p -value = 0.0011; *miR-107*: p -value = 0.0025; *miR-1306*: p -value = 2.1×10^{-5}) and is considered to be involved in AD. Calcium ions are found in an elevated level in tangle-bearing neurons of AD patients compared to healthier neurons [42]. Further, an abnormal increase of intracellular Calcium ion levels in neurites associated with A β deposits was demonstrated in a mouse model of AD [43]. An additional evidence is given by the significant biological processes learning (*miR-103*: p -value = 0.0008; *miR-107*: p -value = 3.3×10^{-5} ; *miR-1306*: p -value = 0.0003), brain development (*miR-103*: p -value = 0.0023; *miR-107*: p -value = 0.0004; *miR-1306*: p -value = 7.1×10^{-6}) and nervous system development (*miR-103*: p -value = 0.0004; *miR-107*: p -value = 0.0004; *miR-1306*: p -value = 6.9×10^{-8}) that the three miRNAs are involved in AD. Mouse models with an overexpression of *ADAM10* showed a positive effect of the α -secretase on learning and memory and mice with a dominant negative mutant form of *ADAM10* had learning deficiencies [12]. Environmental influences occurring during brain development predefine the expression and regulation of APP. As a consequence levels of APP and A β are increased causing AD later in life [44]. The A β fragments forming plaques are of varying length depending on the site of cleavage. The A β_{42} fragment is a ligand for the cellular prion protein, which is important for nervous system development [45]. (See Additional files 2, 3 and 4: List of enriched target genes of miR-103/107/1306 in Gene Ontology).

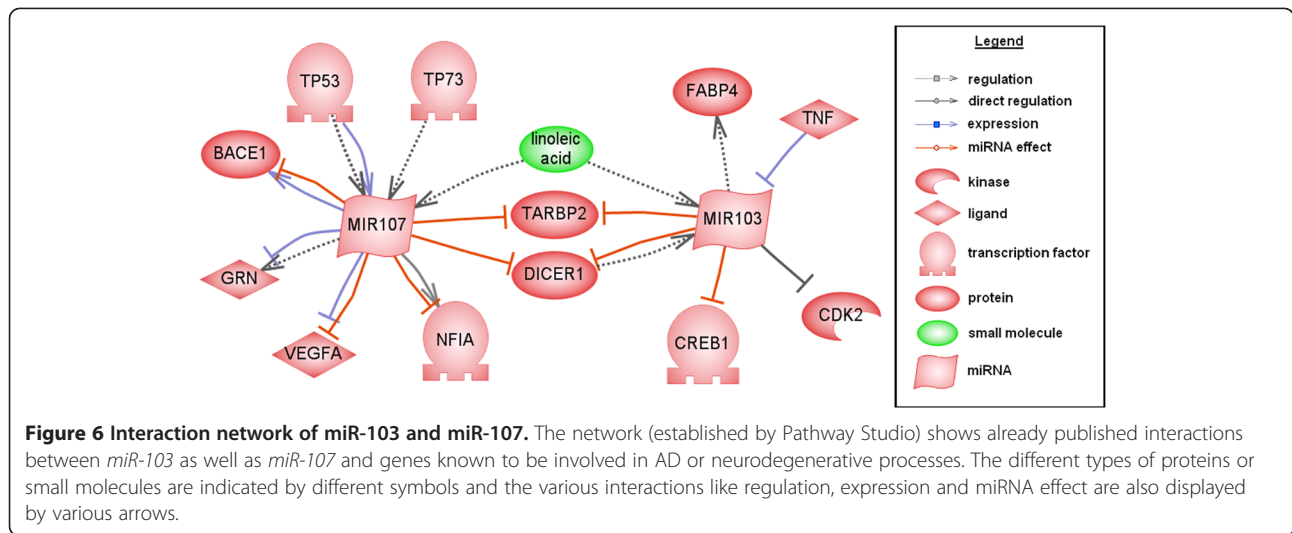
Literature mining

Furthermore, a network (Figure 6) containing already published interactions of *miR-103* and *miR-107* with genes involved in AD or included in the AlzGene database was established using the literature mining tool Pathway Studio. This network allowed confirming the relation of the two miRNAs to AD by their respective target genes. The genes *dicer* 1, ribonuclease type III (*Dicer*) and TAR (HIV-1) RNA binding protein 2 (*TARBP2*) targeted by *miRNA-103* and *miRNA-107* are components of the miRNA-processing complex [46]. Besides those two genes, a link between the two miRNAs is provided by linoleic acid [47], which probably affects AD by increasing the expression of Presenilin1 (*PSEN1*) and A β [48]. Another target gene in the network, Granulin (*GRN*), is regulated by *miR-107* [49] which regulates *BACE1* as well [38]. Therefore both genes might be involved in neurodegenerative diseases especially AD. The tumor suppressors *TP53* as well as *TP73* appear to

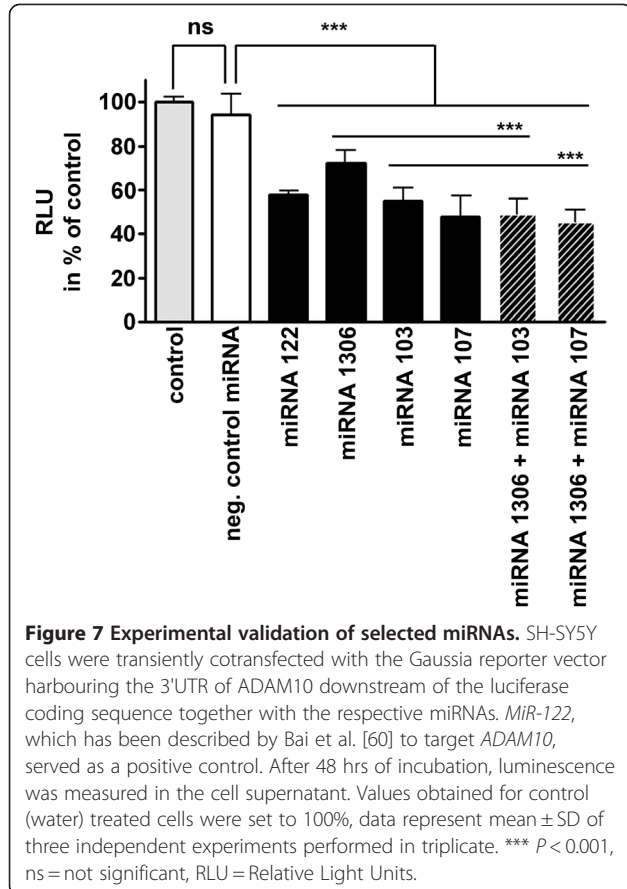
regulate the processing of *miR-107* [46]. *MiR-103* increases the expression level of fatty acid binding protein 4 (*FABP4*) while its expression is reduced by tumor necrosis factor (*TNF*) [50]. All six target genes of the miRNAs 103 and 107 *GRN*, *BACE1*, *TP53*, *TP73*, *FABP4* and *TNF* are included in the AlzGene database, hence putatively playing a role in AD. The four remaining target genes cyclin-dependent kinase 2 (*CDK2*), cAMP responsive element binding protein 1 (*CREB1*), nuclear factor I/A (*NFIA*) and vascular endothelial growth factor A (*VEGFA*) of the network are mentioned in literature to be involved directly or in processes developing AD. *miR-103* directly binds and represses *CDK2* and *CREB1* through 3'UTR binding [51]. *CDK2* is a key regulator in neuronal differentiation with the downregulation of *CDK2* as crucial event [52] and neuronal differentiation is regulated by *PSEN1*, a major key gene of AD [53]. The transcription factor *CREB1* is involved in several types of learning and memory. A direct involvement in AD is seen in some mouse models, where its activity is impaired [54]. *NFIA* is negatively regulated by *miR-107* [55] and plays an important role in the formation of the corpus callosum in the developing brain. The disruption of *NFIA* results in agenesis of the corpus callosum [56], whereas the size of the corpus callosum is significantly reduced in AD patients [57]. The expression of the hypoxia-regulated gene *VEGFA* is decreased by *miR-107* [58]. Additionally, it is known, that polymorphisms within the *VEGFA* promoter region are associated with increased risk for AD, by reducing the neuroprotective effect of *VEGFA* [59]. These findings of the literature and AlzGene database confirm the biological role of the target genes in neurodegenerative processes and hence the involvement of *miR-103* and *miR-107* in AD.

Experimental validation of bioinformatically predicted miRNAs

To demonstrate that the selected miRNAs 1306, 103 and 107 directly regulate *ADAM10* expression by interaction with the 3'UTR of the human gene, we performed cotransfection experiments with a Gaussia reporter vector harbouring the 3'UTR of *ADAM10* downstream of the luciferase coding sequence together with the respective miRNAs. As a positive control we used *miR-122* which has been identified and validated as an important regulator of *ADAM10* in hepatocellular carcinoma by an experimental approach [60]. The programs RNA22, RNAhybrid and miRanda predicted a miR-122 binding site to human *ADAM10* 3'UTR with binding energy -31.2, -25.6 and -21.84 kcal/mol (on average: -26.21 kcal/mol), respectively, comparable to the miR-1306 binding site prediction (see Results and Discussion: "miRNA prediction"). Time resolved measurement revealed that 48 hrs incubation period resulted



in maximal effects of the miRNAs in SH-SY5Y cells (data not shown): while the negative control miRNA had no impact on luciferase activity measured in the cell supernatant, *miR-122* reduced the luminescent signal to 57% as compared to water treated control cells (Figure 7). This effect is even higher than the one observed in the initial publication from Bai et al. (2009)



[60] but might be due to the different reporter enzymes or cell lines used. The three miRNAs identified by bioinformatical approaches and integration of literature mining all showed a significant decreasing effect on the *ADAM10* 3'UTR-reporter construct: *miR-1306* lowered the luminescent signal to 72%, *miR-103* to 55% and *miR-107* to 48% of control. While *miR-103* and *miR-107* target the same DNA sequence within the *ADAM10* 3'UTR (see Figure 2), *miR-1306* has a binding site in closer proximity to the Stop codon. We therefore combined *miR-1306* and *miR-103* or *miR-107*, respectively, but observed no distinct significant synergistic effect (*miR-1306* vs. *miR-1306/103* $p < 0.001$; *miR-1306* vs. *miR-1306/107* $p < 0.001$). This might be due to the applied concentration which was reduced to 50% in the combinations (see methods "3'UTR luciferase reporter assay"). These experimental results suggest an influence of *miR-103*, *miR-107* and *miR-1306* on *ADAM10* expression. Nevertheless, the biological impact of either miRNA has to be elucidated further, e.g. by mRNA and protein measurements. Assessing the effect of the selected miRNAs on pathological features in AD mouse models would also help to understand their distinct role in pathogenesis. However, this study shows that the computational workflow consisting of prediction programs and specific selection criteria is a suitable tool for the identification of miRNAs influencing key genes of diseases such as Alzheimer's disease.

Conclusions

We established a computational approach for the identification of miRNAs putatively influencing the expression of *ADAM10*. A potential functionality of selected miRNAs 103, 107 and 1306 was confirmed by 3'UTR luciferase reporter assay. These results show that the evolutionary conservation of the target gene binding site

facilitates miRNA candidate selection independently from the disease for further experimental validation. Moreover, these experiments underline the reliability of our computational approach, which is a combination of characteristics of the prediction software and specific selection criteria for filtering out false positive predictions: disease relevance, specificity of expression, evolutionary conservation of binding sites and occurrence of multiple binding sites. This workflow can also be applied to key genes of other diseases with adjustment of the selection criteria according to the scientific research interest. Our approach provides a new selection tool for identification and ranking of AD-related miRNAs, but to elucidate a profound pathological role of selected candidates, further experiments have to be done.

Additional files

Additional file 1: List of predicted target genes common in 4 out of 6 DBs. The table shows a list of predicted target genes of miR-103, miR-107, miR-1306. These target genes are either listed in AlzGene DB and target gene of at least one miRNA or target gene of at least two miRNAs (column 3). Beside the gene symbol the Entrez GeneID is given.

Additional file 2: List of enriched target genes of miR-103 in Gene Ontology. The table shows the Gene Ontology entities with enrichment of predicted target genes of miR-103. Additionally a *p*-value and FDR value is given.

Additional file 3: List of enriched target genes of miR-107 in Gene Ontology. The table shows the Gene Ontology entities with enrichment of predicted target genes of miR-107. Additionally a *p*-value and FDR value is given.

Additional file 4: List of enriched target genes of miR-1306 in Gene Ontology. The table shows the Gene Ontology entities with enrichment of predicted target genes of miR-1306. Additionally a *p*-value and FDR value is given.

Abbreviations

Chr: Chromosome; Human: Homo sapiens; Mouse: Mus musculus; Horse: Equus caballus; Dog: Canis lupus familiaris; Chimp: Pan troglodytes; Chicken: Gallus gallus; Rhesus monkey: Macaca mulatta; Zebra fish: Danio rerio; Opossum: Monodelphis domestica; Zebra finch: Taeniopygia guttata.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

RA performed the bioinformatic analyses and wrote the manuscript. RA and DT designed the computational workflow. KE designed the experimental system. SR performed transfection and luciferase reporter assay experiments. SFL and PHK contributed to the planning of the whole project. JH participated in the conception of this work. WW directed the work. DT planned and coordinated the entire project. All authors read and approved the final manuscript.

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References

1. Chekulaeva M, Filipowicz W: Mechanisms of miRNA-mediated post-transcriptional regulation in animal cells. *Curr Opin Cell Biol* 2009, **21**:452–460.
2. Vasudevan S, Steitz JA: AU-rich-element-mediated upregulation of translation by FXR1 and Argonaute 2. *Cell* 2007, **128**:1105–1118.
3. Friedman RC, Farh KK-H, Burge CB, Bartel DP: Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res* 2009, **19**:92–105.
4. Fabian MR, Sonenberg N, Filipowicz W: Regulation of mRNA translation and stability by microRNAs. *Annu Rev Biochem* 2010, **79**:351–379.
5. Hébert SS, De Strooper B: Alterations of the microRNA network cause neurodegenerative disease. *Trends Neurosci* 2009, **32**:199–206.
6. Satoh J: MicroRNAs and their therapeutic potential for human diseases: aberrant microRNA expression in Alzheimer's disease brains. *J Pharmacol Sci* 2010, **114**:269–275.
7. Crews L, Rockenstein E, Masliah E: APP transgenic modeling of Alzheimer's disease: mechanisms of neurodegeneration and aberrant neurogenesis. *Brain Struct Funct* 2010, **214**:111–126.
8. Cole SL, Vassar R: The role of amyloid precursor protein processing by BACE1, the beta-secretase, in Alzheimer disease pathophysiology. *J Biol Chem* 2008, **283**:29621–29625.
9. Lamlich S, Kojro E, Postina R, Gilbert S, Pfeiffer R, Jasionowski M, Haass C, Fahrenholz F: Constitutive and regulated alpha-secretase cleavage of Alzheimer's amyloid precursor protein by a disintegrin metalloprotease. *Proc Natl Acad Sci* 1999, **96**:3922–3927.
10. Kuhn PH, Wang H, Dislich B, Colombo A, Zeitschel U, Ellwart JW, Kremmer E, Rossner S, Lichtenthaler SF: ADAM10 is the physiologically relevant, constitutive alpha-secretase of the amyloid precursor protein in primary neurons. *EMBO J* 2010, **29**:3020–3032.
11. Lichtenthaler SF, Haass C, Steiner H: Regulated intramembrane proteolysis – lessons from amyloid precursor protein processing. *J Neurochem* 2011, **117**:779–796.
12. Postina R, Schroeder A, Dewachter I, Bohl J, Schmitt U, Kojro E, Prinzen C, Endres K, Hiemke C, Blessing M, Flamez P, Dequenne A, Godaux E, van Leuven F, Fahrenholz F: A disintegrin-metalloproteinase prevents amyloid plaque formation and hippocampal defects in an Alzheimer disease mouse model. *J Clin Invest* 2004, **113**:1456–1464.
13. Watanabe Y, Tomita M, Kanai A: Computational methods for microRNA target prediction. *Methods Enzymol* 2007, **427**:65–86.
14. Lewis BP, Burge CB, Bartel DP: Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005, **120**:15–20.
15. Griffiths-Jones S, Saini HK, van Dongen S, Enright AJ: miRBase: tools for microRNA genomics. *Nucleic Acids Res* 2008, **36**:D154–D158.
16. Betel D, Wilson M, Gabow A, Marks DS, Sander C: The microRNA.org resource: targets and expression. *Nucleic Acids Res* 2008, **36**:D149–D153.
17. Krek A, Grün D, Poy MN, Wolf R, Rosenberg L, Epstein EJ, MacMenamin P, da Piedade I, Günsal KC, Stoffel M, Rajewsky N: Combinatorial microRNA target predictions. *Nat Genet* 2005, **37**:495–500.
18. Kertesz M, Iovino N, Unnerstall U, Gaul U, Segal E: The role of site accessibility in microRNA target recognition. *Nat Genet* 2007, **39**:1278–1284.
19. Miranda KC, Huynh T, Tay Y, Ang Y-S, Tam W-L, Thomson AM, Lim B, Rigoutsos I: A pattern-based method for the identification of microRNA binding sites and their corresponding heteroduplexes. *Cell* 2006, **126**:1203–1217.
20. Papadopoulos GL, Reczko M, Simossis VA, Sethupathy P, Hatzigeorgiou AG: The database of experimentally supported targets: a functional update of TarBase. *Nucleic Acids Res* 2009, **37**:D155–D158.

21. Witkos TM, Koscianska E, Krzyzosiak WJ: **Practical Aspects of microRNA Target Prediction.** *Curr Mol Med* 2011, **11**:93–109.
22. Zuker M, Stiegler P: **Optimal computer folding of large RNA sequences using thermodynamics and auxiliary information.** *Nucleic Acids Res* 1981, **9**:133–148.
23. Rehmsmeier M, Steffen P, Hochsmann M, Giegerich R: **Fast and effective prediction of microRNA/target duplexes.** *RNA* 2004, **10**:1507–1517.
24. Krüger J, Rehmsmeier M: **RNAhybrid: microRNA target prediction easy, fast and flexible.** *Nucleic Acids Res* 2006, **34**:W451–W454.
25. Wuchty S, Fontana W, Hofacker IL, Schuster P: **Complete suboptimal folding of RNA and the stability of secondary structures.** *Biopolymers* 1999, **49**:145–165.
26. Enright AJ, John B, Gaul U, Tuschl T, Sander C, Marks DS: **MicroRNA targets in *Drosophila*.** *Genome Biol* 2003, **5**:R1.
27. Cogswell JP, Ward J, Taylor IA, Waters M, Shi Y, Cannon B, Kelnar K, Kempainen J, Brown D, Chen C, Prinjha RK, Richardson JC, Saunders AM, Roses AD, Richards CA: **Identification of miRNA changes in Alzheimer's disease brain and CSF yields putative biomarkers and insights into disease pathways.** *J Alzheimers Dis* 2008, **14**:27–41.
28. Bult CJ, Eppig JT, Kadin JA, Richardson JE, Blake JA, Mouse Genome Database Group: **The Mouse Genome Database (MGD): mouse biology and model systems.** *Nucleic Acids Res* 2008, **36**:D724–D728.
29. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG: **Clustal W and Clustal X version 2.0.** *Bioinformatics* 2007, **23**:2947–2948.
30. Goujon M, McWilliam H, Li W, Valentin F, Squizzato S, Paern J, Lopez R: **A new bioinformatics analysis tools framework at EMBL-EBI.** *Nucleic Acids Res* 2010, **38**:W695–W699.
31. Bertram L, McQueen MB, Mullin K, Blacker D, Tanzi RE: **Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database.** *Nat Genet* 2007, **39**:17–23.
32. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, Sherlock G: **Gene Ontology: tool for the unification of biology.** *Nat Genet* 2000, **25**:25–29.
33. Sethupathy P, Megraw M, Hatzigeorgiou AG: **A guide through present computational approaches for the identification of mammalian microRNA targets.** *Nat Meth* 2006, **3**:881–886.
34. Lu M, Zhang Q, Deng M, Miao J, Guo Y, Gao W, Cui Q: **An analysis of human microRNA and disease associations.** *PLoS One* 2008, **3**:e3420.
35. Noren Hooten N, Abdelmohsen K, Gorospe M, Ejiogu N, Zonderman AB, Evans MK: **microRNA expression patterns reveal differential expression of target genes with age.** *PLoS One* 2010, **5**:e10724.
36. Wang WX, Huang Q, Hu Y, Stromberg AJ, Nelson PT: **Patterns of microRNA expression in normal and early Alzheimer's disease human temporal cortex: white matter versus gray matter.** *Acta Neuropathol* 2011, **121**:193–205.
37. Yao J, Hennessey T, Flynt A, Lai E, Beal MF, Lin MT: **MicroRNA-related cofilin abnormality in Alzheimer's disease.** *PLoS One* 2010, **5**:e15546.
38. Wang WX, Rajeev BW, Stromberg AJ, Ren N, Tang G, Huang Q, Rigoutsos I, Nelson PT: **The expression of microRNA miR-107 decreases early in Alzheimer's disease and may accelerate disease progression through regulation of beta-site amyloid precursor protein-cleaving enzyme 1.** *J Neurosci* 2008, **28**:1213–1223.
39. Augustin R, Lichtenthaler SF, Greeff M, Hansen J, Wurst W, Trümbach D: **Bioinformatics identification of modules of transcription factor binding sites in Alzheimer's disease-related genes by in silico promoter analysis and microarrays.** *International Journal of Alzheimer's Disease* 2011, **2011**:154325.
40. Wang Y, Medvid R, Melton C, Jaenisch R, Blleloch R: **DGCR8 is essential for microRNA biogenesis and silencing of embryonic stem cell self-renewal.** *Nat Genet* 2007, **39**:380–385.
41. Evers LJ, Vermaak MP, Engelen JJ, Curfs LM: **The velocardiofacial syndrome in older age: dementia and autistic features.** *Genet Couns* 2006, **17**:333–340.
42. Nixon RA, Saito KI, Grynspan F, Griffin WR, Katayama S, Honda T, Mohan PS, Shea TB, Beermann M: **Calcium-activated neutral proteinase (calpain) system in aging and Alzheimer's disease.** *Ann N Y Acad Sci* 1994, **747**:77–91.
43. Mattson MP: **ER calcium and Alzheimer's disease: in a state of flux.** *Sci Signal* 2010, **3**:pe10.
44. Zawia NH, Lahiri DK, Cardozo-Pelaez F: **Epigenetics, oxidative stress, and Alzheimer disease.** *Free Radic Biol Med* 2009, **46**:1241–1249.
45. Kim D, Tsai LH: **Bridging physiology and pathology in AD.** *Cell* 2009, **137**:997–1000.
46. Boominathan L: **The tumor suppressors p53, p63, and p73 are regulators of microRNA processing complex.** *PLoS One* 2010, **5**:e10615.
47. Parra P, Serra F, Palou A: **Expression of adipose microRNAs is sensitive to dietary conjugated linoleic acid treatment in mice.** *PLoS One* 2010, **5**:e13005.
48. Liu Y, Yang L, Conde-Knape K, Beher D, Shearman MS, Shachter NS: **Fatty acids increase presenilin-1 levels and gamma-secretase activity in PSwt-1 cells.** *J Lipid Res* 2004, **45**:2368–2376.
49. Wang WX, Wilfred BR, Madathil SK, Tang G, Hu Y, Dimayuga J, Stromberg AJ, Huang Q, Saatman KE, Nelson PT: **miR-107 regulates granulin/progranulin with implications for traumatic brain injury and neurodegenerative disease.** *Am J Pathol* 2010, **177**:334–345.
50. Xie H, Lim B, Lodish HF: **MicroRNAs induced during adipogenesis that accelerate fat cell development are downregulated in obesity.** *Diabetes* 2009, **58**:1050–1057.
51. Liao Y, Lönnnerdal B: **Global microRNA characterization reveals that miR-103 is involved in IGF-1 stimulated mouse intestinal cell proliferation.** *PLoS One* 2010, **5**:e12976.
52. Dobashi Y, Kudoh T, Matsumine A, Toyoshima K, Akiyama T: **Constitutive overexpression of CDK2 inhibits neuronal differentiation of rat pheochromocytoma PC12 cells.** *J Biol Chem* 1995, **270**:23031–23037.
53. Wines-Samuelson M, Handler M, Shen J: **Role of presenilin-1 in cortical lamination and survival of Cajal-Retzius neurons.** *Dev Biol* 2005, **277**:332–346.
54. Puzzo D, Vitolo O, Trinchese F, Jacob JP, Palmeri A, Arancio O: **Amyloid-beta peptide inhibits activation of the nitric oxide/cGMP/cAMP-responsive element-binding protein pathway during hippocampal synaptic plasticity.** *J Neurosci* 2005, **25**:6887–6897.
55. Garzon R, Pichiorri F, Palumbo T, Visentini M, Aqeilan R, Cimmino A, Wang H, Sun H, Volinia S, Alder H, Calin GA, Liu CG, Andreeff M, Croce CM: **MicroRNA gene expression during retinoic acid-induced differentiation of human acute promyelocytic leukemia.** *Oncogene* 2007, **26**:4148–4157.
56. Das Neves L, Duchala CS, Godinho F, Haxhiu MA, Colmenares C, Macklin WB, Campbell CE, Butz KG, Gronostajski RM: **Disruption of the murine nuclear factor I-A gene (Nfia) results in perinatal lethality, hydrocephalus, and agenesis of the corpus callosum.** *Proc Natl Acad Sci* 1999, **96**:11946–11951.
57. Teipel SJ, Bayer W, Alexander GE, Zebuhr Y, Teichberg D, Kulic L, Schapiro MB, Moller H-J, Rapoport SI, Hampel H: **Progression of corpus callosum atrophy in Alzheimer disease.** *Arch Neurol* 2002, **59**:243–248.
58. Yamakuchi M, Lotterman CD, Bao C, Hruban RH, Karim B, Mendell JT, Huso D, Lowenstein CJ: **P53-induced microRNA-107 inhibits HIF-1 and tumor angiogenesis.** *Proc Natl Acad Sci* 2010, **107**:6334–6339.
59. Del Bo R, Scarlato M, Ghezzi S, Martinelli Boneschi F, Fenoglio C, Galbiati S, Virgilio R, Galimberti D, Galimberti G, Crimi M, Ferrarese C, Scarpini E, Bresolin N, Comi GP: **Vascular endothelial growth factor gene variability is associated with increased risk for AD.** *Ann Neurol* 2005, **57**:373–380.
60. Bai S, Nasser MW, Wang B, Hsu SH, Datta J, Kutay H, Yadav A, Nuovo G, Kumar P, Ghoshal K: **MicroRNA-122 inhibits tumorigenic properties of hepatocellular carcinoma cells and sensitizes these cells to sorafenib.** *J Biol Chem* 2009, **284**:32015–32027.

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