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Assessing illicit drug use among adults with schizophrenia

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Abstract

Accurate drug use assessment is vital to understanding the prevalence, course, treatment needs, and outcomes among individuals with schizophrenia because they are thought to remain at long-term risk for negative drug use outcomes, even in the absence of drug use disorder. This study evaluated self-report and biological measures for assessing illicit drug use in the Clinical Antipsychotic Trials of Intervention Effectiveness study (N=1460). Performance was good across assessment methods, but differed as a function of drug type, measure, and race. With the Structured Clinical Interview for DSM-III-R as the criterion, self-report evidenced greater concordance, accuracy and agreement overall, and for marijuana, cocaine, and stimulants specifically, than did urinalysis and hair assays, whereas biological measures outperformed self-report for detection of opiates. Performance of the biological measures was better when self-report was the criterion, but poorer for black compared white participants. Overall, findings suggest that self-report is able to garner accurate information regarding illicit drug use among adults with schizophrenia. Further work is needed to understand the differential performance of assessment approaches by drug type, overall and as a function of race, in this population.

Keywords

assessment; drug use; self-report; biological; schizophrenia; race

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1. Introduction

Over one-quarter of adults with schizophrenia evince *lifetime* drug use disorders (Brady and Sinha, 2005; Meister et al., 2010; Regier et al., 1990; Vincenti et al., 2010), a rate higher than in other groups of adults with serious mental illnesses (SMI) or in the general population. Rates of *current* drug use disorders are similarly high (Fowler et al., 1998) and rates of use are even higher (Swartz et al., 2006a). Adults with schizophrenia who use illicit drugs are at greater risk of adverse sequelae (Drake and Mueser, 2001; Mueser et al., 1998; Schiffer et al., 2010; Swartz et al., 2006b). Therefore, accurate assessment is vital to understanding the prevalence, course, treatment needs, and outcomes in this population.

1.1. Assessment approaches

Studies rely, in the main, on self-report, despite perceived problems related to nondisclosure (Carey and Correia, 1998; Carey et al., 2003; Kilpatrick et al., 2000) and that there has been little evaluation of the accuracy of reported substance use in adults with schizophrenia or other SMI (Møller & Linaker, 2010). Researchers have begun including laboratory tests, such as urinalysis and hair assays, as part of a comprehensive assessment strategy, to overcome this potential bias. Urinalysis, though reliable and valid, has a narrow window of detection, usually from 24 hours to seven or more days (Bellack et al., 2006; Wolff et al., 1999). Radioimmunoassay (RIA) of hair can detect drugs from 2 to 3 days after the most recent use to an indefinite period. RIA of hair is thought to be less intrusive than urinalysis and less vulnerable to countermeasures (Swartz et al., 2003). Acceptable levels of sensitivity and specificity have been found for both biological measures (DuPont & Baumgartner, 1995).

Although inclusion of biological measures is thought to improve detection of substance use over self-report (Allgood et al., 1991; Baumgartner et al., 1989; Bessa et al., 2010; de Beaurepaire et al., 2007; DuPont and Baumgartner, 1995; Kelly and Rogers, 1996; Magura and Kang, 1996; Mieczkowski, 2010), research findings remain equivocal (Haddock et al., 2009; Ledgerwood et al., 2008; Lee et al., 2009; Vitale et al., 2006; Welp et al., 2003; Williams and Nowatzki, 2005; Wolford et al., 1999). In a recent study of adults with schizophrenia, for example, combining self-report, with results of urine testing and RIA of hair increased detection rates from 16% (self-report alone) to 38% (Swartz, et al., 2003). In contrast, self-report outperformed results of urine and blood tests in detecting alcohol and drug use disorders in another study of adults with SMI (Wolford, et al., 1999).

Many factors may affect assessment accuracy, including drug type, recency of use, and race/ ethnicity. For instance, compared to self-report, hair assays proved useful in detecting cocaine and heroin in the general population but less useful for detecting marijuana (Fendrich et al., 2004). Under-reporting appears to increase with recency of use, particularly among black respondents (Fendrich, et al., 2004). Similar race effects have been found regarding concordance between self-reported drug use and hair assays (Ledgerwood, et al., 2008; Vignali et al., 2012). Such racial disparities may be attributable to over-detecting drug metabolites in African Americans' hair (Borges et al., 2003; Cone and Joseph, 1996; Fendrich, et al., 2004; Han et al., 2011; Henderson et al., 1998; Kidwell et al., 2000; Kintz et al., 2000; Ledgerwood, et al., 2008; Welp, et al., 2003); however, findings are mixed (Hoffman, 1999; Kelly et al., 2000; Mieczkowski, 2011; Mieczkowski et al., 2002; Mieczkowski and Newel, 2000), and no data have been presented for adults with schizophrenia.

1.2. Present study

Given the preliminary evidence from, but also the limitations of prior studies, research is needed on whether biological measures, as a part of a comprehensive assessment strategy, increase accuracy over self-report. In addition to the limited research focus on adults with schizophrenia, there also has been limited use of the Structured Clinical Interview for DSM-III-R (SCID) (First et al., 1996), a recognized 'gold standard', as the criterion against which accuracy is measured. Instead, self-report is used sometimes as the criterion (Ledgerwood et al., 2008). No research has examined how drug test performance may differ as a function of these criterion measures. Finally, there has been scant examination of whether measures of *use* contribute to false positives and overidentification of *drug use disorders*.

We examined (1) <u>concordance</u> between self-report, urinalysis, hair assays and the SCID; (2) <u>accuracy</u> of self-report, urinalysis, and hair assays with the *SCID as criterion*; (3) <u>accuracy</u> of urinalysis and hair assays with *self-report as criterion*; (4) <u>incremental validity</u> of urinalysis and hair assays over self-report; and (5) <u>correlates of disagreement</u>. All outcomes were evaluated overall and as a function of race; the latter focus being exploratory, and drawing on general population research that has identified racial effects in drug testing.

2. Methods

2.1. Study design and sample

Data were collected as part of the NIMH Clinical Antipsychotic Trials of Intervention Effectiveness (CATIE) study, a randomized clinical trial investigating the cost-effectiveness of atypical and conventional antipsychotic medications. The CATIE study was conducted at over 50 U.S. sites, including academic medical centers and representative community providers. Only 7% of screened patients were excluded, and the study sample resembled a usual-care, noninterventional study sample in its demographic and clinical characteristics (Swanson et al., 2006). Study design and entry criteria details are presented elsewhere (Stroup et al., 2003). We report findings from baseline assessments (i.e., before randomization and initiation of treatments) of 1460 participants. The CATIE protocol was approved by local IRBs, and participants gave written informed consent prior to enrollment; the University of South Florida IRB approved the current study's protocol.

2.2. Measures & Procedures

2.2.1. Illicit drug use—The <u>SCID</u> (First et al., 1996) was used to assess past month drug abuse or dependence. Assessments were completed by SCID-trained and qualified master's level clinicians.

Subjects also <u>self-reported</u> any use of marijuana, cocaine, opiates, PCP, amphetamines, or other illicit drugs over the past 90 days in a separate research interview. Subjects were coded as positive for self-reported use if they responded in the affirmative to any one of the six self-report questions.

<u>Hair specimens</u> were collected by sites and shipped to PsycheMedics Corporation for RIA. This technique assays drugs and their metabolites transferred from capillary circulation through the hair follicle to the internal hair structure (Baumgartner et al., 1989). A tuft of hair about the diameter of pencil lead and 1.5 inches long was cut from the scalp on the back of the head, a specimen that affords assessment of drug use in the preceding three months. A larger volume was removed from participants with short hair. Hair was taken from the chest, arm, or leg if none was present on the head. A positive test was defined as a result more than three standard deviations from the mean of a comparison sample of drug-free individuals. Initial positives were confirmed using gas chromatography/mass spectrometry. The length and amount of hair removed and assessed from each participant was intended to provide a surveillance window of up to 90 days. However, because of the slow growth rate of hair out of the follicle, detection of very recent use (i.e., between 1 and 7 days) is limited (DuPont and Baumgartner, 1995).

Drug urinalysis was performed with a commercially available rapid multiple immunoassay urine drug test, Triage by Biosite. Urine samples have a shorter surveillance window than do RIA of hair samples. Broadly, urine samples show evidence of drug use between 1 and 4 days (DuPont and Baumgartner, 1995; Verstraete, 2004). This timeframe can vary by chronicity of use and type of drug, with extremely chronic cocaine and marijuana use being detected up the 3 weeks after the most recent use (Verstraete, 2004).

All tests assessed for marijuana, cocaine, opiate, and stimulant use. A composite 'all tested drugs' variable was defined as a positive test for at least one type of drug. We additionally created an 'any laboratory test' variable. A positive test was defined as a positive hair or urine test. Participants who tested positive for a prescribed medication detected in hair or urine were considered not to be using.

2.2.2. Other variables—We included other variables in our regression analyses assessing disagreement (described below): *age* (measured continuously), *sex* (male, female), *race* (white, African American/black), *marital status* (married/living together as married, single/ living alone), *education* (high school graduate or higher, non-high school graduate), total PANSS score (measured continuously), and other drug use (yes, no), all of which were obtained during the research interview.

2.3. Data Analysis

2.3.1. Prevalence and concordance—Differences in prevalence were calculated using chi-square tests. We also calculated detection ratios. McNemar's tests were used to determine if the ratios detected by two different measures were significantly different. Bonferonni corrections were made, based on five comparisons per test per drug (α =0.01).

2.3.2. Performance—With the *SCID as the criterion*, we calculated sensitivity, specificity, and conditional probabilities for self-report, hair and urine, as well 'any laboratory test'. We also calculated the Areas Under the Curve (AUCs) of Receiver Operating Characteristics curves and Cohen's kappas. We calculated the same measures for hair and urine with *self-report as the criterion*.

2.3.3. Incremental validity—Hierarchical logistic regression analyses tested the incremental validity of urine and hair over self-report. Significant chi-square change values reflected model improvement and significant odds ratios indicated contributions of individual factors.

2.3.4. Correlates of disagreement—Multivariable logistic regressions were used to examine the relative effects of correlates on disagreement between assessment methods for cocaine and marijuana, the two most frequently endorsed drugs. It is important to note that disagreement regression models were restricted to white and black subjects given our primary interest in potential race-based differences between these two groups, as well as the relatively small number of participants representing other racial/ethnic categories.

All incremental validity and disagreement models controlled for study site as a random effect (Fleiss et al., 1986; Hosmer and Lemeshow, 2000).

3. Results

Over 70% of participants were male (n=1079). Nearly 60% were white (n=874), 35% black (n=513) and 5% 'other' (n=101). Average age was 40.6 years (SD=11.1; median=42). Onequarter (n=374) did not complete high school, 35.1% (n=512) graduated from high school, and the remaining (39.3%, n=573) had some level of education beyond high school. Four out of five (n=1181) were not married nor cohabitating.

3.1. Prevalence

Table 1 presents prevalence by assessment method, overall and by race. Table 1 also lists the valid N for each assessment method. Black participants evidenced higher rates of cocaine use than white participants. They additionally had higher rates of any drug use across methods, except the SCID. White subjects had higher rates of stimulant use according to three of seven methods.

3.2. Concordance and performance

Table 2 presents performance measures for self-report and the laboratory tests compared to the SCID as criterion. Detection ratios ranged from 0.50 for stimulants (urine) to 6.60 for opiates (any laboratory test). Assessment for 'any drug' classifications, regardless of method, resulted in rates higher than obtained from the SCID. Hair showed the greatest overdetection compared to the SCID. Opiates showed the highest detection ratios across methods.

All AUCs, except self-reported opiate use, ranged between 0.71 and 0.88, indicating good accuracy compared with the SCID (Swets, 1988). The highest AUC for marijuana was found for self-report; 'any laboratory test' had the highest AUCs for both cocaine and opiates; hair had the highest AUC for stimulants. Sensitivity was highest for self-report (0.85) and lowest for urine (0.55) detecting any drug. All approaches were more specific than they were sensitive. Correct classification ranged from 77.0% for 'all tested drugs' (hair) to 99.3% for stimulants (urine). Kappas generally indicated moderate agreement (i.e., κ =0.41–0.60); others, however, only demonstrated fair (i.e., κ =0.21–0.40) or slight agreement (i.e., κ =0.00–0.20) (Landis and Koch, 1977). Assessments of opiate use demonstrated the lowest and assessments of stimulant use, the highest kappas, respectively.

Table 3 presents performance measures, with self-report as the criterion. Detection ratios were generally lower than found with the SCID as criterion. Urine detection was significantly less than self-report (detection ratios < 1), except for opiate use, which was not significantly different than self-report. Hair detected higher rates of use than self-report for cocaine and 'all tested drugs'; 'any laboratory test' detected higher rates for cocaine, opiates and 'all tested drugs'.

The highest AUCs for marijuana and opiate use were found for 'any laboratory test'; hair alone had the highest AUC for both cocaine and stimulant use. Sensitivity was low for most biological tests. Hair for cocaine had the highest sensitivity, whereas urine for opiates and stimulants had the lowest sensitivity. Again, biological measures were more specific than they were sensitive. Correct classification ranged from 79.5% for 'all tested drugs' (hair) to 98.5% for stimulants (any laboratory test). All kappas demonstrated moderate to fair agreement. As before, assessments of opiate use consistently demonstrated the lowest kappas, whereas assessments of stimulant use demonstrated the highest kappas (except urine).

We also stratified analyses by race. With the SCID as criterion, hair produced the highest sensitivity (92.0%) for black participants; however, specificity was poor (66.7%) and only

68.9% were classified correctly. The AUC for hair detecting cocaine use among blacks was 0.79 and κ =0.20. Self-report for black participants appeared more valid for cocaine: sensitivity=84.4%, specificity=91.0%, classified correctly=90.5%, AUC=0.88, and κ =0.56. (Full results not shown, but are available.) Regardless of criterion measure, black participants evinced lower rates of cases classified correctly for biological tests.

3.3. Incremental validity

First we examined whether the biological measures added to the capacity of self-report to detect marijuana use, overall and by race. Self-report was added in Step 1 of each of three models with SCID as the criterion for all, white, and black participants, respectively. All three models were significant (Table 4). In Step 2, laboratory tests were added in one block. Accuracy improved significantly, although there were small increases in R^2 . Across participants, urine and hair demonstrated incremental validity; for white participants, only urine contributed to the model; for black participants, neither urine nor hair were significant factors.

We repeated these analyses for cocaine use (Table 5). All Step 1 models were significant. Adding the laboratory tests significantly improved the models for the entire sample and white, but not black participants. Hair was associated with unique contributions overall and for white, but not black participants. Urine was not a significant factor.

3.4. Correlates of disagreement

We explored disagreement with the SCID and self-report in separate models. Two cocaine models were run: (1) SCID, no; self-report, yes; any laboratory test, yes; and (2) SCID, no; self-report, no; any laboratory test, yes. Three marijuana models were run: (1) SCID, no; self-report, yes; any laboratory test, yes; (2) SCID, no; self-report, no; any laboratory test, yes; and (3) SCID, no; self-report, yes; any laboratory test, no. (Other combinations were endorsed too infrequently.)

Table 6 (first column) shows that older age, male sex and other drug use (i.e., besides cocaine) increased the likelihood of a positive self-report and any positive laboratory test, in the absence of a positive SCID. This model also found negative effects for white compared to black participants and for those who completed at least high school compared to those who did not. The second column shows correlates of disagreement between any positive laboratory test, but a negative SCID and self-report. There were stronger effects for both race and sex in this model, compared to the prior model; however, the age effect was not significant.

Table 7 shows the results for marijuana. The only significant race effect was found for participants who were positive for any laboratory test, but negative for the SCID and self-report: white participants were significantly less likely to be in this classification than black participants.

We repeated these analyses focusing on disagreement between self-report and the laboratory tests (Table 8). There was a significant race effect for both cocaine and marijuana for individuals who did not self-report use, but were positive for urine or hair.

4. Discussion

This study evaluated self-report and biological measures for assessing illicit drug use among adults with schizophrenia. Though results differed somewhat by drug, overall performance was good across assessment methods. Assessments were more specific than they were

sensitive. Results also highlighted differences in performance as a function of assessment method and participant race. We explore these findings in more detail below.

Can adults with SMI accurately self-report substance use (Drake et al., 1990; Goldfinger et al., 1996; Shaner et al., 1993)? Our findings support the validity of self-report data among adults with schizophrenia and provide evidence for their utility over laboratory tests. Self-report demonstrated greater concordance, accuracy and agreement with the SCID overall, and for marijuana, cocaine, and stimulants specifically, than did urinalysis and hair assays. Though biological measures outperformed self-report for detection of opiates, this finding should be interpreted with caution due to low rates of opiate use. Prior investigations (Wolford, et al., 1999) have documented laboratory tests' limited utility in identifying alcohol use disorders among individuals with SMI; the current study extends those results to assessment of illicit drug use among adults with schizophrenia when self-report data are available.

This study is the first to statistically test the utility of biological measures above and beyond self-reported information. Though they contributed statistically to the models, the practical gains were small. Instead, self-report appears sufficient to garner accurate information regarding illicit drug use among adults with schizophrenia. That being said, we are unable to determine how subjects' knowledge of the laboratory tests affected the accuracy of their self-reported use. Indeed, the value of such tests may not be in their contribution of unique information, but rather in their contribution to increased disclosure rates.

Some prior studies have identified racial disparities in assessment accuracy, although none have examined this issue among adults with schizophrenia. Our findings highlighted three effects. First, across assessment methods, higher rates of drug use were detected among black compared to white participants. Second, regardless of whether the SCID or self-report was the criterion, performance of biological measures was better for white than black participants. Third, whereas laboratory results added to the prediction of disordered use among white participants, these tests were not useful for that purpose among black participants.

A unique feature of this study was our use of both the SCID and self-report as criterion measures. As anticipated, measures of use (self-report, laboratory tests) overdetected disordered use *compared to the SCID*. In contrast, urinalysis underdetected marijuana, cocaine, and stimulant use while hair assays typically overdetected use *compared to self-report*. These findings would seem to indicate that the presence of *drug use* in adults with schizophrenia is a poor proxy for *disordered drug use* (Carey, 2002; Carey and Correia, 1998).

Limitations of present study include our inability to look at incremental validity, race effects, and correlates of disagreement for opiates, stimulants and amphetamines due to low base rates. We also were unable to determine rates of inter-rater reliability on the SCID data. There also were 323 cases (22%) who were missing hair data at baseline, a rate higher than any other assessment method. In addition, there were no self-report data regarding intensity or recency of use in the CATIE study. Next, we limited our racial analyses to white and black participants. Future research could expand upon this dichotomy and examine potential differences in other racial and ethnic groups. This might help clarify whether or not potential racial differentially across racial and ethnic groups. Also, it is important to note that willful nondisclosure may vary as a function of social norms or legal status regarding acceptability of drug use, which may affect drug use self-report accuracy and limit generalizability of findings from one setting to another. Future research on the assessment of

illicit drug use among adults with schizophrenia may find utility in a more nuanced focus on other illness-specific factors, as well. Whereas we controlled for total PANSS score, education level and concurrent evidence of any other drug use, other characteristics of the participants (e.g., cognitive deficit, persecutory ideation, antipsychotic medications, etc.), their environment, and the assessors themselves might affect the accuracy of drug tests. Finally, we only can speculate on reasons for the lower accuracy of biological measures.

Because of the implications for research and practice, further work is needed to understand the differential performance of assessment approaches among individuals with schizophrenia. For instance, case reports and other small studies suggest that antipsychotic medications may affect the reliability and validity of biological measures; this potential relationship should be more closely examined with an eye towards refining drug testing in this population. A related issue regarding differential performance of assessment approaches has to do with the concept of a "gold standard". While prior research has identified the SCID as a gold standard assessment approach, and thus, we treated the SCID as that referent measure, it is not without its own limitations (e.g., susceptibility to willful non-disclosure), particularly in research studies where less is known about a participant. Finally, given that a number of people evinced non-disordered drug use, research should seek to provide more nuance regarding differences in course, treatment needs, and outcomes among these non-disordered users compared to those with disordered drug use.

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References

- Allgood, CC.; Sniegoski, LT.; Welch, MJ. The analysis of human hair for drugs of abuse. Paper presented at the In Proceedings of the 39th ASMS Conference on Mass Spectrometry and Allied Topics; Nashville, TN. 1991 May 19–24.
- Baumgartner WA, Hill VA, Bland WH. Hair analysis for drugs of abuse. Journal of Forensic Sciences. 1989; 34:1433–1453.
- Bellack AS, Bennett ME, Gearon JS, Brown CH, Yang Y. A randomized clinical trial of a new behavioral treatment for drug abuse in people with severe and persistent mental illness. Arch Gen Psychiatry. 2006; 63:426–432. [PubMed: 16585472]
- Bessa MA, Mitsuhiro SS, Chalem E, Barros MM, Guinsburg R, Laranjeira R. Underreporting of use of cocaine and marijuana during the third trimester of gestation among pregnant adolescents. Addictive Behaviors. 2010; 35:266–269.10.1016/jaddbeh2009 [PubMed: 19896774]
- Borges CR, Roberts JC, Wilkins DG, Rollins DE. Cocaine, benxoylecgonine, amphetamine, and Nacetylamphetamine binding to melanin subtypes. Journal of Analytic Toxicology. 2003; 27:125– 134.
- Brady KT, Sinha R. Co-occurring mental and substance use disorders: The neurobiological effects of chronic stress. The American Journal of Psychiatry. 2005; 162(8):1483–1493.10.1176/appi.ajp. 162.8.1483 [PubMed: 16055769]
- Carey KB. Clinically useful assessments: substance use and comorbid psychiatric disorders. Behaviour Research and Therapy. 2002; 40(11):1345–1361.10.1016/s0005-7967(02)00039-6 [PubMed: 12384329]
- Carey KB, Carey MP, Chandra PS. Psychometric evaluation of the alcohol use disorders identification test and short drug abuse screening test with psychiatric patients in India. The Journal of Clinical Psychiatry. 2003; 64(7):767–774. [PubMed: 12934976]
- Carey KB, Correia CJ. Severe mental illness and addictions: Assessment considerations. Addictive Behaviors. 1998; 23(6):735–748.10.1016/s0306-4603(98)00063-x [PubMed: 9801713]
- Cone, EJ.; Joseph, RE. The potential for bias in hair testing for drugs of abuse. In: Kintz, P., editor. Drug testing in hair. Boca Raton, FL: CRC Press; 1996. p. 69-93.
- de Beaurepaire R, Lukasiewicz M, Beauverie P, Castéra S, Dagorne O, Espaze R, et al. Comparison of self-reports and biological measures for alcohol, tobacco, and illicit drugs consumption in psychiatric inpatients. European Psychiatry. 2007; 22:540–548.10.1016/j.eurpsy2007.05.001 [PubMed: 17596918]
- Drake RE, Mueser KT. Managing comorbid schizophrenia and substance abuse. Current Psychiatry Reports. 2001; 3:418–422. [PubMed: 11559480]
- Drake RE, Osher FC, Noordsy DL, Hurlbut SC, Teague GB, Beaudett MS. Diagnosis of alcohol use disorders in schizophrenia. Schizophrenia Bulletin. 1990; 16:57–67. [PubMed: 2333482]
- DuPont RL, Baumgartner WA. Drug testing by urine and hair analysis: Complementary features and scientific issues. Forensic Science International. 1995; 70:63–76. [PubMed: 7860037]
- Fendrich M, Johnson TP, Wislar JS, Hubbell A, Spiehler V. The utility of drug testing in epidemiological research: results from a general population survey. Addiction. 2004; 99(2):197– 208.10.1111/j.1360-0443.2003.00632.x [PubMed: 14756712]
- First, MB.; Spitzer, RL.; Gibbon, M.; Williams, JBW. Structured Clinical Interview for Axes I and II DSM-IV Disorders-Patient Edition (SCID-I/P). New York: Biometrics Research Department, New York State Psychiatric Institute; 1996.

- Fleiss JL, Williams JB, Dubro AF. The logistic regression analysis of psychiatric data. Journal of Psychiatric Research. 1986; 20:145–209.
- Fowler IL, Carr VJ, Carter NT, Lewin TJ. Patterns of Current and Lifetime Substance Use in Schizophrenia. Schizophrenia Bulletin. 1998; 24(3):443–455. [PubMed: 9718636]
- Goldfinger SM, Schutt RK, Seidman LJ, Turner WM, Penk WE, Tolomiczenko GS. Alternative measures of substance abuse among homeless mentally illpersons in the cross-section and over time. Journal of Nervous & Mental Disease. 1996; 184:667–672. [PubMed: 8955679]
- Haddock G, Cross Z, Beardmore R, Tarrier N, Moring J, Barrowclough C. Assessing illicit substance use in schizophrenia: The relationship between self report and detection by hair analysis. Schizophrenia Research. 2009; 114:180–181.10.1016/j.schres.2009.07.020 [PubMed: 19713083]
- Han E, Paulus MP, Wittman M, Chung H, Song JM. Hair analysis and self-report of methamphetamine use by methamphetamine dependent individuals. Journal of Chromatography B. 2011; 879:541– 547.10.1016/j.jchromb.2011.01.002
- Henderson GL, Harkey MR, Zhou C, Jones RT, Jacob P. Incorporation of isotopically labeled cocaine into human hair: Race as a factor. Journal of Analytical Toxicology. 1998; 22(2):156–165. [PubMed: 9547413]
- Hoffman BH. Analysis of race effects on drug-test results. Journal of Occupational and Environmental Medicine. 1999; 41:612–616. [PubMed: 10412103]
- Hosmer, D.; Lemeshow, S. Applied logistic regression. 2. New York: John Wiley and Sons; 2000.
- Kelly KS, Rogers R. Detection of misreported drug use in forensic populations: An overview of hair analysis. Bulletin of the American Academy of Psychiatry & the Law. 1996; 24:85–94. [PubMed: 8891324]
- Kelly RC, Mieczkowski T, Sweeney SA. Hair analysis for drugs of abuse. Hair color and race differentials or systematic differences in drug preferences? Forensic Science International. 2000; 107:63–86. [PubMed: 10689562]
- Kidwell DA, Lee EH, DeLauder SF. Evidence for bias in hair testing and procedures to correct bias. Forensic Science International. 2000; 107:39–61. [PubMed: 10689561]
- Kilpatrick B, Howlett M, Sedgwick P, Ghodse AH. Drug use, self-report and urinalysis. Drug and Alcohol Dependence. 2000; 58:111–116. [PubMed: 10669061]
- Kintz P, Cirimele V, Ludes B. Pharmacological criteria that can affect the detection of doping agents in hair. Forensic Science International. 2000; 107:325–334. [PubMed: 10689584]
- Landis JR, Koch GG. The measurement of observer agreement for categorical data. Biometrics. 1977; 33(1):159–174. [PubMed: 843571]
- Ledgerwood DM, Goldberger BA, Risk NK, Lewis CE, Kato Price R. Comparison between self-report and hair analysis of illicit drug use in a community sample of middle-aged men. Addictive Behaviors. 2008; 33(9):1131–1139.10.1016/j.addbeh.2008.04.009 [PubMed: 18547737]
- Lee S, Park, Yang W, Han E, Choe S, Lim M, Chung H. Estimation of the measurement of methamphetamine and amphetamine in hair analysis. Forensic Science International. 2009; 185:59–66.10.1016/j.forsciint.2008.12.012 [PubMed: 19179027]
- Magura S, Kang SY. Validity of self-reported drug use in high risk populations: A meta-analytical review. Substance Use and Misuse. 1996; 31:1131–1153. [PubMed: 8853234]
- Meister K, Burlon M, Rietschel L, Gouzoulis-Mayfrank E, Bock T, Lambert M. Dual diagnosis psychosis and substance use disorders in adolescents, part 1. Fortschritte der Neurologie-Psychiatrie. 2010; 78(2):81–89. [PubMed: 20146152]
- Mieczkowski T. Urinalysis and hair analysis for illicit drugs of driver applicants and drivers in the trucking industry. Journal or Forensic and Legal Medicine. 2010; 17:254–260.10.1016/j.jflm. 2010.02.014 [PubMed: 20569951]
- Mieczkowski T. Assessing the potential for racial bias in hair analysis for cocaine: Examining the relative risk of positive outcomes when comparing urine samples to hair samples. Forensic Science International. 2011; 206:29–34.10.1016/j-forsciint.2010.06.011 [PubMed: 20619981]
- Mieczkowski T, Lersch M, Kruger M. Police drug testing, hair analysis, and the issue of race bias. Criminal Justice Review. 2002; 27(1):124–140.
- Mieczkowski T, Newel R. Statistical examination of hair color as potential biasing factor in hair analysis. Forensic Science International. 2000; 107:13–38. [PubMed: 10689560]

- Møller T, Linaker OM. Using brief self-reports and clinician scales to screen for substance use disorders in psychotic patients. Nordic Journal of Psychiatry. 2010; 64(2):130– 135.10.3109/08039480903274423 [PubMed: 19883188]
- Mueser KT, Drake RE, Wallace MA. Dual diagnosis: A review of etiological theories. Addictive Behaviors. 1998; 23:717–734. [PubMed: 9801712]
- Regier DA, Farmer ME, Rae DS, Locke BZ, Keith SJ, Judd LL, Goodwin FK. Comorbidity of mental disorders with alcohol and other drug abuse. Results from the Epidemiologic Catchment Area (ECA) Study. Journal of the American Medical Association. 1990; 264(19):2511–2518. [PubMed: 2232018]
- Schiffer B, Bernhard WM, Scherbaum N, Forsting M, Wiltfang J, Leygraf N, Gizewski ER. Impulsivity-related brain volume deficits in schizophrenia-addiction comorbidity. Brain: A Journal of Neurology. 2010; 133:3093–3103.10.1093/brain/awq153 [PubMed: 20647266]
- Shaner A, Khalsa ME, Roberts LJ, Wilkins JN, Anglin D, Hsieh SC. Unrecognized cocaine use among schizophrenic patients. American Journal of Psychiatry. 1993; 150:758–762. [PubMed: 8480822]
- Stroup TS, McEvoy JP, Swartz MS, Byerly MJ, Glick ID, Canive JM, McGee MF, Simpson GM, Stevens MC, Lieberman JA. The National Institute of Mental Health Clinical Antipsychotic Trials of Intervention Effectiveness (CATIE) project: Schizophrenia trial design and protocol development. Schizophrenia Bulletin. 2003; 29(1):15–31. [PubMed: 12908658]
- Swanson JW, Swartz MS, Van Dorn RA, Elbogen EB, Wagner HR, Rosenheck R, Stroup S, Lieberman J. A national study of violent behavior in persons with schizophrenia. Archives of General Psychiatry. 2006; 63:490–499. [PubMed: 16651506]
- Swartz MS, Swanson JW, Hannon MJ. Detection of illicit substance use in persons with schizophrenia. Psychiatric Services. 2003; 54:891–895. [PubMed: 12773606]
- Swartz MS, Wagner HR, Swanson JW, Stroup TS, McEvoy JP, Canive JM, Miller DD, Reimherr F, McGee M, Kahn A, Van Dorn RA, Rosenheck RA, Lieberman JA. Substance use in persons with schizophrenia: Baseline prevalence and correlates from the NIMH CATIE study. The Journal of Nervous and Mental Disease. 2006a; 194:164–172. [PubMed: 16534433]
- Swartz MS, Wagner HR, Swanson JW, Stroup TS, McEvoy JP, McGee M, Miller DD, Reimherr F, Khan A, Canive JM, Lieberman JA. Substance use and psychosocial functioning in schizophrenia among new enrollees in the NIMH CATIE Study. Psychiatric Services. 2006b; 57:1110–1116. [PubMed: 16870961]
- Swets J. Measuring the accuracy of diagnostic systems. Science. 1988; 240:1285–1293. [PubMed: 3287615]
- Verstraete AG. Detection times of drugs of abuse in blood, urine, and oral fluid. Therapeutic Drug Monitoring. 2004; 26:200–205. [PubMed: 15228165]
- Vignali C, Stramesi C, Vecchio M, Groppi A. Hair testing and self-report of cocaine use. Forensic Science International. 2012; 215:77–80.10.1016/jforsciint.2011.05.007 [PubMed: 21645979]
- Vincenti A, Ventriglio A, Baldessarini RJ, Talamo A, Fitzmaurice G, Centorrino F. Characteristics and clinical changes during hospitalization in bipolar and psyhotic disorder patients with versus without substance-use disorders. Pharmacopsychiatry. 2010; 43(6):225–232.10.1055/ s-0030-1254153 [PubMed: 20652858]
- Vitale SG, van de Mheen H, van de Wiel A, Garretsen HFL. Substance use among emergency room patients: Is self-report preferable to biochemical markers? Addictive Behaviors. 2006; 31:1661– 1669.10.1016/j.addbeh.2005.12.011 [PubMed: 16446045]
- Welp EAE, Bosman I, Langendam MW, Totté M, Maes RAA, Van Ameijden EJC. Amount of selfreported illicit drug use compared to quantitative hair test results in community-recruited young drug users in Amsterdam. Addiction. 2003; 98(7):987–994.10.1046/j.1360-0443.2003.00421.x [PubMed: 12814504]
- Williams RJ, Nowatzki N. Validity of adolescent self-report of substance use. Substance Use & Misuse. 2005; 40(3):1–13. [PubMed: 15702646]
- Wolff K, Farrell M, Marsden J, Monteiro MG, Ali R, Welch S, Strang J. A review of biological indicators of illicit drug use, practical considerations and clinical usefulness. Addiction. 1999; 94(9):1279–1298.10.1046/j.1360-0443.1999.94912792.x [PubMed: 10615715]

Wolford GL, Rosenberg SD, Drake RE, Mueser KT, Oxman TE, Hoffman D, Vidaver RM, Luckoor R, Carrieri KL. Evaluation of methods for detecting substance use disorder in persons with severe mental illness. Psychology of Addictive Behaviors. 1999; 13(4):313.

Prevalence of illicit drug use by mode of assessment and race for adults with schizophrenia

| Measure | n (%) | White | Black | χ^2 value |
|--|-------------|-------------|-------------|---------------------|
| SCID Drug Abuse (N=1459) | | | | |
| Marijuana | 76 (5.2%) | 38 (4.4%) | 33 (6.4%) | 2.88 |
| Cocaine | 42 (2.9%) | 12 (1.4%) | 27 (5.3%) | 17.87 [†] |
| Opiate | 2 (0.1%) | 2 (0.2%) | 0 (0.0%) | 1.18 |
| Stimulant | 13 (0.9%) | 13 (1.5%) | 0 (0.0%) | 7.71 [†] |
| All tested drugs | 113 (7.8%) | 57 (6.5%) | 50 (9.8%) | 4.67 |
| SCID Drug Dependence (N=1459) | | | | |
| Marijuana | 43 (3.0%) | 21 (2.4%) | 21 (4.1%) | 3.13 |
| Cocaine | 37 (2.5%) | 6 (0.7%) | 28 (5.5%) | 30.73 <i>†</i> |
| Opiate | 7 (0.5%) | 4 (0.5%) | 3 (0.6%) | 0.10 |
| Stimulant | 7 (0.5%) | 5 (0.6%) | 2 (0.4%) | 0.22 |
| All tested drugs | 80 (5.5%) | 34 (3.9%) | 42 (8.2%) | 11.49 [†] |
| Combined SCID Drug Abuse and Dependence (N=1459) | | | | |
| Marijuana | 108 (7.4%) | 53 (6.1%) | 49 (9.6%) | 5.74 |
| Cocaine | 69 (4.7%) | 18 (2.1%) | 46 (9.0%) | 34.98 [†] |
| Opiate | 7 (0.5%) | 4 (0.5%) | 3 (0.6%) | 0.10 |
| Stimulant | 18 (1.2%) | 16 (1.8%) | 2 (0.4%) | 5.25 |
| All tested drugs | 169 (11.6%) | 80 (9.2%) | 80 (15.6%) | 13.09 * |
| Self-report (N=1448) | | | | |
| Marijuana | 247 (17.1%) | 141 (16.2%) | 95 (18.9%) | 1.66 |
| Cocaine | 123 (8.5%) | 39 (4.5%) | 79 (15.7%) | 51.31 [†] |
| Opiate | 20 (1.4%) | 15 (1.7%) | 3 (0.6%) | 3.12 |
| Stimulant | 29 (2.0%) | 22 (2.5%) | 5 (1.0%) | 3.87 |
| All tested drugs | 322 (22.2%) | 171 (19.6%) | 134 (26.6%) | 9.13 [†] |
| <i>Urine</i> (N=1457) | | | | |
| Marijuana | 130 (8.9%) | 67 (7.7%) | 57 (11.1%) | 4.71 |
| Cocaine | 91 (6.3%) | 21 (2.4%) | 64 (12.5%) | 56.99 <i>†</i> |
| Opiate | 27 (1.9%) | 19 (2.2%) | 3 (0.6%) | 5.23 |
| Stimulant | 9 (0.6%) | 7 (0.8%) | 1 (0.2%) | 2.07 |
| All tested drugs | 227 (15.6%) | 100 (11.5%) | 111 (21.7%) | 26.03 [†] |
| Hair | | | | |
| Marijuana (N=990) | 147 (14.9%) | 81 (12.1%) | 61 (22.1%) | 15.43 <i>†</i> |
| Cocaine (N=1133) | 201 (17.7%) | 62 (8.4%) | 130 (37.6%) | 137.80 [†] |
| Opiate (N=1136) | 30 (2.6%) | 16 (2.2%) | 13 (3.7%) | 2.26 |
| Stimulant (N=1134) | 29 (2.6%) | 24 (3.3%) | 2 (0.6%) | 7.30 [†] |
| All tested drugs (N=1137) | 327 (28.8%) | 151 (20.4%) | 165 (47.3%) | 83 147 |
| Any laboratory test (N=1458) | | | | |

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| Measure | n (%) | White | Black | χ^2 value |
|------------------|-------------|-------------|-------------|---------------------|
| Marijuana | 216 (14.8%) | 108 (12.4%) | 100 (19.5%) | 12.96 [†] |
| Cocaine | 228 (15.6%) | 69 (7.9%) | 148 (28.9%) | 107.74 [†] |
| Opiate | 48 (3.3%) | 29 (3.3%) | 14 (2.7%) | 0.37 |
| Stimulant | 30 (2.1%) | 25 (2.9%) | 2 (0.4%) | 10.34 * |
| All tested drugs | 404 (27.7%) | 184 (21.1%) | 201 (39.3%) | 53.15 <i>†</i> |

 $\stackrel{\dagger}{=}$ Bonferonni adjusted p-value (p<0.01)

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Table 2

Measurement performance of drug use assessments for adults with schizophrenia: SCID as criterion

| | | | SCID | 1 Month Drug | Abuse or Dep | endence | |
|---------------------|-------------------|-----------|---------------------------|--------------|--------------|-------------------------------------|---------------|
| Measure | Detection ratio | AUC (SE) | AUC 95%CI | Sensitivity | Specificity | Percent Classified Correctly | Cohen's Kappa |
| Self-report | | | | | | | |
| Marijuana | 2.31° | .87 (.02) | .83–.91 [†] | 86.0% | 88.4% | 88.2% | 0.46 |
| Cocaine | 1.80° | .87 (.03) | .8293 † | 79.4% | 95.0% | 94.2% | 0.54 |
| Opiate | 2.80° | .67 (.13) | .39–.88 | 28.6% | 98.8% | 95.2% | 0.14 |
| Stimulant | 1.66 | .88 (.06) | <i>‡</i> 66.− <i>11</i> . | 77.8% | %0.66 | 96.8% | 0.59 |
| All tested drugs | 1.91° | .86 (.02) | .82–.89 $\mathring{\tau}$ | 85.1% | 86.0% | 85.9% | 0.51 |
| Urine | | | | | | | |
| Marijuana | 1.20 | .74 (.03) | .68–.80 <i>†</i> | 53.7% | 94.7% | 91.4% | 0.44 |
| Cocaine | 1.34 | .73 (.04) | .65–.81 \mathring{r} | 50.0% | 95.9% | 93.6% | 0.40 |
| Opiate | $3.80 ^{\circ}$ | .74 (.13) | .4999 | 50.0% | 98.3% | 98.1% | 0.17 |
| Stimulant | 0.50 | .71 (.08) | .55–.86 † | 41.2% | %6.66 | 99.3% | 0.53 |
| All tested drugs | 1.34° | .73 (.02) | .68–.77 † | 55.4% | 89.6% | 85.5% | 0.39 |
| Hair | | | | | | | |
| Marijuana | $2.01 ^{\circ}$ | .73 (.04) | .66–.80 $^{+}$ | 58.3% | 88.5% | 86.1% | 0.32 |
| Cocaine | 3.31° | .85 (.03) | .78–.91 <i>†</i> | 84.1% | 84.9% | 84.9% | 0.25 |
| Opiate | 5.20° | .74 (.16) | .42–1.00 | 50.0% | 97.5% | 97.3% | 0.11 |
| Stimulant | 2.17° | .87 (.07) | .74–.99 <i>†</i> | 75.0% | 98.5% | 98.2% | 0.52 |
| All tested drugs | 2.48° | .75 (.02) | ⁺70–.79 [†] | 75.4% | 77.2% | 77.0% | 0.32 |
| Any laboratory test | | | | | | | |
| Marijuana | 2.00 [†] | .78 (.03) | $.7283^{ t}$ | 65.7% | 89.3% | 87.4% | 0.38 |
| Cocaine | 3.32° | .78 (.03) | .72–.85 $^{\pm}$ | 69.6% | 87.0% | 86.2% | 0.27 |
| Opiate | 6.60° | .77 (.12) | .5499 | 57.1% | 97.0% | 96.8% | 0.14 |
| Stimulant | 1.75° | .88 (.06) | ⁺76–.99 [†] | 76.5% | 98.8% | 98.6% | 0.55 |
| All tested drugs | 2.39 t | .78 (.02) | .74–.81 $^{\acute{\tau}}$ | 76.3% | 78.6% | 78.4% | 0.34 |

 $$ mathematical adjusted p-value (p<0.01) \\ \uparrow = Bonferonni adjusted p-value (p<0.01)$

\$watermark-text

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Measurement performance of drug use assessments for adults with schizophrenia: Self-report as criterion

| | | | | Setf | -report | | |
|----------------------------------|---------------------|-----------|------------------------|-------------|---------------|------------------------------|---------------|
| Measure | Detection ratio | AUC (SE) | AUC 95%CI | Sensitivity | Specificity | Percent Classified Correctly | Cohen's Kappa |
| Urine | | | | | | | |
| Marijuana | 0.52° | .70 (.02) | .66–.74 [†] | 41.9% | 97.8% | 88.3% | 0.49 |
| Cocaine | 0.74° | .72 (.03) | .6778 [†] | 47.2% | 97.6% | 93.0% | 0.51 |
| Opiate | 1.36 | .67 (.07) | .47–.76 $^{\div}$ | 25.0% | 98.5% | 97.7% | 0.20 |
| Stimulant | 0.30° | .62 (.06) | .5075 | 25.0% | %6 .66 | 98.4% | 0.37 |
| All tested drugs | $0.70 ^{\circ}$ | .72 (.02) | .68–.76 <i>†</i> | 49.5% | 94.1% | 84.3% | 0.49 |
| Hair | | | | | | | |
| Marijuana | 0.87 | .70 (.03) | .65–.75 <i>†</i> | 48.8% | 91.9% | 84.6% | 0.43 |
| Cocaine | 2.08° | .86 (.02) | .81–.90 $\acute{\tau}$ | 83.7% | 87.7% | 87.3% | 0.44 |
| Opiate | 1.86 | .64 (.08) | .48–.79 | 29.4% | 97.8% | 97.1% | 0.20 |
| Stimulant | 1.30 | .84 (.06) | $.7295^{\circ}$ | 68.0% | 98.9% | 98.3% | 0.62 |
| All tested drugs | 1.30° | .76 (.02) | $.7279^{\circ}$ | 68.8% | 82.5% | 79.5% | 0.46 |
| Any laboratory test | | | | | | | |
| Marijuana | 0.87 | .75 (.02) | .7179 | 56.3% | 93.7% | 87.3% | 0.53 |
| Cocaine | 1.84° | .81 (.02) | .76–.85 $^{\div}$ | 71.5% | 89.5% | 87.9% | 0.44 |
| Opiate | 2.36 † | .74 (.07) | .6088 | 50.0% | 97.3% | 96.9% | 0.28 |
| Stimulant | 1.05 | .82 (.06) | .7193 | 64.3% | 99.2% | 98.5% | 0.61 |
| All tested drugs | 1.25° | .77 (.02) | $.7480^{\circ}$ | 6.69 | 84.3% | 81.1% | 0.50 |
| $\dot{\tau}$ =Bonferonni adjuste | d p-value (p<0.01) | | | | | | |

Logistic Regression Analyses Testing Incremental Validity for Assessment of Marijuana Use

|) |) | | | | • | | | 2 | | | | |
|----------------------|-------------|-----------------|--|----------------------|----------|----------------------------|--|----------------------------|----------|----------|---|---------------------|
| | | I II V | Participants (n | 1 = 983) | | White | e Participants | (n=669) | | Black | K Participants | (n=271) |
| Step 1 | Mc | ¢ til ləbc | $\chi^2(2) = 176.98$ | ***, $R^2 = 0.41$ | Mc | del fit χ | $^{2}(2) = 143.77$ | ***, $R^2 = 0.50$ | M | odel fit | $\chi^2(2) = 31.68^*$ | **, $R^2 = 0.26$ |
| | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI |
| Self-Report | 3.73 *** | 0.35 | 41.69 | 21.19–82.02 | 4.73 *** | 0.61 | 113.74 | 34.22–378.03 | 2.65 *** | 0.50 | 14.12 | 5.32–37.50 |
| Step 2 | W | odel fit 2 Z | $\chi^2(4) = 203.47$ $\Delta \chi^2(2) = 26.45$ | $^{***}, R^2 = 0.46$ | Mc | Δ del fit λ | $\chi^2(4) = 158.16^{-2}$ $\chi^2(2) = 14.39$ | $^{***}_{***}, R^2 = 0.55$ | W | odel fit | $\chi^{2}(4) = 36.92^{*}$ $\Delta \chi^{2}(2) = 5.2$ | $^{**}, R^2 = 0.30$ |
| | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI |
| Self-Report | 3.02 *** | 0.38 | 20.43 | 9.76-42.79 | 3.97 *** | 0.65 | 52.72 | 14.69–189.18 | 2.29 *** | 0.53 | 9.90 | 3.52–27.83 |
| Urine | 1.29 *** | 0.35 | 3.63 | 1.82 - 7.23 | 1.06^* | 0.44 | 2.89 | 1.22-6.83 | 1.15 | 0.63 | 3.16 | 0.92 - 10.90 |
| Hair | 0.68^* | 0.34 | 1.97 | 1.01 - 3.85 | 0.81 | 0.44 | 2.25 | 0.95–5.31 | 0.36 | 0.57 | 1.44 | 0.47-4.37 |
| Notes. | | | | | | | | | | | | |
| $^{*}_{P<.05.}$ | | | | | | | | | | | | |
| $^{**}_{P<.01.}$ | | | | | | | | | | | | |
| $^{***}_{P<.001.}$ | | | | | | | | | | | | |
| R^2 values are l | Vagelkerke. | | | | | | | | | | | |

Logistic Regression Analyses Testing Incremental Validity for Assessment of Cocaine Use

|) |) | - | |) | • | | | | | | | |
|----------------------|---------------------|-----------------|---|----------------------------------|----------|-----------------------|-----------------------------------|------------------------------------|----------|----------|--|----------------------|
| | | I II V | Participants (n | (=1125) | | White | e Participants | (n=736) | | Blac | k Participants | (n=341) |
| Step 1 | Mc | λdel fit χ | $\chi^2(2) = 159.25$ | ***, $R^2 = 0.48$ | Mc | del fit | $\chi^2(2) = 53.82^{-1}$ | ***, $R^2 = 0.43$ | M | odel fit | $\chi^2(2) = 71.26^3$ | ***, $R^2 = 0.47$ |
| | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI |
| Self-Report | 4.56 ^{***} | 0.44 | 95.69 | 40.34-226.97 | 4.75 *** | 0.70 | 115.13 | 29.23-453.47 | 4.17 *** | 0.65 | 64.68 | 17.98–232.67 |
| Step 2 | Mc | χ til ləbc | $\chi^{2}(4) = 169.92$ $\Delta \chi^{2}(2) = 10.6$ | $e^{**}, R^2 = 0.51$ e^{**} | Mc | odel fit _∕ | $\chi^2(4) = 63.96^{\frac{1}{2}}$ | $^{***}_{4}, R^2 = 0.51_{4}^{***}$ | W | odel fit | $\chi^{2}(4) = 76.35^{-1}$ $\Delta \chi^{2}(2) = 5.1$ | $^{***}, R^2 = 0.50$ |
| | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI | β | S.E. | Odds Ratio | Odds Ratio 95% CI |
| Self-Report | 3.40 *** | 0.55 | 30.02 | 10.24-88.03 | 4.04 *** | 0.83 | 56.56 | 11.11–288.03 | 3.31 *** | 0.80 | 27.48 | 5.76-131.10 |
| Urine | 0.62 | 0.45 | 1.85 | 0.76-4.49 | -1.48 | 0.92 | 0.23 | 0.04 - 1.39 | 1.12 | 09.0 | 3.05 | 0.95 - 9.80 |
| Hair | 1.43^{*} | 0.60 | 4.16 | 1.29–13.47 | 2.63 ** | 0.85 | 13.86 | 2.62–73.37 | 0.63 | 0.99 | 1.87 | 0.27-12.95 |
| Notes. | | | | | | | | | | | | |
| $^{*}_{P<.05.}$ | | | | | | | | | | | | |
| $^{**}_{P<.01.}$ | | | | | | | | | | | | |
| $^{***}_{P<.001.}$ | | | | | | | | | | | | |
| R^2 values are l | Nagelkerke. | | | | | | | | | | | |

Cross-site multivariable models: Correlates of cocaine disagreement between modes of assessment for adults with schizophrenia who did not meet SCID criteria

| | Coc | aine |
|--------------------------------|--|---|
| | Self-report, yes; any laboratory test, yes | Self-report, no; any laboratory test, yes |
| | OR (95% CI) | OR (95% CI) |
| Age | 1.01 (1.00 – 1.02) ** | 1.00 (0.99 – 1.01) |
| Male | 1.27 (1.01 – 1.59)* | 1.73 (1.28 – 2.36) *** |
| White | 0.55 (0.45 - 0.68) *** | 0.31 (0.23 – 0.41) *** |
| Cohabitation | 1.13 (0.89 – 1.44) | 1.13 (0.82 – 1.57) |
| High school or higher | 0.77 (0.62 – 0.95)* | 0.67 (0.50 – 0.88)** |
| PANSS total score | 1.00 (0.99 – 1.00) | 1.00 (1.00 – 1.01) |
| Evidence of any other drug use | 2.41 (1.95 – 2.97) *** | 1.80 (1.37 – 2.37) *** |

p<0.05;

** p<0.01

*** p<0.001

\$watermark-text

Cross-site multivariable models: Correlates of marijuana disagreement between modes of assessment for adults with schizophrenia who did not meet SCID criteria

| | | Marijuana | |
|--------------------------------|--|---|--|
| | Self-report, yes; any laboratory test, yes | Self-report, no; any laboratory test, yes | Self-report, yes; any laboratory test, no |
| | OR (95% CI) | OR (95% CI) | OR (95% CI) |
| Age | $0.99 (0.98 - 1.00)^*$ | 1.00 (0.99 – 1.01) | 0.97 (0.95 -0.98)*** |
| Male | 1.18 (0.88 – 1.57) | 0.98 (0.73 – 1.30) | 0.96 (0.72 -1.29) |
| White | 1.09 (0.83 – 1.42) | 0.59 (0.45 – 0.77) *** | 1.22 (0.92 -1.60) |
| Cohabitation | 1.01 (0.74 – 1.39) | 1.25 (0.92 – 1.70) | 0.86 (0.62 -1.18) |
| High school or higher | 1.15 (0.87 – 1.53) | 0.73 (0.55 – 0.96)* | 1.12 (0.85 -1.48) |
| PANSS total score | 0.99 (0.99 – 1.00) | 1.00 (0.99 – 1.00) | 1.00 (0.99 -1.01) |
| Evidence of any other drug use | 2.87 (2.18 – 3.79) *** | 1.33 (0.99 – 1.78) | 1.75 (1.32 –2.33) **** |

_____p<0.05;

** p<0.01

*** p<0.001

\$watermark-text

Cross-site multivariable models: Correlates of disagreement between self-report and combined labs for adults with schizophrenia

| | C | ocaine | Ma | rijuana |
|--------------------------------|--|--|--|--|
| | Self-report, yes; any laboratory test, no | Self-report, no; any laboratory test, yes | Self-report, yes; any laboratory test, no | Self-report, no; any laboratory test, yes |
| | OR (95% CI) | OR (95% CI) | OR (95% CI) | OR (95% CI) |
| Age | 1.00 (0.99–1.01) | 1.00 (0.99–1.01) | 0.96 (0.95–0.98)*** | 1.00 (0.99–1.01) |
| Male | 1.07 (0.85–1.34) | 1.64 (1.20–2.23)** | 1.11 (0.82–1.51) | 1.04 (0.78–1.40) |
| White | 0.82 (0.66–1.00) | 0.32 (0.24–0.42)*** | 1.05 (0.79–1.40) | 0.52 (0.40–0.68) *** |
| Cohabitation | 0.72 (0.56–0.93)* | 1.12 (0.81–1.55) | 0.79 (0.56–1.11) | 1.30 (0.95–1.78) |
| High school or higher | 1.02 (0.82–1.27) | 0.68 (0.51–0.90)** | 1.12 (0.84–1.51) | 0.63 (0.48–0.83) *** |
| PANSS total score | 1.00 (1.00-1.01) | 1.00 (1.00–1.01) | 1.00 (1.00–1.01) | 1.00 (0.99–1.01) |
| Evidence of any other drug use | 2.16 (1.75–2.66)*** | 1.93 (1.46–2.54)*** | 2.57 (1.92–3.43)*** | 1.21 (0.90–1.63) |

* p<0.05;

** p<0.01

*** p<0.001