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Vascular effects of prostacyclin: does activation of PPAR δ play a role?

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Abstract

Prostacyclin (PGI₂) is a potent vasodilator that exerts multiple vasoprotective effects in the cardiovascular system. The effects of PGI₂ are mediated by activation of the cell membrane G-protein coupled PGI₂-receptor (IP receptor). More recently, however, it has been suggested that PGI₂ might also serve as an endogenous ligand and activator of nuclear peroxisome proliferator-activated receptor- δ (PPAR δ). Consistent with this concept, studies designed to define pharmacological properties of stable PGI₂ analogues revealed that beneficial effects of these compounds appear to be mediated, in part, by activation of PPAR δ . This review discusses emerging evidence regarding contribution of PPAR δ activation to vasoprotective and regenerative functions of PGI₂ and stable analogues of PGI₂.

PGI₂

Pharmacology of PGI₂ has been extensively studied since 1976, when PGI₂ was identified as a major product of arachidonic acid metabolism within the vasculature (1). In the cardiovascular system, PGI₂ is a potent vasodilator, and in some vascular beds, it functions as an endothelium-derived relaxing factor (2). In addition, PGI₂ exerts an inhibitory effect on aggregation of platelets (1,2). PGI₂ also inhibits white blood cells adhesion and proliferation of smooth muscle cells, thus preventing the development of atherosclerosis (3,4). Beneficial vascular effects of PGI₂ produced in the endothelium are in many respects similar to the effects of endothelium-derived nitric oxide (NO). Indeed, PGI₂ may compensate for the loss of NO thereby preserving endothelial function in conditions associated with impairment of NO signaling (5). It has also been recognized that biosynthesis of PGI₂ is an important determinant of regenerative functions in the cardiovascular system including angiogenesis and repair of injured endothelium (6–9). In this regard, accumulating evidence continues to substantiate an important contribution of PGI₂ to the regenerative capacity of endothelial progenitor cells (EPCs; 9–12).

PGI₂ is generated by cytosolic phospholipase A₂- α (cPLA₂- α)-induced mobilization of arachidonic acid which serves as a substrate for cyclooxygenase (COX) enzyme activity. Two isoforms, COX-1 and COX-2, are coupled to prostacyclin synthase (PGIS), enzyme responsible for synthesis of PGI₂. Interestingly, in vascular endothelial cells, enzymes required for production of PGI₂ are localized at endoplasmic reticulum and around the

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nuclear membrane (13–15), thereby suggesting that locally generated PGI₂ might be involved in nuclear signaling. Notably, COX-1, COX-2, and PGIS have been detected on both inner and outer surfaces of the nuclear membrane (14). An increase in intracellular calcium is required for activation of cPLA₂. Moreover, elevated intracellular calcium also causes translocation of cPLA₂-α to the nuclear envelope, thus enabling mobilization of arachidonic acid and subsequent production of PGI₂ in close proximity to nucleus (16; Figure 1). Consistent with this concept, colocalization of translocated cPLA₂-α with COX-2 has been demonstrated in perinuclear region of endothelial cells (17). Recognition of ability of PGI₂ (and stable analogues of PGI₂) to activate nuclear PPARδ provided functional explanation for the nuclear localization of enzymes responsible for production of PGI₂ (18,19). However, in the cardiovascular system, ongoing efforts are only beginning to define the stimuli (e.g. shear stress) that can activate the PGI₂ nuclear signaling pathway under physiological or pathological conditions (20). The exact nature of the agonists and the physical stimuli that can activate and translocate cPLA₂ to couple perinuclear generation of PGI₂ with activation of nuclear receptor(s) remains to be determined. Therefore, in this review we will discuss existing evidence regarding pharmacological relevance of PPARδ activation in mediation of vascular effects of PGI₂ and stable analogues of PGI₂. We will also address implications of this signaling pathway for understanding of the vascular effects of clinically relevant drugs that may affect production of PGI₂.

Vascular protective effects of PGI₂

In mice, genetic inactivation of PGIS causes significant alterations in vascular wall architecture including thickening of arterial media attended by increased arterial blood pressure (21). Moreover, phenotypic changes in the cardiovascular system substantially progress as PGIS null mice become older. These findings suggest that, in the cardiovascular system, loss of PGI₂ exacerbates phenotypic characteristics of aging.

The vasoprotective function of PGI₂ was also corroborated by studies of IP-deficient mice. Under basal conditions, IP-deficient mice had normal blood pressure and did not suffer from spontaneous thrombosis (22,23). However, deletion of IP receptors predisposes mice to an exaggerated response to thrombotic stimuli, accelerated atherosclerosis, and augmented intimal hyperplasia after vascular injury (22,23). Moreover, hypertensive response to a high-salt diet is exacerbated in IP-deficient mice (24). These findings indicate that production of PGI₂ and activation of IP receptors are essential mechanisms responsible for preservation of normal vascular function. Additional support for vasoprotective function of PGI₂ is provided by the results of gene delivery studies demonstrating that elevation of PGI₂ biosynthesis by expression of recombinant COX-1 and PGIS attenuates aberrant remodeling of arterial wall after injury, exerts beneficial effect on pulmonary hypertension, and prevents arterial thrombosis in ischemic cerebrovascular disease (25–28). Traditionally, protective vascular effects of PGI₂ have been ascribed to the activation of IP receptors located on cell membrane. However, more recently, it has been recognized that activation of PPARδ represents a previously unappreciated nuclear signaling mechanism that might contribute to vasoprotective and regenerative effects of PGI₂.

Vascular protective effects of PPARδ

PPARδ is a nuclear receptor that functions as a ligand-activated transcription factor. Ligand-induced activation of PPARδ causes formation of heterodimers with a retinoid X receptor and subsequent binding to specific PPAR responsive elements in promoter regions of target genes (29,30; Figure 1). PPARδ is ubiquitously expressed in various tissues, including vascular endothelial and smooth muscle cells (7). Vascular effects of PPARδ activation involve: a) direct effects induced by activation of PPARδ in blood vessel wall, and b) indirect effects mediated by systemic alterations in glucose, lipid, and lipoprotein

metabolism (31). PPAR δ agonists exert anti-atherosclerotic effects in murine models of hypercholesterolemia (32,33). In the vascular endothelium, major direct effects of PPAR δ activation are anti-inflammatory effects, prevention of apoptosis, and stimulation of angiogenesis (32–38). In smooth muscle cells, PPAR δ is responsible for inhibition of migration, and neointimal formation however controversial results have been reported regarding the effect on smooth muscle cells proliferation (39,40). Moreover, activation of PPAR δ in smooth muscle cells exerts anti-inflammatory effects (41) and attenuates apoptotic cell death induced by oxidized low-density lipoprotein (42). PGI $_2$ released by shear stress from endothelium may activate PPAR δ in smooth muscle cells thereby promoting conversion of synthetic to contractile phenotype (20). Generally, the effects on vascular wall mediated by activation of PPAR δ are considered beneficial, with the exception of angiogenesis, which under certain conditions might contribute to tumor growth (31–45).

Preclinical studies in insulin-resistant primates have demonstrated that a PPAR δ agonist, GW501516, decreases plasma triglyceride, LDL cholesterol, and increases levels of HDL cholesterol (46). Based on these observations, clinical development of PPAR δ agonists has been mostly directed towards treatment of dyslipidemia (47–50). Prevention of dyslipidemia and the beneficial effects of PPAR δ activation on insulin resistance and glucose homeostasis protect vascular endothelium from well-established risk factors responsible for initiation of endothelial dysfunction and progression of vascular disease, including atherosclerosis. In this regard, it is interesting to note that recent findings demonstrate that stable analogue of PGI $_2$, beraprost, exerts beneficial effects on metabolic syndrome and dyslipidemia (51,52). Currently, the exact mechanisms underlying these effects are poorly understood, but could be in part mediated by activation of nuclear receptor(s) including PPAR δ . Thus, both direct effects of PPAR δ activation on blood vessel wall and indirect effects mediated by normalization of metabolic functions are essential for protection of vascular wall (31). Interventions designed to increase production of endogenous PPAR δ ligands or treatment with selective PPAR δ agonists are currently being studied as novel therapeutic strategies. However, efforts in drug development designed to harness therapeutic effects of PPAR δ agonists have yet to produce a drug approved for specific therapeutic application in general population.

PGI $_2$: a putative endogenous ligand for PPAR δ

Vascular endothelium has the highest production of PGI $_2$ under basal conditions as well as during activation of arachidonic acid metabolism by vascular injury (22,53). Although smooth muscle cells and adventitia may generate PGI $_2$, the levels in these layers are lower as compared to endothelium. Consistent with role of PGI $_2$ in nuclear signaling, enzymes required for synthesis of PGI $_2$ in endothelium localize around the nucleus (Figure 1; 15). Prior studies have provided evidence that under *in vitro* conditions PPAR δ can be activated by several different products of arachidonic acid metabolism and stable analogues of PGI $_2$ (54,55). However, inability of exogenous PGI $_2$ itself to activate PPAR δ under some *in vitro* conditions (55) is most likely result of PGI $_2$ instability in neutral and acidic buffer. PGI $_2$ is quickly hydrolyzed (30–120 s) to a pharmacologically inactive product, 6-ketoPGF $_{1\alpha}$. Nevertheless, relevant to the concept of PGI $_2$ as an endogenous PPAR δ ligand in endothelium, several studies have demonstrated that stable analogues of PGI $_2$ [including carbaprostacyclin (cPGI $_2$) and iloprost] behave as PPAR δ ligands (18,55). The first evidence supporting the role of PGI $_2$ as an endogenous ligand for PPAR δ *in vivo* was reported in studies of mechanisms responsible for embryo implantation (18). These findings provided impetus for re-assessment of signaling mechanisms that may help to explain beneficial effects of PGI $_2$ in different tissues including cells of blood vessel wall. As mentioned above, studies with stable analogues of PGI $_2$ consistently demonstrated that these compounds activate PPAR δ , thereby providing support for the importance of endogenous

PGI₂ in activation of PPAR δ . In agreement with this concept, a more recent study presented the crystal structure of the PPAR δ ligand-binding domain bound to iloprost thus establishing evidence for direct interaction between iloprost with PPAR δ (56).

It appears that significantly higher concentrations of PGI₂ analogues are required for activation of nuclear PPAR δ receptor as compared to concentrations that are able to stimulate cell membrane IP receptors (57). Interestingly, cicaprost (stable PGI₂ analogue) is a highly selective IP agonist that does not activate PPAR δ even in very high (μ M) concentrations (9,18,55). This pharmacological property of cicaprost has been successfully employed in studies designed to dissect contribution of IP receptors and PPARs to the vascular effects of stable PGI₂ analogues (9,58). Although iloprost and cPGI₂ stimulated angiogenesis *in vivo*, cicaprost had no effect, suggesting that activation of PPARs might be the mechanism underlying the angiogenic effects of PGI₂ analogues (58).

We would also like to point out that although available evidence is consistent with the concept that PGI₂ signaling might depend in part on activation of PPAR δ , direct *in vivo* evidence supporting this hypothesis is currently missing. In this regard, future studies in IP-receptor-deficient mice may help to define the effects of endogenous PGI₂ dependent on activation of PPAR δ . Of note, reported lack of PPAR δ ligands effects on embryonic development in IP-receptor deficient mice (60), suggests that, in some cells, activation of PPAR δ might depend on intact function of IP receptors. Several prior studies demonstrated that drugs causing an increase in cyclic AMP or activation of protein kinase A (PKA) enhance basal and stimulated activity of PPAR δ (61–64). Because activation of IP receptors is coupled to increased production of cyclic AMP, it is possible that PGI₂-induced elevation of cyclic AMP might participate in regulation of activity of PPAR δ . Because complex mechanisms underlying PPAR δ signaling involve dissociation of co-repressor proteins and recruitment of co-activator proteins, the exact molecular targets affected by elevation of cyclic AMP or activation of PKA are currently unknown. Thus, additional studies are required to determine the mechanisms underlying interaction between IP and PPAR δ signaling in blood vessel walls.

Although the majority of studies suggest that PGI₂ is a ligand for PPAR δ , there are also reports demonstrating an ability of PGI₂ to activate PPAR α . Previous *in vitro* studies established that stable analogues of PGI₂, including iloprost and cPGI₂ (but not cicaprost), may also behave as ligands for PPAR α (55). Consistent with this concept, it has been demonstrated that PGI₂ released from endothelium by shear stress causes synthetic-to contractile phenotypic modulation in smooth muscle cells by activation of both PPAR α and PPAR δ (20). In addition, it has been suggested that the angiogenic effects of PGI₂ in some experimental conditions are dependent on activation of PPAR α (59). Relevant to this concept, crystal structures of PPAR α ligand domains bound to iloprost have been reported, thus providing structural basis for the recognition of PPAR α by iloprost (56). Thus, it appears that PGI₂ might influence vascular function by activation of PPAR α . The exact conditions and mechanisms responsible for activation of this signaling mechanism remain to be determined.

Angiogenesis

The importance of arachidonic acid metabolism via the COX pathway in angiogenesis was recognized more than 20 years ago (65,66). Initially, it was observed that pro-angiogenic proteins, including angiogenin and acidic and basic fibroblast growth factors (aFGF and bFGF), stimulate production of PGI₂ and that this effect is an important component of their ability to promote angiogenesis. These studies were followed by reports demonstrating that vascular endothelial growth factor (VEGF) is also a potent stimulator of PGI₂ production (6,67). By contrast, it was demonstrated that a stable PGI₂ analogue, SM-10902, promotes

wound healing by stimulation of angiogenesis (68). Further analysis supported the role of PGI₂ as possible ligand for nuclear receptors, leading to speculation that activation of PPAR δ in endothelial cells could be important for PGI₂ signaling (69). Moreover, in an *in vivo* study designed to examine contribution of nuclear receptors to the stimulatory effect of stable PGI₂ analogues on angiogenesis, the authors found that iloprost and cPGI₂ upregulated expression of VEGF mRNA and protein (58). Inactivation of VEGF signaling inhibited the pro-angiogenic effect of iloprost and cPGI₂. Based on prior studies demonstrating that iloprost and cPGI₂ are ligands for PPAR δ (and in some tissues for PPAR α), the authors proposed that activation of PPAR(s) is the most likely mechanism of the observed angiogenic effects (58). This concept was further extended and refined by studies in human EPCs (9). Since specific molecular markers for identification of EPCs have not been established, studies were performed on so-called “late outgrowth” EPCs. Obtained findings suggest that these cells possess intrinsically high production of PGI₂ (9). One possibility is that the angiogenic function of EPCs is dependent on PGI₂-induced activation of PPAR δ . Indeed, *in vitro* and *in vivo* analysis of angiogenic function of EPCs in response to endogenous PGI₂ or treatment with iloprost pointed to PPAR δ as a major molecular target (9). Further research revealed that the angiogenic effect of PPAR δ activation in human EPCs is dependent on increased production of tetrahydrobiopterin, an enzymatic cofactor (70). The regenerative effect of tetrahydrobiopterin was independent of endothelial nitric oxide synthase (eNOS) activation thus suggesting that tetrahydrobiopterin has effects on EPCs that could not be explained by its role as a cofactor for eNOS (70).

Existing evidence supports the concept that PPAR δ (unlike PPAR α and PPAR γ) is pro-angiogenic (38). Treatment of cultured human endothelial cells with GW501516, a selective PPAR δ agonist, stimulates proliferation and increases expression and production of VEGF (36). Consistent with *in vitro* observations, activation of PPAR δ *in vivo* also causes angiogenesis (36). Other target genes that might be responsible for PPAR δ -induced stimulation of angiogenesis have been identified including angiopoietin-like protein 4, calcineurin, cyclin-dependent kinase inhibitor 1c (Cdkn 1c), and Cl⁻ intracellular channel protein 4 (CLIC4; 38). However, PPAR δ activation has an inhibitory effect on some anti-angiogenic proteins including thrombospondin-1 and cellular retinol-binding protein-1 (CRBP1; 38). More recent studies indicate that PPAR δ activation enhances regenerative and angiogenic capacity of EPCs (9,70–72).

Efforts to harness potentially beneficial vascular (and metabolic) effects of PPAR δ agonists have been hampered by the tumorigenic effects of this signal transduction pathway. Although the exact role of PGI₂/PPAR δ signaling in pathogenesis in cancer remains to be determined (73,74), existing literature suggests that tumor angiogenesis could be stimulated by activation of PPAR δ (31, 43–45). This issue has to be approached with high degree of caution, and requires very careful monitoring of patients entering clinical trials designed to test therapeutic safety and efficacy of compounds that may activate PPAR δ .

Pulmonary hypertension

Stable analogues of PGI₂ are essential drugs used in therapy of pulmonary hypertension. It is generally accepted that the beneficial effects of these compounds are, for the most part, mediated by activation of IP receptors on vascular smooth muscle cells and the resulting decrease in pulmonary vascular resistance (75). In addition, these salutary effects are dependent on the ability of PGI₂ analogues to arrest and reverse pathological remodeling of pulmonary vascular wall. Emerging experimental evidence suggests that some of the long-term benefits could be mediated by activation of PPAR δ receptors. For instance, activation of PPAR δ in the lungs inhibits proliferation of fibroblasts and the subsequent aberrant vascular remodeling responsible for narrowing of pulmonary blood vessel diameter (57). In addition, more recent findings indicate that, in children with pulmonary hypertension, the

PGI₂ analogue treprostinil increases the number and angiogenic potential of EPCs (12), thereby stimulating endothelial repair and formation of new blood vessels. These observations may help to explain clinically observed long-term therapeutic effects of PGI₂ analogues that could not be attributed to pulmonary vasodilatation in patients resistant to the acute vasodilator effect of PGI₂ (75,76).

Adverse cardiovascular effects of COX inhibitors

Non-steroidal anti-inflammatory drugs (NSAIDs) exert their therapeutic effects by inhibiting COX isoforms. However, long-term use of NSAIDs as anti-inflammatory agents and analgesics is associated with adverse cardiovascular effects (77). Existing evidence suggests that older patients with cardiovascular risk factors are particularly prone to develop adverse effects including high blood pressure, edema, and congestive heart failure. Moreover, the detrimental effects of these drugs on cardiovascular function predispose treated patients to development of stroke and myocardial infarction (77). Although the mechanisms underlying adverse effects are quite complex and incompletely understood, inactivation of PGI₂ synthesis is considered a major mechanism contributing to development of adverse cardiovascular effects of COX inhibitors (78). In this regard, recognition of nuclear PPAR δ receptors as potentially important target for PGI₂ signaling may provide new insights. For example, emerging evidence suggests that loss of PGI₂/PPAR δ signaling causes apoptosis of endothelial cells (79) and impairment of regenerative capacity of EPCs (9,72). In addition, aging is associated reduced expression of PPAR δ (80). This may increase the vulnerability of an aging population to drugs that interfere with activation of PPAR δ . However, the exact contribution of diminishing PGI₂/PPAR δ signaling to adverse cardiovascular effects of COX inhibitors in humans is unknown and remains to be determined.

Concluding remarks

Existing evidence is consistent with the concept that stable analogues of PGI₂ might exert vascular effects in part by activation of PPAR δ . More recent findings suggest that this signaling pathway may also contribute to vascular regenerative functions of these compounds. However, direct *in vivo* evidence demonstrating ability of endogenous PGI₂ to behave as PPAR δ ligand in vascular tissue remains to be established. Further improvements in understanding of PGI₂/PPAR δ signaling in the human cardiovascular system will certainly help to optimize a number of therapeutic interventions dependent on modulation of arachidonic acid metabolism and subsequent activation of nuclear receptor(s). Future efforts in this area might also provide important new insights into molecular mechanisms underlying cardiovascular effects of widely used COX inhibitors.

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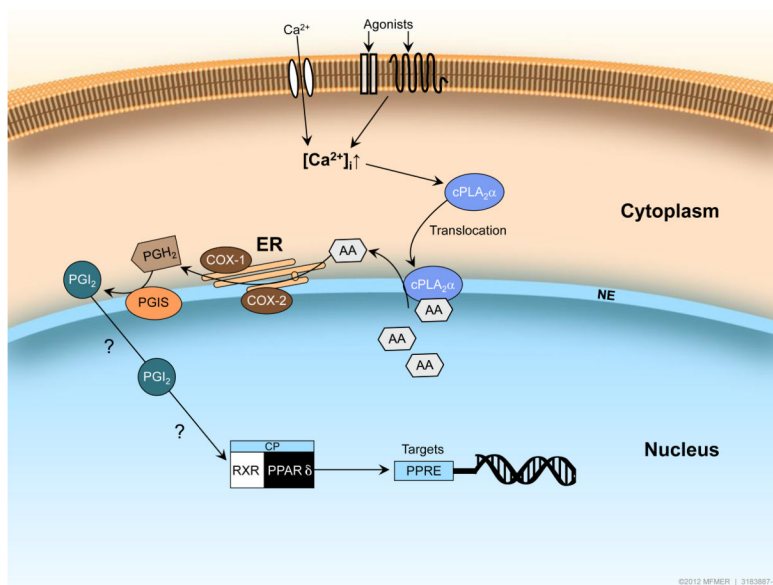


Figure 1. Hypothetical model of the cellular localization of enzymes responsible for PGI₂ synthesis and subsequent activation of PPARδ. Increases in intracellular Ca²⁺ levels cause activation and translocation of cPLA₂α from the cytoplasm to the nuclear envelope. (Complexity of cPLA₂α activation has been simplified to include only activation by increased Ca²⁺ levels). Mobilization of arachidonic acid from phospholipids by cPLA₂α provides substrate for enzyme activity of COX-1 or COX-2 and PGIS resulting in production of PGI₂ and subsequent activation of nuclear receptor PPARδ. PPARδ-RXR heterodimers undergo a conformational shift and this causes dismissal of the co-repressor complex in exchange for co-activator proteins thereby resulting in enhanced PPARδ target gene expression. Exact nature of agonists and physical stimuli responsible for PGI₂-induced activation of PPARδ remains to be determined (denoted by ? in the figure). cPLA₂=cytosolic phospholipase A₂; ER = endoplasmic reticulum; NE = nuclear envelope; AA = arachidonic acid; PGI₂ = prostacyclin; PGH₂ = prostaglandin H₂; PGIS = prostacyclin synthase; RXR = retinoid X receptor; CP = co-activator proteins; PPRE = PPARδ responsive element.