

Cultivation and Characterization of Cornea Limbal Epithelial Stem Cells on Lens Capsule in Animal Material-Free Medium

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Abstract

A simple, reproducible, animal-material free method for cultivating and characterizing cornea limbal epithelial stem cells (LESCs) on human lens capsule (LC) was developed for future clinical transplantation. The limbal tissue explants (2×2×0.25 mm) were harvested from 77 cadavers and expanded *ex vivo* on either cell culture plates or LC in medium containing human serum as the only growth supplement. Cell outgrowth at the edge of the explants was observed within 24 hours of cultivation and achieved viable outgrowth (>97% viability as measured by MTT assay and flow cytometry) within two weeks. The outgrowing cells were examined by genome-wide microarray including markers of stemness (p63 α , ABCG2, CK19, Vimentin and Integrin α 9), proliferation (Ki-67), limbal epithelial cells (CK 8/18 and 14) and differentiated cornea epithelial cells (CK 3 and 12). Immunostaining revealed the non-hematopoietic, -endothelial and -mesenchymal stem cell phenotype of the LESCs and the localization of specific markers *in situ*. Cell adhesion molecules, integrins and lectin-based surface carbohydrate profiling showed a specific pattern on these cells, while colony-formation assay confirmed their clonal potency. The LESCs expressed a specific surface marker fingerprint (CD117/c-kit, CXCR4, CD144/VE-Cadherin, CD146/MCAM, CD166/ALCAM, and surface carbohydrates: WGA, ConA, RCA, PNA and AIL) which can be used for better localization of the limbal stem cell niche. In summary, we report a novel method combining the use of a medium with human serum as the only growth supplement with LC for cultivating, characterizing and expanding cornea LESCs from cadavers or alternatively from autologous donors for possible treatment of LESCs deficiency.

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Introduction

Cornea limbal epithelial stem cells (LESC) have been described to exist within special niches located at the basal cell layer of the limbal epithelium at the corneo-scleral junction [1–4]. The role of the LESCs is in renewal of healthy [5,6] and regeneration of injured [7,8] corneal epithelium. Besides infection and injury, corneal diseases can also affect the LESCs and their renewal potency resulting in serious visual problems. Any imbalance of the wound-healing process can result in an increased corneal vascularization and decreased transparency [5,9,10].

The recovery of the corneal epithelium arises mainly from the LESCs continually giving rise to transient amplifying cells (TACs) which migrate centripetally and superficially while becoming more and more differentiated [3,11,12]. Some proofs of existence of

corneal epithelial stem cells centripetal to the limbus have also been reported based on their colony-forming potential [13].

Due to lack of corneal donor tissue or decreased chance for graft survival after penetrating keratoplasty, an autologous or homologous expansion of human LESCs has been proposed in cases of limbal stem cell deficiency (LSCD) [10]. Many attempts have been made for using human amniotic membrane (HAM) and other bioscaffolds as carriers for transplanting LESCs [14–19]. The HAM has some obvious biodegradable and immunosuppressive properties during transplantation, although its thickness and variable transparency have been used as a counter argument for its use. In addition, using feeder cells or complex media containing growth factors and animal materials has raised the safety issue of transferring prion or yet unknown diseases [20,21]. The growth of LESCs using animal-material free medium on HAM has only recently been described for transplantation purposes [22].

The use of human lens capsule (LC) as a bioscaffold for growing limbal stem-like cells has been introduced earlier [14]. We explored use of LC for *ex vivo* cultivation of LESC in human serum as the only growth supplement due to its high transparency and small thickness, as well as low immune- or xenogenic factors insulating ability [23]. Cornea LESC grown under such conditions were characterized by genome-wide microarray and immunostaining for markers of stemness (tumor/transformation-related protein 63 (p63/TP63), ATP-binding cassette sub-family G member 2 (ABCG2), cytokeratin (CK/KRT) 19, Vimentin (Vim) and Integrin (Itg/ITG) $\alpha 9$), proliferation- (Ki-67/MKI67), limbal epithelial- (CK 8/18 and 14) and differentiated corneal epithelial- (CK 3 and 12) markers [4,22,24]. Phenotyping covered the exclusion of hematopoietic, endothelial and mesenchymal stem cell markers [25] as well as surface Itgs, cell-adhesion molecules (CAMs) and broad lectin-based surface carbohydrate marker profiling [4,26]. We propose a simple, reproducible, animal-material free method for *ex vivo* expansion and characterization of cornea auto- or allografts of LESC on LC for the treatment of LSCD.

Results

Cultivation and Viability of Human Cornea LESC

Human cornea limbal tissue explants were harvested from cadavers within 12 hours from biologic death and cultured on either cell culture plates or human LC. Cell culture-plated grafts showed cell outgrowth with epithelial morphology and intact cytoskeleton within 24 hours of cultivation (**Figure 1 A1–2**). Cell proliferation was observed over another 2 weeks till it reached confluence. Similarly, grafts grown on human LC showed cell outgrowth (**Figure 1 A3–4**) and formed stratified epithelial layer within 7 days of cultivation (**Figure 1 B1–2**). Under both growth conditions and use of medium containing human serum as the only growth supplement, the cell viability of the outgrowing LESC was >97% at the two checkpoints - 7 and 14 days of cultivation, as measured by the 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay (data not shown). Accordingly, the percentage of early apoptotic (<2% annexin-Fluorescein Isothiocyanate (FITC)⁺) and late apoptotic (<1% annexin-FITC⁺/Propidium iodide⁺) cells remained low under both growth conditions (**Figure 1C**) up to day 14.

Transcriptional Profiling in Human Cornea LESC

Transcriptional profiling of the LESC was carried out using a microarray in three different donors. Intensity profiles of the log₂ transformed signal values of the 28869 transcripts were obtained, out of which 955 and 875 transcripts had a more than 2 fold change (FC) increase and decrease in expression, respectively ($n = 3$, $p < 0.01$), between the cultured LESC and differentiated corneal epithelial cells. This indicates a relatively high transcriptional difference between the two cell types. **Figure 2**, **Table 1** and **Table S1** show the heatmap and the functional clustering of 67 genes selected on the basis of their high or low FC or previously documented relation to LESC ($n = 3$, $p < 0.01$). These genes were mostly involved in ion-, nucleotide- or protein binding, as well as receptor or enzyme activities. Among the general epithelial markers, limbal epithelium recognizing markers (KRT8/KRT18 and KRT14) could be distinguished, along the ones specific for differentiated corneal epithelium (KRT3/12). KRT8 and KRT14 showed similar or slightly higher expression levels in the limbal tissue-derived cells as compared to the differentiated control epithelium (FC: 4.0 and 1.9, respectively) indicating the commitment of LESC towards the corneal epithelium lineage. Mean-

while, the specific differentiated corneal epithelial markers KRT3 and KRT12 decreased expression (FC: -31.0 and -5.8, respectively), probably due to an earlier differentiation state or preserved multipotency of these cells (**Table 1**). Higher expression of at least two orders of magnitude was found in the putative stem cell markers of LESC (KRT19 (FC: 6.0) and VIM (FC: 4.4) compared to the differentiated control epithelial cells, strengthening their stem-like character. The high proliferation capacity of the cultured LESC was also confirmed by higher expression of the proliferation-specific marker MKI67 (FC: 3.0) (**Table 1**).

Expression of Epithelial-, Stemness- and Proliferation Specific Markers in LESC Grown on Human LC Measured by Immunofluorescence Staining

To validate the expression of previously identified genes at the protein level, LESC grown on human LC were stained by fluorescent labelled specific antibodies (**Figure 3**). CK19 showed a scattered cytoplasmic staining throughout the outgrowing cell sheet, representing the corneal phenotype of the cultured cells. The expression of ABCG2, a putative marker of stemness was also observed in both the cell membrane and cytoplasm of LESC. Strong staining and co-localization of the proliferation marker Ki-67 and CK8/18 was present in some cells (**Figure 1A, insert**), further confirming the proliferating and differentiating potential of these cells, respectively. The nuclear protein p63 α and Vim, both markers of stemness, showed co-localization and positivity in most of the LESC grown on human LCs.

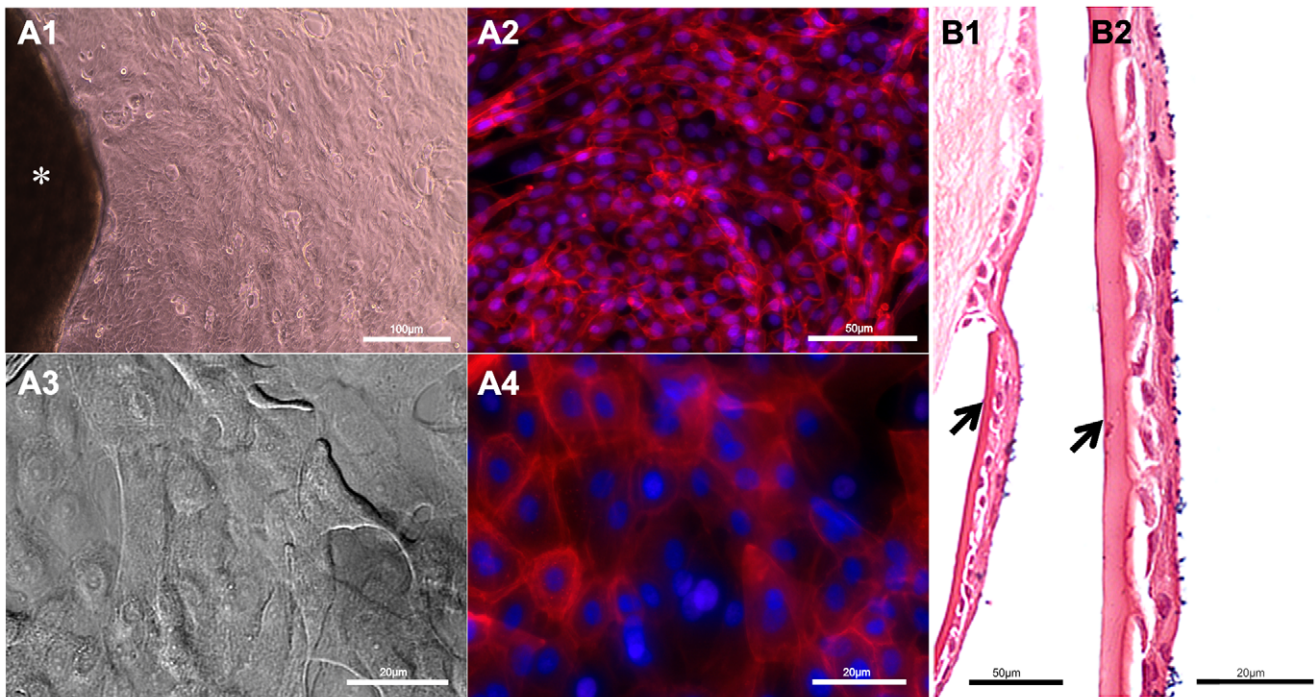
Phenotyping of the Cell Surface Markers on LESC

To define the phenotype of the outgrowing cells, a flow cytometric analysis with well-known stem cell surface markers corresponding to hematopoietic, endothelial and mesenchymal lineages was carried out on the LESC grown on human LC (summary of the results and flow cytometry histograms are shown in **Table 2** and **Figure S1, respectively**).

No common hematopoietic cell surface markers were detected on the outgrowing cells: CD45, CD34, CD133 and human leukocyte antigens (HLA)-DR. LESC expressed slightly, but significantly higher CD14 when compared to bone marrow derived mesenchymal stem cells (bmMSCs) ($p < 0.05$). A small population of LESC showed C-X-C chemokine receptor type 4 (CXCR4) and CD117/c-kit positivity characteristic for migrating and early progenitor or pluripotent stem cells, respectively, which is not characteristic for bmMSCs ($p = 0.0059$ and $p = 0.0332$). High CD47 expression of cultivated LESC was similar to that of bmMSCs demonstrating the viability and immunocompetence of both cell types.

Regarding the endothelial-related markers, no CD31/Platelet endothelial cell adhesion molecule (PECAM) and vascular endothelial growth factor receptor 2 (VEGFR2)/Kinase insert domain receptor (KDR) could be detected, showing no endothelial-related contamination of the cell culture. When compared to bmMSCs, more cells in the LESC culture expressed CD144/vascular endothelial (VE)-Cadherin ($p = 0.0321$) and CD104/Itg $\beta 4$ ($p = 0.0458$).

Significant differences were also found between LESC and bmMSC in the most important MSC markers: a very small population of LESC expressed CD90/Thymocyte differentiation antigen 1 (Thy-1) and less than half of them were CD105/Endoglin positive, unlike bmMSCs ($p = 0.000032$ and $p = 0.0006$, respectively). The expression of CD73, CD147/Neurothelin and platelet-derived growth factor receptor β (PDGF-R β) showed no significant difference as compared to bmMSCs.



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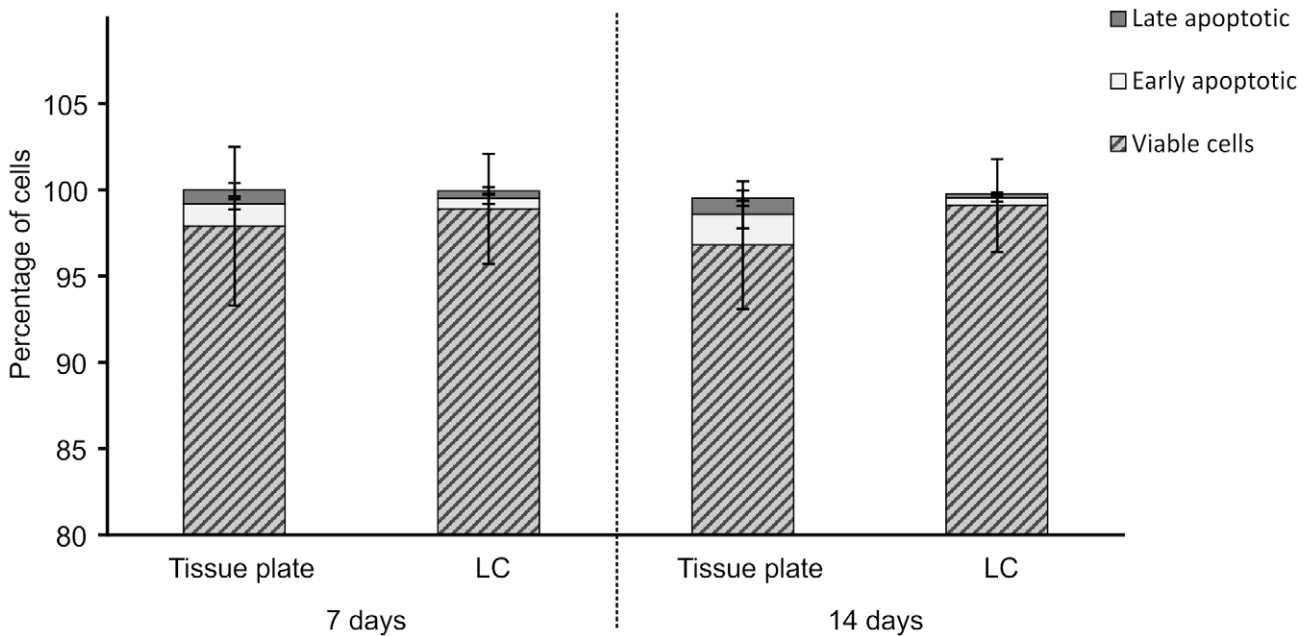


Figure 1. Cultivation and viability of LESC. Limbal graft (*) cultured on cell culture plate (A) or human LC (B) showing outgrowth of cells with epithelial morphology within 24 hrs of cultivation (image shown represents a 3 day cell outgrowth, A1 and A3 are bright field images, A2 and A4 are immunofluorescent images of actin cytoskeleton (red) and nucleus (blue)). Hematoxylin & Eosin staining of LESC grown on LC (arrows) forming stratified epithelial layer at day 7 (B1 and B2). Cell viability and death of the cultured LESC (viable cells (striped bar), early apoptotic or annexin V-FITC⁺ cells (light gray bar); late apoptotic or annexin V-FITC/propidium iodide⁺ cells (dark gray bar)) (C). Data shown are mean ± S.D (n=3, Scale bars: 100 μm A1, 50 μm A2, 20 μm A3–4; 50 μm B1, 20 μm B2). doi:10.1371/journal.pone.0047187.g001

Due to their importance in the attachment of the extracellular matrix (ECM) and maintenance of growth supporting environment, next we tested the presence of CAMs and Itgs. Significant differences were found in the expression pattern of Itgs on LESC:

CD44/homing-associated cell adhesion molecule (H-CAM) was expressed at lower (p = 0.00052), while CD49b/Itg α2 (p = 0.038) and CD49f/Itg α6 (p = 0.008) at higher levels than on the surface of bmMSCs. The expression of CD29/Itg β1, CD49a/Itg α1,

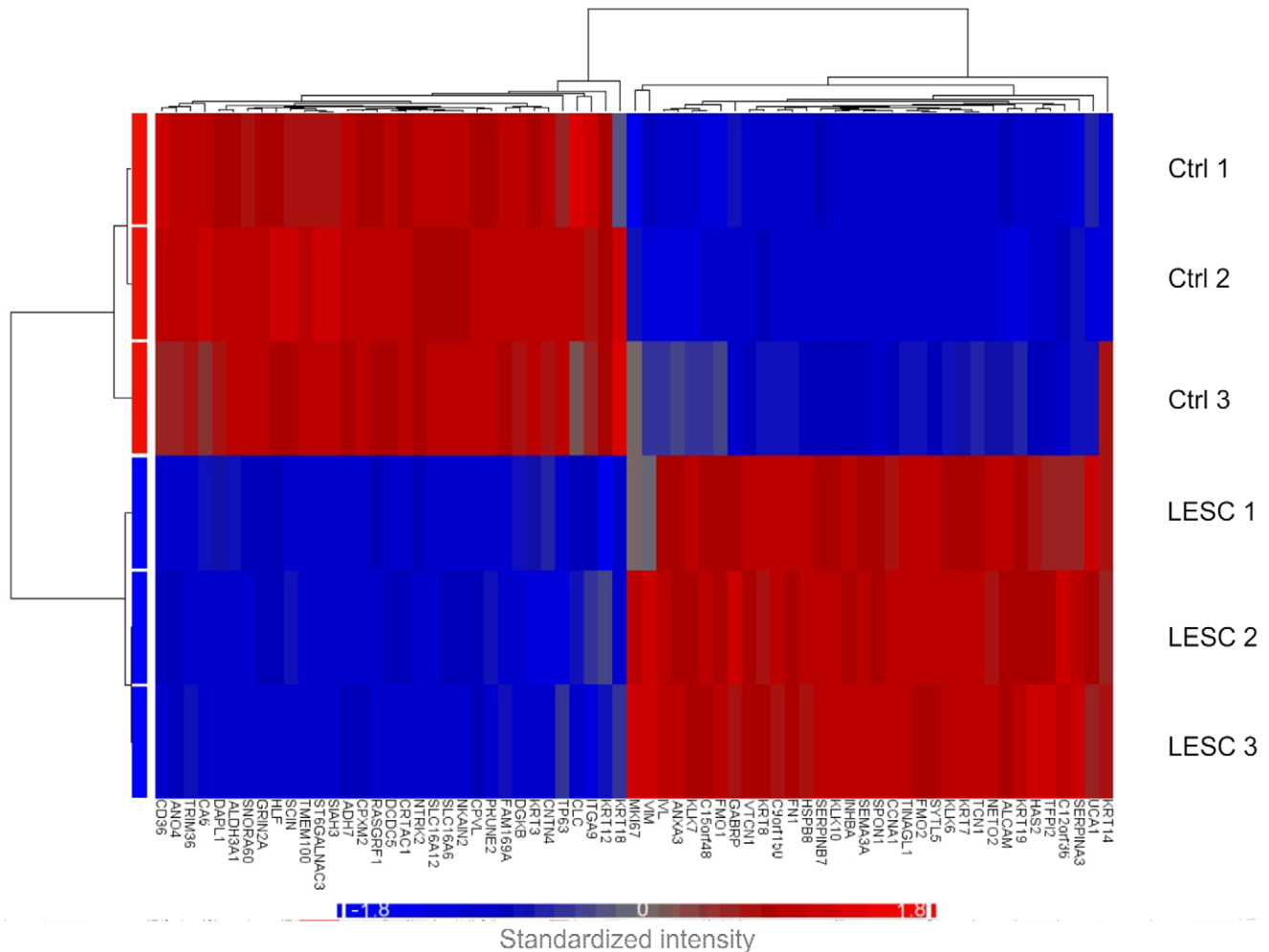


Figure 2. Transcriptional profiling in L ESCs. Heatmap of the transcripts and functional clustering of 67 genes selected on the basis of their high or low FC or previously documented relation to L ESCs ($n=3$, $p<0.01$). Red and blue colors indicate high and low expression, respectively. doi:10.1371/journal.pone.0047187.g002

CD56/neural cell adhesion molecule (NCAM), CD146/melanoma cell adhesion molecule (MCAM) and CD166/activated leukocyte cell adhesion molecule (ALCAM) were similar in L ESCs and bmMSCs.

To exclude a possible lens epithelium origin of some of the significantly different markers found on L ESCs, surface profiling of lens epithelial cells (LECs) was carried out also. LECs similar to L ESCs were CD34⁻ and CD45⁻, expressed no CD144/VE-Cadherin and showed higher CD44/H-CAM (50.43±29.28%), similar CD146/M-CAM (74.22±2.23%) or lower expression of CD166/ALCAM (74.19±46.07%) compared to cultured L ESCs ($n=3$). Immunostaining of human limbal sections further confirmed the presence and localization of these L ESC markers *in situ* (Table 3 and Figure S2).

Profiling of the Carbohydrate Surface Markers on L ESCs

The membrane of stem cells is characterized by typical carbohydrate patterns which can change during differentiation [26,27]. Lectin-based screening of the most common terminal carbohydrates of cell surface glycolipids and glycoproteins was carried on the outgrowing L ESCs (Figure 4 and Table 4). The surface of these cells contained high amounts of sialic acid stained by Wheat germ agglutinin (WGA) (Median = 1423.19±8.08). The

majority of the cells (51.59±3.1%) showed very strong Concanavalin A (ConA, Fluorescence Intensity Median (FI_{med}) = 2125.02±25.99) positivity due to the presence of branched α -mannosidic structures. Ricinus communis agglutinin (RCA), Jacalin (AIL) and Peanut agglutinin (PNA), which bind to galactose and/or N-acetylgalactosamine, were all positive on L ESCs, although lower fluorescence intensity could be detected by PNA ($FI_{med} = 185.75 \pm 1.06$) showing a small amount of T-antigen present as opposed to RCA ($FI_{med} = 850.79 \pm 14.96$) and AIL ($FI_{med} = 687.85 \pm 7.61$). Ulex europaeus agglutinin I (UEA)-lectin exhibited moderate fluorescence intensity on 61.1±1.97% of the cells, only indicating low levels of detectable fucose molecules on a subset of L ESCs.

Colony-forming Potential of L ESCs

In order to test whether the expanded L ESCs resemble the pluripotency signature reflected by the gene and protein expression levels of putative stem cell markers, their colony forming potential was tested. The L ESCs were dissociated and cultivated at clonal density (3000 cells/cm²) on Gelatin, Fibronectin and MethoCult coated plates. All epithelial sheets tested (both cell culture plates- and human LC-grown, $n=4$) were capable of forming epithelial holoclone-like colonies on Gelatin and Fibro-

Table 1. Transcripts and functional clustering of selected genes in LESC compared to differentiated corneal epithelium with high or low FC or previously documented relation to LESC (n = 3, p < 0.01).

Gene symbol	Gene description	Fold change	Regulation	Molecular function
KRT14	Keratin 14	2	Up	Structural constituent of cytoskeleton
SERPINA3	Serpin peptidase inhibitor, clade A (alpha-1 antiproteinase), member 3	21	Up	DNA binding
KRT19	Keratin 19	6	Up	Structural constituent of cytoskeleton
ALCAM	Activated leukocyte cell adhesion molecule	20	Up	Receptor binding
KRT7	Keratin 7	31	Up	Structural molecule activity
KLK6	Kallikrein-related peptidase 6	71	Up	Serine-type endopeptidase activity
FMO2	Flavin containing monooxygenase 2 (non-functional)	75	Up	Monooxygenase activity
SEMA3A	Sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3A	40	Up	Receptor activity
KLK10	Kallikrein-related peptidase 10	29	Up	Serine-type endopeptidase activity
SERPINB7	Serpin peptidase inhibitor, clade B (ovalbumin), member 7	29	Up	Serine-type endopeptidase inhibitor activity
FN1	Fibronectin 1	75	Up	Extracellular matrix structural constituent
KRT8	Keratin 8	4	Up	Structural molecule activity
KLK7	Kallikrein-related peptidase 7	57	Up	Serine-type endopeptidase activity
VIM	Vimentin	4	Up	Structural constituent of cytoskeleton
MKI67	Antigen identified by monoclonal antibody Ki-67	3	Up	Nucleotide binding
KRT18	Keratin 18	1	Down	Structural molecule activity
KRT12	Keratin 12	6	Down	Structural molecule activity
ITGA9	Integrin, alpha 9	1	Down	Receptor activity
TP63	Tumor protein p63	1	Down	DNA binding
KRT3	Keratin 3	31	Down	Structural molecule activity
NTRK2	Neurotrophic tyrosine kinase, receptor, type 2	30	Down	Nucleotide binding
CRTAC1	Cartilage acidic protein 1	72	Down	Calcium ion binding
DCDC5	Doublecortin domain containing 5	43	Down	Tubulin binding
RASGRF1	Ras protein-specific guanine nucleotide-releasing factor 1	20	Down	Guanyl-nucleotide exchange factor activity
CPXM2	Carboxypeptidase X (M14 family), member 2	25	Down	Metalloprotease activity
ADH7	Alcohol dehydrogenase 7 (class IV), mu or sigma polypeptide	64	Down	Alcohol dehydrogenase (NAD) activity
ALDH3A1	Aldehyde dehydrogenase 3 family, member A1	30	Down	Aldehyde dehydrogenase (NAD) activity
DAPL1	Death associated protein-like 1	33	Down	Epithelial differentiation or apoptosis
CA6	Carbonic anhydrase VI	33	Down	Carbonate dehydratase activity

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nectin as previously described [28,29] (**Figure 5A**). Large and small colonies were visible within 7 days of culture and exhibited active cytoskeleton and smooth-outline appearance on Gelatin and Fibronectin surfaces, but not on MethoCult coated plates (**Figure 5B, C**).

Discussion

An animal material-free method for *ex vivo* expansion of cornea LESC from cadavers or autologous donors would be a safe way for tissue regeneration in cases of chemical, thermal or microbial injuries of the cornea as well as diseases such as Steven-Johnson syndrome. So far, most standard protocols for cultivating cornea LESC included animal materials such as fetal bovine serum (FBS) and/or exogenous growth factors, hormones and cholera toxin in the growth media [30,31]. Increasing concentrations of FBS (1–

20%) have been shown to stimulate cornea limbal progenitor cells into clonal proliferation [32].

Different carriers have also been used for supporting the growth of LESC ranging from synthetic biopolymers to natural materials such as HAM [15], anterior LC [14], fibrin matrix [17] and temperature-responsive polymers [16]. Although HAM has the advantage of containing growth, anti-angiogenic and anti-inflammatory factors that can prevent or decrease fibrosis in the healing tissue [33] or it can be sized and used as a surface for cells and biologic patch material [34], it is not transparent and thin enough.

Anterior human LC can be obtained most commonly from uneventful capsulorrhexis during cataract surgery or alternatively, from cadaver eyes at Cornea Banks. Although LC is of limited size, its transparency and thinness are superior to other biomaterials. It is the later properties upon which we decided to cultivate LESC on human LC [14]. In addition to the previously reported use of human LC for expansion of limbal-like stem cells

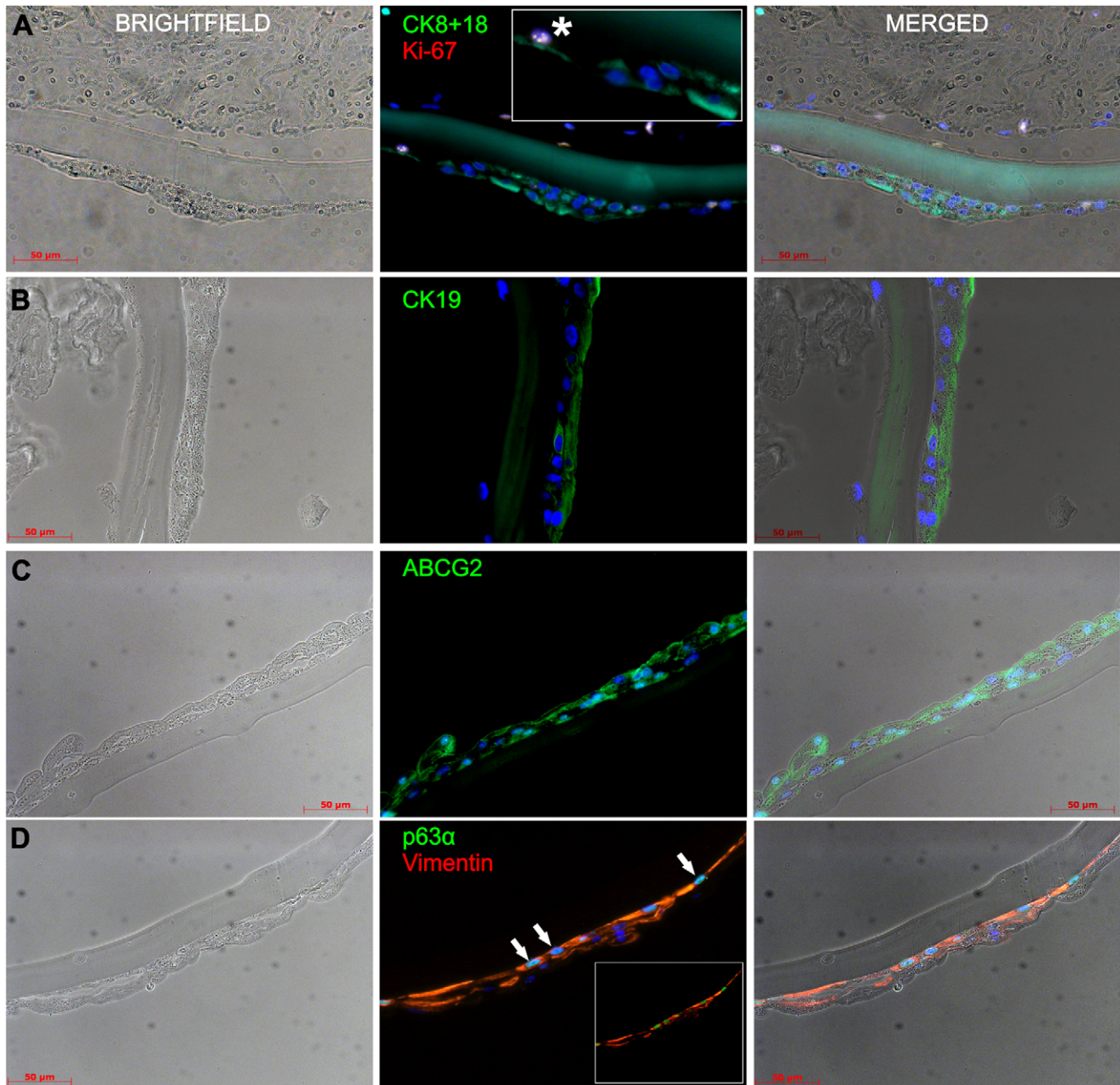


Figure 3. Expression of epithelial-, stemness- and proliferation specific markers in LSCs grown on human LC measured by immunofluorescence staining. Immunohistochemistry was performed to detect the (co)-expression of CK8/18/Ki-67, CK19, ABCG2, Vim/p63 α in the LSCs grown on human LC (Left column: bright field-; Center: immunofluorescent; Right column: merged image; Colors on the text correspond to the color of the marker examined, while all nuclei are stained blue with DAPI; Insert: shows co-localization of CK8+18 and Ki-67 and (*) refers to the different staining pattern in the region shown; Arrows: show expression of Vimentin in the basal cells; the images are representative of at least 3 independent experiments, scale bar: 50 μ m). doi:10.1371/journal.pone.0047187.g003

[14], we combined the use of such bioscaffold with the use of human serum as the only growth supplement for cultivating LSCs.

Although there are no definitive markers for identifying adult stem cells in general, or corneal stem cells in particular [35], characterization by a consensus-based panel of expression markers for LSCs [4], together with some additional, yet not described putative stem cell markers were used here for the detailed characterization of these cells.

Both cell culture plate- and LC-grown LSCs showed low death rate (<3%) over the 2 weeks cultivation period, achieving stratified epithelial-like cell growth. Presence of limbal epithelial markers CK14 and CK8/CK18 confirmed the corneo-conjunctival- and, in particular, the limbal epithelial- origin of the LSCs. CK8 being a differentiating/migrating marker also showed the transition potential of these cells towards differentiated corneal epithelium [35–39]. Together with the low expression of differentiated cornea epithelial markers - CK3/12 [2,40], and the high proliferative potential of the LSCs (MKI67 expression),

Table 2. Expression of hematopoietic, endothelial, stemness and adhesion molecules on LESC.

		LESC	bmMSC	
Hematopoietic	CD14	12.13±4.85	0.38±0.1	*
Monocyte markers	CD34	0±0	0±0	
	CD45	0±0	0±0	
	CD47	98.98±0.10	96.97±0.81	
	CD133	0±0	0±0	
	CD117/c-kit	17.98±4.53	0±0	*
	CXCR4	27.81±4.41	0±0	**
	HLA-DR	0±0	0±0	
Endothelial markers	CD31/PECAM	0±0	0±0	
	CD144/VE-Cadherine	81.92±3.57	41.55±9.57	*
	VEGFR2/KDR	0±0	0±0	
	CD104/Integrin β4	75.87±5.18	38.49±10.31	*
MSC	CD73	87.91±1.24	90.59±1.80	
Fibroblast markers	CD90/Thy-1	12.25±4.29	90.13±0.96	***
	CD105/Endoglin	42.09±4.91	81.90±1.96	***
	CD147/Neurothelin	97.13±0.33	75.21±7.81	
	PDGF Rβ	54.93±1.68	75.36±7.80	
Integrins and CAMs	CD29/Integrin β1	97.01±0.40	92.77±1.65	
	CD44/H-CAM	16.55±4.95	87.90±2.48	***
	CD49a/Integrin α1	71.73±6.09	71.42±7.15	
	CD49b/Integrin α2	91.16±1.27	60.55±7.19	*
	CD49f/Integrin α6	68.38±8.18	0±0	***
	CD56/NCAM	2.17±1.03	24.68±7.57	
	CD146/MCAM	82.40±3.11	87.28±2.18	
CD166/ALCAM	98.02±0.20	86.57±6.26		

The expression of different groups of surface antigens on the LESC was compared to those found on bmMSCs. A small population of the LESC expressed higher CD14, CD117/c-kit, CXCR4 which are markers of special progenitor cell types. Although the LESC expressed most of the MSC-like markers, just a minor population expressed CD90/Thy-1 and CD105 which determine the MSC phenotype. BmMSCs lack CD49f/Itg α9 which is strongly expressed on LESC (the data represent percentage of positive cells within the total LESC culture shown as mean ± S.D., n=7; p<0.05 *, p<0.01 **, p<0.001 ***).

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it can be initially concluded that the outgrowing cells are undifferentiated limbal epithelial cells.

The multipotency of the LESC was confirmed by the increased expression of putative stem cell markers: p63α [41,42], ABCG2 [43,44], CK19 [4,45] and Vim [4,45,46]. These four markers have been described as being expressed mainly in the basal limbal epithelium [4], although Donisi et al. [47] and Sacchetti et al. [45] described lack of CK19 expression in these cells. In addition, ITG α9 has been described as being responsible for cell migration during injury [46].

Genome-wide profiling of the LESC provided a list of high and low expressed genes that have already been demonstrated, genes that are novel or possess yet unidentified function in LESC [48]. Serine proteinase inhibitor 3A (SERPINA3) being overexpressed in the LESC in our gene array (FC: 21.1) has been investigated previously for its anti-angiogenic and anti-inflammatory effects during corneal injury [49]. Semaphorin 3A (SEMA3A) (FC: 40.2) has been shown to be involved in the development of mouse cornea and differentiation of cornea epithelial cells [50]. Fibro-

Table 3. *In situ* immunohistochemical characteristics of the basal (B) and apical cells (A), and the stroma (S) in human cornea limbal sections.

Antibody specificity	B	A	S
CD34	–	–	+
CD45	–	–	+
CD144/VE-Cadherine	+	–	+
CD44/H-CAM	–	few cells	+
CD146/MCAM	few cells	–	+
CD166/ALCAM	few cells	–	+

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nectin 1 (FN1) (FC: 74.9) is involved in cell adhesion and migration processes during wound healing [51].

Considering the fact that the slow-cycling stem cells in the limbal region represent less than 10% of the limbal basal cells [52] and the finding of stem- and proliferative/migratory cell markers on our LESC, very likely the outgrowing cells represent a mixture of stem- and TACs rather than pure limbal stem cells.

The LESC grown on human LC are of non-hematogenous origin. They are viable cells (99% of them are CD47⁺), express markers of early pluripotency (18% are CD117/c-kit⁺) and possess migratory capacity (28% are CXCR4⁺) very much needed during resolution of corneal injuries. In addition, the outgrowing cells carry CD104/Itg β4⁺ (76%), a marker found in basal cells of limbal and corneal epithelium [53], and also, CD144/VE-Cadherin⁺ (82%), a yet not described basal limbal cell-marker. Exclusion of the markers present on the surface of both LESC and LECs, further strengthened CD144/VE-Cadherin as a new putative LESC marker, while *in situ* immunostaining of human limbal sections confirmed its presence and localization in the basal cell layer of the limbus. In addition, yet uncharacterized putative markers of LESC could be localized in the limbal epithelium: CD44/H-CAM in the apical layer, and CD146/MCAM and CD166/ALCAM in the basal layer.

The presence of different plasma membrane Itgs tested here: α1, α2, α6 and β1, resembles a previously reported positivity found mainly in basal limbal, but also in basal corneal cells [4,53]. In addition, previously undefined CAMs on the surface of LESC were detected and confirmed *in situ* as putative markers of LESC: CD44/H-CAM in the apical layer, and CD146/MCAM and CD166/ALCAM in the basal layer of the limbus.

Since our LESC had a generally lower expression of MSC-surface markers (CD73, CD90/Thy-1, CD105/Endoglin, PDGF-Rβ) than in bmMSCs, and a distinct or significantly different pattern of CD117/c-kit, CXCR4, Itgs α2, α6 and β4 expression (**Table 2**), very likely these cells are pluripotent and capable of migration.

Indeed, the presence of pluripotent cells in the cultured LESC could be confirmed by the formation of small and large colonies of cells with intact cytoskeleton on Gelatin and Fibronectin surfaces, and absence of colonies on MethoCult surfaces, thus excluding the hematopoietic (myeloid or erythroid) differentiation potential of these cells.

To assess and profile the carbohydrates present in glycolipids and glycoproteins on the surface of LESC, a comprehensive lectin-based screening for 14 carbohydrate structures (1 sialic acid, 5 N-acetylglucosamines, 1 mannose, 6 galactose and 1 fucose) was carried out. To our present knowledge, limbal epithelial cells have been shown to express unsialylated galactose

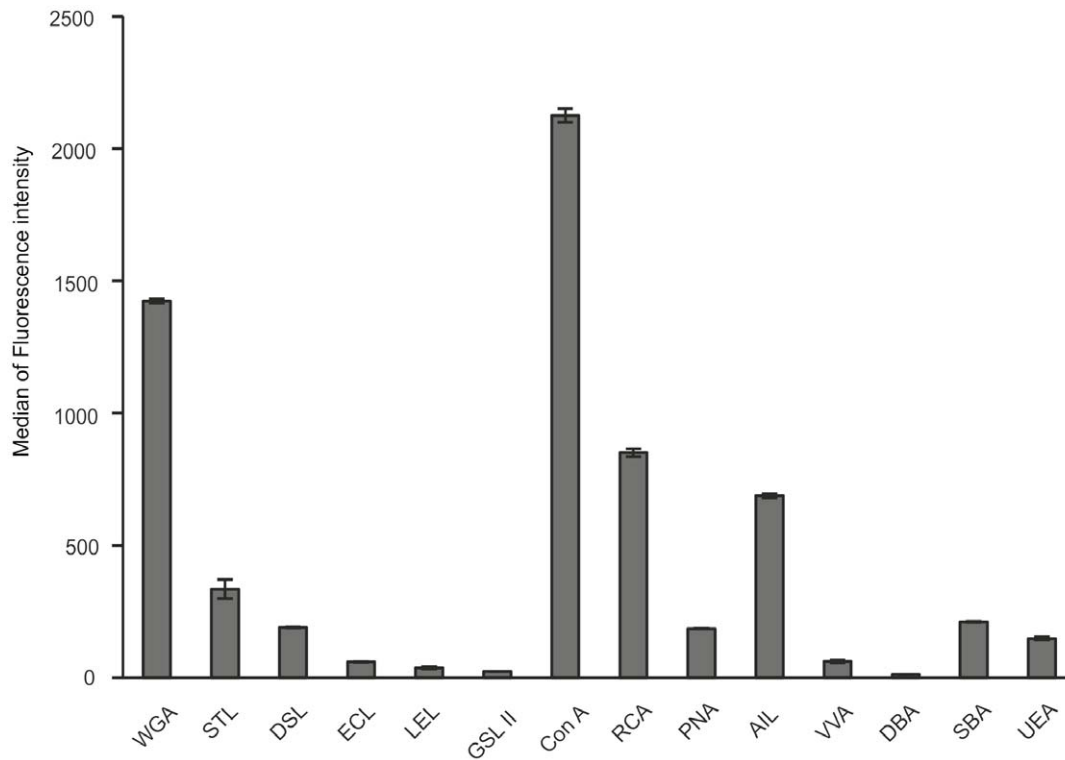


Figure 4. Expression of carbohydrate molecules on the surface of L ESCs. Lectins-based staining of carbohydrate specific molecules on the surface of L ESCs. For abbreviations used see Table 2. (Data shown are mean \pm S.D. of the median of fluorescence intensity, n = 3). doi:10.1371/journal.pone.0047187.g004

Table 4. Lectin-based staining of surface carbohydrate molecules on L ESCs.

	Lectin	Cells (%)	Median of FI	Affinity
Sialic acid	WGA	97.37 \pm 0.33	1423.19 \pm 8.08	GlcNAc β 1-4GlcNAc β 1-4GlcNAc. Neu5Ac (sialic acid)
N-acetylglucosamine binding lectins	STL	97.68 \pm 1.64	335.38 \pm 35.63	GlcNAc oligomers
	DSL	98.94 \pm 0.23	190.04 \pm 1.23	(b-1.4) linked N-acetylglucosamine oligomers
	ECL	95.64 \pm 2.79	59.45 \pm 0.16	galactosyl (b-1.4) N-acetylglucosamine
	LEL	83.39 \pm 14.23	38.15 \pm 4.22	N-acetylglucosamine oligomers
	GSL II	82.52 \pm 2.66	24.63 \pm 0.27	alpha- or beta-linked N-acetylglucosamine
Mannose binding lectins	ConA	51.59 \pm 3.10	2125.02 \pm 26.00	high-mannose type. hybrid type and biantennary complex type N-Glycans
Galactose N-acetylglucosamine binding lectins	RCA	98.28 \pm 0.35	850.79 \pm 14.96	Gal β 1-4GlcNAc β 1-R
	PNA	97.92 \pm 0.51	185.75 \pm 1.06	Gal β 1-3GalNAc α 1-Ser/Thr (T-Antigen)
	AIL	98.99 \pm 0.01	687.85 \pm 7.61	(Sia)Gal β 1-3GalNAc α 1-Ser/Thr (T-Antigen)
	VVA	94.49 \pm 0.38	61.49 \pm 5.32	alpha- or beta-linked terminal N-acetylglucosamine
	DBA	89.93 \pm 2.54	12.24 \pm 0.10	N-acetylglucosamine
	SBA	97.11 \pm 0.60	211.11 \pm 1.32	a- or b-linked N-acetylglucosamin
Fucose binding lectins	UEA	61.10 \pm 1.97	148.39 \pm 5.91	Fuc α 1-2Gal-R

The L ESCs surface contained high amount of sialic acid, N-acetylglucosamine and galactose molecules. Just around half of the cells contained mannose and two thirds contained fucose molecules, showing subpopulations within the LESC cell culture. These carbohydrate molecules determine the ECM-binding and immunological properties of the cells. **WGA**: Wheat germ agglutinin (*Triticum vulgare*), **STL**: Potatoe lectin (*Solanum tuberosum*), **DSL**: Datura stramonium lectin (*Datura stramonium*), **ECL**: Erythrina cristagalli lectin (*Erythrina cristagalli*), **LEL**: Tomato lectin (*Lycopersicon esculentum*), **GSL II**: Griffonia (Bandeiraea) simplicifolia lectin II (*Griffonia simplicifolia*), **ConA**: Concanavalin A (*Canavalia ensiformis*), **RCA**: Ricinus communis Agglutinin (*Ricinus communis*), **PNA**: Peanut agglutinin (*Arachis hypogaea*), **AIL**: Jacalin (*Artocarpus integrifolia*), **VVA**: Hairy vetch agglutinin (*Vicia villosa*), **DBA**: Horse gram lectin (*Dolichos biflorus*), **SBA**: Soy bean agglutinin (*Glycine max*), **UEA**: Ulex europaeus agglutinin (*Ulex europaeus*) (Data shown are mean \pm S.D., n = 3). doi:10.1371/journal.pone.0047187.t004

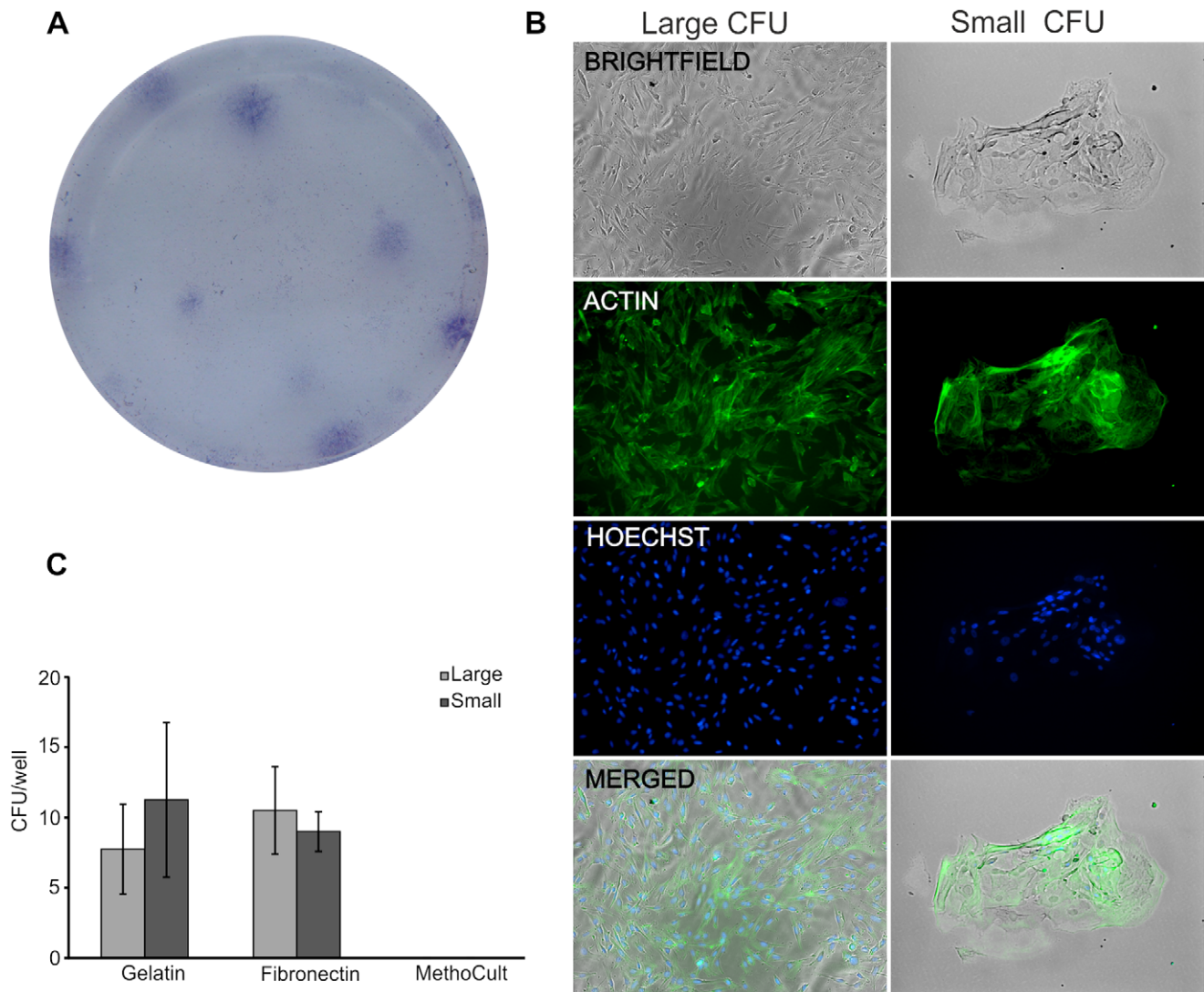


Figure 5. Colony-forming potential of LESC. The LESC were cultured at clonal density of 3000 cells/cm² and early epithelial holoclone-like colony formation was recorded at day 7 of culture. LESC formed colony forming units on Gelatin and Fibronectin coated plates as stained by crystal violet (0.5% w/v) (A). The colony forming unit (CFU)-forming cells were stained for actin (phalloidin-FITC, green) and Hoechst 33342 (blue, nuclear). The CFUs could be divided into two groups: large CFUs containing >50 cells, and small CFUs containing <50 LSCs. No significant difference in the CFU types on Gelatin and Fibronectin matrices was found, while MethoCult matrix-grown LESC formed no colonies at all (B). (Data shown are mean ± S.D., n=3).

doi:10.1371/journal.pone.0047187.g005

residues on their cell surface recognized by PNA and lack α -2,3-bound sialic acid [54]. Our LESC had a lower median fluorescence intensity for PNA as compared to the median values of binding detected for WGA, ConA, RCA and AIL although 98% of the cells were positive for PNA.

The distinct surface marker fingerprint of the LESC and the 5 surface carbohydrate markers (WGA, ConA, RCA, PNA and AIL) distinguished on these cells, point out a mixed population of slowly proliferating limbal stem cells and highly proliferating, migrating and potentially differentiating TACs in the outgrowth cultures. Sorting out these two cell populations and running a differential gene expression screening (currently undertaken in our lab) would probably give better insight and understanding of the function of these cells. From transplantation point of view, having a mixture of both stem-like- and undifferentiated/

TACs would be a highly desirable condition towards successful corneal transplantation.

In conclusion, we hereby show that cornea LESC can be consistently expanded *ex vivo* on human LC using a medium containing human serum as the only growth supplement. Cells isolated and cultivated in such a way are viable; they preserve their pluripotency as confirmed by their positivity for p63 α , ABCG2, CK19, Vim and Itg α 9, and low expression of CK3/12. The presence of differentiation properties of our cultured cells (positivity for CK8/18 and CK14) shows the directional differentiating potential into corneal epithelium *in situ*. Additional markers of pluripotency in LESC have been described here that can be added to the future recognition of these cells and as indicative factors for positive clinical outcome after corneal transplantation.

Materials and Methods

Limbal Tissue Harvesting

All tissue collection complied with the Guidelines of the Helsinki Declaration and was approved by the Regional and Institutional Research Ethics Committee at the University of Debrecen, Hungary (DE OEC: 3094–2010). Limbal tissue collection was done from cadavers only and Hungary follows the EU Member States' Directive 2004/23/EC on presumed consent practice for tissue collection [55]. Limbal tissue was harvested from 77 cadavers (44 males and 33 females, age 70.5 ± 9.3 years) within 12 hours of biologic death. In brief, after a thorough povidone iodide eye wash, the conjunctiva was incised and separated from the limbal junction consequently, a $2 \times 2 \times 0.25$ mm rectangular shape limbal graft was dissected away and towards the cornea, respectively, at the 12 o'clock position. The depth of the graft was kept superficial or within the epithelial layer; multiple grafts were collected from a single eye. The graft dissection was performed by lamellar knife placed tangential to the surface being cut.

Culture Medium and Cultivation Conditions

Corneal epithelial cell culture medium consisted of Dulbecco-modified Eagle's medium (DMEM, Sigma-Aldrich, St. Louis, MO, USA) supplemented with 20% human AB serum (Human serum Type AB, PAA, Pasching, Austria), 200 mM/mL L-glutamine (Sigma-Aldrich), 10,000 U/mL penicillin- 10 mg/mL streptomycin (Sigma-Aldrich). The orientation of the graft was epithelial side up in $1,91 \text{ cm}^2$ tissue culture plates. Limbal tissues were proliferated *in vitro* on human lens capsules that were obtained from uneventful capsulorhexis during cataract surgery and pretreated with 0,025% trypsin-EDTA (PAA, Pasching, Austria) (20 minutes, 37°C). For grafting on LCs, a drop of medium was used to smooth-out the capsule first and then the limbal tissue was placed in the middle of the capsule. Following adherence to the lens capsule and/or the culture plate, the graft was cultivated in total of 1 mL medium. Feeding of the cells occurred on every alternate day. The growth of the cells was monitored under phase contrast microscope regularly. Only grafts which had cell outgrowth within 24 hours were used further to decrease the chance of fibroblast contamination.

Assay for Cell Death Analysis

Cell death was assessed by the Annexin-V-FITC Apoptosis Detection Kit (MBL, Woburn, MA, USA) according to manufacturer's recommendations; proportion of stained Annexin-V⁺ and Annexin-V⁺/Propidium iodide⁺ cells was determined by fluorescence activated cell sorter (FACS) analysis on FACSCalibur flow cytometer (BD Biosciences Immunocytometry Systems, San Jose, CA, USA) and data were analyzed using WinMDI freeware (Joseph Trotter, La Jolla, CA, USA).

Microarray and Data Analysis

Microarray analyses were performed using the Affymetrix GeneChip Human Gene 1.0 ST Arrays (Affymetrix, Santa Clara, CA) which contains more than 28,000 gene transcripts. 150 ng of total RNA was subjected to Ambion WT Expression Kit (Ambion) and GeneChip WT Terminal Labeling Kit (Affymetrix, Santa Clara, CA, USA) following the manufacturers' protocols for whole genome gene expression analysis. The arrays were washed and stained using FS-450 fluidics station (Affymetrix). Signal intensities were detected by Hewlett Packard Gene Array Scanner 3000 7G (Hewlett Packard, Palo Alto, CA, USA). The scanned images were processed using GeneChip Command Console Software (AGCC) (Affymetrix) and the CEL files were imported into Partek

Genomics Suite software (Partek, Inc. MO, USA). Robust microarray analysis (RMA) was applied for normalization. Gene transcripts with maximal signal values of less than 32 across all arrays were removed to filter for low and non-expressed genes, reducing the number of gene transcripts to 23190. Differentially expressed genes between groups were identified using one-way ANOVA analysis in Partek Genomics Suite Software. Clustering analysis was made using the clustering analysis module in Partek Genomics Suite Software.

Histological and Immunofluorescent Analysis

LESCs grown on the surface of cell culture-grade glass-cover slips or human LCs, as well as full thickness cornea limbal grafts were fixed in 4% paraformaldehyde for 20 min, room temperature. The LC-grown samples and full thickness limbal grafts were dehydrated and embedded in paraffin after which $3 \mu\text{m}$ thick longitudinal sections were obtained for staining with Hematoxylin and Eosin (H&E) according to standard laboratory protocols. Alternatively, immunofluorescent labelling with anti- p63alpha, ABCG2, CK19, CK8/18, Vim and Ki-67 antibodies was used for visualization under a ZEISS Axio Observer.Z1 (ZEISS, Oberkochen, Germany) fluorescent microscope (list of primary antibodies used is provided in **Table S2**). Similarly, immunofluorescent labelling with anti- CD34, CD45, CD144/VE-Cadherine, CD144/H-CAM, CD146/MCAM and CD166/ALCAM antibodies was used for staining the full thickness limbal grafts.

Phenotyping of Cells and Comparison to bmMSCs

To analyse the phenotype of the isolated corneal limbal cells multicolour flow cytometric analysis was used. FITC, R-phycoerythrin (PE) and allophycocyanin (APC) conjugated antibodies were used to measure the expression of CD34, CD44, CD45, CD49f/Itg $\alpha 6$, CD73, CD106, CD144, CD147 (all from BD Biosciences, San Jose, CA, USA); CD49a/Itg $\alpha 1$ (Biolegend, San Diego, CA, USA), CD14, CD29/Itg $\beta 1$, CD31, CD36, CD47, CD49b/Itg $\alpha 2$, CD54, CD56/NCAM, CD69, CD90/Thy-1, CD104, CD105, CD117/c-kit, CD146/MCAM, CD166/ALCAM, CXCR4, HLA-DR, PDGF-R β , VEGFR2 (all from R&D Systems, Minneapolis, MN, USA) and CD133 molecules (Miltenyi Biotech, Gladbach, Germany) (for further details refer to **Table S2**). Samples were measured by FACSCalibur flow cytometer (BD Biosciences Immunocytometry Systems) and data were analyzed using WinMDI freeware (Joseph Trotter, La Jolla, CA, USA). For comparison, bmMSCs were used – their availability and isolation protocol were based on another unrelated study carried by V.Z. and R.E.

Lectin Staining of Cells

Lectin screening of isolated LESCs were performed by Lectin kits from Vector Labs (Burlingame, CA). For detecting certain carbohydrate structures, the following lectins were tested: for sialic acid (**WGA**: Wheat germ agglutinin (*Triticum vulgare*)); for N-acetylglucosamines (**STL**: Potatoe lectin (*Solanum tuberosum*), **DSL**: Datura stramonium lectin (*Datura stramonium*), **ECL**: Erythrina cristagalli lectin (*Erythrina cristagalli*), **LEL**: Tomato lectin (*Lycopersicon esculentum*), **GSL II**: Griffonia (Bandeiraea) simplicifolia lectin II (*Griffonia simplicifolia*)); for mannose (**ConA**: Concanavalin A (*Canavalia ensiformis*)); for galactose N-acetylgalactosamines (**RCA**: Ricinus communis Agglutinin (*Ricinus communis*), **PNA**: Peanut agglutinin (*Arachis hypogaea*), **AAL**: Jacalin (*Artocarpus integrifolia*), **VVA**: Hairy vetch agglutinin (*Vicia villosa*), **DBA**: Horse gram lectin (*Dolichos biflorus*), **SBA**: Soy bean agglutinin (*Glycine max*) and for fucose (**UEA**: Ulex europaeus agglutinin (*Ulex europaeus*)).

The lectins were diluted in Lectin dilution buffer; the rest of the staining procedure was similar to the staining described for the FACS analysis.

Colony-forming Assay

To check the colony forming properties of LESC, cells were seeded at a 3000 cells/cm² density into 6 well plates, coated with various matrices. The wells were pre-coated for 30 minutes with either 0.1% Gelatin (Sigma-Aldrich), 10 ng/mL Fibronectin (BD Biosciences) or MethoCult (Stem Cell Technologies, Vancouver, Canada) before the cells were added. Standard growth medium for the LESC was used and changed every other day. At day 7, the samples were fixed in 4% paraformaldehyde and stained with H&E. The colonies were stained with crystal violet (0.5% w/v) against actin with phalloidin-FITC and the nucleus with Hoechst 33342. Examination was carried out under an Olympus IX81 inverted microscope with MT20 station (Olympus, Münster, Germany), and acquired and analysed by a ScanR (Olympus) software.

Statistical Analysis

Each experiment was performed at least three times and each sample was tested in triplicates. Data are expressed as mean \pm S.D. or SEM. Statistically significant differences were determined by paired student-t tests. $p < 0.05$ *, $p < 0.01$ **, $p < 0.001$ ***.

References

- Davanger M, Evensen A (1971) Role of the pericorneal papillary structure in renewal of corneal epithelium. *Nature* 229: 560–561.
- Schermer A, Galvin S, Sun TT (1986) Differentiation-related expression of a major 64K corneal keratin in vivo and in culture suggests limbal location of corneal epithelial stem cells. *J Cell Biol* 103: 49–62.
- Tseng SC (1989) Concept and application of limbal stem cells. *Eye (Lond)* 3 (Pt 2): 141–157.
- Schlötzer-Schrehardt U, Kruse FE (2005) Identification and characterization of limbal stem cells. *Exp Eye Res* 81: 247–264.
- Notara M, Alatza A, Gilfillan J, Harris AR, Levis HJ, et al. (2010) In sickness and in health: Corneal epithelial stem cell biology, pathology and therapy. *Exp Eye Res* 90: 188–195.
- Xie HT, Chen SY, Li GG, Tseng SC (2012) Isolation and expansion of human limbal stromal niche cells. *Invest Ophthalmol Vis Sci* 53: 279–286.
- Chen JJ, Tseng SC (1991) Abnormal corneal epithelial wound healing in partial-thickness removal of limbal epithelium. *Invest Ophthalmol Vis Sci* 32: 2219–2233.
- Mort RL, Ramaesh T, Kleinjan DA, Morley SD, West JD (2009) Mosaic analysis of stem cell function and wound healing in the mouse corneal epithelium. *BMC Dev Biol* 9: 4.
- Dua HS, Azuara-Blanco A (2000) Limbal stem cells of the corneal epithelium. *Surv Ophthalmol* 44: 415–425.
- Rama P, Matuska S, Paganoni G, Spinelli A, De Luca M, et al. (2010) Limbal stem-cell therapy and long-term corneal regeneration. *N Engl J Med* 363: 147–155.
- Kinoshita S, Friend J, Thoft RA (1981) Sex chromatin of donor corneal epithelium in rabbits. *Invest Ophthalmol Vis Sci* 21: 434–441.
- Sun T-T, Tseng SC, Lavker RM (2010) Location of corneal epithelial stem cells. *Nature* 463: E10–E11.
- Majo F, Rochat A, Nicolas M, Jaoude GA, Barrandon Y (2008) Oligopotent stem cells are distributed throughout the mammalian ocular surface. *Nature* 456: 250–254.
- Galal A, Perez-Santonja JJ, Rodriguez-Prats JL, Abad M, Alio J (2007) Human anterior lens capsule as a biologic substrate for the ex vivo expansion of limbal stem cells in ocular surface reconstruction. *Cornea* 26: 473–478.
- Grueterich M, Tseng SC (2002) Human limbal progenitor cells expanded on intact amniotic membrane ex vivo. *Arch Ophthalmol* 120: 783–790.
- Nishida K, Yamato M, Hayashida Y, Watanabe K, Maeda N, et al. (2004) Functional bioengineered corneal epithelial sheet grafts from corneal stem cells expanded ex vivo on a temperature-responsive cell culture surface. *Transplantation* 77: 379–385.
- Rama P, Bonini S, Lambiase A, Golisano O, Paterna P, et al. (2001) Autologous fibrin-cultured limbal stem cells permanently restore the corneal surface of patients with total limbal stem cell deficiency. *Transplantation* 72: 1478–1485.

Supporting Information

Figure S1 Histograms of the expression of hematopoietic (A), endothelial (B), stemness (C) and adhesion (D) molecules on LESC shown in Table 1.

(TIF)

Figure S2 *In situ* immunohistochemical staining of human cornea limbal sections for the presence and localization of LESC markers found by flow cytometry.

(TIF)

Table S1 Additional transcripts and functional clustering of selected genes in LESC compared to differentiated corneal epithelium with high or low FC or previously documented relation to LESC (n=3, p<0.01).

(PDF)

Table S2 Details of the antibodies used for immunohistochemistry and/or flow cytometry.

(PDF)

Author Contributions

Conceived and designed the experiments: RA ZV GP. Performed the experiments: RA ZV KC MCM EOJ OKO GP. Analyzed the data: RA ZV KC MCM OKO GP. Contributed reagents/materials/analysis tools: BN ER LF AB. Wrote the paper: RA ZV GP.

33. Plummer CE (2009) The use of amniotic membrane transplantation for ocular surface reconstruction: A review and series of 58 equine clinical cases (2002–2008). *Vet Ophthalmol* 12: 17–24.
34. Gomes J, Romano A, Santos M, Dua H (2005) Amniotic membrane use in ophthalmology. *Curr Opin Ophthalmol* 16: 233–240.
35. O'Sullivan F, Clynes M (2007) Limbal stem cells, a review of their identification and culture for clinical use. *Cytotechnology* 53: 101–106.
36. Barnard Z, Apel AJG, Harkin DG (2001) Phenotypic analyses of limbal epithelial cell cultures derived from donor corneoscleral rims. *Clin Experiment Ophthalmol* 29: 138–142.
37. Kurpakus MA, Maniaci MT, Esco M (1994) Expression of keratins K12, K4 and K14 during development of ocular surface epithelium. *Curr Eye Res* 13: 805–814.
38. Merjava S, Neuwirth A, Mandys V, Jirsova K (2009) Cytokeratins 8 and 18 in adult human corneal endothelium. *Exp Eye Res* 89: 426–431.
39. Merjava S, Brejchova K, Vernon A, Daniels JT, Jirsova K (2011) Cytokeratin 8 is expressed in human corneconjunctival epithelium, particularly in limbal epithelial cells. *Invest Ophthalmol Vis Sci* 52: 787–794.
40. Pajoohesh-Ganji A, Stepp MA (2005) In search of markers for the stem cells of the corneal epithelium. *Biol Cell* 97: 265–276.
41. Arpitha P, Prajna NV, Srinivasan M, Muthukkaruppan V (2005) High expression of p63 combined with a large N/C ratio defines a subset of human limbal epithelial cells: implications on epithelial stem cells. *Invest Ophthalmol Vis Sci* 46: 3631–3636.
42. Parsa R, Yang A, McKeon F, Green H (1999) Association of p63 with proliferative potential in normal and neoplastic human keratinocytes. *J Invest Dermatol* 113: 1099–1105.
43. Chen Z, de Paiva CS, Luo L, Kretzer FL, Pflugfelder SC, et al. (2004) Characterization of putative stem cell phenotype in human limbal epithelia. *Stem Cells* 22: 355–366.
44. Zhou S, Schuetz JD, Bunting KD, Colapietro AM, Sampath J, et al. (2001) The ABC transporter *Bcrp1/ABCG2* is expressed in a wide variety of stem cells and is a molecular determinant of the side-population phenotype. *Nat Med* 7: 1028–1034.
45. Sacchetti M, Lambiase A, Cortes M, Sgrulletta R, Bonini S, et al. (2005) Clinical and cytological findings in limbal stem cell deficiency. *Graefes Arch Clin Exp Ophthalmol* 243: 870–876.
46. Stepp MA, Zhu L, Sheppard D, Cranfill RL (1995) Localized distribution of alpha 9 integrin in the cornea and changes in expression during corneal epithelial cell differentiation. *J Histochem Cytochem* 43: 353–362.
47. Donisi PM, Rama P, Fasolo A, Ponzin D (2003) Analysis of limbal stem cell deficiency by corneal impression cytology. *Cornea* 22: 533–538.
48. Turner HC, Budak MT, Akinci MAM, Wolosin JM (2007) Comparative Analysis of Human Conjunctival and Corneal Epithelial Gene Expression with Oligonucleotide Microarrays. *Invest Ophthalmol Vis Sci* 48: 2050–2061.
49. Liu X, Lin Z, Zhou T, Zong R, He H, et al. (2011) Anti-angiogenic and anti-inflammatory effects of SERPINA3K on corneal injury. *PLoS One* 6: e16712.
50. Ko JA, Mizuno Y, Yanai R, Chikama T, Sonoda KH (2010) Expression of semaphorin 3A and its receptors during mouse corneal development. *Biochem Biophys Res Commun* 403: 305–309.
51. Nishida T, Nakagawa S, Awata T, Ohashi Y, Watanabe K, et al. (1983) Fibronectin promotes epithelial migration of cultured rabbit cornea in situ. *J Cell Biol* 97: 1653–1657.
52. Lavker RM, Dong G, Cheng SZ, Kudoh K, Cotsarelis G, et al. (1991) Relative proliferative rates of limbal and corneal epithelia. Implications of corneal epithelial migration, circadian rhythm, and suprabasally located DNA-synthesizing keratinocytes. *Invest Ophthalmol Vis Sci* 32: 1864–1875.
53. Stepp MA, Spurr-Michaud S, Gipson IK (1993) Integrins in the wounded and unwounded stratified squamous epithelium of the cornea. *Invest Ophthalmol Vis Sci* 34: 1829–1844.
54. Wolosin JM, Wang Y (1995) Alpha-2,3 sialylation differentiate the limbal and corneal epithelial cell phenotypes. *Invest Ophthalmol Vis Sci* 36: 2277–2286.
55. EUR-lex website. Available: <http://eur-lex.europa.eu/Notice.do?mode= dbl&lng1 = en,hu&lang = &lng2 = bg,cs,da,de,el,en,es,et,fi,fr,-hu,it,lt,lv,mt,nl,pl,pt,ro,sk,sl,sv,&val = 506440:cs&page = &hwords = null> Accessed 2012 09 14.