ORIGINAL ARTICLE

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A Comparison between the Antimicrobial Effects of Triple Antibiotic Paste and Calcium Hydroxide Against *Entrococcus Faecalis*

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Introduction: The purpose of this study was to determine the *in vitro* antimicrobial ability against *Entrococcus (E.) faecalis* of triple antibiotic paste and its components compared with calcium hydroxide mixtures.

Materials and Methods: An agar well diffusion assay and MIC method were used to determine the efficacy of the experimental medicaments in removing E. faecalis. Medicaments were divided into 9 groups; triple antibiotic powder with saline or chlorhexidine, metronidazole, ciprofloxacin, minocycline antibiotics were also separately tested (with normal saline), and $Ca(OH)_2$ (plus normal saline or 2% chlorhexidine). These medicaments were evaluated at four concentrations of 25, 50, 100 and 200 µgram per mL in an agar well diffusion test. The diameters of the growth inhibition zones for each group were recorded and compared. The minimum inhibitory concentrations (MIC) of tested medicaments that are required to kill E. faecalis were also determined. The differences between groups were analyzed by Kruskal-Wallis and Mann-Whitney U tests.

Results: The largest inhibition zones were observed for the triple antibiotic mixture/saline, triple antibiotic mixture/2% chlorhexidine and minocycline/saline, and the smallest for Ca(OH)₂/saline, Ca(OH)₂/2% chlorhexidine. Concentration increases produced greater antibacterial effects in all groups. The MIC determination method showed similar results.

Conclusion: The results suggest that the triple antibiotic paste with either 2% chlorhexidine or normal saline would be the preferred medicament against *E. faecalis* and, among its three components, minocycline has the greatest antibacterial effect.

Keywords: Antibiotics; Antimicrobial Efficacy; Calcium Hydroxide; *Entrococcus Faecalis*; Intracanal Medicaments

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Introduction

There have been many studies that prove that bacteria is the major cause of pulpal and periapical diseases [1-4]. Because of the complex nature of the root canal system and the presence of many inaccessible areas, a combination of mechanical instrumentation and irrigation is necessary to decrease the amount of bacteria/micro-organisms in the root canal system [5]. However, chemo-

mechanical preparation is often not enough, and many bacteria may remain in the root canal system [6-8].

The use of inter-appointment medicaments may reduce microorganisms that remain in root canals. Ca(OH)₂ is widely used in endodontics, but this common endodontic medicament does not sufficiently affect *Entrococcus* (E.) faecalis, the most important bacterium in resistant infections [9-11]. E. faecalis is an enteric

Table 1. The details of medicaments tested against *E. faecalis*

Groups	Description
1	Triple antibiotic paste + Normal Saline
2	Triple antibiotic paste + 2% Chlorhexidine
3	Minocycline + Normal Saline
4	Metronidazole + Normal Saline
5	Ciprofloxacin + Normal saline
6	Ca(OH) ₂ + Normal Saline
7	Ca(OH) ₂ + Chlorhexidine 2%
8	Talc powder + Chlorhexidine 2%
9	Talc powder + Normal Saline

facultative gram positive bacterium which can grow independently in the root canal without the assistance of other bacteria This microorganism has the ability to invade and live in the dentinal tubules [4-12].

Recently, triple antibiotic paste (a mixture of metronidazole, ciprofloxacin and minocycline) has been used as an intracanal medicament for disinfecting the root canal during regenerative procedures [13-16]. Bose *et al.*, in a retrospective study, showed that Ca(OH)₂ and triple antibiotic paste, because of their antibacterial properties, can aid further development of the pulp dentin complex when used as an intracanal medicament in immature necrotic teeth [17].

Sato *et al.* showed triple antibiotic paste can destroy the bacteria in deep areas of the root canal system [18]. In a study on dogs, the results indicated the effectiveness of triple antibiotic paste in the disinfection of immature teeth with apical periodontitis [19]. To date, there is insufficient documentation regarding the antimicrobial effect of triple antibiotic paste in eliminating *E. faecalis*. The purpose of this *in vitro* study was to provide further insight into the efficacy of the antibiotic paste in eradicating *E. faecalis*.

Materials and Methods

Agar well diffusion assay method

The medicaments tested were in powdered form. They were as follows: the antibiotic mixture, the individual components of the triple antibiotic mixture-metronidazole (Flagyl, Winthrop Pharmaceuticals, UK), ciprofloxacin (Bayer plc, UK) and minocycline (Expanscience Laboratories, Paris, France), Ca(OH)₂ powder (Golchay, Tehran, Iran) and talc powder (Royal Minerals, India). The triple antibiotic mixture, Ca(OH)₂ and talc powder were mixed with either

normal saline (NaCl 0.9%, Darupakhsh, Tehran, Iran) or 2% chlorhexidine (Consepsis, Ultradent Inc., South Jordan, UT. USA). Each component of the triple antibiotic paste was just mixed with normal saline. Talc powder mixed with normal saline served as the control group, because a pilot study showed it has no antibacterial activity against *E. faecalis* (Table 1).

All the medicaments were tested at concentrations of 25, 50, 100 and 200 µgram per mL in normal saline/2% chlorhexidine.

The triple antibiotic paste was prepared for each group with identical amount of the three antibiotic powders (mg) and then mixed with 1 mL normal saline or 2% chlorhexidine.

The bacterium tested was *E. faecalis* (ATCC 11700). Pure *E. faecalis* was grown on BHI agar plates (Himedia Laboratories, Mumbai, India). The microorganisms were inoculated into tubes containing 5 mL 0.9% sterile saline solution. The suspension was adjusted by using McFarland tubes to match the turbidity to 1.5×108 cfu/mL. The BHI agar plates were flooded with the test suspension. Then 4 wells (4 mm depth ×6 mm in diameter) were cut in the agar for each concentration of medicaments. A sterile spatula was used to place the pastes into each well.

The plates were then incubated at 37°C under appropriate atmospheric conditions (80% N2, 10% CO2, 10% H2) for 7 days under anaerobic conditions in a CO2 incubator (Mart Microbiology B. V., Netherlands). The diameters of the zones of bacterial growth inhibition around the wells containing the test substances were then recorded after the period of incubation. The inhibitory zone was determined in millimeters by measuring the shortest distance between the outer margin of the well and initial microbial growth.

Each experiment was performed six times and the means and standard deviations of the inhibitory zones were calculated. The differences between groups were analysed by Kruskal-Wallis and Mann-Whitney U tests. P values less than 0.05 were considered as significant.

MIC determination method

A reference bacterial strain *E. faecalis* (ATCC 11700) obtained from the American type culture collection was used. The bacteria was cultured in Brain heart infusion (BHI agar) culture medium, stored in a tube and incubated at

Table2. The means (SD) of agar well diffusion assay-(mm) after 7 days from 9 separate groups

Group/concentration	25 μg	50 μg	100 μg	200 μg
group 1	33.83 (0.40) ^a	35.66 (0.51) ^a	36.50 (0.54) ^a	38.16 (0.75) ^a
group 2	35.66 (0.56) a	36.66 (0.51) ^a	37.50 (1.24) ^a	39.66 (1.03 ^a
group 3	34.83 (0.75) ^a	35.83 (0.75) ^a	37.83 (0.75) ^a	39.33 (0.81) ^a
group 4	18.83 (0.75) ^a	$20.83 (0.75)^a$	26.16 (0.93) ^a	29.50 (0.54) ^a
group 5	26.66 (1.50) a	28.66 (0.51) ^a	29.83 (0.75) ^a	$34.00(0.63)^a$
group 6	0.00(0.00)	0.00(0.00)	4.33 (4.75)	9.00 (4.47) a
group 7	0.00(0.00)	0.00(0.00)	$6.66(5.16)^a$	$11.66 (1.03)^a$
group 8	0.00(0.00)	0.00(0.00)	1.66 (4.08)	6.50 (5.04)
group 9	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)

 $[\]overline{a}$: Indicates a significant difference (P < 0.05) compared with the control group (group 9) according to Mann-Withney U test

37°C for 7 days. Microbial cells were resuspended in saline to obtain a final concentration of 1.5×108 cfu/mL, similar to that of tube No. 0.5 on the MacFarland scale. A total of 0.1 mL of each pure suspension was used to obtain a mixture of the test microorganisms.

The experimental medicaments (groups 1–7) were prepared similar to the agar diffusion test. To determine the MIC, six fold dilutions were made for each group.

After dilution of the medicaments, inoculums of 0.09 mL from the experimental suspension were added. These were incubated at 37°C for 7 days.

Microbial growth was evaluated by two methods: turbidity of the culture medium and subculture in a specific nutrient broth. In the latter method, the samples were transferred from tubes with no bacterial growth to nutrient agar plates by means of an inoculating loop, and lack of bacterial growth was confirmed. The lowest concentration at which no bacterial growth was seen was determined to be the MIC of that medicament.

The MIC for each medicament was determined and the experiments were repeated four times. Then the mean values and standard deviations were calculated. Comparisons were made among the MIC values of different medicaments using the Kruskal-Wallis test and Mann-Whitney as post-hoc test.

Results

Agar well diffusion assay method

The means and standard deviations of the diameters of the growth inhibition zones for each concentration of the preparations are presented in Table 2.

The range of inhibitory values between experimental groups varied broadly and showed significant differences (P<0.05).

Each experimental group was compared to the talc/saline control (group 9) separately. Overall, triple antibiotic paste/saline, triple antibiotic paste/2% chlorhexidine and minocycline/saline (groups 1, 2, 3) had the largest zones of growth inhibition in the well diffusion assay at all concentrations (Figure 1). Metronidazole and ciprofloxacin (groups 4, 5) had significant inhibitory effects on E. faecalis at all concentrations compared to the control group (P<0.05).

Ca(OH)₂/saline was effective against *E. faecalis* only at 200 μ gm/mL and Ca(OH)₂/2% chlorhexidine showed inhibitory effect even at concentrations of 100 and 200 μ gm/mL (P<0.05).

Talc/2% chlorhexidine showed minimal inhibitory effects at concentrations of 100 and 200 μ gm/mL, which were not significantly different from the control group (P<0.05).

Increasing the concentrations of the tested medicaments produced inhibition zones with larger diameters in all groups, (P<0.05); *i.e* they had greater inhibitory effect on E. faecalis.

MIC determination method

The minimum inhibitory concentration (MIC) was defined as the lowest concentration able to inhibit any visible bacterial growth.

MIC's of the tested medicaments required to kill the bacterium are shown in Table 3.

The most effective medicament against E. faecalis was triple antibiotic powder/normal saline, with a MIC equal to 77.5 µgram per mL. Triple antibiotic powder/ 2% chlorhexidine and minocycline/normal saline had similar MIC

Table 3. The means (SD) of MICs of tested medicaments

Group	MIC(μg/mL)
group 1	77.5(22.50)
group 2	325(225.00)
group 3	325(225.00)
group 4	550(259.80)
group 5	3250(2250.00)
group 6	
group 7	195000(135000.00)

values, equivalent to 325 µgram per mL.

The least effective was group 7 (Ca(OH)₂/2% chlorhexidine), which had a MIC equal to 195000 µgram per mL. The results also showedthat group 6 (Ca(OH)₂+normal saline) was not active against *E. faecalis* even at the highest concentration used in this study.

Statistical analysis showed a significant difference between group 7 [Ca(OH)₂/2% chlorhexidine 2%] and the other medicaments. Also, group 5 (ciprofloxacin/normal saline) was significantly less effective than groups 1-4.

Discussion

The agar diffusion test we used in this study is useful for evaluating and comparing the in vitro antimicrobial activities of medicaments before performing more advanced tests; many studies have used this method for evaluations of antibacterial effects of various endodontic materials.[20-22]. The results obtained from this test must be interpreted with caution, as this assay may not demonstrate the full clinical potential of the material being tested. The MIC test is also a research tool to determine the in vitro activity of new medicaments and antimicrobials. Asna ashari et al. used two techniques, including zone of inhibition (ZI) and minimum inhibitory concentration (MIC), to evaluate the antimicrobial effects of MTAD, sodium hypochlorite (NaOCl) and their combination on endodontic microorganisms. Similarly, in the current study we used these methods to compare the antimicrobial effects of intracanal medicaments [22].

The bacterial species *E. faecalis* was selected as representing an organism commonly isolated from the root canals of teeth that have been previously root filled [23]. There have been several studies which applied *E. faecalis* as a target microorganism to

evaluate the effects of antibacterial agents [24-26]. In fact, previously treated failed teeth are nine times more likely to be infected with *E. faecalis* than primary endodontic infections [27]. *E. faecalis* is also resistant to calcium hydroxide, a commonly used intracanal medicament, especially when a high pH is not maintained [28-30].

Some studies have shown promising effects of chlorhexidine in a 2% gel or liquid form to reduce or completely eliminate E. faecalis from the root canal space and dentinal tubules [31-33]. Turk et al. showed calcium hydroxide mixed with 2% chlorhexidine digluconate was more effective against E. faecalis than calcium hydroxide mixed with other vehicles [34]. Therefore we utilised chlorhexidine as one of the vehicles for this study. To ascertain which substance had the highest antibacterial effect, each component was independently mixed with normal saline. The results showed the triple antibiotic powder, either mixed with normal saline or 2% chlorhexidine, produced the largest zone of inhibition against E. faecalis. The minocycline/normal saline combination was very effective activity against E. faecalis. Although metronidazole/normal saline and ciprofloxacin/normal saline showed antibacterial efficacies significantly higher than the control group, their ability to kill E. faecalis was less than minocycline and the triple antibiotic paste, according to both the agar diffusion and MIC tests. It can be concluded that minocycline is the most effective component of the triple antibiotic paste against E. faecalis, as adding the two other antibiotics to the mixture did not considerably increase the diameter of its zones of inhibition.

Pinheiro *et al.* reported most of the 21 microbial isolates from root canals of filled teeth with persistent periapical lesions were susceptible to tetracycline and doxycycline, which is supports the outcome of the present study that showed minocycline is the most effective component against *E. faecalis* [35]. Antibacterial activity of chlorhexidine may be attributed to its chemical characteristics.

In the present study, the antimicrobial effect of the triple antibiotic paste and its three components against *E. faecalis* were compared to calcium hydroxide, either mixed with

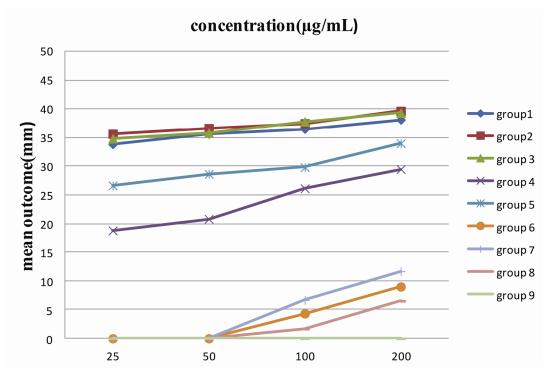


Figure 1. Comparison of groups 1-9 in different concentrations

normal saline or 2% chlorhexidine. Considering the low antibacterial capability of talc powder/chlorhexidine and Ca(OH)₂ chlorhexidine, and the high ability of triple antibiotic powder/normal saline in this study, we conclude the ability of the former is related to its mixed antibiotic contents, not to the vehicle's properties.

The results of the present study also indicated that calcium hydroxide in normal saline or 2% chlorhexidine had some antibacterial efficacy against *E. faecalis*, but only at concentrations of 100 and 200 µgram per mL. The zones of inhibition generated by groups containing calcium hydroxide were much smaller than the zones of the antibiotic-containing groups, concurring with previous literature that calcium hydroxide is not an effective intracanal medicament when root canals are infected by *E. Faecalis* [28-30].

The study showed that the higher the concentrations (25 µgram per mL to 200 µgram per mL) of antibiotics or calcium hydroxide, the greater its' antibacterial efficacy. A similar trend was reported by Blanseet *et al.* for different mixtures of calcium hydroxide [36].

E. faecalis is the most frequent species in failed root canal therapy cases [9-11]; therefore

the findings of the present study support the use of triple antibiotic paste as a potential intracanal medicament in the treatment of endodontic failures. It is noteworthy to mention that triple antibiotic paste is utilised in primary infections of open apex teeth, in which *E. faecalis* is not a prevalent bacteria [13-15,37].

In vitro tests may not show the full clinical potential of a tested material, as dentin can interact with endodontic disinfectants [38]. We suggest future investigations repeat the efficacy of the triple antibiotic paste in reducing CFUs in a dentin diffusion model, using clinically relevant periods of disinfection and deep testing of dentin.

Conclusion

Under the limitations of this study, the triple antibiotic paste is very effective against *E. faecalis* and can be considered as a more powerful root canal medicament compared to calcium hydroxide pastes. The results also show that minocycline is the most effective component of the triple antibiotic paste.

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Conflict of Interest: 'none declared'.

References

- [1] Kakehashi S, Stanley HR, Fitzgerald RJ. The Effects of Surgical Exposures of Dental Pulps in Germ-Free and Conventional Laboratory Rats. Oral Surg Oral Med Oral Pathol. 1965;20(3):340-9.
- [2] Moller AJ, Fabricius L, Dahlen G, Sundqvist G, Happonen RP. Apical periodontitis development and bacterial response to endodontic treatment. Experimental root canal infections in monkeys with selected bacterial strains. Eur J Oral Sci. 2004;112(3):207-15.
- [3] Dahlen G, Fabricius L, Heyden G, Holm SE, Moller AJ. Apical periodontitis induced by selected bacterial strains in root canals of immunized and nonimmunized monkeys. Scand J Dent Res. 1982;90(3):207-16.
- [4] Fabricius L, Dahlen G, Holm SE, Moller AJ. Influence of combinations of oral bacteria on periapical tissues of monkeys. Scand J Dent Res. 1982;90(3):200-6.
- [5] Abbott PV. Medicaments: aids to success in endodontics. Part 1. A review of the literature. Aust Dent J. 1990;35(5):438-48.
- [6] Peters LB, van Winkelhoff AJ, Buijs JF, Wesselink PR. Effects of instrumentation, irrigation and dressing with calcium hydroxide on infection in pulpless teeth with periapical bone lesions. Int Endod J. 2002;35(1):13-21.
- [7] Bystrom A, Sundqvist G. Bacteriologic evaluation of the efficacy of mechanical root canal instrumentation in endodontic therapy. Scand J Dent Res. 1981;89(4):321-8.
- [8] Orstavik D, Kerekes K, Molven O. Effects of extensive apical reaming and calcium hydroxide dressing on bacterial infection during treatment of apical periodontitis: a pilot study. Int Endod J. 1991;24(1):1-7.

- [9] Siqueira JF, Jr., Lopes HP. Mechanisms of antimicrobial activity of calcium hydroxide: a critical review. Int Endod J. 1999;32(5):361-9.
- [10] Portenier I, Haapasalo H, Orstavik D, Yamauchi M, Haapasalo M. Inactivation of the antibacterial activity of iodine potassium iodide and chlorhexidine digluconate against Enterococcus faecalis by dentin, dentin matrix, type-I collagen, and heat-killed microbial whole cells. J Endod. 2002;28(9):634-7.
- [11] Orstavik D. Root canal disinfection: a review of concepts and recent developments. Aust Endod J. 2003;29(2):70-4.
- [12] Love RM. Enterococcus faecalis--a mechanism for its role in endodontic failure. Int Endod J. 2001;34(5):399-405.
- [13] Nosrat A, Seifi A, Asgary S. Regenerative endodontic treatment (revascularization) for necrotic immature permanent molars: a review and report of two cases with a new biomaterial. J Endod. 2011;37(4):562-7.
- [14] Thibodeau B, Teixeira F, Yamauchi M, Caplan DJ, Trope M. Pulp revascularization of immature dog teeth with apical periodontitis. J Endod. 2007;33(6):680-9.
- [15] Petrino JA, Boda KK, Shambarger S, Bowles WR, McClanahan SB. Challenges in regenerative endodontics: a case series. J Endod. 2010;36(3):536-41.
- [16] Jung IY, Lee SJ, Hargreaves KM. Biologically based treatment of immature permanent teeth with pulpal necrosis: a case series. J Endod. 2008;34(7):876-87.
- [17] Bose R, Nummikoski P, Hargreaves K. A retrospective evaluation of radiographic outcomes in immature teeth with necrotic root canal systems treated with regenerative endodontic procedures. J Endod. 2009;35(10):1343-9.
- [18] Sato I, Ando-Kurihara N, Kota K, Iwaku M, Hoshino E. Sterilization of infected root-canal dentine by topical application of a mixture of ciprofloxacin, metronidazole and minocycline in situ. Int Endod J. 1996;29(2):118-24.
- [19] Windley W, 3rd, Teixeira F, Levin L, Sigurdsson A, Trope M. Disinfection of immature teeth with a triple antibiotic paste. J Endod. 2005;31(6):439-43.
- [20] Asgary S, Akbari Kamrani F, Taheri S. Evaluation of antimicrobial effect of MTA, calcium hydroxide, and CEM cement. Iran Endod J. 2007;2(3):105-9.

- [21] Razmi H, Ashofteh Yazdi K, Jabalameli F, Parvizi S. Antimicrobial effects of AH26 sealer/antibiotic combinations against Enterococcus faecalis. Iran Endod J. 2008;3(4):107-12.
- [22] Asna Ashari M, Fayaz F, Moezzi Ghadim N, Alim Marvesti L, Mehrabi Y. Evaluation of the antimicrobial effects of MTAD, NaOCl against selected endodontic pathogens. Iran Endod J. 2009;4(2):63-68.
- [23] Siqueira JF, Jr. Aetiology of root canal treatment failure: why well-treated teeth can fail. Int Endod J. 2001;34(1):1-10.
- [24] Ahangari Z, Mohammad Samiee D, Yolmeh MA. Antimicrobial activity of three root canal irrigants on Enterococcus Faecalis: An in vitro study. Iran Endod J. 2008;3(2):33-37.
- [25] Sharifian MR, Shokouhinejad N, Aligholi M, Emaneini M, Alizadeh M. Antibacterial substantivity of Carvacrol and sodium hypochlorite in infected bovine root dentin. Iran Endod J. 2009;4(2):45-8.
- [26] Mehrvarzfar P, Akhavan H, Rastgarian H, Akhlagi NM, Soleymanpour R, Ahmadi A. An in vitro comparative study on the antimicrobial effects of bioglass 45S5 vs. calcium hydroxide on Enterococcus faecalis. Iran Endod J. 2011;6(1):29-33.
- [27] Rocas IN, Siqueira JF, Jr., Santos KR. Association of Enterococcus faecalis with different forms of periradicular diseases. J Endod. 2004;30(5):315-20.
- [28] Lin YH, Mickel AK, Chogle S. Effectiveness of selected materials against Enterococcus faecalis: part 3. The antibacterial effect of calcium hydroxide and chlorhexidine on Enterococcus faecalis. J Endod. 2003;29(9):565-6.
- [29] McHugh CP, Zhang P, Michalek S, Eleazer PD. pH required to kill Enterococcus faecalis in vitro. J Endod. 2004;30(4):218-9.

- [30] Tronstad L, Andreasen JO, Hasselgren G, Kristerson L, Riis I. pH changes in dental tissues after root canal filling with calcium hydroxide. J Endod. 1981;7(1):17-21.
- [31] Vahdaty A, Pitt Ford TR, Wilson RF. Efficacy of chlorhexidine in disinfecting dentinal tubules in vitro. Endod Dent Traumatol. 1993;9(6):243-8.
- [32] Gomes BP, Souza SF, Ferraz CC, Teixeira FB, Zaia AA, Valdrighi L, Souza-Filho FJ. Effectiveness of 2% chlorhexidine gel and calcium hydroxide against Enterococcus faecalis in bovine root dentine in vitro. Int Endod J. 2003;36(4):267-75.
- [33] Basrani B, Santos JM, Tjaderhane L, Grad H, Gorduysus O, Huang J, Lawrence HP, Friedman S. Substantive antimicrobial activity in chlorhexidine-treated human root dentin. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2002;94(2):240-5.
- [34] Turk BT, Sen BH, Ozturk T. In vitro antimicrobial activity of calcium hydroxide mixed with different vehicles against Enterococcus faecalis and Candida albicans. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2009;108(2):297-301.
- [35] Pinheiro ET, Gomes BP, Drucker DB, Zaia AA, Ferraz CC, Souza-Filho FJ. Antimicrobial susceptibility of Enterococcus faecalis isolated from canals of root filled teeth with periapical lesions. Int Endod J. 2004;37(11):756-63.
- [36] Blanscet ML, Tordik PA, Goodell GG. An agar diffusion comparison of the antimicrobial effect of calcium hydroxide at five different concentrations with three different vehicles. J Endod. 2008;34(10):1246-8.
- [37] Lenzi R, Trope M. Revitalization procedures in two traumatized incisors with different biological outcomes. J Endod. 2012;38(3):411-4.
- [38] Haapasalo M, Qian W, Portenier I, Waltimo T. Effects of dentin on the antimicrobial properties of endodontic medicaments. J Endod. 2007;33(8):917-25.