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Neurometabolites in schizophrenia and bipolar disorder – A systematic review and meta-analysis

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Abstract

This meta-analysis evaluates alterations of neurometabolites in schizophrenia and bipolar disorder. PubMed was searched to find controlled studies evaluating N-acetylaspartate (NAA), Choline (Cho) and Creatine (Cr) assessed with ¹H-MRS (proton magnetic resonance spectroscopy) in patients with schizophrenia and bipolar disorder up to September 2010. Random effects meta-analyses were conducted to estimate pooled standardized mean differences. I² statistic was used to quantify inconsistencies. Subgroup analyses were conducted to explore potential explanations for inconsistencies. 146 studies with 5643 participants were included in the systematic review. NAA levels were affected in schizophrenia and bipolar disorder. Decreased levels in the basal ganglia and frontal lobe were the most consistent findings in schizophrenia, decreased levels in the basal ganglia were the most consistent findings in bipolar disorder. Cho and Cr levels were not altered in either disorder. Findings for Cr were most consistent in the thalamus, frontal lobe and dorsolateral prefrontal cortex in schizophrenia and the basal ganglia and frontal lobe in bipolar disorder. Findings for Cho were most consistent in the thalamus, frontal lobe and anterior cingulate cortex in schizophrenia and basal ganglia in bipolar disorder. Large, carefully designed studies are needed to better estimate the extent of alterations in neurometabolites.

Keywords

Magnetic resonance spectroscopy (MRS); N-acetylaspartate; Creatine (Cr); Choline (Cho)

1. INTRODUCTION

Emil Kraepelin was the first to establish dementia praecox and manic-depressive insanity as dichotomous model, which has been utilized in conceptualization of schizophrenia and

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bipolar disorder ever since (Crow, 1990a; Heckers, 2008). However, emerging data suggest a relationship between schizophrenia and bipolar disorder, even a conceptualization of these disorders as polar ends of a continuum of the same mental illness is proposed (Crow, 1990b).

Family studies and twin studies have shown co-aggregation between schizophrenia and bipolar disorder. Shared genetic susceptibility has been reported in both candidate gene studies and whole-genome linkage analyses (Dalby et al. 1986; Bramon and Sham, 2001; Craddock and Owen, 2005; Lichtenstein et al., 2009; Purcell et al., 2009; Van Snellenberg and de Candia, 2009). A meta-analysis in magnetic resonance imaging (MRI) studies suggested gray matter reductions in paralimbic structures implicated in emotional processing in bipolar disorder, with a more extensive reduction in schizophrenia, not only affecting paralimbic structures, but also limbic and neocortical structures (Ellison-Wright and Bullmore, 2010).

Magnetic Resonance Spectroscopy (MRS) is increasingly being applied to characterize tissue-based chemical or metabolic abnormalities in psychiatric disorders. It is a non-invasive technique that measures chemical composition of tissues, energy metabolism, neurotransmitter levels, and neuronal integrity *in vivo*. It detects magnetic resonance signals produced by atomic nuclei located within molecules in living tissue. Quantification of MRS signal amplitude can provide an estimate for concentrations of signal generating molecules. (Kreis et al., 1993; Dager et al., 2008; Alger, 2010). The peak integral is proportional to the number of resonating nuclei. However, metabolite quantification is affected by high variability as the signal has low sensibility and multiple processing steps are performed. To take these variations in account, a reference signal is obtained. The reference signal is generally generated by a metabolite, water or a chemical compound in a phantom object. The reference signal often is classified as either “internal” or “external”, with internal meaning that the reference signal is generated by a metabolite or by water within the brain, and external meaning that the reference signal is generated by a phantom that is outside the brain. Resulting metabolite levels are then reported in two different ways. The first method is to report metabolite ratios, referring to Creatine (Cr) (e.g. NAA/Cr); the second method is to report absolute concentrations. These absolute concentrations are referenced to either brain water content or an external metabolic phantom with known metabolite concentrations and usually reported as mmol/l or institutional units (i.u.) (reviewed in Bagory et al., 2007).

¹H-MRS (proton magnetic resonance spectroscopy) is the most widely applied technique studying alterations of neurometabolites in psychiatric disorders. Metabolites measured with ¹H-MRS include N-acetyl aspartate (NAA), a metabolite that is thought to reflect neuronal integrity and is exclusively found in the brain; Cr, a putative marker of phosphate metabolism; Trimethylamines/ choline containing compounds (Cho), indicating breakdown of cell membranes and cellular turnover; and neurotransmitters such as Gamma-aminobutyric acid, glutamine, glutamate, with glutamate being the most abundant amino acid and excitatory neurotransmitter in the brain (Miller, 1991; Keshavan et al. 2000).

Abnormalities of neurometabolites in various regions of the brain have been implicated in the pathophysiology of both schizophrenia and bipolar disorder. Meta-analytic evidence in schizophrenia suggests that NAA may be reduced in the hippocampus and the frontal lobe (grey and white matter). Similar findings were reported in a systematic review performed in bipolar disorder, showing decreased NAA levels in euthymic bipolar patients in the hippocampus and frontal lobe as well. (Steen et al., 2005; Yildiz-Yesiloglu and Ankerst, 2006). Decrease of NAA is thought reflect neuronal or axonal loss or mitochondrial dysfunction (Meyerhoff et al., 1993; Sager et al., 2001), implying structural abnormalities on a molecular/neuronal level in both disorders. Cr has long thought to be a stable

neurometabolite, and has been widely used as internal reference in MRS studies. However, several studies found reduced Cr levels in the dorsolateral prefrontal cortex, hippocampus and basal ganglia in both bipolar disorder and schizophrenia (Deicken et al., 2003; Ohrmann et al. 2005; Frey et al., 2007; Ruesch et al., 2008). These reports not only suggest alterations in the cellular energy metabolism but also question the validity of using Cr as internal reference. Conflicting results are also found for Cho; levels have been reported to be decreased in some studies in the basal ganglia, hippocampus and DLPFC in schizophrenia while others report an increase (Maier et al., 1996; Stanley et al. 1996; Bustillo et al., 2002; Ohrmann et al., 2005; Bustillo et al., 2008; Ruesch et al., 2008). In bipolar disorder, different studies suggest an increase, decrease or no change in Cho levels in the DLPFC, increased and decreased levels in the hippocampus and increased or unchanged levels in the ACC (Deicken et al., 2003; Michael et al., 2003; Brambilla et al., 2005; Frye et al., 2007a; Iosifescu et al. 2009; Colla et al., 2009).

We sought to systematically review all controlled studies of brain metabolite levels measured by ^1H -MRS, to estimate the extent to which NAA, Cr, and Cho are altered in schizophrenia and bipolar disorder, to seek explanations (other than chance) by which studies in this field have yielded inconsistent results, and to examine if data supports the conception of a continuum or a dichotomy of bipolar disorder and schizophrenia.

2. METHODS

2.1 Eligibility Criteria

Eligible studies were clinical trials evaluating brain metabolites with ^1H -MRS in patients with schizophrenia, schizoaffective disorder, schizophreniform disorder, or bipolar disorder according to DSM-III, DSM-III-R, or DSM-IV criteria.

Studies published in languages other than English, reported neurometabolites other than NAA, Cho or Cr, did not include a healthy control group, postmortem studies and studies enrolling adolescent subjects (younger than age 18) or geriatric subjects (older than 65) were excluded.

2.2 Literature Search

NVK and MAR performed a literature search in PubMed for ^1H -MRS studies in schizophrenia and bipolar disorder for the timeframe up to September 2010 using the following key words: “Schizophrenia”, “Bipolar Disorder”, “Manic depressive disorder”, and “Magnetic Resonance Spectroscopy (MRS)”. The reference lists of included studies were inspected for additional eligible studies.

2.3 Study Selection

NVK and MAR reviewed titles and abstracts retrieved from the search and selected potentially eligible studies for full text review. Full text articles were then requested and assessed for eligibility. Figure 1 describes the outcomes at each level of our study identification process.

2.4. Author Contact

If relevant data were not reported in the article, we attempted to contact authors via e-mail to obtain this information. If no initial response was received, a second e-mail was sent two weeks later. If we did not receive a response then, the study was excluded from the meta-analysis. We contacted 76 authors for additional information. We received information from 13% of the authors. A total of 146 studies were included in the systematic review.

2.5. Data Extraction

We extracted the following data from each study: year of publication, number of participants (patients and healthy controls), illness duration (first episode vs. chronic illness), mood state (manic, depressed, euthymic), use of psychotropic medication (currently on medication vs. off medication vs. never treated and or minimally treated), MRS data acquisition parameters (MRSI vs. single-voxel (SV), field strength, location of voxel placement, use of a internal or external reference), absolute metabolite levels (mM or institutional units), and metabolite ratios.

2.6. Outcome Measures

Our primary outcome variables were absolute metabolite levels reported as mmol/l or i.u. (NAA, Cr, and Cho) as well as metabolite ratios (NAA/Cr and Cho/Cr) in the following regions of the brain: frontal lobe (given that the frontal lobe is a large and functionally complex region, we decided to analyze the ACC and DLPFC separately when authors specified that they studied these specific regions), parietal lobe, temporal lobe, occipital lobe, hippocampus, thalamus, basal ganglia, and cerebellum.

2.7. Statistical Analyses

2.7.1. Meta-analyses—We conducted meta-analyses for studies including subjects with bipolar disorder and schizophrenia meeting criteria as outlined above across all mentioned regions of the brain; however, not all regions had adequate number of studies to conduct a formal meta-analysis. If individual study results were reported separately for the left and right hemisphere, the left hemisphere data was included in the analysis, as it is the dominant hemisphere in most subjects. (Analyses were then re-run with right hemisphere data and compared with the initial analysis, given there was no significant difference between results of left and right hemisphere, this data is not shown).

To avoid double counting of the control group we only included subjects with chronic schizophrenia and not first episode psychosis when results were reported separately for both groups, but only one control group was reported. Chronic schizophrenia subjects were included because diagnostic stability likely is higher in this group.

To limit heterogeneity, eight studies that reported values (either in the original publication or via author contact) were excluded from analysis with absolute metabolite data, because they reported peak areas and did not report a internal water reference or external phantom of metabolites for spectra obtained (Buckley et al., 1994; Nasrallah et al., 1994; Shioiri et al., 1996; Kegeles et al., 2000; Ohara et al., 2000; Delamillieure et al., 2002; Szulc et al., 2007; Tang et al., 2007). One study was excluded because it reported relaxation times as opposed to concentrations (Ongur et al., 2010a). Eight studies were excluded because they reported the same subjects, or clearly had high overlap in subjects that have been published in other studies already included in the meta-analysis (Maier et al., 1995; Deicken et al., 1999; Deicken et al., 2000; van Elst et al., 2005; Ohrmann et al., 2007; Theberge et al., 2007; Ongur et al., 2008; Wood et al., 2009).

Data were analyzed with Review Manager 5.0.25 (Collaboration, 2008). We expected a high level of heterogeneity in the studies included in this meta-analysis; therefore, we conducted meta-analyses using the DerSimonian and Laird random-effects model to estimate effect sizes as standardized mean difference and its 95% confidence interval (CI) (DerSimonian and Laird, 1986).

We further quantified the extent to which observed inconsistency corresponded to between-study differences using the I^2 statistic which measures the percentage of total variation

across studies due to methodological or treatment heterogeneity rather than chance. Inconsistency is low when I^2 is less than 25%, moderate when I^2 is between 25% and 75%, and high when I^2 is greater than 75% (Higgins et al., 2003).

2.7.2. Quantitative comparison of schizophrenia and bipolar disorder—We performed secondary meta-analyses, grouping both studies conducted in bipolar disorder and schizophrenia together in the same analysis. To assess differences in metabolite levels based on diagnosis, we then did a subgroup analysis with subgroups being defined as bipolar disorder and schizophrenia.

2.7.3. Subgroup analyses—*A priori* hypotheses examining potential heterogeneity across studies included differences in magnetic resonance field strength (low field strength: 1.5T and 2T vs. high field strength: 3T and 4T), duration of illness (first episode vs. chronic disease), mood state (manic vs. depressed vs. euthymic), and medication status (current treatment vs. off medication vs. never/minimal treatment). A subgroup analysis performs separate meta-analyses in each subgroup. SMD in each subgroup is obtained, I^2 statistics are performed to assess heterogeneity within the subgroup and χ^2 statistics are done to explore differences between subgroups. Subgroup analyses were performed if inconsistency was moderate to high, and if there were at least two studies to include in each subgroup.

2.7.4. Assessment of publication bias—We planned to conduct a funnel plot to explore publication bias, but did not have an adequate number of studies to perform analysis. When only a limited number of studies are included, accurate identification of publication bias is practically due to chance. Resulting problems are subjectivity in the visual interpretation of the results, technical feasibility, and remaining uncertainty (Lau et al., 2006).

3. RESULTS

3.1. Study Characteristics

Table 1a and 1b give the characteristics of the studies. A total of 4182 subjects (2067 patients and 2115 healthy controls) were included in the 103 studies conducted in patients with schizophrenia, with a median study size of 36 participants (range: 13–115). In the 43 studies with bipolar disorder, 1461 subjects (738 patients and 721 healthy controls) were included, with a median study size of 34 participants (range: 13–64).

84% of studies investigating schizophrenia included chronically ill patients, with 80% of studies including patients currently on antipsychotic medications. 33% of studies researching bipolar disorder included subjects in various mood states, 45% included euthymic patients only, three studies depressed patients only, and five studies included exclusively manic subjects. 45% of studies were conducted in patients currently on psychotropic medications.

A total of 104 studies used MRS with 1.5T field strength, eight studies with 2T, 21 studies with 3T, and 13 studies with 4T field strength.

3.2. Meta-Analyses

In schizophrenia, available data allowed conducting meta-analyses of absolute metabolite levels (NAA, Cr, and Cho) in the following regions of the brain: frontal lobe ($n= 11$) (separate analyses shown for ACC ($n= 10$) and DLPFC ($n= 6$)), hippocampus ($n= 7$), thalamus ($n= 8$), and basal ganglia ($n= 6$). Data were insufficient to perform analyses in both temporal lobe and cerebellum.

Meta-analyses for NAA/Cr ratios were performed in the hippocampus ($n= 8$), thalamus ($n= 9$), basal ganglia ($n= 8$), frontal lobe ($n= 16$), and temporal lobe ($n= 7$). Analyses for Cho/Cr ratios were performed in all the aforementioned regions except temporal lobe, due to lack of sufficient number of studies to include. There were not a sufficient number of studies to conduct meta-analysis in the ACC.

In bipolar disorder, sufficient data were available to conduct meta-analyses of absolute metabolite levels (NAA, Cr, and Cho) in the frontal lobe ($n= 7$) (separate analyses shown for ACC ($n= 5$) and DLPFC ($n= 5$)), hippocampus ($n= 4$), and basal ganglia ($n= 4$). Data were insufficient to perform analyses in the temporal lobe and thalamus.

Meta-analyses for NAA/Cr were conducted in the hippocampus ($n= 4$), basal ganglia ($n= 7$), ACC ($n= 5$), and DLPFC ($n= 6$). For Cho/Cr ratios analyses were conducted in the hippocampus ($n= 3$), basal ganglia ($n= 6$), ACC ($n= 6$), and DLPFC ($n= 5$). There were not sufficient numbers of studies to conduct meta-analyses in the thalamus, frontal lobe, and temporal lobe.

3.2.1. Absolute metabolite value data—Results in schizophrenia demonstrated significantly decreased levels of NAA in the thalamus [SMD = -0.62 (CI -1.12 to -0.13); $p= 0.01$], and frontal lobe [SMD = -0.44 (CI -0.65 to -0.23); $p< 0.001$]. Cr and Cho levels did not differ in any of the regions investigated (Figure 2a). NAA data were consistent over studies in basal ganglia and the frontal lobe. Cr data were consistent in frontal lobe, DLPFC and thalamus. Cho data were consistent in the thalamus, frontal lobe and ACC. Inconsistency was moderate to high in all metabolite levels in the hippocampus. Inconsistencies were evident also in thalamus (NAA), basal ganglia (Cr, Cho), ACC (NAA, Cr) and DLPFC (NAA, Cho).

In bipolar disorder (Figure 2b), results showed significantly decreased levels of NAA in the basal ganglia compared to healthy controls [SMD = -0.44 (CI -0.83 to -0.06); $p= 0.02$]. Increased NAA levels approached significance in the DLPFC [SMD = 0.24 (CI 0.00 to 0.49); $p= 0.05$]. Cr and Cho levels did not differ in any of the regions investigated (Figure 2b). Results were consistent over studies in the basal ganglia for all metabolites and in the frontal lobe for Cr levels. Results showed moderate to high inconsistency in all metabolite levels in hippocampus, ACC, and DLPFC. Inconsistency for Cho in the frontal lobe was moderate.

3.2.2. Metabolite ratio data—Results in schizophrenia demonstrated lower levels of NAA/Cr in the hippocampus ($p< 0.01$), thalamus ($p< 0.01$), frontal lobe ($p< 0.01$), and temporal lobe ($p< 0.01$), but not in the basal ganglia or the DLPFC. Cho/Cr was significantly lower in the hippocampus ($p= 0.03$). No differences were found in any other region. Inconsistency between studies was low for NAA/Cr in the thalamus and frontal lobe and for Cho/Cr in the hippocampus. However, results for ratio data were moderately inconsistent in the hippocampus and basal ganglia, and highly inconsistent in the temporal lobe (Table 2a).

In bipolar disorder, significantly lower NAA/Cr ratios were seen in the hippocampus ($p< 0.01$). Inconsistency was low for Cho/Cr only in the hippocampus and DLPFC (Table 2b).

3.2.3. Absolute versus ratio data—In schizophrenia, data for NAA levels and NAA/Cr ratios were relatively consistent in areas examined. Absolute Cho levels were not affected in patients with schizophrenia, but Cho/Cr levels in the hippocampus were significantly lower in schizophrenia compared to healthy controls.

In bipolar disorder, absolute NAA levels were found to be significantly decreased in the basal ganglia, but these findings could not be confirmed with NAA/Cr ratio data. NAA/Cr ratios were only decreased in the hippocampus, while absolute NAA levels were not decreased in the hippocampus.

3.2.4. Quantitative comparison of schizophrenia and bipolar disorder—We found significant lower NAA levels in the hippocampus ($\chi^2= 21.96$; $df= 1$; $p< 0.001$) and DLPFC ($\chi^2= 11.84$; $df= 1$; $p< 0.001$) in subjects with schizophrenia compared to subjects with bipolar disorder. Cho levels in the hippocampus ($\chi^2= 10.17$; $df= 1$; $p< 0.001$) and Cr levels in the ACC ($\chi^2= 7.35$; $df= 1$; $p< 0.001$) were also significantly lower in schizophrenia than in bipolar disorder. No significant differences in metabolite levels between the disorders were found in any other region studied.

3.2.5. Subgroup analyses

3.2.5.1. First episode vs. chronic schizophrenia: In schizophrenia, all studies that were conducted in first episode subjects enrolled them while off medication. Further, all studies that were conducted in subjects with chronic schizophrenia enrolled subjects currently treated with medication. We were therefore unable to attribute differences between subgroups to either duration of illness or medication status alone.

While initial analyses in schizophrenia showed a significant decrease of NAA in the thalamus with moderate inconsistency, subgroup analyses demonstrated that the decrease was attributable to studies conducted in chronic schizophrenia/subjects on medication [SMD= -0.77 , $p<0.01$]. In first episode psychosis/subjects of medication, there was no significant decrease of NAA compared to healthy controls [SMD= -0.13 ; $p= 0.86$]. Differences between subgroups were significant [$\chi^2= 10.90$; $p< 0.01$]. Inconsistencies found in the basal ganglia and DLPFC could not be explained by difference in illness duration. Data were not sufficient to conduct subgroup analyses in the hippocampus and ACC.

3.2.5.2. Manic vs depressive episode vs euthymic mood state: Data were insufficient to perform subgroup analyses based on mood states.

3.2.5.3. Medication status: For studies conducted in subjects with schizophrenia, results are as described above. Data was insufficient to perform subgroup analyses based on medication status in bipolar disorder.

3.2.5.4. Field strength: In the thalamus, studies conducted with low field strength demonstrated significant decrease of absolute NAA concentrations in schizophrenia (SMD= -0.89 , $p< 0.01$) with inconsistencies across studies remaining moderate, while studies conducted with high field strength did not show any difference compared to healthy controls, with low inconsistency (SMD= -0.17 , $p= 0.86$). There was a significant difference between studies conducted with low vs. high field strength [$\chi^2= 11.03$; $p< 0.01$]. This means that studies conducted with high field strength did not find differences, and these findings were consistent across studies. Findings with low field strength did show decrease in NAA but inconsistency was higher between studies. Inconsistencies in hippocampus and ACC could not be explained by differences in field strength, SMDs between studies conducted with high vs. low field strength did not significantly differ, heterogeneity between studies in subgroups remained high. Data was not sufficient to perform subgroup analyses in basal ganglia and DLPFC.

While none of the subgroups significantly differed in from healthy controls, there was a significant difference between studies conducted with low vs high field strength in the ACC

in bipolar disorder. (NAA (absolute values): $\chi^2= 5.82$; $p= 0.02$, Cr (absolute values): $\chi^2= 16.02$; $p< 0.01$). When only studies with low field strength vs studies with high field strength were grouped together, inconsistency between studies was low. This means that inconsistencies between groups can be explained by differences in field strength. Inconsistencies in the frontal lobe in both absolute and ratio data could not be explained by differences in field strength. Data was not sufficient to perform subgroup analyses in the hippocampus and DLPFC.

4. DISCUSSION

4.1. Findings

While our meta-analysis failed to reveal significant abnormalities in either Cr or Cho levels, we found several abnormalities in NAA levels in schizophrenia and bipolar disorder.

NAA levels appear to be globally decreased (hippocampus, thalamus, frontal and temporal lobe) in patients with schizophrenia compared with healthy controls, which is consistent with previous reports of a subtle decrease of NAA of about 5% overall (Steen, et al., 2005). Brugger previously suggested decreased NAA levels in the thalamus as trait marker of schizophrenia, as he did not find any significant difference between first episode psychosis and chronic schizophrenia (Brugger et al., 2011). However, our analysis suggests that NAA levels in the thalamus are only decreased in chronic schizophrenia but not first episode patients. These differences in results may be partly accounted for by inclusion of a study in pediatric population (O'Neill et al., 2004) and a study published in Turkish language (Basoglu et al., 2006) by Brugger, but at this time it is unclear if findings can be generalized to first episode patients.

NAA levels in bipolar disorder appear to be decreased in the basal ganglia. Data for the DLPFC show that NAA levels in bipolar disorder are increased compared to healthy controls. While the finding of an increase in NAA appears counterintuitive, a recent study suggests a close correlation between NAA and glutamate levels in healthy subjects (Waddell et al., 2011). Glutamate levels are consistently found to be elevated in bipolar disorder (Yuksel and Ongur, 2010), which could imply that the increase of NAA may be related to an increase of glutamate levels. Our understanding of the physiology/ pathophysiology of NAA remains limited, relationships between and interactions with other neurometabolites and resulting pathological implications are poorly researched. In conclusion, alterations of NAA levels may reflect a much more complex underlying process than simply neuronal viability.

In a comparison of effect sizes of alterations in metabolite levels we found significant differences in NAA in the DLPFC between schizophrenia and bipolar disorder. While levels in bipolar disorder were significantly increased compared to healthy controls, decrease in NAA did not reach significance in schizophrenia. However, a bidirectional effect appears to be emerging. While not as clear, a similar pattern is noticeable in NAA and Cho in the hippocampus. No statistically significant increase in NAA in the hippocampus in bipolar disorder was found, but levels were decreased at a trend level in schizophrenia; results differed significantly between the two groups. Cho in the hippocampus and Cr in the ACC did not significantly differ from healthy controls in schizophrenia and bipolar disorder. However, Cho in the hippocampus and Cr in the ACC were significantly lower in schizophrenia and bipolar disorder. It appears that, although alterations in neurometabolites share several commonalities, there appear to be several differences, which stands in contrast to the conceptualization of schizophrenia and bipolar disorder as a continuum of the same disorder. However, the validity of these findings is limited, as there was no direct comparison in metabolite levels. Also, the total number of subjects reported in studies conducted in schizophrenia (1329 patients and 1394 healthy controls in 103 studies) differs

from the ones conducted bipolar disorder (738 patients and 721 healthy controls in 43 studies). While this may be affecting outcomes, subgroup analysis compares effect sizes between groups. Effect size calculations take into account the number of subjects included in the analysis. Many studies did not specify that they have matched their samples for age and gender, which may have confounded results. To date, only a few studies have directly examined neurometabolite level differences between the disorders. Ongur did not find differences in Cr between subjects with schizophrenia and bipolar disorder (Ongur et al., 2009). In the ACC, NAA/Cr was found to be lower in schizophrenia and Cho/Cr was higher when compared to bipolar disorder (Sarramea et al., 2000); others reported decrease in NAA that were similar in both disorders (Molina et al., 2007). More data needs to become available before firm conclusions can be drawn.

4.2. Limitations

The evidence of alterations in brain metabolite levels consists of small studies in different regions of the brain with different patient populations receiving different or no treatment for their disorder. To detect a 10% change in NAA levels with 80% power, a sample size of 39 subjects and 39 controls was suggested; assuming these parameters, no studies conducted in bipolar disorder and only five studies in schizophrenia were adequately powered (Steen et al., 2005). While we attempted to ameliorate the effect of publication and outcome reporting bias by contacting authors, but only 13% provided requested data.

To avoid double counting effects, we excluded studies that included subjects reported elsewhere from analysis. However, it is possible that data remains in analyses that have been published in two sources. For the same reason, we decided to only include one reported dataset when two different sets of schizophrenic/bipolar subjects (e.g. first episode and chronic schizophrenia, manic and euthymic subjects) but only one healthy control group were included in a study. While this approach may lead to decreased power in overall analysis, we believe it is crucial to do so in order to avoid an overstating of the precision of results (Senn, 2009).

To reduce inconsistencies we excluded studies from data analysis if no internal or external concentration reference was used in data acquisition.

We have not attempted to weigh studies by methodological quality, as there is no objective way to assess this from published study methodology descriptions. However, it should be noted that studies conducted with magnets with high field strength that have a priori defined measures of spectral quality, spectral fitting and partial volume correction and include subjects that are not on medications with healthy controls that are matched by age, gender and socioeconomic status are considered to be of highest methodological quality.

We only examined NAA, Cr, and Cho in our systematic review. Since high field strength magnets are available, glutamate, glutamine, and GABA have become metabolites of interest. These metabolites have been implicated in the pathophysiology of both schizophrenia and bipolar disorder (Ongur, et al., 2008; Bustillo et al., 2010; Ongur et al., 2010b; Reid et al., 2010). Given the limited number of studies conducted examining glutamate, we decided that a meta-analytical approach would be premature and not include glutamate, glutamine and GABA in our analysis.

High remaining inconsistency in different areas of the brain makes results harder to interpret as factors that have not been controlled for could have a significant influence on the results. Further data will need to be obtained to clarify these findings, and other potential confounding factors need to be explored. Inconsistencies may be partly attributable to a number of variables that differ in the populations studied.

One of the variables that differ between populations and potentially confounds results is medication status. Many studies enrolled subjects currently treated with antipsychotic or mood stabilizers, potentially confounding our results. 80% of all studies conducted in schizophrenia included subjects on medication, while only 45% of subjects in bipolar trials were currently treated. Modulation of neurometabolite levels with both classes of drugs were demonstrated in the rat model. A consistent upregulation of NAA, with distinct regional patterns of activation depending on the agent given was the most robust finding (McLoughlin et al., 2009). In humans, antipsychotic medications are reported to increase NAA levels, even after a short period of treatment (Bertolino et al., 2001; Szulc et al., 2005). While some studies reported fewer effects on NAA levels in typical antipsychotics than atypical antipsychotics, others were unable to replicate these findings (Braus et al., 2001; Braus et al., 2002; Szulc, et al., 2007; Bustillo et al., 2008). Valproic acid was found to increase NAA/Cr in bipolar patients; lithium, but not valproic acid, was reported to increase NAA in euthymic bipolar patients (Silverstone et al., 2003; Atmaca et al., 2007). We were unable to draw conclusions on the influence of psychotropic medication on neurometabolites as data was not sufficient to do subgroup analyses.

Another potentially confounding clinical factor is mood state of subjects included. A small study suggested that Cr in the frontal lobe was decreased when the same subjects were in a depressed state compared to euthymia (Hamkawa et al., 1999). Severity of depression was positively correlated with elevation of Cho levels in the cingulate, suggesting a mood state dependent alteration (Moore et al., 2000). A recent systematic review found alteration of glutamate levels in bipolar disorder are increased independent of mood state (Yuksel and Ongur, 2010). Our analysis did not allow drawing conclusions about mood state dependent alterations of neurometabolites, as not enough data was available to conduct subgroup analysis.

Several technical variations that may contribute to remaining inconsistencies. Differences in voxel size and voxel placement as well as grey and white matter contributions to selected voxels across studies may have contributed to inconsistencies. Studies used various definitions for areas of interest. Especially in studies with voxel placement within the frontal lobe, authors often identified their area of interest globally as “frontal lobe”. Others placed and defined their area of interest specifically within the ACC or DLPFC, both located within the frontal lobe. Where areas of interest within the frontal lobe were identified, we decided to analyze them separately, in an attempt to avoid overgeneralization of results in this functionally complex region. However, a potential limitation is that studies that actually did place their voxel completely or partly in one of these areas but did not state this in their method section were included in the general frontal lobe analysis which may have skewed the results. Further, MRS quantification is affected by signal to noise ratio (SNR), the quality of spectra from some regions is higher than others. Results show the moderate to high inconsistency in all metabolite levels in hippocampus, inconsistencies were evident also in thalamus, which may be attributable to lower SNR. T1/T2 is different at different field strength and affects metabolite quantification, it has also been demonstrated that T2 changes are associated with psychotic disorders, which may affect outcomes (Ongur et al. 2010a).

Some studies acquired single-voxel MRS while others used MRSI. Single-voxel MRS benefits from ease of implementation and quantitation, with only a single spectrum to process. MRSI has the advantage of acquiring data from multiple voxels in a single measurement, allowing investigation of different brain regions and tissue types. However, the spectrum at a given voxel will have contributions from neighboring voxels, and it is difficult to get a good shim and water suppression over a larger volume.

^1H -MRS studies generally report a combined NAA and N-acetylaspartyl-glutamate (NAAG) peak, this peak is composed of approximately 90% NAA and 10% NAAG (Pouwels et al., 1997; Edden et al., 2007). NAAG is a peptide that is synthesized from NAA and glutamate and abundant in the nervous system, NAAG has been implied to be altered in both schizophrenia and bipolar disorder. It is possible that some of the alterations in NAA levels can be contributed to actual abnormalities of NAAG.

Use of a concentration reference is necessary to account for spatial field inhomogeneities as well as field variations across scanning sessions. Metabolite ratios provide a measure of relative concentrations and are easy to obtain, many studies used Cr as an internal reference. However, this method has the disadvantage of not definitively distinguishing between the numerator and denominator metabolite changes. To calculate metabolite concentrations, an internal or external reference is needed. Some studies used an unsuppressed internal water reference spectrum. Another approach is the use of an external reference solution, but this method is sensitive to inhomogeneities because the reference solution is separated from the volume of interest in the brain.

4.3. Research implications

We found a global decrease of NAA levels in schizophrenia. Of the regions studied, altered NAA levels in bipolar disorder seem to be limited to the basal ganglia and frontal lobe, with decreased levels in the ACC and increased levels in the DLPFC. Cr and Cho levels do not appear to be significantly affected any of the studied areas in either schizophrenia or bipolar disorder. While absolute metabolite levels and ratio data appeared to be consistent in schizophrenia, data were more conflictive in bipolar disorder.

Heterogeneity of data remains large in both schizophrenia and bipolar disorder. We were able to explain some of the inconsistencies. Specifically, high field strength magnets appear to detect more subtle alterations in metabolite levels, but factors that have not been controlled for are probably accounting for the majority of inconsistent findings. Likely confounding factors are not only variability in the patient populations studied but also data acquisition parameters.

Large, carefully designed studies are needed to better estimate the extent of alterations in brain metabolite levels in patient and to determine if MRS could be established as a tool to help differentiate schizophrenia from bipolar disorder.

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DISCLOSURE/ CONFLICT OF INTEREST

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REFERENCES

- Alger JR. Quantitative proton magnetic resonance spectroscopy and spectroscopic imaging of the brain: a didactic review. *Topics in Magnetic Resonance Imaging*. 2010; 21(2):115–128. [PubMed: 21613876]
- Amaral JA, Tamada RS, Issler CK, Caetano SC, Cerri GG, de Castro CC, Lafer B. A ^1H MRS study of the anterior cingulate gyrus in euthymic bipolar patients. *Human Psychopharmacology*. 2006; 21(4): 215–220. [PubMed: 16783812]
- Ando K, Takei N, Matsumoto H, Iyo M, Isoda H, Mori N. Neural damage in the lenticular nucleus linked with tardive dyskinesia in schizophrenia: a preliminary study using proton magnetic resonance spectroscopy. *Schizophrenia Research*. 2002; 57(2–3):273–279.

- Atmaca M, Yildirim H, Ozdemir H, Poyraz AK, Tezcan E, Ogur E. Hippocampal 1H MRS in first-episode bipolar I patients. *Progress in Neuropsychopharmacology and Biological Psychiatry*. 2006; 30(7):1235–1239.
- Atmaca M, Yildirim H, Ozdemir H, Ogur E, Tezcan E. Hippocampal 1H MRS in patients with bipolar disorder taking valproate versus valproate plus quetiapine. *Psychological Medicine*. 2007; 37(1): 121–129. [PubMed: 17094813]
- Auer DP, Wilke M, Grabner A, Heidenreich JO, Bronisch T, Wetter TC. Reduced NAA in the thalamus and altered membrane and glial metabolism in schizophrenic patients detected by 1H-MRS and tissue segmentation. *Schizophrenia Research*. 2001; 52(1–2):87–99. [PubMed: 11595395]
- Aydin K, Uçok A, Cakir S. Quantitative proton MR spectroscopy findings in the corpus callosum of patients with schizophrenia suggest callosal disconnection. *American Journal of Neuroradiology*. 2007; 28(10):1968–1974. [PubMed: 17898202]
- Aydin K, Uçok A, Guler J. Altered metabolic integrity of corpus callosum among individuals at ultra high risk of schizophrenia and first-episode patients. *Biological Psychiatry*. 2008; 64(9):750–757. [PubMed: 18486106]
- Bagory M, Durand-Dubief F, Ibarrola D, Confavreux C, Sappey-Marinié D. "Absolute" quantification in Magnetic Resonance Spectroscopy: validation of a clinical protocol in multiple sclerosis. *Conference Proceedings IEEE Engineering in Medicine and Biology Society*. 2007; 2007:3458–34661.
- Bartha R, Williamson PC, Drost DJ, Malla A, Carr TJ, Cortese L, Canaran G, Rylett RJ, Neufeld RW. Measurement of glutamate and glutamine in the medial prefrontal cortex of never-treated schizophrenic patients and healthy controls by proton magnetic resonance spectroscopy. *Archives of General Psychiatry*. 1997; 54(10):959–965. [PubMed: 9337777]
- Bartha R, al-Semaan YM, Williamson PC, Drost DJ, Malla AK, Carr TJ, Densmore M, Canaran G, Neufeld RW. A short echo proton magnetic resonance spectroscopy study of the left mesial-temporal lobe in first-onset schizophrenic patients. *Biological Psychiatry*. 1999; 45(11):1403–1411. [PubMed: 10356621]
- Basoglu C, Cetin M, Oner O, Ebrinc S, Semiz UB, Kandilcioglu H, Silit E, Kizilkaya E. [Comparison of right thalamus and temporal cortex metabolite levels of drug-naïve first-episode psychotic and chronic schizophrenia in patients]. *Turkish Journal of Psychiatry*. 2006; 17(2):85–91. [PubMed: 16755408]
- Bertolino A, Nawroz S, Mattay VS, Barnett AS, Duyn JH, Moonen CT, Frank JA, Tedeschi D, Weinberger DR. Regionally specific pattern of neurochemical pathology in schizophrenia as assessed by multislice proton magnetic resonance spectroscopic imaging. *American Journal of Psychiatry*. 1996; 153(12):1554–1563. [PubMed: 8942451]
- Bertolino A, Breier A, Callicott JH, Adler C, Mattay VS, Shapiro M, Frank JA, Pickar D, Weinberger DR. The relationship between dorsolateral prefrontal neuronal N-acetylaspartate and evoked release of striatal dopamine in schizophrenia. *Neuropsychopharmacology*. 2000a; 22(2):125–132. [PubMed: 10649825]
- Bertolino A, Esposito G, Callicott JH, Mattay VS, Van Horn JD, Frank JA, Berman KF, Weinberger DR. Specific relationship between prefrontal neuronal N-acetylaspartate and activation of the working memory cortical network in schizophrenia. *American Journal of Psychiatry*. 2000b; 157(1):26–33. [PubMed: 10618009]
- Bertolino A, Callicott JH, Mattay VS, Weidenhammer KM, Rakow R, Egan MF, Weinberger DR. The effect of treatment with antipsychotic drugs on brain N-acetylaspartate measures in patients with schizophrenia. *Biological Psychiatry*. 2001; 49(1):39–46. [PubMed: 11163778]
- Bertolino A, Frye M, Callicott JH, Mattay VS, Rakow R, Shelton-Repella J, Post R, Weinberger DR. Neuronal pathology in the hippocampal area of patients with bipolar disorder: a study with proton magnetic resonance spectroscopic imaging. *Biological Psychiatry*. 2003a; 53(10):906–913. [PubMed: 12742678]
- Bertolino A, Sciota D, Brudaglio F, Altamura M, Blasi G, Bellomo A, Antonucci N, Callicott JA, Goldberg TE, Scarabino T, Weinberger DR, Nardini M. Working memory deficits and levels of N-acetylaspartate in patients with schizophreniform disorder. *American Journal of Psychiatry*. 2003b; 160(3):483–489. [PubMed: 12611829]

- Bhagwagar Z, Wylezinska M, Jezard P, Evans J, Ashworth F, Sule A, Matthews PM, Cowen PJ. Reduction in occipital cortex gamma-aminobutyric acid concentrations in medication-free recovered unipolar depressed and bipolar subjects. *Biological Psychiatry*. 2007; 61(6):806–812. [PubMed: 17210135]
- Block W, Bayer TA, Tepest R, Traber F, Rietschel M, Muller DJ, Schulze TG, Honer WG, Maier W, Schild HH, Falkai P. Decreased frontal lobe ratio of N-acetyl aspartate to choline in familial schizophrenia: a proton magnetic resonance spectroscopy study. *Neuroscience Letters*. 2000; 289(2):147–151. [PubMed: 10904141]
- Brambilla P, Stanley JA, Nicoletti MA, Sassi RB, Mallinger AG, Frank E, Kupfer DJ, Keshavan MS, Soares JC. 1H Magnetic resonance spectroscopy study of dorsolateral prefrontal cortex in unipolar mood disorder patients. *Psychiatry Research*. 2005; 138(2):131–139. [PubMed: 15766636]
- Bramon E, Sham PC. The common genetic liability between schizophrenia and bipolar disorder: a review. *Current Psychiatry Reports*. 2001; 3(4):332–337. [PubMed: 11470041]
- Braus DF, Ende G, Weber-Fahr W, Demirakca T, Henn FA. Favorable effect on neuronal viability in the anterior cingulate gyrus due to long-term treatment with atypical antipsychotics: an MRSI study. *Pharmacopsychiatry*. 2001; 34(6):251–253. [PubMed: 11778146]
- Braus DF, Ende G, Weber-Fahr W, Demirakca T, Tost H, Henn FA. Functioning and neuronal viability of the anterior cingulate neurons following antipsychotic treatment: MR-spectroscopic imaging in chronic schizophrenia. *European Neuropsychopharmacology*. 2002; 2(2):145–152. [PubMed: 11872332]
- Brugger S, Davis JM, Leucht S, Stone JM. Proton magnetic resonance spectroscopy and illness stage in schizophrenia—a systematic review and meta-analysis. *Biological Psychiatry*. 2011; 69(5):495–503. [PubMed: 21145039]
- Buckley PF, Moore C, Long H, Larkin C, Thompson P, Mulvany F, Redmond O, Stack JP, Ennis JT, Waddington JL. 1H-magnetic resonance spectroscopy of the left temporal and frontal lobes in schizophrenia: clinical, neurodevelopmental, and cognitive correlates. *Biological Psychiatry*. 1994; 36(12):792–800. [PubMed: 7893844]
- Bustillo JR, Lauriello J, Rowland LM, Jung RE, Petropoulos H, Hart BL, Blanchard J, Keith SJ, Brooks WM. Effects of chronic haloperidol and clozapine treatments on frontal and caudate neurochemistry in schizophrenia. *Psychiatry Research*. 2001; 107(3):135–149. [PubMed: 11566430]
- Bustillo JR, Lauriello J, Rowland LM, Thomson LM, Petropoulos H, Hammond R, Hart B, Brooks WM. Longitudinal follow-up of neurochemical changes during the first year of antipsychotic treatment in schizophrenia patients with minimal previous medication exposure. *Schizophrenia Research*. 2002a; 58(2–3):313–321.
- Bustillo JR, Rowland LM, Lauriello J, Petropoulos H, Hammond R, Hart B, Brooks WM. High choline concentrations in the caudate nucleus in antipsychotic-naïve patients with schizophrenia. *American Journal of Psychiatry*. 2002b; 159(1):130–133. [PubMed: 11772701]
- Bustillo JR, Rowland LM, Jung R, Brooks WM, Qualls C, Hammond R, Hart B, Lauriello J. Proton magnetic resonance spectroscopy during initial treatment with antipsychotic medication in schizophrenia. *Neuropsychopharmacology*. 2008; 33(10):2456–2466. [PubMed: 18094668]
- Bustillo JR, Rowland LM, Mullins P, Jung R, Chen H, Qualls C, Hammond R, Brooks WM. 1H-MRS at 4 tesla in minimally treated early schizophrenia. *Molecular Psychiatry*. 2010; 15(6):629–636. [PubMed: 19918243]
- Callicott JH, Egan MF, Bertolino A, Mattay VS, Langheim FJ, Frank JA, Weinberger DR. Hippocampal N-acetyl aspartate in unaffected siblings of patients with schizophrenia: a possible intermediate neurobiological phenotype. *Biological Psychiatry*. 1998; 44(10):941–950. [PubMed: 9821558]
- Callicott JH, Bertolino A, Egan MF, Mattay VS, Langheim FJ, Weinberger DR. Selective relationship between prefrontal N-acetylaspartate measures and negative symptoms in schizophrenia. *American Journal of Psychiatry*. 2000a; 157(10):1646–1651. [PubMed: 11007719]
- Callicott JH, Bertolino A, Mattay VS, Langheim FJ, Duyn J, Coppola R, Goldberg TE, Weinberger DR. Physiological dysfunction of the dorsolateral prefrontal cortex in schizophrenia revisited. *Cerebral Cortex*. 2000b; 10(11):1078–1092. [PubMed: 11053229]

- Cecil KM, Lenkinski RE, Gur RE, Gur RC. Proton magnetic resonance spectroscopy in the frontal and temporal lobes of neuroleptic naive patients with schizophrenia. *Neuropsychopharmacology*. 1999; 20(2):131–140. [PubMed: 9885793]
- Cecil KM, DelBello MP, Morey R, Strakowski SM. Frontal lobe differences in bipolar disorder as determined by proton MR spectroscopy. *Bipolar Disorders*. 2002; 4(6):357–365. [PubMed: 12519095]
- Choe BY, Kim KT, Suh TS, Lee C, Paik IH, Bahk YW, Shinn KS, Lenkinski RE. 1H magnetic resonance spectroscopy characterization of neuronal dysfunction in drug-naive, chronic schizophrenia. *Academic Radiology*. 1994; 1(3):211–216. [PubMed: 9419488]
- Choe BY, Suh TS, Shinn KS, Lee CW, Lee C, Paik IH. Observation of metabolic changes in chronic schizophrenia after neuroleptic treatment by in vivo hydrogen magnetic resonance spectroscopy. *Investigative Radiology*. 1996; 31(6):345–352. [PubMed: 8761867]
- Colla M, Schubert F, Bubner M, Heidenreich JO, Bajbouj M, Seifert F, Luborzewski A, Heuser I, Kronenberg G. Glutamate as a spectroscopic marker of hippocampal structural plasticity is elevated in long-term euthymic bipolar patients on chronic lithium therapy and correlates inversely with diurnal cortisol. *Molecular Psychiatry*. 2009; 14(7):696–704. 647. [PubMed: 18347601]
- Collaboration TC. Review Manager (RevMan) [computer program]. Version 5.0. 2008
- Craddock N, Owen MJ. The beginning of the end for the Kraepelinian dichotomy. *British Journal of Psychiatry*. 2005; 186:364–366. [PubMed: 15863738]
- Crow TJ. The continuum of psychosis and its genetic origins. The sixty-fifth Maudsley lecture. *British Journal of Psychiatry*. 1990a; 156:788–797. [PubMed: 2207509]
- Crow, TJ. The question of a genetic continuum for schizophrenia and affective disorder Depression in Schizophrenia. DeLisi, LE., editor. Washington, DC: American Psychiatric Press; 1990b. p. 81-98.
- Dager SR, Friedman SD, Parow A, Demopoulos C, Stoll AL, Lyoo IK, Dunner DL, Renshaw PF. Brain metabolic alterations in medication-free patients with bipolar disorder. *Archives of General Psychiatry*. 2004; 61(5):450–458. [PubMed: 15123489]
- Dager SR, Corrigan NM, Richards TL, Posse S. Research applications of magnetic resonance spectroscopy to investigate psychiatric disorders. *Topics in Magnetic Resonance Imaging*. 2008; 19(2):81–96. [PubMed: 19363431]
- Dalby JT, Morgan D, Lee ML. Schizophrenia and mania in identical twin brothers. *Journal of Nervous and Mental Disease*. 1986; 174(5):304–308. [PubMed: 3701318]
- Deicken RF, Zhou L, Corwin F, Vinogradov S, Weiner MW. Decreased left frontal lobe N-acetylaspartate in schizophrenia. *American Journal of Psychiatry*. 1997a; 154(5):688–690. [PubMed: 9137129]
- Deicken RF, Zhou L, Schuff N, Weiner MW. Proton magnetic resonance spectroscopy of the anterior cingulate region in schizophrenia. *Schizophrenia Research*. 1997b; 27(1):65–71. [PubMed: 9373896]
- Deicken RF, Zhou L, Schuff N, Fein G, Weiner MW. Hippocampal neuronal dysfunction in schizophrenia as measured by proton magnetic resonance spectroscopy. *Biological Psychiatry*. 1998; 43(7):483–488. [PubMed: 9547926]
- Deicken RF, Pegues M, Amend D. Reduced hippocampal N-acetylaspartate without volume loss in schizophrenia. *Schizophrenia Research*. 1999; 37(3):217–223. [PubMed: 10403193]
- Deicken RF, Johnson C, Eliaz Y, Schuff N. Reduced concentrations of thalamic N-acetylaspartate in male patients with schizophrenia. *American Journal of Psychiatry*. 2000; 157(4):644–647. [PubMed: 10739431]
- Deicken RF, Eliaz Y, Feiwell R, Schuff N. Increased thalamic N-acetylaspartate in male patients with familial bipolar I disorder. *Psychiatry Research*. 2001a; 106(1):35–45. [PubMed: 11231098]
- Deicken RF, Feiwell R, Schuff N, Soher B. Evidence for altered cerebellar vermis neuronal integrity in schizophrenia. *Psychiatry Research*. 2001b; 107(3):125–134. [PubMed: 11566429]
- Deicken RF, Pegues MP, Anzalone S, Feiwell R, Soher B. Lower concentration of hippocampal N-acetylaspartate in familial bipolar I disorder. *American Journal of Psychiatry*. 2003; 160(5):873–882. [PubMed: 12727690]

- Delamillieure P, Constans J, Fernandez J, Brazo P, Dollfus S. Proton magnetic resonance spectroscopy (1H-MRS) of the thalamus in schizophrenia. *European Psychiatry*. 2000a; 15(8):489–491. [PubMed: 11175927]
- Delamillieure P, Fernandez J, Constans JM, Brazo P, Benali K, Abadie P, Vasse T, Thibaut F, Courtheoux P, Petit M, Dollfus S. Proton magnetic resonance spectroscopy of the medial prefrontal cortex in patients with deficit schizophrenia: preliminary report. *American Journal of Psychiatry*. 2000b; 157(4):641–643. [PubMed: 10739430]
- Delamillieure P, Constans JM, Fernandez J, Brazo P, Benali K, Courtheoux P, Thibaut F, Petit M, Dollfus S. Proton magnetic resonance spectroscopy (1H MRS) in schizophrenia: investigation of the right and left hippocampus, thalamus, and prefrontal cortex. *Schizophrenia Bulletin*. 2002; 28(2):329–339. [PubMed: 12693438]
- DerSimonian R, Laird N. Meta-analysis in clinical trials. *Controlled Clinical Trials*. 1986; 7(3):177–188. [PubMed: 3802833]
- Edden RAE, Pomper MG, Barker PB. In vivo differentiation of N-acetyl aspartyl glutamate from N-acetyl aspartate at 3 Tesla. *Magnetic resonance in medicine*. 2007; (6):977–982. [PubMed: 17534922]
- Ellison-Wright I, Bullmore E. Anatomy of bipolar disorder and schizophrenia: a meta-analysis. *Schizophrenia Research*. 2010; 117(1):1–12. [PubMed: 20071149]
- Eluri R, Paul C, Roemer R, Boyko O. Single-voxel proton magnetic resonance spectroscopy of the pons and cerebellum in patients with schizophrenia: a preliminary study. *Psychiatry Research*. 1998; 84(1):17–26. [PubMed: 9870414]
- Ende G, Braus DF, Walter S, Weber-Fahr W, Soher B, Maudsley AA, Henn FA. Effects of age, medication, and illness duration on the N-acetyl aspartate signal of the anterior cingulate region in schizophrenia. *Schizophr Research*. 2000; 41(3):389–395.
- Ende G, Braus DF, Walter S, Henn FA. Lower concentration of thalamic n-acetylaspartate in patients with schizophrenia: a replication study. *American Journal of Psychiatry*. 2001; 158(8):1314–1316. [PubMed: 11481168]
- Ende G, Braus DF, Walter S, Weber-Fahr W, Henn FA. Multiregional 1H-MRSI of the hippocampus, thalamus, and basal ganglia in schizophrenia. *European Archives of Psychiatry and Clinical Neuroscience*. 2003; 253(1):9–15.
- Ende G, Hubrich P, Walter S, Weber-Fahr W, Kammerer N, Braus DF, Henn FA. Further evidence for altered cerebellar neuronal integrity in schizophrenia. *American Journal of Psychiatry*. 2005; 162(4):790–792. [PubMed: 15800155]
- Fannon D, Simmons A, Tennakoon L, O'Ceallaigh S, Sumich A, Doku V, Shew C, Sharma T. Selective deficit of hippocampal N-acetylaspartate in antipsychotic-naïve patients with schizophrenia. *Biological Psychiatry*. 2003; 54(6):587–598. [PubMed: 13129653]
- Frey BN, Folgierini M, Nicoletti M, Machado-Vieira R, Stanley JA, Soares JC, Kapczinski F. A proton magnetic resonance spectroscopy investigation of the dorsolateral prefrontal cortex in acute mania. *Human Psychopharmacology*. 2005; 20(2):133–139. [PubMed: 15648094]
- Frey BN, Stanley JA, Nery FG, Monkul ES, Nicoletti MA, Chen HH, Hatch JP, Caetano SC, Ortiz O, Kapczinski F, Soares JC. Abnormal cellular energy and phospholipid metabolism in the left dorsolateral prefrontal cortex of medication-free individuals with bipolar disorder: an in vivo 1H MRS study. *Bipolar Disorders*. 2007; 9(Suppl 1):119–127. [PubMed: 17543030]
- Friedman SD, Dager SR, Parow A, Hirashima F, Demopoulos C, Stoll AL, Lyoo IK, Dunner DL, Renshaw PF. Lithium and valproic acid treatment effects on brain chemistry in bipolar disorder. *Biological Psychiatry*. 2004; 56(5):340–348. [PubMed: 15336516]
- Frye MA, Watzl J, Banakar S, O'Neill J, Mintz J, Davanzo P, Fisher J, Chirichigno JW, Ventura J, Elman S, Tsuang J, Walot I, Thomas MA. Increased anterior cingulate/medial prefrontal cortical glutamate and creatine in bipolar depression. *Neuropsychopharmacology*. 2007a; 32(12):2490–2499. [PubMed: 17429412]
- Frye MA, Thomas MA, Yue K, Binesh N, Davanzo P, Ventura J, O'Neill J, Guze B, Curran JG, Mintz J. O. Reduced concentrations of N-acetylaspartate (NAA) and the NAA-creatine ratio in the basal ganglia in bipolar disorder: a study using 3-Tesla proton magnetic resonance spectroscopy. *Psychiatry Research*. 2007b; 154(3):259–265. [PubMed: 17346949]

- Fujimoto T, Nakano T, Takano T, Takeuchi K, Yamada K, Fukuzako T, Akimoto H. Proton magnetic resonance spectroscopy of basal ganglia in chronic schizophrenia. *Biological Psychiatry*. 1996; 40(1):14–18. [PubMed: 8780850]
- Fukuzako H, Takeuchi K, Hokazono Y, Fukuzako T, Yamada K, Hashiguchi T, Obo Y, Ueyama K, Takigawa M, Fujimoto T. Proton magnetic resonance spectroscopy of the left medial temporal and frontal lobes in chronic schizophrenia: preliminary report. *Psychiatry Research*. 1995; 61(4):193–200. [PubMed: 8748464]
- Fukuzako H, Kodama S, Fukuzako T, Yamada K, Doi W, Sato D, Takigawa M. Subtype-associated metabolite differences in the temporal lobe in schizophrenia detected by proton magnetic resonance spectroscopy. *Psychiatry Research*. 1999; 92(1):45–56. [PubMed: 10688159]
- Fukuzako H. Heritability heightens brain metabolite differences in schizophrenia. *Journal of Neuropsychiatry and Clinical Neuroscience*. 2000; 12(1):95–97.
- Galinska B, Szulc A, Tarasow E, Kubas B, Dzienis W, Czernikiewicz A, Walecki J. Duration of untreated psychosis and proton magnetic resonance spectroscopy (1H-MRS) findings in first-episode schizophrenia. *Medical Science Monitor*. 2009; 15(2):CR82–CR88. [PubMed: 19179972]
- Gimenez M, Junque C, Perez M, Vendrell P, Baeza I, Salamero M, Merdader JM, Bernardo M. Basal ganglia N-acetylaspartate correlates with the performance in the procedural task 'Tower of Hanoi' of neuroleptic-naive schizophrenic patients. *Neuroscience Letters*. 2003; 347(2):97–100. [PubMed: 12873737]
- Goto N, Yoshimura R, Moriya J, Kakeda S, Ueda N, Ikenouchi-Sugita A, Umene-Nakano W, Hayashi K, Oonari N, Korogi Y, Nakamura J. Reduction of brain gamma-aminobutyric acid (GABA) concentrations in early-stage schizophrenia patients: 3T Proton MRS study. *Schizophrenia Research*. 2009; 112(1–3):192–193. [PubMed: 19464152]
- Hagino H, Suzuki M, Mori K, Nohara S, Yamashita I, Takahashi T, Kurokawa K, Matsui M, Watanabe N, Seto H, Kurachi M. Proton magnetic resonance spectroscopy of the inferior frontal gyrus and thalamus and its relationship to verbal learning task performance in patients with schizophrenia: a preliminary report. *Psychiatry and Clinical Neurosciences*. 2002; 56(5):499–507. [PubMed: 12193238]
- Hajek T, Bernier D, Slaney C, Propper L, Schmidt M, Carrey N, MacQueen G, Duffy A, Alda M. A comparison of affected and unaffected relatives of patients with bipolar disorder using proton magnetic resonance spectroscopy. *Journal of Psychiatry and Neuroscience*. 2008; 33(6):531–540. [PubMed: 18982176]
- Hamakawa H, Kato T, Murashita J, Kato N. Quantitative proton magnetic resonance spectroscopy of the basal ganglia in patients with affective disorders. *European Archives of Psychiatry and Clinical Neurosciences*. 1998; 248(1):53–58.
- Hamakawa H, Kato T, Shioiri T, Inubushi T, Kato N. Quantitative proton magnetic resonance spectroscopy of the bilateral frontal lobes in patients with bipolar disorder. *Psychological Medicine*. 1999; 29(3):639–644. [PubMed: 10405085]
- Heckers S. Making progress in schizophrenia research. *Schizophrenia Bulletin*. 2008; 34(4):591–594. [PubMed: 18492660]
- Heimberg C, Komoroski RA, Lawson WB, Cardwell D, Karson CN. Regional proton magnetic resonance spectroscopy in schizophrenia and exploration of drug effect. *Psychiatry Research*. 1998; 83(2):105–115. [PubMed: 9818736]
- Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. *British Medical Journal*. 2003; 327(7414):557–560. [PubMed: 12958120]
- Iosifescu DV, Moore CM, Deckersbach T, Tilley CA, Ostacher MJ, Sachs GS, Nierenberg AA. Galantamine-ER for cognitive dysfunction in bipolar disorder and correlation with hippocampal neuronal viability: a proof-of-concept study. *CNS Neuroscience and Therapeutics*. 2009; 15(4):309–319. [PubMed: 19889129]
- Jakary A, Vinogradov S, Feiwell R, Deicken RF. N-acetylaspartate reductions in the mediodorsal and anterior thalamus in men with schizophrenia verified by tissue volume corrected proton MRSI. *Schizophrenia Research*. 2005; 76(2–3):173–185. [PubMed: 15949650]
- Kato T, Hamakawa H, Shioiri T, Murashita J, Takahashi Y, Takahashi S, Inubushi T. Choline-containing compounds detected by proton magnetic resonance spectroscopy in the basal ganglia in

- bipolar disorder. *Journal of Psychiatry and Neuroscience*. 1996; 21(4):248–254. [PubMed: 8754593]
- Kaufman RE, Ostacher MJ, Marks EH, Simon NM, Sachs GS, Jensen JE, Renshaw PF, Pollack MH. Brain GABA levels in patients with bipolar disorder. *Progress in Neuropsychopharmacology and Biological Psychiatry*. 2009; 33(3):427–434.
- Kegeles LS, Shungu DC, Anjilvel S, Chan S, Ellis SP, Xanthopoulos E, Malaspina D, Gorman JM, Mann JJ, Laruelle M, Kaufmann CA. Hippocampal pathology in schizophrenia: magnetic resonance imaging and spectroscopy studies. *Psychiatry Research*. 2000; 98(3):163–175. [PubMed: 10821999]
- Keshavan MS, Stanley JA, Pettegrew JW. Magnetic resonance spectroscopy in schizophrenia: methodological issues and findings--part II. *Biological Psychiatry*. 2000; 48(5):369–380. [PubMed: 10978720]
- Klar AA, Ballmaier M, Leopold K, Hake I, Schaefer M, Bruhl R. Interaction of hippocampal volume and N-acetylaspartate concentration deficits in schizophrenia: a combined MRI and 1H-MRS study. *Neuroimage*. 2010; 53(1):51–57. [PubMed: 20541020]
- Kreis R, Ernst T, Ross BD. Development of the human brain: in vivo quantification of metabolite and water content with proton magnetic resonance spectroscopy. *Magnetic Resonance in Medicine*. 1993; 30(4):424–437. [PubMed: 8255190]
- Lau J, Ioannidis JP, Terrin N, Schmid CH, Olkin I. The case of the misleading funnel plot. *British Medical Journal*. 2006; 333(7568):597–600. [PubMed: 16974018]
- Lichtenstein P, Yip BH, Bjork C, Pawitan Y, Cannon TD, Sullivan PF, Hultman CM. Common genetic determinants of schizophrenia and bipolar disorder in Swedish families: a population-based study. *Lancet*. 2009; 373(9659):234–239. [PubMed: 19150704]
- Lim KO, Adalsteinsson E, Spielman D, Sullivan EV, Rosenbloom MJ, Pfefferbaum A. Proton magnetic resonance spectroscopic imaging of cortical gray and white matter in schizophrenia. *Archives of General Psychiatry*. 1998; 55(4):346–352. [PubMed: 9554430]
- Lutkenhoff ES, van Erp TG, Thomas MA, Therman S, Manninen M, Huttunen MO, Kaprio J, Lonngvist J, O'Neill J, Cannon TD. Proton MRS in twin pairs discordant for schizophrenia. *Molecular Psychiatry*. 2010; 15(3):308–318. [PubMed: 18645571]
- Maier M, Ron MA, Barker GJ, Tofts PS. Proton magnetic resonance spectroscopy: an in vivo method of estimating hippocampal neuronal depletion in schizophrenia. *Psychological Medicine*. 1995; 25(6):1201–1209. [PubMed: 8637950]
- Maier M, Ron MA. Hippocampal age-related changes in schizophrenia: a proton magnetic resonance spectroscopy study. *Schizophrenia Research*. 1996; 22(1):5–17. [PubMed: 8908686]
- Malhi GS, Ivanovski B, Wen W, Lagopoulos J, Moss K, Sachdev P. Measuring mania metabolites: a longitudinal proton spectroscopy study of hypomania. *Acta Psychiatrica Scandinavia Supplement*. 2007; 434:57–66.
- Martinez-Granados B, Brotons O, Martinez-Bisbal MC, Celda B, Marti-Bonmati L, Aguilar EJ, Gonzalez JC, Sanjuan J. Spectroscopic metabolomic abnormalities in the thalamus related to auditory hallucinations in patients with schizophrenia. *Schizophrenia Research*. 2008; 104(1–3): 13–22. [PubMed: 18650068]
- McLoughlin GA, Ma D, Tsang TM, Jones DN, Cilia J, Hill MD, Robbins MJ, Benzel IM, Maycox PR, Holmes E, Bahn S. Analyzing the effects of psychotropic drugs on metabolite profiles in rat brain using 1H NMR spectroscopy. *Journal of Proteome Research*. 2009; 8(4):1943–1952. [PubMed: 19714815]
- Meyerhoff DJ, MacKay S, Bachman L, Poole N, Dillon WP, Weiner MW, Fein G. Reduced brain N-acetylaspartate suggests neuronal loss in cognitively impaired human immunodeficiency virus-seropositive individuals: in vivo 1H magnetic resonance spectroscopic imaging. *Neurology*. 1993; 43(3 Pt 1):509–515. [PubMed: 8450992]
- Michael N, Erfurth A, Ohrmann P, Gossling M, Arolt V, Heindel W, Pfleiderer B. Acute mania is accompanied by elevated glutamate/glutamine levels within the left dorsolateral prefrontal cortex. *Psychopharmacology (Berl)*. 2003; 168(3):344–346. [PubMed: 12684737]
- Michael N, Erfurth A, Pfleiderer B. Elevated metabolites within dorsolateral prefrontal cortex in rapid cycling bipolar disorder. *Psychiatry Research*. 2009; 172(1):78–81. [PubMed: 19239983]

- Miller BL. A review of chemical issues in ¹H NMR spectroscopy: N-acetyl-L-aspartate, creatine and choline. *NMR in biomedicine*. 1991; 4(2):47–52. [PubMed: 1650241]
- Molina V, Sanchez J, Reig S, Sanz J, Benito C, Santamarta C, Pascau J, Sarramea F, Gisbert JD, Misiego JM, Palomo T, Desco M. N-acetyl-aspartate levels in the dorsolateral prefrontal cortex in the early years of schizophrenia are inversely related to disease duration. *Schizophrenia Research*. 2005; 73(2–3):209–219. [PubMed: 15653263]
- Molina V, Sanz J, Sarramea F, Luque R, Benito C, Palomo T. No association between dorsolateral prefrontal gray matter deficit and N-acetyl aspartate ratios in schizophrenia. *Neuropsychobiology*. 2006; 54(3):171–178. [PubMed: 17230035]
- Molina V, Sanchez J, Sanz J, Reig S, Benito C, Leal I, Sarramea F, Rebolledo R, Palomo T, Desco M. Dorsolateral prefrontal N-acetyl-aspartate concentration in male patients with chronic schizophrenia and with chronic bipolar disorder. *European Psychiatry*. 2007; 22(8):505–512. [PubMed: 17904824]
- Moore CM, Breeze JL, Gruber SA, Babb SM, Frederick BB, Villafuerte RA, Stoll AL, Hennen J, Yurgelun-Todd DA, Cohen BM, Renshaw PF. Choline, myo-inositol and mood in bipolar disorder: a proton magnetic resonance spectroscopic imaging study of the anterior cingulate cortex. *Bipolar Disorders*. 2000; 2(3 Pt 2):207–216. [PubMed: 11249799]
- Moore CM, Bonello CM, Sherwood AR, Cohen BM, Renshaw PF, Yurgelun-Todd DA. Mesial temporal lobe Cho to Cr(PCr) ratio asymmetry in chronic schizophrenics. *Schizophrenia Research*. 2002; 57(1):35–42. [PubMed: 12165374]
- Moore GJ, Bebchuk JM, Hasanat K, Chen G, Seraji-Bozorgzad N, Wilds IB, Faulk MW, Koch S, Glitz DA, Jolkovsky L, Manji HK. Lithium increases N-acetyl-aspartate in the human brain: in vivo evidence in support of bcl-2's neurotrophic effects? *Biological Psychiatry*. 2000; 48(1):1–8. [PubMed: 10913502]
- Nasrallah HA, Skinner TE, Schmalbrock P, Robitaille PM. Proton magnetic resonance spectroscopy (¹H MRS) of the hippocampal formation in schizophrenia: a pilot study. *British Journal of Psychiatry*. 1994; 165(4):481–485. [PubMed: 7804662]
- O'Neill J, Levitt J, Caplan R, Asarnow R, McCracken JT, Toga AW, Alger JR. ¹H MRSI evidence of metabolic abnormalities in childhood-onset schizophrenia. *Neuroimage*. 2004; 21(4):1781–1789. [PubMed: 15050598]
- Ohara K, Isoda H, Suzuki Y, Takehara Y, Ochiai M, Takeda H, Igarashi Y, Ohara K. Proton magnetic resonance spectroscopy of the lenticular nuclei in bipolar I affective disorder. *Psychiatry Research*. 1998; 84(2–3):55–60. [PubMed: 10710163]
- Ohara K, Isoda H, Suzuki Y, Takehara Y, Ochiai M, Takeda H, Hattori K, Igarashi Y, Ohara K. Proton magnetic resonance spectroscopy of lenticular nuclei in simple schizophrenia. *Progress in Neuropsychopharmacology and Biological Psychiatry*. 2000; 24(4):507–519.
- Ohrmann P, Siegmund A, Suslow T, Spitzberg K, Kersting A, Arolt V, Heindel W, Pleiderer B. Evidence for glutamatergic neuronal dysfunction in the prefrontal cortex in chronic but not in first-episode patients with schizophrenia: a proton magnetic resonance spectroscopy study. *Schizophrenia Research*. 2005; 73(2–3):153–157. [PubMed: 15653258]
- Ohrmann P, Siegmund A, Suslow T, Pedersen A, Spitzberg K, Kersting A. Cognitive impairment and in vivo metabolites in first-episode neuroleptic-naive and chronic medicated schizophrenic patients: a proton magnetic resonance spectroscopy study. *Journal of Psychiatric Research*. 2007; 41(8):625–634. [PubMed: 16949099]
- Ohrmann P, Kugel H, Bauer J, Siegmund A, Kolkebeck K, Suslow T, Wiedl KH, Rothermundt M, Arolt V, Pedersen A. Learning potential on the WCST in schizophrenia is related to the neuronal integrity of the anterior cingulate cortex as measured by proton magnetic resonance spectroscopy. *Schizophrenia Research*. 2008; 106(2–3):156–163. [PubMed: 18799290]
- Olbrich HM, Valerius G, Rusch N, Buchert M, Thiel T, Hennig J, Ebert D, Van Elst LT. Frontolimbic glutamate alterations in first episode schizophrenia: evidence from a magnetic resonance spectroscopy study. *World Journal of Biological Psychiatry*. 2008; 9(1):59–63. [PubMed: 17853298]
- Omori M, Murata T, Kimura H, Koshimoto Y, Kado H, Ishimori Y, Ito H, Wada Y. Thalamic abnormalities in patients with schizophrenia revealed by proton magnetic resonance spectroscopy. *Psychiatry Research*. 2000; 98(3):155–162. [PubMed: 10821998]

- Ongur D, Jensen JE, Prescott AP, Stork C, Lundy M, Cohen BM, Renshaw PF. Abnormal glutamatergic neurotransmission and neuronal-glia interactions in acute mania. *Biological Psychiatry*. 2008; 64(8):718–726. [PubMed: 18602089]
- Ongur D, Prescott AP, Jensen JE, Cohen BM, Renshaw PF. Creatine abnormalities in schizophrenia and bipolar disorder. *Psychiatry Research*. 2009; 172(1):44–48. [PubMed: 19239984]
- Ongur D, Prescott AP, Jensen JE, Rouse ED, Cohen BM, Renshaw PF. T2 relaxation time abnormalities in bipolar disorder and schizophrenia. *Magnetic Resonance in Medicine*. 2010a; 63(1):1–8. [PubMed: 19918902]
- Ongur D, Prescott AP, McCarthy J, Cohen BM, Renshaw PF. Elevated gamma-aminobutyric acid levels in chronic schizophrenia. *Biological Psychiatry*. 2010b; 68(7):667–670. [PubMed: 20598290]
- Pae CU, Choe BY, Joo RH, Lim HK, Kim TS, Yoo SS, Choi BG, Kim JJ, Lee SJ, Lee C, Paik IH, Lee CU. Neuronal dysfunction of the frontal lobe in schizophrenia. *Neuropsychobiology*. 2004; 50(3):211–215. [PubMed: 15365217]
- Pajonk FG, Wobrock T, Gruber O, Scherk H, Berner D, Kaizl I, Kierer A, Mueller S, Oest M, Meyer T, Backens M, Schneider-Axmann T, Thornton AE, Honer WG, Falkai P. Hippocampal plasticity in response to exercise in schizophrenia. *Archives of General Psychiatry*. 2010; 67(2):133–143. [PubMed: 20124113]
- Port JD, Unal SS, Mrazek DA, Marcus SM. Metabolic alterations in medication-free patients with bipolar disorder: a 3T CSF-corrected magnetic resonance spectroscopic imaging study. *Psychiatry Research*. 2008; 162(2):113–121. [PubMed: 18164911]
- Premkumar P, Parbhakar VA, Fannon D, Lythgoe D, Williams SC, Kuipers E, Kumari V. N-acetyl aspartate concentration in the anterior cingulate cortex in patients with schizophrenia: a study of clinical and neuropsychological correlates and preliminary exploration of cognitive behaviour therapy effects. *Psychiatry Research*. 2010; 182(3):251–260. [PubMed: 20488677]
- Pouwels PJ, Frahm J. Differential distribution of NAA and NAAG in human brain as determined by quantitative localized proton MRS. *NMR in biomedicine*. 1997; 10(2):73–78. [PubMed: 9267864]
- Purcell SM, Wray NR, Stone JL, Visscher PM, O'Donovan MC, Sullivan PF, Sklar P. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature*. 2009; 460(7256):748–752. [PubMed: 19571811]
- Reid MA, Stoeckel LE, White DM, Aysar KB, Bolding MS, Akella NS, Knowlton RC, den Hollander JA, Lahti AC. Assessments of function and biochemistry of the anterior cingulate cortex in schizophrenia. *Biological Psychiatry*. 2010; 68(7):625–633. [PubMed: 20570244]
- Rowland LM, Spieker EA, Francis A, Barker PB, Carpenter WT, Buchanan RW. White matter alterations in deficit schizophrenia. *Neuropsychopharmacology*. 2009; 34(6):1514–1522. [PubMed: 19052539]
- Rusch N, Tebartz van Elst L, Valerius G, Buchert M, Thiel T, Ebert D, Henning J, Olbrich HM. Neurochemical and structural correlates of executive dysfunction in schizophrenia. *Schizophrenia Research*. 2008; 99(1–3):155–163. [PubMed: 17616347]
- Sager TN, Topp S, Torup L, Hanson LG, Egestad B, Moller A. Evaluation of CA1 damage using single-voxel 1H-MRS and un-biased stereology: Can non-invasive measures of N-acetyl-aspartate following global ischemia be used as a reliable measure of neuronal damage? *Brain Research*. 2001; 892(1):166–175. [PubMed: 11172761]
- Sanches RF, Crippa JA, Hallak JE, de Sousa JP, Araujo D, Santos AC, Zuardi AW. Proton magnetic resonance spectroscopy of the frontal, cingulate and perirolandic cortices and its relationship to skin conductance in patients with schizophrenia. *Brazilian Journal of Medical and Biological Research*. 2008; 41(12):1132–1141. [PubMed: 19148378]
- Sarramea F, Luque R, Prieto D, Sau P, Albert C, Leal I. Biochemical changes in the cingulum in patients with schizophrenia and chronic bipolar disorder. *European Archives of Psychiatry and Clinical Neuroscience*. 2008; 258(7):394–401. [PubMed: 18437276]
- Scherk H, Backens M, Schneider-Axmann T, Kemmer C, Usher J, Reith W, Falkai P, Gruber O. Neurochemical pathology in hippocampus in euthymic patients with bipolar I disorder. *Acta Psychiatrica Scandinavica*. 2008; 117(4):283–288.

- Scherk H, Backens M, Schneider-Axmann T, Kraft S, Kemmer C, Usher J, Reith W, Falkai P, Meyer J, Gruber O. Dopamine transporter genotype influences N-acetyl-aspartate in the left putamen. *World Journal of Biological Psychiatry*. 2009a; 10(4 Pt 2):524–530. [PubMed: 17965994]
- Scherk H, Backens M, Schneider-Axmann T, Usher J, Kemmer C, Reith W, Falkai P, Gruber O. Cortical neurochemistry in euthymic patients with bipolar I disorder. *World Journal of Biological Psychiatry*. 2009b; 10(4):285–294. [PubMed: 19921970]
- Senaratne R, Milne AM, MacQueen GM, Hall GB. Increased choline-containing compounds in the orbitofrontal cortex and hippocampus in euthymic patients with bipolar disorder: a proton magnetic resonance spectroscopy study. *Psychiatry Research*. 2009; 172(3):205–209. [PubMed: 19386476]
- Senn SJ. Overstating the evidence: double counting in meta-analysis and related problems. *BMC Medical Research Methodology*. 2009; 9:10. [PubMed: 19216779]
- Sharma R, Venkatasubramanian PN, Barany M, Davis JM. Proton magnetic resonance spectroscopy of the brain in schizophrenic and affective patients. *Schizophrenia Research*. 1992; 8(1):43–49. [PubMed: 1329928]
- Shimizu E, Hashimoto K, Ochi S, Fukami G, Fujisaki M, Koike K, Okamura N, Ohgake S, Koizumi H, Matsuzawa D, Zhang L, Watanabe H, Nakazato M, Shinoda N, Komatsu N, Morita F, Iyo M. Posterior cingulate gyrus metabolic changes in chronic schizophrenia with generalized cognitive deficits. *Journal of Psychiatric Research*. 2007; 41(1–2):49–56. [PubMed: 15993895]
- Shioiri T, Hamakawa H, Kato T, Murashita J, Fujii K, Inubushi T, Takahasi S. Proton magnetic resonance spectroscopy of the basal ganglia in patients with schizophrenia: a preliminary report. *Schizophrenia Research*. 1996; 22(1):19–26. [PubMed: 8908687]
- Shirayama Y, Obata T, Matsuzawa D, Nonaka H, Kanazawa Y, Yoshitome E, Ikehira H, Hashimoto K, Iyo M. Specific metabolites in the medial prefrontal cortex are associated with the neurocognitive deficits in schizophrenia: a preliminary study. *Neuroimage*. 2010; 49(3):2783–2790. [PubMed: 19850131]
- Sigmundsson T, Maier M, Toone BK, Williams SC, Simmons A, Greenwood K, Ron MA. Frontal lobe N-acetylaspartate correlates with psychopathology in schizophrenia: a proton magnetic resonance spectroscopy study. *Schizophrenia Research*. 2003; 64(1):63–71. [PubMed: 14511802]
- Silverstone PH, Wu RH, O'Donnell T, Ulrich M, Asghar SJ, Hanstock CC. Chronic treatment with both lithium and sodium valproate may normalize phosphoinositol cycle activity in bipolar patients. *Human Psychopharmacology*. 2002; 17(7):321–327. [PubMed: 12415549]
- Silverstone PH, Wu RH, O'Donnell T, Ulrich M, Asghar SJ, Hanstock CC. Chronic treatment with lithium, but not sodium valproate, increases cortical N-acetylaspartate concentrations in euthymic bipolar patients. *International Clinical Psychopharmacology*. 2003; 18(2):73–79. [PubMed: 12598817]
- Stanley JA, Williamson PC, Drost DJ, Rylett RJ, Carr TJ, Malla A, Thompson RT. An in vivo proton magnetic resonance spectroscopy study of schizophrenia patients. *Schizophrenia Bulletin*. 1996; 22(4):597–609. [PubMed: 8938914]
- Steel RM, Bastin ME, McConnell S, Marshall I, Cunningham-Owens DG, Lawrie SM, Johnstone EC, Best JJ. Diffusion tensor imaging (DTI) and proton magnetic resonance spectroscopy (1H MRS) in schizophrenic subjects and normal controls. *Psychiatry Research*. 2001; 106(3):161–170. [PubMed: 11382538]
- Steen RG, Hamer RM, Lieberman JA. Measurement of brain metabolites by 1H magnetic resonance spectroscopy in patients with schizophrenia: a systematic review and meta-analysis. *Neuropsychopharmacology*. 2005; 30(11):1949–1962. [PubMed: 16123764]
- Szulc A, Galinska B, Tarasow E, Dzienis W, Kubas B, Konarzewska B, Walecki J, Alathiaki AS, Czernikiewicz A. The effect of risperidone on metabolite measures in the frontal lobe, temporal lobe, and thalamus in schizophrenic patients. A proton magnetic resonance spectroscopy (1H MRS). 2005; 38(5):214–219.
- Szulc A, Galinska B, Tarasow E, Kubas B, Dzienis W, Konarzewska B, Poplawska R, Bibulowicz D, Simonieko K, Walecki J. N-acetylaspartate (NAA) levels in selected areas of the brain in patients with chronic schizophrenia treated with typical and atypical neuroleptics: a proton magnetic resonance spectroscopy (1H MRS) study. *Medical Science Monitor*. 2007; 13(Supplement 1):17–22. [PubMed: 17507880]

- Tanaka Y, Obata T, Sassa T, Yoshitome E, Asai Y, Ikehira H, Suhara T, Okubo Y, Nishikawa T. Quantitative magnetic resonance spectroscopy of schizophrenia: relationship between decreased N-acetylaspartate and frontal lobe dysfunction. *Psychiatry and Clinical Neurosciences*. 2006; 60(3):365–372. [PubMed: 16732755]
- Tang CY, Friedman J, Shungu D, Chang L, Ernst T, Stewart D, Hajianpour A, Carpenter D, Ng J, Mao X, Hof PR, Buchsbaum MS, Davis K, Gorman JM. Correlations between Diffusion Tensor Imaging (DTI) and Magnetic Resonance Spectroscopy (1H MRS) in schizophrenic patients and normal controls. *BMC Psychiatry*. 2007; 7:25. [PubMed: 17578565]
- Tayoshi S, Sumitani S, Taniguchi K, Shibuya-Tayoshi S, Numata S, Iga J, Nakataki M, Ueno S, Harada M, Omori T. Metabolite changes and gender differences in schizophrenia using 3-Tesla proton magnetic resonance spectroscopy (1H-MRS). *Schizophrenia Research*. 2009; 108(1–3): 69–77. [PubMed: 19097753]
- Theberge J, Bartha R, Drost DJ, Menon RS, Malla A, Takhar J, Neufeld RW, Rogers J, Pavlosky W, Schaefer B, Densmore M, Al-Semaan Y, Williamson PC. Glutamate and glutamine measured with 4.0 T proton MRS in never-treated patients with schizophrenia and healthy volunteers. *American Journal of Psychiatry*. 2002; 159(11):1944–1946. [PubMed: 12411236]
- Theberge J, Al-Semaan Y, Williamson PC, Menon RS, Neufeld RW, Rajakumar N, Schaefer B, Densmore M, Drost DJ. Glutamate and glutamine in the anterior cingulate and thalamus of medicated patients with chronic schizophrenia and healthy comparison subjects measured with 4.0-T proton MRS. *American Journal of Psychiatry*. 2003; 160(12):2231–2233. [PubMed: 14638596]
- Theberge J, Al-Semaan Y, Drost DJ, Malla AK, Neufeld RW, Bartha R, Manchanda R, Mennon R, Densmore M, Schaefer B, Williamson PC. Duration of untreated psychosis vs. N-acetylaspartate and choline in first episode schizophrenia: a 1H magnetic resonance spectroscopy study at 4.0 Tesla. *Psychiatry Research*. 2004a; 131(2):107–114. [PubMed: 15313517]
- Theberge J, Al-Semaan Y, Jensen JE, Williamson PC, Neufeld RW, Menon RS, Schaefer B, Densmore M, Drost DJ. Comparative study of proton and phosphorus magnetic resonance spectroscopy in schizophrenia at 4 Tesla. *Psychiatry Research*. 2004b; 132(1):33–39. [PubMed: 15546701]
- Theberge J, Williamson KE, Aoyama N, Drost DJ, Manchanda R, Malla AK, Northcott S, Menon RS, Neufeld RW, Rajakumar N, Pavlosky W, Densmore M, Schaefer B, Williamson PC. Longitudinal grey-matter and glutamatergic losses in first-episode schizophrenia. *British Journal of Psychiatry*. 2007; 191:325–334. [PubMed: 17906243]
- Tibbo P, Hanstock CC, Asghar S, Silverstone P, Allen PS. Proton magnetic resonance spectroscopy (1H-MRS) of the cerebellum in men with schizophrenia. *Journal of Psychiatry and Neuroscience*. 2000; 25(5):509–512. [PubMed: 11109301]
- Tunc-Skarka N, Weber-Fahr W, Hoerst M, Meyer-Lindenberg A, Zink M, Ende G. MR spectroscopic evaluation of N-acetylaspartate's T2 relaxation time and concentration corroborates white matter abnormalities in schizophrenia. *Neuroimage*. 2009; 48(3):525–531. [PubMed: 19573608]
- van Elst LT, Valerius G, Buchert M, Thiel T, Rusch N, Bubl E, Hennig J, Ebert D, Olbrich HM. Increased prefrontal and hippocampal glutamate concentration in schizophrenia: evidence from a magnetic resonance spectroscopy study. *Biological Psychiatry*. 2005; 58(9):724–730. [PubMed: 16018980]
- Van Snellenberg JX, de Candia T. Meta-analytic evidence for familial coaggregation of schizophrenia and bipolar disorder. *Archives of General Psychiatry*. 2009; 66(7):748–755. [PubMed: 19581566]
- Waddell KW, Zanjani-pour P, Pradhan S, Xu L, Welch EB, Joers JM, Martin PR, Avison MJ, Gore JC. Anterior cingulate and cerebellar GABA and Glu correlations measured by (1)H J-difference spectroscopy. *Magnetic Resonance Imaging*. 2011; 29(1):19–24. [PubMed: 20884148]
- Weber-Fahr W, Ende G, Braus DF, Bachert P, Soher BJ, Henn FA, Buchel C. A fully automated method for tissue segmentation and CSF-correction of proton MRSI metabolites corroborates abnormal hippocampal NAA in schizophrenia. *Neuroimage*. 2002; 16(1):49–60. [PubMed: 11969317]

- Winsberg ME, Sachs N, Tate DL, Adalsteinsson E, Spielman D, Ketter TA. Decreased dorsolateral prefrontal N-acetyl aspartate in bipolar disorder. *Biological Psychiatry*. 2000; 47(6):475–481. [PubMed: 10715353]
- Wobrock T, Kamer T, Roy A, Vogeley K, Schneider-Axmann T, Wagner M, Maier W, Rietschel M, Schulze TG, Scherk H, Schild HH, Block W, Traber F, Tepest R, Honer WG, Falkai P. Reduction of the internal capsule in families affected with schizophrenia. *Biological Psychiatry*. 2008; 63(1):65–71. [PubMed: 17574215]
- Wood SJ, Berger G, Velakoulis D, Phillips LJ, McGorry PD, Yung AR, Desmon P, Pantelis C. Proton magnetic resonance spectroscopy in first episode psychosis and ultra high-risk individuals. *Schizophrenia Bulletin*. 2003; 29(4):831–843. [PubMed: 14989417]
- Wood SJ, Yucel M, Wellard RM, Harrison BJ, Clarke K, Fornito A, Velakoulis D, Pantelis C. Evidence for neuronal dysfunction in the anterior cingulate of patients with schizophrenia: a proton magnetic resonance spectroscopy study at 3 T. *Schizophrenia Research*. 2007; 94(1–3): 328–331. [PubMed: 17574388]
- Wood SJ, Berger GE, Wellard RM, Proffitt T, McConchie M, Velakoulis D. A 1H-MRS investigation of the medial temporal lobe in antipsychotic-naive and early-treated first episode psychosis. *Schizophrenia Research*. 2008; 102(1–3):163–170. [PubMed: 18456460]
- Wood SJ, Berger GE, Wellard RM, Proffitt TM, McConchie M, Berk M, McGorry PD, Pantelis C. Medial temporal lobe glutathione concentration in first episode psychosis: a 1H-MRS investigation. *Neurobiology of Disease*. 2009; 33(3):354–357. [PubMed: 19118629]
- Wu RH, O'Donnell T, Ulrich M, Asghar SJ, Hanstock CC, Silverstone PH. Brain choline concentrations may not be altered in euthymic bipolar disorder patients chronically treated with either lithium or sodium valproate. *Annals of General Hospital Psychiatry*. 2004; 3(1):13. [PubMed: 15283867]
- Yamasue H, Fukui T, Fukuda R, Kasai K, Iwanami A, Kato N, Kato T. Drug-induced parkinsonism in relation to choline-containing compounds measured by 1H-MR spectroscopy in putamen of chronically medicated patients with schizophrenia. *International Journal of Neuropsychopharmacology*. 2003; 6(4):353–360. [PubMed: 14604450]
- Yildiz-Yesiloglu A, Ankerst DP. Neurochemical alterations of the brain in bipolar disorder and their implications for pathophysiology: a systematic review of the in vivo proton magnetic resonance spectroscopy findings. *Progress in Neuropsychopharmacology and Biol Psychiatry*. 2006; 30(6): 969–995.
- Yuksel C, Ongur D. Magnetic resonance spectroscopy studies of glutamate-related abnormalities in mood disorders. *Biological Psychiatry*. 2010; 68(9):785–794. [PubMed: 20728076]
- Yurgelun-Todd DA, Renshaw PF, Gruber SA, Ed M, Waternaux C, Cohen BM. Proton magnetic resonance spectroscopy of the temporal lobes in schizophrenics and normal controls. *Schizophrenia Research*. 1996; 19(1):55–59. [PubMed: 9147496]

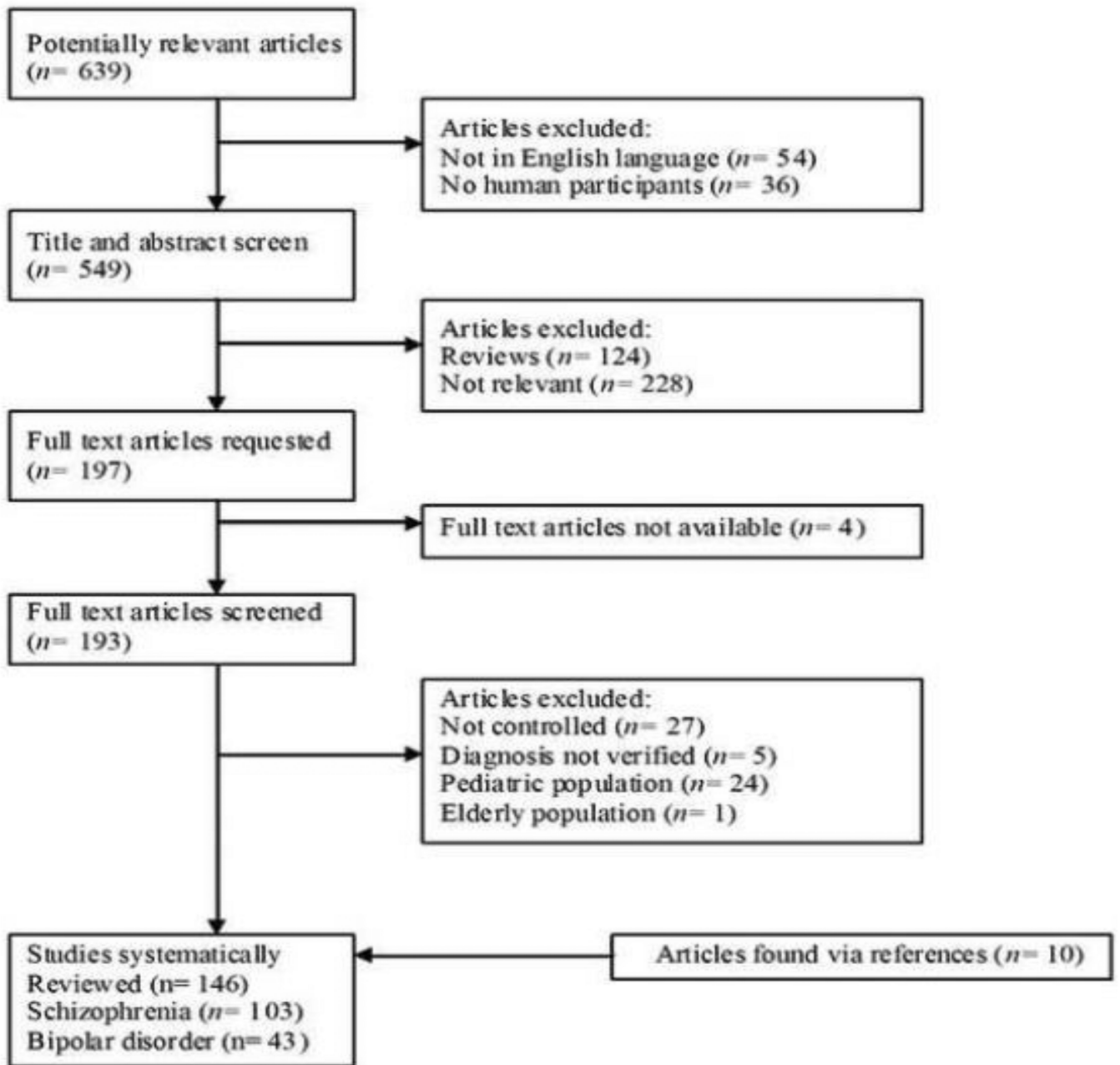
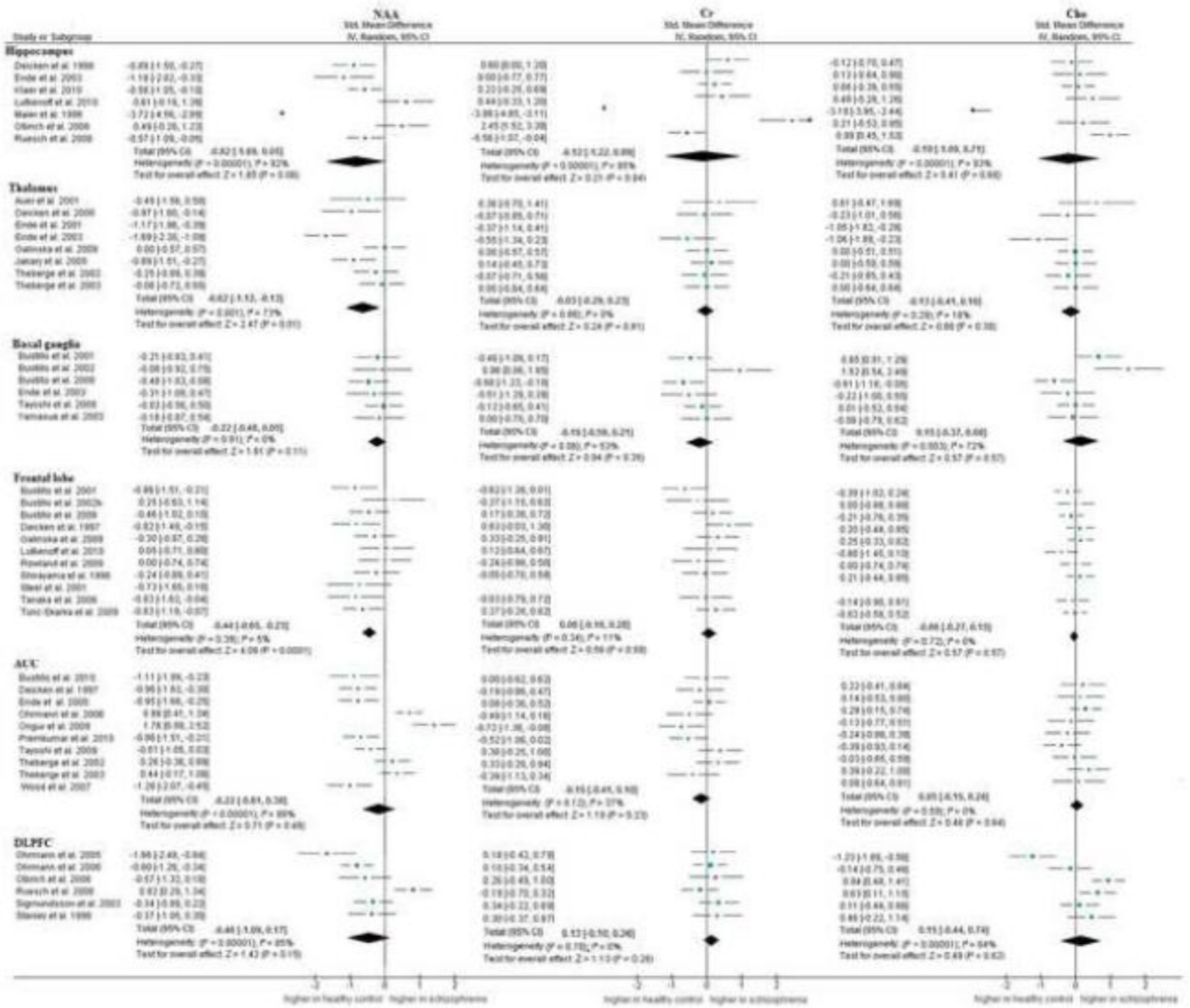


Fig. 1.



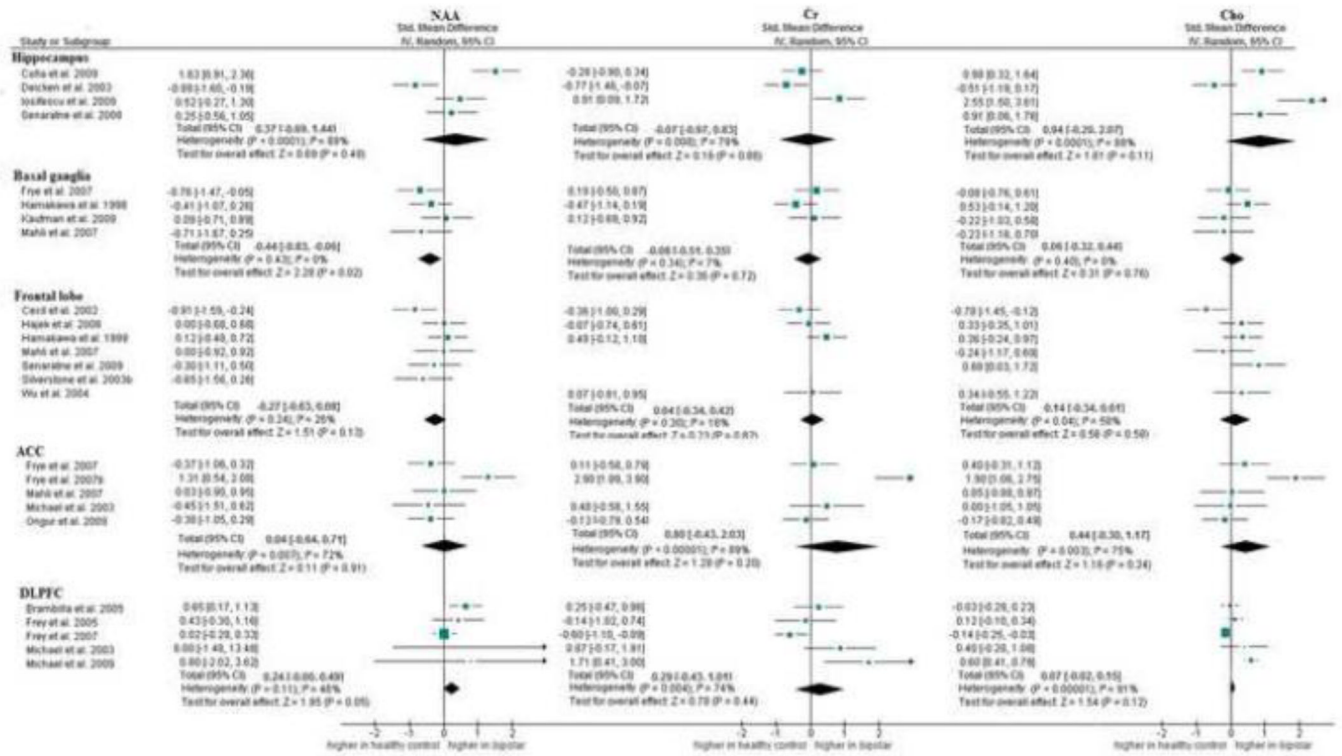


Fig. 2.

Table 1

a: Studies included in systematic review, schizophrenia

Author	Year	n sz/hc	Illness duration	Antipsych otic treatment	MRS	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Ando et al.	2002	7/7	chronic	current	SV	1.5	peak area		lenticular nucleus		incomplete*
Auer et al.	2001	32/17	chronic	current	SV	1.5	institutional units	internal water	thalamus, parietal white matter	X	
Aydin et al.	2007	28/28	chronic	current	SV	1.5	concentration	Internal water	cingulate gyrus	X	
Aydin et al.	2008	14/30	first episode	never	SV	1.5	concentration	internal water	whole brain	X	
Bartha et al.	1997	10/10	first episode	never	SV	1.5	concentration	internal water	medial prefrontal cortex, ACC		
Bartha et al.	1999	11/11	first episode	never/minimal	SV	1.5	concentration	internal water	left mesial temporal lobe	incomplete	incomplete
Bertolino et al.	1996	10/10	chronic	current	MRSI	1.5	peak area		hippocampus, DLPFC, thalamus, putamen, superior temporal gyrus, orbitofrontal cortex, posterior cingulate, ACC, occipital cortex, centrum semiovale, prefrontal white matter		
Bertolino et al.	2000a	13/13	chronic	varies	MRSI	1.5	peak area		DLPFC		
Bertolino et al.	2000b	9/7	chronic	never/off	MRSI	1.5	peak area		DLPFC		
Bertolino et al.	2003b	24/24	first episode	minimal	MRSI	1.5	peak area		hippocampus, DLPFC		incomplete
Bloek et al.	2000	25/19	chronic	current	SV	1.5	peak area		frontal lobe, BG		X
Buckley et al.	1994	28/20	varies	varies	SV	1.5	?concentration		frontal lobe, temporal lobe	X	
Bustillo et al.	2001	19/21	chronic	current	SV	1.5	concentration	internal water	frontal lobe, caudate nucleus	X	
Bustillo et al.	2002a	11/11	first episode	never	SV	1.5	concentration	internal water	frontal lobe, occipital lobe	X	
Bustillo et al.	2002b	10/10	first episode	never/minimal	SV	1.5	concentration	Internal water	caudate nucleus	X	
Bustillo et al.	2008	32/21	early	never/minimal	SV	1.5	concentration	internal water	frontal lobe, occipital lobe, cerebellum, caudate nucleus	X	
Bustillo et al.	2010	14/10	early	never/minimal	SV	4	concentration	internal water	frontal white matter, thalamus, ACC	incomplete	
Callicott et al.	1998	47/66	chronic	current	MRSI	1.5	peak area		mesial temporal cortex, hippocampus		
Callicott et al.	2000a	13/18	chronic	unclear	MRSI	1.5	peak area		superior temporal gyrus, DLPFC, ACC, posterior cingulate, occipital cortex, frontal white matter,		

a: Studies included in systematic review, schizophrenia

Author	Year	n sz/hc	Illness duration	Antipsychotic treatment	MRS	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Callicott et al.	2000b	36/73	chronic	varies	MRSI	1.5	peak area		putamen, hippocampus, thalamus		
Choe et al.	1994	23/10	chronic	never	SV	1.5	peak area		DLPFC, hippocampus		incomplete
Choe et al.	1996	55/20	chronic	off	SV	1.5	peak area		prefrontal white matter		incomplete
Cecil et al.	1999	8/14	first episode	never	SV	1.5	peak area		prefrontal cortex		incomplete
Deicken et al.	1997a	24/15	chronic	current	MRSI	1.5	concentration	internal water	DLPFC, temporal lobe	X	
Deicken et al.	1997b	26/16	chronic	varies	MRSI	1.5	concentration	internal water	frontal lobe	X	
Deicken et al.	1998	30/18	chronic	varies	MRSI	1.5	concentration	internal water	ACC	X	
Deicken et al.	1999	23/18	chronic	current	MRSI	1.5	concentration	internal water	hippocampus	X	X
Deicken et al.	2000	17/10	chronic	current	MRSI	1.5	concentration	internal water	hippocampus	X	
Deicken et al.	2001	20/15	chronic	current	MRSI	1.5	concentration	internal water	thalamus	X	incomplete
Delamillieure et al.	2000a	27/24	unclear	unclear	SV	1.5	peak area	internal water	cerebellum	X	
Delamillieure et al.	2000b	17/21	chronic	current	SV	1.5	peak area		thalamus		incomplete
Delamillieure et al.	2002	17/14	chronic	current	SV	1.5	peak area		medial prefrontal cortex	X*	incomplete
Eluri et al.	1998	12/8	chronic	current	SV	1.5	peak area		prefrontal cortex, thalamus, hippocampus	X*	X*
Ende et al.	2000	19/16	chronic	current	MRSI	1.5	absolute integral values		cerebellum, pons		incomplete
Ende et al.	2001	15/15	chronic	current	MRSI	1.5	absolute integral values		ACC	X	
Ende et al.	2003	13/13	chronic	current	MRSI	1.5	absolute integral values		thalamus	X	
Ende et al.	2005	13/14	chronic	unclear	MRSI	1.5	absolute integral values	internal water	thalamus, hippocampus, BG	X	incomplete
Fannon et al.	2003	11/25	first episode	off	SV	1.5	?concentration	internal water	cerebellum, dentate nucleus		incomplete
Fujimoto et al.	1996	14/12	chronic	current	SV	2	peak area	internal water	prefrontal cortex, hippocampus, BG		incomplete
Fukuzako et al.	1995	15/15	chronic	current	SV	2	peak area		BG		X
Fukuzako et al.	1999	40/40	chronic	current	SV	2	peak area		frontal lobe, medial temporal lobe		X
Fukuzako et al.	2000	64/51	chronic	current	SV	2	peak area		medial temporal lobe		incomplete
Galinska et al.	2009	30/19	first episode	never	SV	1.5	?concentration	internal water	hippocampus		incomplete
Gimenez et al.	2003	11/11	first episode	never	SV	1.5	peak area		frontal lobe, temporal lobe, thalamus	X*	incomplete
									striatum		

a: Studies included in systematic review, schizophrenia

Author	Year	n sz/hc	Illness duration	Antipsychotic treatment	MRS	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Goto et al.	2009	18/18	unclear	current	SV	3			frontal lobe, BG, parieto-occipital lobe		incomplete*
Hagino et al.	2002	13/13	chronic	current	SV	1.5	peak area		inferior frontal cortex, thalamus		X
Heimberg et al.	1998	24/39	chronic	varies	SV	1.5	peak area	internal water	BG, frontal cortex, temporal cortex, thalamus		incomplete
Jakary et al.	2005	22/22	chronic	current	MRSI	1.5	concentration	internal water	thalamus	X	
Klaer et al.	2010	29/44	chronic	current	SV	3	concentration	Internal water	hippocampus	X	
Kegeles et al.	2000	10/10	chronic	varies	SV	1.5	peak area		hippocampus	X*	X
Lim et al.	1998	10/9	chronic	current	MRSI	1.5	concentration	AA, Cho and Cr	grey matter, white matter	X	incomplete
Lutkenhoff et al.	2010	14/13	chronic	unclear	SV	3	concentration	internal water	mesial prefrontal cortex, prefrontal white matter, hippocampus	X	
Maier et al.	1995	25/32	chronic	unclear	SV	1.5	concentration	internal water	hippocampus	X*	
Maier et al.	1996	26/38	chronic	current	SV	1.5	concentration	internal water	hippocampus	X	
Martinez-Granados et al.	2008	49/37	chronic	current	MRSI	1.5	peak area		thalamus		X
Molina et al.	2005	17/15	chronic	current	SV	1.5	peak area		DLPFC		X*
Molina et al.	2006	34/20	chronic	current	SV	1.5	peak area		DLPFC		X*
Molina et al.	2007	11/10	chronic	current	SV	1.5	peak area		DLPFC		X*
Moore et al.	2002	20/20	chronic	current	SV	1.5	peak area		mesial temporal lobe		incomplete
Nasrallah et al.	1994	11/11	chronic	current	SV	1.5	peak area		hippocampus		incomplete
Ohara et al.	2000	10/10	chronic	varies	SV	1.5	peak area		lenticular nucleus		X
Ohrmann et al.	2005	21/21	chronic	current	SV	1.5	concentration	internal water	DLPFC	X	
Ohrmann et al.	2007	20/20	chronic	current	SV	1.5	concentration	internal water	DLPFC	X	
Ohrmann et al.	2008	43/37	chronic	current	SV	1.5	concentration	internal water	ACC, DLPFC	X	
Olbrieh et al.	2008	9/32	first episode	naive	SV	2	concentration	internal water	DLPFC, hippocampus	X	
Omori et al.	2000	20/16	chronic	varies	SV	1.5	peak area		thalamus, frontal lobe		incomplete
Ongur et al.	2010b	21/19	chronic	current	SV	4	concentration	internal water	ACC, parieto-occipital cortex	X	
Ongur et al.	2009	15/22	chronic	current	SV	4	institutional units	internal water	ACC, parieto-occipital cortex	incomplete	
Pae et al.	2004	24/20	varies	naive/off	SV	1.5	peak area		frontal lobe		incomplete

a: Studies included in systematic review, schizophrenia

Author	Year	n sz/hc	Illness duration	Antipsych otic treatment	MRS	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Pajonk et al.	2010	8/8	chronic	current	SV	1.5	peak area	internal water	hippocampus	X	incomplete
Premkumar et al.	2010	30/15	chronic	current	SV	1.5	concentration	internal water	ACC	X	X*
Reid et al.	2010	26/23	chronic	current	SV	3	concentration	internal water	ACC	X	X*
Rowland et al.	2009	20/11	chronic	current	SV	3	concentration	internal water	frontal lobe, inferior parietal lobe	X	X*
Ruesch et al.	2008	29/31	chronic	current	SV	2	concentration	internal water	hippocampus, DLPFC	X*	X*
Sanches et al.	2008	38/38	chronic	current	SV	1.5	peak area	internal water	frontal lobe, ACC, perirhinal fissure	X*	X*
Sarramea et al.	2008	14/15	chronic	current	SV	1.5	peak area	internal water	cingulate gyrus	X	X
Sharma et al.	1992	4/9	chronic	current	SV	1.5	peak area	internal water	basal ganglia, occipital cortex	incomplete	incomplete
Shimizu et al.	2007	19/18	chronic	current	SV	1.5	peak area	internal water	posterior cingulate gyrus	incomplete	incomplete
Shirayama et al.	2010	19/18	chronic	current	SV	3	concentration	internal water	medial prefrontal cortex	X	X
Shioiri et al.	1996	21/21	chronic	current	SV	1.5	peak area	internal water	BG	X	X
Sigmundsson et al.	2003	25/26	chronic	current	SV	1.5	concentration	internal water	DLPFC	X	X*
Stanley et al.	1996	13/25	first episode	never	SV	1.5	concentration	internal water	DLPFC	X*	X*
Steel et al.	2001	10/10	chronic	current	SV	2	institutional units	internal water	frontal lobe	incomplete	incomplete
Szulc et al.	2007	58/21	chronic	current	SV	1.5	?concentration	internal water	frontal lobe, temporal lobe, thalamus	X	incomplete
Tanaka et al.	2006	14/13	chronic	current	SV	1.5	concentration	NAA phantom	frontal lobe	X	incomplete
Tang et al.	2007	40/42	chronic	varies	SV	3	peak area	internal water	DLPFC, medial temporal cortex, occipital cortex	incomplete	incomplete
Tayoshi et al.	2009	30/25	chronic	current	SV	3	concentration	internal water	ACC, BG	incomplete*	X*
Theberge et al.	2002	21/21	first episode	never	SV	4	concentration	internal water	ACC, thalamus	X*	X*
Theberge et al.	2003	21/21	chronic	current	SV	4	concentration	internal water	ACC, thalamus	X*	X*
Theberge et al.	2004a	9/8	chronic	current	SV	4	concentration	internal water	ACC, thalamus	X*	X*
Theberge et al.	2004b	18/18	first episode	never	SV	4	concentration	internal water	ACC, thalamus	incomplete*	incomplete*
Theberge et al.	2007	13/16	first episode	never	SV	4	concentration	internal water	ACC, thalamus	X*	X*
Tibbo et al.	2000	12/12	chronic	current	SV	3	peak area	internal water	cerebellum	X	incomplete
Tunc-Skarka et al.	2009	23/29	chronic	varies	SV	3	concentration	internal water	frontal white matter	X	X*

a: Studies included in systematic review, schizophrenia

Author	Year	n sz/hc	Illness duration	Antipsych otic treatment	MRS	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
van Elst et al.	2005	21/32	chronic	current	SV	2	concentration	internal water	DLPFC, hippocampus	X	
Weber-Fahr et al.	2002	15/15	chronic	current	MRSI	1.5	concentration	internal water	hippocampus		
Wobrock et al.	2008	14/24	chronic	current	SV	1.5	relative concentration		internal capsule		X
Wood et al.	2003	56/21	first episode	current	SV	1.5	peak area		temporal lobe, DLPFC		X
Wood et al.	2007	15/14	chronic	current	SV	3	concentration	internal water	ACC	X*	
Wood et al.	2008	19/19	chronic	current	SV	3	concentration	internal water	temporal lobe	X*	
Wood et al.	2009	30/18	first episode	never/minimal	SV	3	concentration	internal water	temporal lobe	X*	
Yamasue et al.	2003	16/15	chronic	current	SV	1.5	concentration	in vitro spectra	putamen	X	X*
Yurgelun-Todd et al.	1996	16/14	chronic	current	SV	1.5	peak area		mesial temporal lobe		incomplete

b: Studies included in systematic review, bipolar disorder

Author	Year	n bp/hc	Mood state	Mood stabilizing treatment	MRS	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Amaral et al.	2006	13/15	euthymic	current	SV	1.5	ratio		ACC		incomplete
Atmaca et al.	2006	12/12	unclear	unclear	MRSI	1.5	ratio		hippocampus		X
Atmaca et al.	2007	30/10	varies	varies	MRSI	1.5	ratio		hippocampus		X
Bertolino et al.	2003a	17/17	varies	varies	MRSI	1.5	ratio		Thalamus, putamen, hippocampus, inferior frontal gyrus, DLPFC, ACC, posterior cingulate, centrum semiovale, prefrontal white matter, superior temporal gyrus		incomplete
Bhagwagar et al.	2007	16/18	euthymic	off meds	SV	3	ratio		parieto-occipital cortex		incomplete
Brambilla et al.	2005	10/32	unclear	varies	SV	1.5	concentration	internal water	DLPFC	X	X
Cecil et al.	2002	17/21	Manic/mixed	varies	SV	1.5	concentration	internal water	frontal lobe grey and white matter	X	
Colla et al.	2009	21/19	Manic/mixed	current	SV	3	concentration	internal water	hippocampus	X	
Dager et al.	2004	32/26	depressed/ mixed	off meds	MRSI	1.5	concentration	internal water	thalamus, putamen, cingulate gyrus, caudate nucleus, frontal white matter, parietal white matter, occipital lobe		
Deicken et al.	2001	15/15	euthymic	varies	MRSI	1.5	institutional units	internal water	thalamus	X	
Deicken et al.	2003	15/20	euthymic	varies	SV	1.5	institutional units	internal water	hippocampus	X	
Frey et al.	2005	10/10	manic/mixed	varies	SV	1.5	concentration	internal water	DLPFC	X	

b: Studies included in systematic review, bipolar disorder

Author	Year	n bp/hc	Mood state	Mood stabilizing treatment	MRSI	Test a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Frey et al.	2007	32/32	varies	off meds	SV	1.5	concentration	internal water	DLPFC	X	
Friedman et al.	2004	21/12	varies	off meds	MRSI	1.5	concentration	internal water	grey matter, white matter		
Frye et al.	2007a	16/17	manic	current	SV	3	concentration	internal water	ACC, basal ganglia, occipito- parietal white matter	X	incomplete
Frye et al.	2007b	23/12	depressed	varies	SV	1.5	concentration	internal water	ACC	X	incomplete
Hamakawa et al.	1998	18/20	varies	varies	SV	1.5	concentration	external phantom	basal ganglia	X	X
Hamakawa et al.	1999	23/20	varies	varies	SV	1.5	concentration	external phantom	frontal lobe	X	
Hajek et al.	2008	14/21	unclear	unclear	SV	1.5	concentration	internal water	dorsal frontal lobe	X	
Iosifescu et al.	2009	18/10	euthymic	current	SV	4	institutional units	internal water	hippocampus	X	
Kato et al.	1996	19/19	euthymic	varies	SV	1.5	ratio		basal ganglia		incomplete
Kaufman et al.	2009	13/11	varies	current	MRSI	4	concentration	internal water	basal ganglia, whole brain	X	
Michael et al.	2003	8/8	manic	unclear	SV	1.5	concentration	internal water	ACC, DLPFC	X*	
Michael et al.	2009	6/8	varies	current	SV	1.5	concentration	internal water	DLPFC	X	
Mahli et al.	2007	9/9	hypomanic/euthymic	current	SV	1.5	concentration	internal water	ACC, basal ganglia, frontal white matter	incomplete	
Molina et al.	2007	13/10	euthymic	current	SV	1.5	ratio		DLPFC		X*
Moore et al.	2000	12/9	depressed	off meds	SV	1.5	arbitrary units	internal water	frontal lobe, parietal lobe, occipital lobe, temporal lobe		
Moore et al.	2000b	9/14	depressed	current	MRSI	1.5	ratio		ACC		X
Ohara et al.	1998	10/10	euthymic	varies	SV	1.5	ratio		basal ganglia		
Ongur et al.	2008	15/22	manic	current	SV	4	arbitrary units	internal water	ACC, parieto-occipital cortex	X	
Ongur et al.	2009	15/22	manic	current	SV	4	arbitrary units	internal water	ACC, parieto-occipital cortex		
Ongur et al.	2010a	15/20	manic	varies	SV	4	relaxation times	internal water	ACC, parieto-occipital cortex	X	
Port et al.	2008	21/21	varies	off meds	MRSI	3	concentration	internal water	basal ganglia		
Sarramea et al.	2008	15/17	euthymic	varies	SV	1.5	ratio		cingulum		X
Scherk et al.	2008	13/13	euthymic	varies	SV	1.5	ratio		hippocampus, thalamus, putamen		X
Scherk et al.	2009a	30/16	euthymic	current	SV	1.5	ratio		putamen		X
Scherk et al.	2009b	33/29	euthymic	current	SV	1.5	ratio		ACC, DLPFC		X
Senaratne et al.	2009	12/12	euthymic	current	SV	3	concentration	internal water	hippocampus, orbitofrontal lobe, occipital lobe	incomplete	
Sharma et al.	1992	4/9	manic	current	SV	1.5	ratio		basal ganglia, occipital cortex		X
Silverstone et al.	2002	14/18	euthymic	current	SV	3	ratio	external standard	temporal lobe		incomplete

b: Studies included in systematic review, bipolar disorder

Author	Year	n bp/hc	Mood state	Mood stabilizing treatment	MRS	Tesl a	Data processing	use of reference	Region studied	Absolute value data available	Ratio data available
Silverstone et al.	2003	25/18	euthymic	current	SV	3	ratio		frontal lobe, temporal lobe		incomplete
Silverstone et al.	2003	9/11	euthymic	current	SV	3	concentration	internal water	frontal lobe, temporal lobe	incomplete	
Winsberg et al.	2000	20/20	euthymic	off meds	SV	1.5	ratio		DLPPC		X
Wu et al.	2004	25/18	euthymic	current	SV	3	concentration	internal water	frontal lobe, temporal lobe	incomplete	

* data obtained from author directly

X: data is available and included in meta-analysis. Studies with no X have not reported actual values

Incomplete: only some data available

Table 2

a: Meta-analysis with ratio data, schizophrenia						
Region	number of studies	Metabolite	SMD [95% CI]	p value	Heterogeneity (I ²)	
Hippocampus	8	NAA/Cr	-0.72 [-1.20, -0.25]	<0.01	74%	
	5	Cho/Cr	-0.28 [-0.54, -0.02]	0.03	0%	
Thalamus	9	NAA/Cr	-0.37 [-0.58, -0.17]	<0.01	6%	
	6	Cho/Cr	-0.02 [-0.34, 0.30]	0.91	42%	
Basal ganglia	8	NAA/Cr	-0.16 [-0.46, 0.13]	0.28	32%	
	6	Cho/Cr	0.13 [-0.22, 0.48]	0.47	37%	
Frontal lobe	16	NAA/Cr	-0.22 [-0.39, -0.06]	<0.01	0%	
	13	Cho/Cr	0.09 [-0.24, 0.41]	0.61	68%	
DLPFC	3	NAA/Cr	0.14 [-0.72, 1.00]	0.75	86%	
	2	Cho/Cr	-0.15 [-0.73, 0.42]	0.60	58%	
Temporal lobe	7	NAA/Cr	-0.64 [-1.09, -0.19]	<0.01	77%	

b: Meta-analysis with ratio data, bipolar disorder						
Region	number of studies	Metabolite	SMD [95% CI]	p value	Heterogeneity (I ²)	
Hippocampus	4	NAA/Cr	-0.96 [-1.37, -0.55]	<0.01	0%	
	3	Cho/Cr	-0.37 [-0.84, 0.11]	0.13	0%	
Basal ganglia	7	NAA/Cr	-0.02 [-0.55, 0.51]	0.95	69%	
	6	Cho/Cr	-0.04 [-0.62, 0.54]	0.89	70%	
ACC	5	NAA/Cr	-0.59 [-1.18, 0.01]	0.06	75%	
	6	Cho/Cr	0.00 [-0.00, 0.01]	0.52	43%	
DLPFC	6	NAA/Cr	-0.03 [-0.42, 0.36]	0.88	49%	
	5	Cho/Cr	0.05 [-0.23, 0.33]	0.74	6%	