

Published in final edited form as:

Tissue Antigens. 2010 July; 76(1): 31–34. doi:10.1111/j.1399-0039.2010.01460.x.

African Americans Exhibit a Predominant Allele in the Midst of Extensive *KIR2DL1* Allelic Diversity

LiHua Hou, **Minghua Chen**, **Bo Jiang**, **Jennifer Ng**, and **Carolyn Katovich Hurley** Departments of Pediatrics and Oncology, CW Bill Young Marrow Donor Recruitment and Research Program, Georgetown University Medical Center, Washington, DC

Abstract

KIR2DL1 alleles were identified by DNA sequencing of the coding region from amplified genomic DNA from 100 random African Americans. The majority of individuals (97%) carried a KIR2DL1 locus. Allele KIR2DL1*00302 was found in 68% of individuals but KIR2DL1*00401, *002, *00303, *006, and *007 were also frequent. Eleven new alleles were described: KIR2DL1*00403, *01101,*01102*012, *013N, *014, *015, *016, *017,*018, and *019. Nine of the novel alleles encoded amino acid substitutions located throughout the receptor; one allele carried a stop codon in the exon encoding the first extracellular domain.

Introduction

The natural killer (NK) cell receptor KIR2DL1 is a cell surface inhibitory receptor that recognizes a subset of HLA-C allelic products carrying Lys80 (1,2). Recognition of its ligand results in signals that prevent the NK cell from responding to normal cells. Loss of its HLA ligand on virally infected or malignant cells removes the inhibitory signal and may result in cytotoxicity or in cytokine production (3). The gene encoding KIR2DL1 is located in a cluster of similar genes in the Leukocyte Receptor Complex (4) although some *KIR* haplotypes lack the *KIR2DL1* gene entirely (5). Fifteen alleles encoding ten variant receptors have been identified for *KIR2DL1* (6,7). Studies of a similar receptor, KIR3DL1, have demonstrated that allelic variants may differ in their level of expression at the cell surface and ligand binding affinity (8). The purpose of this study was to evaluate *KIR2DL1* allelic diversity in a population of random African American individuals.

Materials and Methods

Genomic DNA was isolated from 100 unique and unrelated African Americans from the human variation panel obtained from the National Institute of General Medical Sciences (NIGMS) Human Genetics Resource Center DNA and Cell Line Repository (http://ccr.coriell.org/nigms/) using a QIAamp® DNA Blood Mini Kit (Qiagen, Valencia, CA). Testing for the presence or absence of *KIR2DL1* used polymerase chain reaction (PCR) primers and reaction conditions previously described (9,10,11). A primer pair, described by Gomez-Lozano et al., specific for *KIR2DS1* and *KIR2DL1*004*, *007, and *010 was a useful control (12). For sequencing of exons 1 through 9, the strategy based on PCR amplification of genomic DNA followed by DNA sequence analysis has been described (13) with the following exception: The antisense PCR primer used to obtain the amplicon including exon 4 through exon 9 was 2DL1-E89R

(TGTGAGGAAGATCGATGCCCTAAG). The latter, described by Murdoch et al (14), was

useful to identify a 3'untranslated region variation distinguishing KIR2DL1*0040101 from *0040102. Allele assignments were obtained by comparison with a KIR2DL1 cDNA reference library from ImmunoPolymorphismDatabase (IPD)-KIR Release 2.1.0 (6). In this report, the numbering of nucleotides and codons is based on IPD-KIR unless noted. Novel alleles were isolated using allele specific amplification, cloning and/or by using haplotype specific extraction (2DL1-435G; HaploPrep, Qiagen, Valencia, CA). Sequences were submitted to GenBank and novel allele designations were assigned by the KIR subcommittee of the World Health Organization Nomenclature Committee for Factors of the HLA System (7). Confirmatory sequences of previously described alleles were submitted to the IPD-KIR database and include: KIR2DL1*00303 (Cell GM17132; GenBank GQ406045), KIR2DL1*0040102 (GM17116; GQ844297); KIR2DL1*006 (GM17177; GQ406047), KIR2DL1*007 (GM17178; GQ406043), and KIR2DL1*010 (GM17108, GQ406044). The first four alleles had been previously identified in AfroCaribbean or African American individuals (15,13); the last allele in an individual of European ancestry (14). A phylogenetic tree of KIR2DL1 was constructed using a maximum likelihood technique with a molecular clock (DNAMLK version 3.66). Since the 5' sequence is unknown for some alleles, the tree was based on the nucleotide sequence from codon -17 through codon 327 (16).

HLA allele identification

To identify the *HLA-A,-B,-C* alleles carried by each individual, PCR primers were used to amplify each locus as previously described (17). Applied Biosystems Big Dye terminator chemistry and an Applied Biosystems Models 3730xl DNA analyzer (PE Applied Biosystems, Foster City, CA) were used to obtain the sequences of both strands of exons 2 and 3. IMGT/HLA database 2.21.0 was used for the interpretation of sequencing results. *DRB1* alleles were amplified and identified using the One Lambda LABType® SSO HD Kit (version RSSOH2B1_002, One Lambda, Canoga Park, California) following manufacturer's protocols. Alleles identical in exons 2 and 3 (class I) or exon 2 (*DRB1*) were not resolved. For those class I samples yielding alternative genotypes, either allele specific sequencing primers, allele specific PCR amplification, or HaploPrep (Qiagen, Valencia, CA) was used to link polymorphisms and to identify the specific allele combination. [In-house primer sequences used for all loci are available at www.dodmarrow.org.] Supplemental Table 1 provides the HLA data.

Results and Discussion

Three individuals (3%) showed a complete absence of the KIR2DL1; another study observed 2% of African Americans negative for this locus (18). Forty two individuals carried a single KIR2DL1 coding sequence and 55 were heterozygous. Of the 13 previously described alleles that differ within the coding region of the gene, eight were observed (Table 1). KIR2DL1*00302 was the most common allele being found in 68 individuals. Two other frequent alleles included KIR2DL1*00401 (23 individuals) and KIR2DL1*002 (13 individuals),. These alleles are the ones most frequently observed in studies of other populations (e.g., Japanese (8), Irish (19)). The G to A transition in the 3' untranslated region of KIR2DL1*00401 at nucleotide 34 following the stop codon which distinguishes KIR2DL1*0040101 from *0040102 was observed in this population. We identified the G variant (KIR2DL1*0040101) in 14 individuals and the A variant (KIR2DL1*0040102) in nine individuals; one individual carried both alleles. The A variant was initially observed in an Afro-Carribean population (15). Other alleles observed in multiple individuals, KIR2DL1*006(11 individuals), KIR2DL1*00303 (8 individuals) and KIR2DL1*007(7 individuals), have been previously observed only in populations of African origin (15,19,13).

Eleven novel alleles were characterized. Four of the novel alleles were found in multiple individuals: KIR2DL1*01102 (2 individuals), KIR2DL1*012 (5 individuals), KIR2DL1*014 (3 individuals), and KIR2DL1*019 (2 individuals). One allele carried a synonymous substitution of the common allele KIR2DL1*00401 creating KIR2DL1*00403. Five novel alleles, KIR2DL1*01101, KIR2DL1*01102, KIR2DL1*012, KIR2DL1*013N, and KIR2DL1*019 (codon 114), carried substitutions that were found in other alleles at the locus. For example, KIR2DL1*012 carried a substitution altering the first domain at codon 16 altering a proline to an arginine. Twelve of the 15 KIR2DL1 alleles encode arginine; the other three encode proline at this position. Five novel alleles introduced polymorphism at codons which were previously conserved in all or the majority of alleles altering the amino acid: KIR2DL1*014, KIR2DL1*015, KIR2DL1*016, KIR2DL1*017, KIR2DL1*018, KIR2DL1*019 (codon 226). Four novel alleles, KIR2DL1*01101, KIR2DL1*01102, KIR2DL1*013N, and KIR2DL1*019, exhibited two substitutions. KIR2DL1*013N is interesting because the second substitution in the exon encoding the first extracellular domain creates a termination codon. Other truncated alleles previously described carry termination codons in the exon encoding the second extracellular domain: KIR2DS3*003N (substitution), KIR3DL1*024N (deletion of nucleotide), and KIR3DS1*049N (deletion of nucleotide) (20,21).

The majority of the new alleles described carried nonsynonymous substitutions. The substitutions altered the amino acid sequence of the signal peptide (altered in 2 alleles), domain 1 (1 allele), domain 2 (2 alleles), transmembrane region (2 alleles) and cytoplasmic tail (3 alleles). Only one of the substitutions in the two external domains of KIR introduced a novel amino acid; the other two are observed in other alleles. Based on the crystal structure, none of the extracellular variations appear to impact residues that contact the HLA-C/peptide complex (2). In the cytoplasmic tail, *KIR2DL1*018* (CCA/Pro) has a substitution at codon 282 that was conserved in most alleles (ACA/Thr). This residue falls within the membrane proximal immunoreceptor inhibitory motif (ITIM) motif, adjacent to the tyrosine which is phosphorylated during signaling (22,23). While there is some latitude in amino acids at this position in ITIM motifs, the substitution of a cyclic amino acid (Pro) with properties that affect secondary protein stucture may impact signaling for this form of the receptor.

Figure 1 shows a phylogenetic tree showing the relationships among all of the reported *KIR2DL1* alleles. There are two major clusters of alleles, each containing frequent alleles in the African American population. The novel alleles found in African Americans are found in both of these clusters. The number of *KIR2DL1* alleles observed in this population of 100 individuals is remarkable; 19 alleles with 68% of individuals carrying *KIR2DL1*00302*. In contrast, studies of European American (n=75) or Northern Irish (n=140) populations observed 7 alleles with *KIR2DL1*00302* being found in 63–66% of individuals (13,19).

Identification of HLA-C alleles in the 97 *KIR2DL1* positive African Americans showed that 25 individuals (26%) did not carry an HLA-C allele that would serve as a ligand for KIR2DL1, that is, an HLA-C allelic product with Lys80 (Group 2). In these individuals without the cognate ligand, NK cells expressing, as their only inhibitory KIR receptor, KIR2DL1, would not be "licensed" and are likely to be hyporeactive (24).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This research is supported by funding from the Office of Naval Research N00014-07-2108. The views expressed in this article are those of the authors and do not reflect the official policy or position of the Department of the Navy, the Department of Defense, or the U.S. government. The phylogetic tree was created by Dr. Dongying Wu. We would like to thank him.

References

- Mandelboim O, Reyburn HT, Vales-Gomez M, Pazmany L, Colonna M, Borsellino G, Strominger JL. Protection from lysis by natural killer cells of group 1 and 2 specificity is mediated by residue 80 in human histocompatibility leukocyte antigen C alleles and also occurs with empty major histocompatibility complex molecules. J Exp.Med. 1996; 184:913–922. [PubMed: 9064351]
- Fan QR, Long EO, Wiley DC. Crystal structure of the human natural killer cell inhibitory receptor KIR2DLI-HLA-Cw4 complex. Nature Immunology. 2001; 2:452–460. [PubMed: 11323700]
- 3. Lanier LL. NK cell recognition. Annu.Rev Immunol. 2005; 23:225-274. [PubMed: 15771571]
- 4. Trowsdale J. Genetic and functional relationships between MHC and NK receptor genes. Immunity. 2001; 15:363–374. [PubMed: 11567627]
- Hsu KC, Chida S, Dupont B, Geraghty DE. The killer cell immunoglobulin-like receptor (KIR) genomic region: gene-order, haplotypes and allelic polymorphism. Immunological Reviews. 2002; 190:40–52. [PubMed: 12493005]
- Robinson J, Waller MJ, Stoehr P, Marsh SG. IPD--the Immuno Polymorphism Database. Nucleic Acids Res. 2005; 33:D523–D526. [PubMed: 15608253]
- Marsh SGE, Parham P, Dupont B, Geraghty DE, Trowsdale J, Middleton D, Vilches C, Carrington M, Witt C, Guethlein LA, Shilling H, Garcia CA, Hsu KC, Wain H. Killer-cell immunoglobulin-like receptor (KIR) nomenclature report, 2002. Tissue Antigens. 2003; 62:79–86. [PubMed: 12859599]
- 8. Yawata M, Yawata N, Draghi M, Little AM, Partheniou F, Parham P. Roles for HLA and KIR polymorphisms in natural killer cell repertoire selection and modulation of effector function. J Exp.Med. 2006; 203:633–645. [PubMed: 16533882]
- Hou L, Steiner NK, Chen M, Belle I, Kubit AL, Ng J, Hurley CK. Limited Allelic Diversity of Stimulatory Two Domain Killer Immunoglobulin-Like Receptors. Human Immunology. 2008; 69:174–178. [PubMed: 18396209]
- Uhrberg M, Valiante NM, Shum BP, Shilling HG, Lienert-Weidenbach K, Corliss B, Tyan D, Lanier LL, Parham P. Human diversity in killer cell inhibitory receptor genes. Immunity. 1997; 7:753–763. [PubMed: 9430221]
- Vilches C, Castano J, Gomez-Lozano N, Estefania E. Facilitation of KIR genotyping by a PCR-SSP method that amplifies short DNA fragments. Tissue Antigens. 2007; 70:415–422. [PubMed: 17854430]
- 12. Gomez-Lozano N, Vilches C. Genotyping of human killer-cell immunoglobulin-like receptor genes by polymerase chain reaction with sequence-specific primers: An update. Tissue Antigens. 2002; 59:184–193. [PubMed: 12074708]
- 13. Hou LH, Steiner NK, Chen M, Belle I, Ng J, Hurley CK. KIR2DL1 allelic diversity: four new alleles characterized in a bone marrow transplant population and three families. Tissue Antigens. 2007; 69:250–254. [PubMed: 17493149]
- 14. Murdoch S, Seoud M, Kircheisen R, Mazhar B, Slim R. Detailed gene and allele content analysis of three homozygous KIR haplotypes. Tissue Antigens. 2006; 68:72–77. [PubMed: 16774543]
- Artavanis-Tsakonas K, Eleme K, McQueen KL, Cheng NW, Parham P, Davis DM, Riley EM. Activation of a subset of human NK cells upon contact with Plasmodium falciparum-infected erythrocytes. J Immunol. 2003; 171:5396–5405. [PubMed: 14607943]
- Felsenstein J. Evolutionary trees from DNA sequences: a maximum likelihood approach. J Mol Evol. 1981; 17:368–376. [PubMed: 7288891]
- 17. Tu B, Mack SJ, Lazaro A, Lancaster A, Thomson G, Cao K, Chen M, Ling G, Hartzman R, Ng J, Hurley CK. HLA-A, -B, -C, -DRB1 allele and haplotype frequencies in an African American population. Tissue Antigens. 2007; 69:73–85. [PubMed: 17212710]

18. Du Z, Gjertson DW, Reed EF, Rajalingam R. Receptor-ligand analyses define minimal killer cell Ig-like receptor (KIR) in humans. Immunogenetics. 2007; 59:1–15. [PubMed: 17103212]

- 19. Meenagh A, Gonzalez A, Sleator C, McQuaid S, Middleton D. Investigation of killer cell immunoglobulin-like receptor gene diversity, KIR2DL1 and KIR2DS1. Tissue Antigens. 2008; 72:383–391. [PubMed: 18643963]
- Luo L, Du Z, Sharma SK, Cullen R, Spellman S, Reed EF, Rajalingam R. Chainterminating natural mutations affect the function of activating KIR receptors 3DS1 and 2DS3. Immunogenetics. 2007; 59:779–792. [PubMed: 17646980]
- 21. Norman PJ, Abi-Rached L, Gendzekhadze K, Korbel D, Gleimer M, Rowley D, Bruno D, Carrington CV, Chandanayingyong D, Chang YH, Crespi C, Saruhan-Direskeneli G, Fraser PA, Hameed K, Kamkamidze G, Koram KA, Layrisse Z, Matamoros N, Mila J, Park MH, Pitchappan RM, Ramdath DD, Shiau MY, Stephens HA, Struik S, Verity DH, Vaughan RW, Tyan D, Davis RW, Riley EM, Ronaghi M, Parham P. Unusual selection on the KIR3DL1/S1 natural killer cell receptor in Africans. Nat Genet. 2007; 39:1092–1099. [PubMed: 17694054]
- 22. Burshtyn DN, Scharenberg AM, Wagtmann N, Rajagopalan S, Berrada K, Yi T, Kinet JP, Long EO. Recruitment of tyrosine phosphatase HCP by the killer cell inhibitor receptor. Immunity. 1996; 4:77–85. [PubMed: 8574854]
- Ravetch JV, Lanier LL. Immune inhibitory receptors. Science. 2000; 290:84–89. [PubMed: 11021804]
- 24. Anfossi N, Andre P, Guia S, Falk CS, Roetynck S, Stewart CA, Breso V, Frassati C, Reviron D, Middleton D, Romagne F, Ugolini S, Vivier E. Human NK cell education by inhibitory receptors for MHC class I. Immunity. 2006; 25:331–342. [PubMed: 16901727]

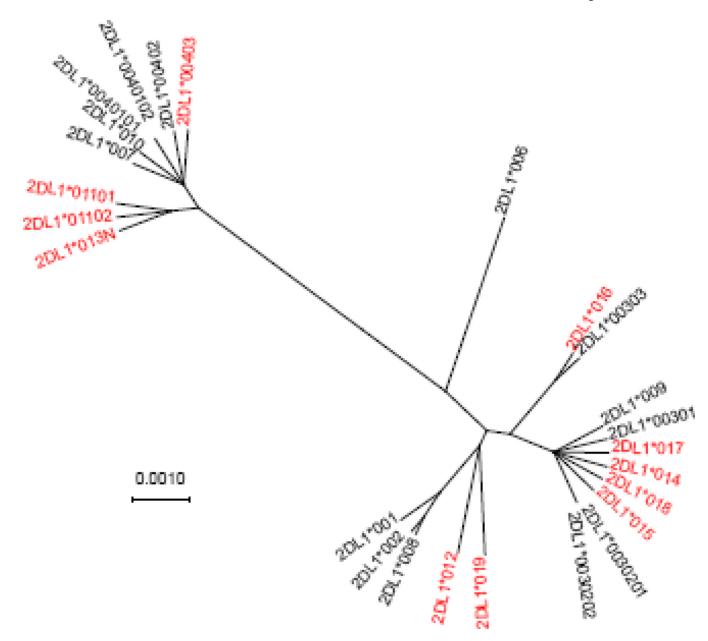


Figure 1. Phylogenetic tree of *KIR2DL1* alleles. Alleles shown in red were described in this study. Previously described *KIR2DL1*005* was not included because it is missing a portion of the nucleotide sequence encoding the signal peptide.

Table 1 Frequency of *KIR2DL1* Alleles in an African American Panel (n=100)

KIR2DL1 Allele	Number Of Individuals Positive (%)	KIR2DL1 Allele	Number Of Individuals Positive (%)
*001	2 (2)	*008	0
*002	13 (13)	*009	0
*00301	0	*010	1(1)
*00302 ^a	68 (68)	*01101 <i>b</i>	1 (1)
*00303	8 (8)	*01102 ^b	2 (2)
*0040101	14 (14)	*012b	5 (5)
*0040102	9 (9)	*013N ^b	1 (1)
*00402	0	*014 ^b	3 (3)
*00403 <i>b</i>	1 (1)	*015 ^b	1 (1)
*005	0	*016 ^b	1 (1)
*006	11 (11)	*017 ^b	1 (1)
*007	7 (7)	*018 b	1 (1)
		*019b	2 (2)

 $[^]a\mathit{KIR2DL1*0030201} \text{ and } \mathit{KIR2DL1*0030202} \text{ were not individually distinguished in this study}.$

 $^{^{}b}$ Novel allele described in this study.

Hou et al.

Table 2

Novel KIR2DL1 alleles observed in African Americans

2DL1*00403 2L		Nucleotide Codon (Amino Acid) ^a		Number	
	2DL1*00401	254 GCG (A) => GCT (A)		EU930369	GM17109
2DL1*01101 2L	2DL1*00401	-17 GTC (V) \Rightarrow TTC (F), 221 CGA (R) \Rightarrow AGA (R)	Signal peptide	EU930368	GM17159
2DL1*01102 2I	2DL1*00401	-17 GTC (V) \Rightarrow TTC (F), 186 TAC (Y) \Rightarrow TAT (Y)	Signal peptide	FJ655851	GM17200 b
2DL1*012 2I	2DL1*001	16 CCC (P) \Rightarrow CGC (R)	Domain 1	EU930370	GM17119 <i>b</i>
2DL1*013N 2L	2DL1*00401	-17 GTC (V) => TTC (F), 35 GAA (E) => TAA (Stop)	Truncated	EU930371	GM17120
2DL1*014 2I	2DL1*00302	179 GGC (G) \Rightarrow AGC (S)	Domain 2	EU930372	GM17123 b
2DL1*015 2L	2DL1*00302	$275 \mathrm{GAC}\mathrm{(D)} \Rightarrow \mathrm{GAA}\mathrm{(E)}$	Cytoplasmic	EU930373	GM17124
2DL1*016 2L	2DL1*00303	296 CGC (R) => TGC (C)	Cytoplasmic	FJ655852	GM17126
2DL1*017 2L	2DL1*00302	221 CGA (R) => CAA (Q)	Transmembrane	FJ655853	GM17131
2DL 1 *018 2L	2DF11*00302	282 ACA (T) => CCA (P)	Cytoplasmic	FJ655854	GM17153
2DL1*019 2I	2DL1*00303	114 CTG (L) => CCG (P), 226 CTG (L) => GTG (V)	Domain 2 Transmembrane	FJ655855	GM17160 b

 2 Most similar known allele => novel allele

 $\frac{b}{(M17102)}$ GM17102, GM17198; KR2DL1*012, GM17198; KR2DL1*012, GM17141, GM17142, GM17166, GM17185; KR2DL1*014, GM17174; KR2LD1*019, GM17102.

Page 8