

Review

The evolution and comparative neurobiology of endocannabinoid signalling

Maurice R. Elphick*

School of Biological and Chemical Sciences, Queen Mary University of London, Mile End Road, London E1 4NS, UK

CB₁- and CB₂-type cannabinoid receptors mediate effects of the endocannabinoids 2-arachidonoyl-glycerol (2-AG) and anandamide in mammals. In canonical endocannabinoid-mediated synaptic plasticity, 2-AG is generated postsynaptically by diacylglycerol lipase alpha and acts via presynaptic CB₁-type cannabinoid receptors to inhibit neurotransmitter release. Electrophysiological studies on lampreys indicate that this retrograde signalling mechanism occurs throughout the vertebrates, whereas system-level studies point to conserved roles for endocannabinoid signalling in neural mechanisms of learning and control of locomotor activity and feeding. CB₁/CB₂-type receptors originated in a common ancestor of extant chordates, and in the sea squirt *Ciona intestinalis* a CB₁/CB₂-type receptor is targeted to axons, indicative of an ancient role for cannabinoid receptors as axonal regulators of neuronal signalling. Although CB₁/CB₂-type receptors are unique to chordates, enzymes involved in biosynthesis/inactivation of endocannabinoids occur throughout the animal kingdom. Accordingly, non-CB₁/CB₂-mediated mechanisms of endocannabinoid signalling have been postulated. For example, there is evidence that 2-AG mediates retrograde signalling at synapses in the nervous system of the leech *Hirudo medicinalis* by activating presynaptic transient receptor potential vanilloid-type ion channels. Thus, postsynaptic synthesis of 2-AG or anandamide may be a phylogenetically widespread phenomenon, and a variety of proteins may have evolved as presynaptic (or postsynaptic) receptors for endocannabinoids.

Keywords: cannabinoid; anandamide; 2-AG; invertebrate; vertebrate; CRIP1a

1. PREFACE

On 29 March 2001, a review titled ‘The neurobiology and evolution of cannabinoid signalling’ was published in *Philosophical Transactions of the Royal Society B* [1]. It was the first review on cannabinoid signalling to be published in this journal. Since then, the field of research on cannabinoid signalling has grown exponentially. Accordingly, this review is one of 15 reviews that collectively form an entire journal issue devoted to ‘Endocannabinoids in nervous system health and disease’. Writing of the 2001 review required a survey of approximately 3000 articles, which was challenging but feasible. Ten years later, by the end of 2011, the PubMed database had over 11 000 articles that could be found using the search term ‘cannabinoid*’. Clearly, it is no longer feasible to comprehensively review this field of research in a journal article. Even a large book devoted to cannabinoid biology could not cover the range of papers on this topic. Therefore, it is necessary in a review such as this to focus on a specific aspect of cannabinoid biology and the theme here is ‘The evolution and comparative neurobiology of endocannabinoid signalling’, focusing largely on articles that have been published since 2001.

*m.r.elphick@qmul.ac.uk

One contribution of 15 to a Theme Issue ‘Endocannabinoids in nervous system health and disease’.

In discussing the evolution and comparative neurobiology of endocannabinoid signalling, it is necessary to first provide an overview of current understanding of mechanisms of endocannabinoid signalling in the group of animals in which this system was discovered—the mammals. It is fitting therefore that on 29 March 2001, three landmark experimental papers were also published that transformed our understanding of endocannabinoid signalling in the mammalian nervous system. Independently, three research groups obtained evidence that postsynaptic depolarization of principal neurons in the hippocampus or cerebellar cortex triggers postsynaptic synthesis of endocannabinoids, which then act presynaptically to cause CB₁-mediated inhibition of neurotransmitter release [2–4]. Thus, a mechanism of synaptic plasticity mediated by retrograde endocannabinoid signalling was discovered. The concept that endocannabinoids might act as retrograde synaptic signalling molecules had been proposed three years earlier, on the basis of our neuroanatomical observations [5], and a model of this putative signalling mechanism was presented in the 2001 review article [1]. But it was the three other papers published on 29 March 2001 that converted a hypothesis into a textbook principle. Within a decade, the field of research on endocannabinoid signalling has moved from a marginal position to the centre stage of twenty-first century neuroscience. So, looking back, 29 March 2001 can be thought of

as a turning point for cannabinoid research and indeed it has been referred to as a *dies mirabilis* for the field [6].

2. INTRODUCTION TO ENDOCANNABINOID SIGNALLING

(a) *Discovery of CB₁ and CB₂ cannabinoid receptors*

The existence of cannabinoid receptors in the brain was first inferred from the stereoselective pharmacological actions of Δ^9 -tetrahydrocannabinol (Δ^9 -THC), the psychoactive constituent of cannabis and other cannabinoid-type compounds [7]. However, demonstration of the existence of specific cannabinoid binding sites in the brain using the radiolabelled cannabinoid ³H-CP-55,940 provided the first solid evidence that cannabinoid receptors exist in the brain [8]. Molecular characterization of a protein that confers cannabinoid binding sites on rodent brain cell membranes provided the definitive proof of a receptor and revealed a 473-residue G-protein-coupled receptor (GPCR) [9], which is now referred to as CB₁. This nomenclature distinguishes CB₁ from a related GPCR known as CB₂, which is predominantly associated with immune cells [10]. Thus, in humans and other mammals, there are two G-protein-coupled cannabinoid receptors, CB₁ and CB₂, and analysis of CB₁-knockout mice and CB₂-knockout mice indicates that these two receptors are largely responsible for mediating the pharmacological effects of Δ^9 -THC in mammals [11–13].

(b) *Endocannabinoids and enzymes involved in endocannabinoid biosynthesis and inactivation*

The discovery of CB₁ and CB₂ pointed to the existence of endogenous ligands for these receptors and two such ‘endocannabinoids’ have been identified—*N*-arachidonylethanolamide (‘anandamide’) and *sn*-2-arachidonoylglycerol (2-AG) [14–16].

2-AG is synthesized in the brain by the enzyme diacylglycerol lipase (DAGL)α, which catalyses cleavage of 2-AG from arachidonic acid containing diacylglycerols (DAGs) [17–19]. A second DAGL that is related to DAGLα based on sequence similarity has been identified and is known as DAGLβ [17]. However, while DAGLβ can catalyse the formation of 2-AG *in vitro* [17], comparative analysis of the brain content of 2-AG in DAGLα- and DAGLβ-knockout mice indicates that the contribution of DAGLβ to 2-AG biosynthesis in adult brain is much less significant compared with DAGLα [18,19]. 2-AG is inactivated by the enzyme monoacylglycerol lipase (MAGL), which cleaves 2-AG into arachidonic acid and glycerol [20–22]. Approximately, 85 per cent of brain 2-AG hydrolase activity is attributable to MAGL, while the remaining 15 per cent is largely attributed to the α/β hydrolases ABH6 and ABH12 [23].

The mechanisms by which anandamide is synthesized in the brain are not yet fully characterized. *In vitro* studies suggested that anandamide may be synthesized by a two-step enzymatic pathway wherein a Ca²⁺-activated *N*-acyltransferase transfers a *sn*-1 arachidonoyl acyl group of a phospholipid onto the amine of phosphatidylethanolamine (PE) to generate

N-acyl PE (NAPE) and then NAPE is converted by a phospholipase D (NAPE-PLD) into anandamide and phosphatidic acid [24–27]. However, the levels of anandamide in brains from NAPE-PLD-knockout mice are not significantly different from wild-type mice, arguing against a role for NAPE-PLD in anandamide biosynthesis in the brain. The levels of long-chain saturated *N*-acylethanolamines (NAEs) are substantially reduced in NAPE-PLD-knockout mice though, indicating that the primary function of NAPE-PLD in the brain may be in biosynthesis of these molecules [28]. The physiological roles of long-chain saturated NAEs in the brain are unknown, but localization of NAPE-PLD in the axons and axon terminals of sub-populations of neurons in the brain has provided a neuroanatomical framework for further investigation of this issue [29].

Other enzymatic pathways have also been implicated in the biosynthesis of anandamide [30–35] but, as yet, definite proof that these are involved in *in vivo* production of anandamide in the brain has not been forthcoming. It is possible that multiple and potentially interacting pathways are involved, which may make it difficult to pinpoint roles for particular enzymes.

While our knowledge of mechanisms of anandamide biosynthesis in the brain remains incomplete, enzymes that catalyse inactivation of anandamide have been identified. In 1996, Cravatt *et al.* [36] identified an enzyme known as fatty acid amide hydrolase (FAAH), which converts anandamide to arachidonic acid and ethanolamine, and subsequent analysis of FAAH-knockout mice and mice treated with selective FAAH inhibitors have demonstrated that FAAH has a major role in regulation of anandamide levels in the brain [37,38]. In humans, but not in rodents, there is a second FAAH-like enzyme, which is known as FAAH-2 [39]. Analysis of the biochemical properties of FAAH-2 reveals that it associated with lipid droplets in cells and hydrolyses anandamide at rates 30–40% of those of FAAH [40]. Furthermore, cyclooxygenase-2 (COX-2) also contributes to the metabolism of anandamide in neurons and other cell types [41,42].

Lastly, evidence for and against the existence of proteins involved in transport of endocannabinoids has been reported [43,44] and recently it was proposed that a catalytically silent isoform of FAAH (FAAH-like anandamide transporter or FLAT) may drive anandamide transport into neurons [45].

(c) *Putative regulators of cannabinoid receptor signalling*

The existence of proteins that regulate the activity of GPCRs is well established. These include proteins such as GPCR kinases, which phosphorylate serine and threonine residues in GPCR C-terminal tails following G-protein dissociation, and arrestins, which bind to C-terminally phosphorylated GPCRs and then block interaction with G-proteins and mediate receptor internalization [46]. However, these are generic GPCR-interacting proteins that regulate the activity of many GPCRs. In addition to these generic

GPCR-interacting proteins, other proteins that interact only with specific GPCRs have been identified. For example, the melanocortin receptor accessory protein mediates targeting of MC2-type melanocortin receptors to the cell surface in adrenal cells [47–49].

The first report of candidate cannabinoid receptor interacting proteins (CRIPs) was published in 2007 [50]. Deletion of the C-terminal region of the CB₁ receptor had been found to alter CB₁ signalling [51], and it was postulated that accessory proteins binding to this region of the receptor may modulate CB₁ activity. Using a polypeptide corresponding to the C-terminal 55 residues of the CB₁ receptor as bait, a yeast two-hybrid screen was used to identify potential interacting partner proteins expressed in human brain. A 128-residue protein was identified as a positive hit, and analysis of its sequence revealed that it is encoded by a gene containing four exons (1, 2, 3a and 3b) that is subject to alternative splicing, with exons 1, 2 and 3b encoding the 128-residue protein and exons 1, 2 and 3a encoding a 164-residue protein [50]. Biochemical evidence that both the 164-residue protein and the 128-residue protein interact with the C-terminal tail of CB₁ was obtained and, accordingly, these two proteins were named cannabinoid receptor interacting protein 1a (CRIP1a) and cannabinoid receptor interacting protein 1b (CRIP1b), respectively. Furthermore, co-expression of CRIP1a or CRIP1b with CB₁ in superior cervical ganglion neurons revealed that CRIP1a, but not CRIP1b, suppresses CB₁-mediated tonic inhibition of voltage-gated Ca²⁺ channels, providing evidence of a role for CRIP1a in regulation of CB₁ signalling [50]. More recently, it has been reported that co-expression of CRIP1a with CB₁ receptors in cultured cortical neurons alters the actions of cannabinoids in a neuroprotection assay, inhibiting the neuroprotective effect of a CB₁ agonist (WIN55,212-2) and conferring responsiveness to the CB₁ antagonist SR141716 as a neuroprotective agent [52]. These data provide further evidence that CRIP1a may regulate CB₁ signalling. However, as yet, evidence that CRIP1a regulates CB₁ signalling *in vivo* has not been reported and for this we may have to await the characterization of CRIP1a gene-knockout mice.

(d) Endocannabinoid signalling as a mediator of synaptic plasticity in the nervous system

Thus far, a catalogue of proteins that act as cannabinoid receptors or regulators of cannabinoid receptor signalling or catalyse biosynthesis/inactivation of endocannabinoids has been presented. However, from a neurobiological perspective, our interest is in understanding how these proteins work together at the cellular level to enable neurophysiological mechanisms to operate. The term ‘cannabinoid or endocannabinoid signalling’ first appears in the literature in 1998 [5,53] but prior to this, much was already known about the distribution of the CB₁ receptor in the brain and the effects of cannabinoids on neurotransmitter release. On the basis of an analysis of the distribution of cannabinoid binding sites (using ³H-C-P-55,940 autoradiography) combined with lesion

studies and analysis of patterns of CB₁ gene expression (using mRNA *in situ* hybridization), it was concluded that the CB₁ receptor is targeted to the axons and axon terminals of neurons in the brain [54–57]. This was then confirmed by a series of immunocytochemical studies published in 1998 [5,58,59]. This presynaptic targeting of CB₁ receptors in neurons was consistent with electrophysiological studies demonstrating that cannabinoids cause inhibition of neurotransmitter release [60]. Furthermore, evidence that endocannabinoids are released in response to neuronal stimulation was reported [61], which suggested that endocannabinoids act as intercellular (not intracellular) signalling molecules. A logical extrapolation of these anatomical and physiological observations was that endocannabinoids are synthesized postsynaptically and act as retrograde synaptic signalling molecules [5], which was subsequently proved to be correct.

Depolarization of principal neurons in several brain regions causes CB₁-mediated inhibition of presynaptic release of the excitatory neurotransmitter glutamate (depolarization-induced suppression of excitation or DSE) and/or CB₁-mediated inhibition of presynaptic release of the inhibitory neurotransmitter GABA (depolarization-induced suppression of inhibition or DSI) [2–4]. DSE and DSI are not observed in DAGL α -knockout mice, indicating that 2-AG mediates these mechanisms of synaptic plasticity [18,19]. DAGL α is concentrated postsynaptically in dendritic spines that are apposed to CB₁-expressing axon terminals [62], which is consistent with the notion that 2-AG is synthesized postsynaptically but acts presynaptically. The 2-AG degrading enzyme MAGL is localized presynaptically in the axons of neurons, most notably in glutamatergic neurons [20,63] and the duration of DSI and DSE in MAGL-knockout mice is prolonged when compared with wild-type mice, indicating that MAGL controls the temporal dynamics of 2-AG/CB₁-mediated retrograde synaptic signalling [64,65]. Accordingly, the MAGL inhibitor JZL184 also prolongs the duration of DSI and DSE in mice [66].

Endocannabinoid signalling also mediates long-term depression (LTD) of synaptic transmission. For example, stimulation of cortical glutamatergic input to the striatum causes activation of postsynaptic metabotropic glutamate receptors, leading to endocannabinoid/CB₁-mediated LTD of transmission at excitatory cortico-striatal synapses [67]. Endocannabinoid/CB₁-mediated LTD has subsequently been reported in other brain regions and there is evidence that, as with DSE and DSI, it is postsynaptic formation of 2-AG that mediates this particular form of long-term synaptic plasticity [68].

The physiological roles of anandamide as an endogenous agonist for CB₁ receptors in the central nervous system are currently less well characterized when compared with 2-AG. Evidence that anandamide may also mediate retrograde signalling at synapses has also been reported [69] and it has been suggested that anandamide may mediate tonic endocannabinoid signalling, thereby performing a role that is distinct from the transient and stimulated release of 2-AG [70]. Furthermore, there is evidence

that anandamide may mediate mechanisms of synaptic plasticity via CB₁-independent molecular pathways. Thus, postsynaptic elevation of intracellular anandamide levels is thought to cause LTD via a mechanism mediated by the cation channel transient receptor potential vanilloid 1 (TRPV1), which results in internalization of postsynaptic AMPA-type glutamate receptors [6,69,71].

While our knowledge and understanding of the roles of endocannabinoid signalling at the synaptic level have improved dramatically over the last decade, there is still much work to be done in linking processes at this level to the systems level. The CB₁ receptor is widely distributed in the brain but not all neurons express CB₁, so why do particular neural pathways in the brain use endocannabinoid signalling to regulate synaptic transmission, while others do not? Proximate answers to this question will surely emerge as we learn more about the patterns of electrical activity that trigger synthesis of endocannabinoids in different regions of the brain and the net behavioural consequences of this. However, ultimate answers will only be obtained by comparative analysis of the physiological roles of the endocannabinoid system, which may shed light on how over evolutionary timescales the endocannabinoid system has been recruited as a regulator of neural processes in different lineages. Some roles of the endocannabinoid system in brain function may be ancient and highly conserved; other roles may have evolved more recently as neural adaptations that are unique to particular lineages. If we are to understand endocannabinoid signalling, it will be necessary to explore the physiological roles of this system throughout the animal kingdom, and already important insights are beginning to emerge from comparative studies on non-mammalian animals, as discussed below.

3. THE PHYLOGENETIC DISTRIBUTION AND EVOLUTION OF ENDOCANNABINOID SIGNALLING

Canonical endocannabinoid signalling in the mammalian nervous system, as it is currently understood, could be characterized as a process in which postsynaptic formation of 2-AG by DAGL α in response to depolarization-induced Ca²⁺ elevation or activation of metabotropic receptors coupled via G-proteins to phospholipase C (PLC) causes inhibition of neurotransmitter release when 2-AG binds to presynaptic CB₁ receptors, with the spatial and temporal dynamics of this signalling mechanism being controlled by presynaptic degradation of 2-AG by MAGL. Thus, in investigating the evolutionary origins of endocannabinoid signalling, one could specifically investigate the phylogenetic distribution DAGL α , MAGL and CB₁-type receptors. However, this would be a rather narrowly defined view of endocannabinoid signalling in the nervous system. It is true to say that at present our understanding of the physiological role of anandamide as an endogenous ligand for CB₁ receptors is incomplete by comparison with 2-AG. Nevertheless, the phylogenetic distribution of enzymes involved or implicated in anandamide biosynthesis or inactivation is of interest. Likewise, it is important to investigate the

phylogenetic distribution of proteins implicated as regulators of CB₁ signalling such as CRIP1a and CRIP1b, because this may inform understanding of their proposed functions. While CB₁ is by far the most abundant cannabinoid receptor in the mammalian nervous system, there is evidence that CB₂ may have important roles in neural functions [72] and therefore the phylogenetic distribution of CB₂ receptors is also of interest from a neurobiological perspective.

(a) *The phylogenetic distribution of CB₁/CB₂-type cannabinoid receptors*

As mediators of the pharmacological effects of Δ^9 -THC and the physiological actions of endocannabinoids, the G-protein-coupled cannabinoid receptors CB₁ and CB₂ are the focal points for a phylogenetic survey of endocannabinoid signalling. CB₁ and CB₂ share more sequence similarity with each other (approx. 44%) than with any other mammalian GPCRs, indicating that they originated by duplication of a common ancestral gene (i.e. they are paralogs). Furthermore, the relatively low level of sequence similarity shared by CB₁ and CB₂ receptors in mammals is suggestive of an evolutionarily ancient gene duplication. Analysis of the phylogenetic distribution of CB₁ and CB₂ receptors indicates that the gene duplication that gave rise to these two receptors occurred in a common ancestor of extant vertebrates, probably concurrently with a whole-genome duplication event. Thus, CB₁ and CB₂ receptor genes can be found in the genomes of non-mammalian tetrapod vertebrates (amphibians, e.g. *Xenopus tropicalis*; birds, e.g. *Gallus gallus*) and in bony fish (e.g. the zebrafish *Danio rerio*) [73,74]. Interestingly, in teleosts, duplicate copies of CB₁ or CB₂ genes are found, attributable to a genome duplication in a common ancestor of teleosts followed by subsequent lineage-specific retention/loss of duplicate genes. Thus, in the zebrafish *D. rerio*, there is one CB₁ gene and two CB₂ genes, whereas in the puffer fish *Fugu rubripes*, there are two CB₁ genes and one CB₂ gene. However, the functional significance of the differential retention of duplicate CB₁ or CB₂ genes in different teleost lineages is currently unknown [73,74].

To date, there are no published reports of CB₁ and CB₂ genes in the most basal of the extant vertebrate orders—the chondrichthyes (e.g. sharks and rays) and the agnathans (e.g. lampreys and hagfish). However, unpublished genome sequence data are available for the elephant shark *Callorhynchus milii* (<http://eshark.genome.imcb.a-star.edu.sg/>) and the sea lamprey *Petromyzon marinus* (http://genome.wustl.edu/genomes/view/petromyzon_marinus), and in both species, a gene encoding a CB₁-type receptor can be found. Interestingly, a CB₂-type receptor gene is not evident in the currently available genome sequence data, which may simply reflect incomplete sequence data or perhaps more interestingly may reflect loss of CB₂ receptor genes in these basal vertebrates.

Genes encoding CB₁/CB₂-type receptors have been found in the invertebrate groups that are most closely related to the vertebrates (urochordates, e.g. CiCBR in *Ciona intestinalis*; cephalochordates, e.g. BfCBR in

Branchiostoma floridae) but not in the non-chordate invertebrate phyla [73,75–78]. Thus, it appears that CB₁/CB₂-type receptors are unique to the phylum Chordata and, as such, they have a rather restricted phylogenetic distribution in the animal kingdom.

(b) The phylogenetic distribution of diacylglycerol lipases

The antiquity of DAGLs is evident in the strategy that led to the discovery of the mammalian enzymes DAGL α and DAGL β —the sequence of a DAGL originally identified in the bacterium *Penicillium* was used to identify related proteins in BLAST searches of the human genome sequence [17]. This indicates that DAGLs are an ancient enzyme family that originated in prokaryotes. Submission of human DAGL α and human DAGL β as query sequences in BLAST searches of the GenBank protein database reveals orthologues of both isoforms in deuterostomian invertebrates and protostomian invertebrates. Thus, the gene duplication that gave rise to DAGL α or DAGL β dates back at least as far as the common ancestor of extant bilaterian animals.

(c) The phylogenetic distribution of monoacylglycerol lipase

MAGL was originally discovered on account of its role in fat metabolism [79] and subsequently, it was proposed that MAGL may regulate 2-AG levels in the brain [20]. Submission of human MAGL as a query sequence in BLAST searches of the GenBank protein database reveals orthologues in a wide range of animal species, including deuterostomian invertebrates, protostomian invertebrate and basal invertebrates such as cnidarians (*Nematostella vectensis*) and placozoans (*Trichoplax adhaerens*). Therefore, MAGL was present in the common ancestor of extant animals. However, there has been loss of MAGL in some lineages; for example, in *Drosophila* and other insects. Interestingly, MAGL is also found in poxviruses, which is probably a consequence of horizontal gene transfer from host species [80].

(d) The phylogenetic distribution of NAPE-PLD as an enzyme implicated in anandamide biosynthesis

Although analysis of NAPE-PLD-knockout mice indicates that NAPE-PLD is not responsible for synthesis of the bulk of anandamide in the brain [28], this does not rule out the possibility that NAPE-PLD participates in anandamide biosynthesis in other organs and organisms. Therefore, it is of interest to determine the phylogenetic distribution of NAPE-PLD with respect to the evolution of endocannabinoid signalling. Orthologues of NAPE-PLD are found throughout the animal kingdom, in non-mammalian vertebrates, deuterostomian invertebrates (e.g. the sea urchin *Strongylocentrotus purpuratus*), protostomian invertebrates (e.g. the crustacean *Daphnia pulex* and the nematode *Caenorhabditis elegans*) and basal invertebrates such as the cnidarian *N. vectensis* and the placozoan *T. adhaerens*. However, as with MAGL, there has been loss of NAPE-PLD in some lineages. For example, orthologues of NAPE-PLD are not present in *Drosophila* and other

insects, the urochordate *C. intestinalis* and the cephalochordate *B. floridae*. The functional significance of NAPE-PLD loss in some animal lineages is currently unknown. However, biochemical analysis of species that lack NAPE-PLD may provide useful new insights on NAPE-PLD-independent mechanisms of *N*-acylethanolamine biosynthesis.

(e) The phylogenetic distribution of FAAH and FAAH-2

Analysis of the phylogenetic distribution of FAAH and FAAH2 indicates that the gene duplication that gave rise to these related proteins probably predates the origins of the first animals with nervous systems. However, in addition to the loss of FAAH2 in rodents (see above), there are other examples of lineage-specific loss of FAAH or FAAH2. For example, only a FAAH2 orthologue is found in *Drosophila* and other insects.

(f) The phylogenetic distribution of CRIP1a and CRIP1b

Analysis of the phylogenetic distribution of CRIP1a and CRIP1b in mammals reveals that, while CRIP1a is found throughout the mammals, CRIP1b may be unique to catarrhine primates. For example, orthologues of human CRIP1b can be found in the chimpanzee *Pan troglodytes*, the gibbon *Nomascus leucogenys* and the rhesus monkey *Macaca mulatta*. Thus, it appears that exon 3b of the human CRIP1 gene, which is unique to CRIP1b, may have originated relatively recently in mammalian evolution. The functional significance of this is unknown and it will be interesting to investigate the roles of CRIP1b in brain function.

Unlike the restricted phylogenetic distribution of CRIP1b, CRIP1a has a much wider phylogenetic distribution that extends throughout much of the animal kingdom. Indeed, orthologues of CRIP1a can be found in basal invertebrates such as the cnidarian *N. vectensis*, indicating that CRIP1a is very ancient protein with origins dating back to the first animals with nervous systems. Accordingly, orthologues of human CRIP1a are found throughout the vertebrates and in deuterostomian invertebrates (e.g. in the cephalochordate *B. floridae* and in the hemichordate *Saccoglossus kowalevskii*) and protostomian invertebrates (e.g. in the insect *Bombus impatiens* and in the nematode *C. elegans*). This contrasts with the much more restricted phylogenetic distribution of CB₁/CB₂-type cannabinoid receptors, which, as highlighted above, are only found in vertebrates and invertebrate chordates. What this suggests is that CRIP1a is evolutionarily much more ancient than the CB₁ receptor protein that it is thought to interact with. We can infer from this that CRIP1a must have other physiological roles in cells, in addition to its proposed interaction with CB₁ receptors.

4. COMPARATIVE NEUROBIOLOGY OF ENDOCANNABINOID SIGNALLING

(a) Neurobiology of CB₁/CB₂-type endocannabinoid signalling in non-mammalian vertebrates

Given that a great deal is now known about the role of endocannabinoid-CB₁ signalling in mediating

retrograde signalling at synapses in the mammalian brain, it is pertinent to pose the question: is there evidence that endocannabinoid-CB₁-mediated retrograde signalling operates at synapses in the central nervous systems of non-mammalian vertebrates? Addressing this question may shed light on the evolutionary origin of this particular mechanism of synaptic plasticity. Not surprisingly, direct evidence from electrophysiological studies comparable to those carried out on rodent brain slices is sparse. The strongest evidence can be found in an impressive series of studies investigating the roles of endocannabinoid signalling in the spinal neuronal network that controls swimming in the lamprey *Lampetra fluviatilis*. Collectively, the data obtained indicate that 2-AG is synthesized postsynaptically by neurons in the spinal locomotor network and acts presynaptically to inhibit both excitatory and inhibitory neurotransmission via CB₁-mediated mechanisms. Furthermore, nitric oxide and endocannabinoid signalling interact to regulate the frequency/amplitude of the locomotor rhythm by differentially modulating excitatory and inhibitory inputs to motoneurons [81–84]. Thus, it appears that 2-AG/CB₁-mediated regulation of excitatory and inhibitory neurotransmission is a highly conserved mechanism throughout the vertebrates. Consistent with this hypothesis, recent electrophysiological studies have demonstrated that endocannabinoid-CB₁ signalling mediates DSE and metabotropic glutamate receptor-induced LTD in area X of the zebra finch brain [85]. Furthermore, immunocytochemical analysis of CB₁ expression in the nervous systems of non-mammalian vertebrates reveals patterns of expression consistent with axonal targeting and presynaptic sites of action [86–89].

Given the key role that DAGL α has in postsynaptic formation of 2-AG as a mediator of retrograde synaptic signalling in the mammalian central nervous system (CNS), it would be interesting to determine whether DAGL α is located in the somatodendritic compartment of neurons postsynaptic to CB₁-expressing axons in non-mammalian vertebrates. However, while the existence of DAGL α in non-mammalian vertebrates is confirmed by comparative analysis of genome sequence data (see above), detailed neuroanatomical analyses of DAGL α expression in the CNS of non-mammalian vertebrates have not yet been conducted.

It is perhaps not surprising that the physiological roles of 2-AG/CB₁-mediated endocannabinoid signalling at the sub-cellular/cellular level are conserved throughout the vertebrates. Are, however, the roles of endocannabinoid signalling also conserved at the system level, for example with respect to the regions of the CNS where the CB₁ receptor is expressed and the physiological/behavioural processes that the endocannabinoid signalling system regulates? To address this question, we must look to a currently rather limited number of neuroanatomical and behavioural studies of the cannabinoid system in non-mammalian vertebrates.

Developmental analysis of the zebrafish *D. rerio* reveals CB₁ mRNA expression in cells located in the presumptive preoptic area of the diencephalon at 24 h post-fertilization, and by 48 h, expression is observed in the telencephalon, the hypothalamus, the tectum and the hindbrain (ventral to cerebellum).

In adult zebrafish, CB₁ mRNA expression is observed in the anterior region of the telencephalon and in the periventricular medial zone and central zone of the dorsal telencephalon. Expression is also evident in the hypothalamus and posterior tuberculum (diencephalon) and in the torus longitudinalis (mesencephalon) [90]. Complementing the use of *in situ* hybridization techniques by Lam *et al.* for analysis of CB₁ mRNA expression in *D. rerio*, Cottone *et al.* have used immunocytochemical techniques to investigate the distribution of the CB₁ protein in the cichlid *Pelvicachromis pulcher* [86,91]. Immunostained neurons and/or fibres were observed in several brain regions, including the telencephalon, the prefrontal preoptic nucleus, the lateral infundibular lobes of the hypothalamus, the pretectal central nucleus and the posterior tuberculum.

In amphibians, the distribution of CB₁ mRNA in the brain of the rough-skinned newt *Taricha granulosa* has been examined using mRNA *in situ* hybridization methods, revealing a widespread pattern of expression with CB₁ mRNA detected in the telencephalon (olfactory bulb, the pallium and amygdala), the diencephalon (preoptic area and thalamus), the mesencephalon (tegmentum and tectum) and the hindbrain (cerebellum and stratum griseum) [92]. Complementing the use of *in situ* hybridization techniques by Hollis *et al.*, for analysis of CB₁ mRNA expression in *Taricha*, Cesa *et al.* have used immunocytochemical techniques to investigate the distribution of CB₁ in the brain of *Xenopus laevis*, revealing CB₁-immunoreactive cells and/or fibres in the olfactory bulbs, dorsal and medial pallium, striatum, amygdala, thalamus, hypothalamus, mesencephalic tegmentum and cerebellum [87]. CB₁-immunoreactivity is also present in the dorsal and central fields of the *Xenopus* spinal cord, regions that correspond to laminae I–IV and X of the mammalian spinal cord [88].

In birds, CB₁ expression has been analysed in the brain of the chick *Gallus gallus* [93], the zebra finch *Taeniopygia guttata* [89] and the budgerigar *Melopsittacus undulatus* [94], revealing some patterns of expression that are strikingly similar to findings in mammals [56,95]. For example, high levels of CB₁ expression are observed in the hippocampus and amygdala and, as in mammals, in the cerebellar cortex, the CB₁ gene is expressed in granule cells and the receptor protein is targeted to parallel fibres in the molecular layer.

Detailed descriptions of the distribution of CB₁ receptor expression in the CNS provide valuable frameworks for further investigation of the roles of the endocannabinoid signalling system in non-mammalian vertebrates. However, the number of species analysed thus far are too few to enable any meaningful general conclusions on how the neuroarchitecture of the cannabinoid signalling system has been shaped by lineage-specific changes in brain organization over evolutionary time scales. Nevertheless, the expression of CB₁ in so many different brain regions suggests that endocannabinoid signalling has been a fundamental and widely employed mechanism of synaptic plasticity throughout more than 400 million years of vertebrate brain evolution. Moreover, there is evidence that at least some of the physiological/behavioural roles of endocannabinoid signalling that

have been discovered in mammals are also applicable to non-mammalian vertebrates, suggesting evolutionarily ancient origins.

Some of the most striking actions of CB₁ cannabinoid receptor agonists in mammals are dose-dependent modulatory effects on locomotor activity [96]. These behavioural effects are consistent with abundant expression of the CB₁ receptor in brain regions involved in initiation (basal ganglia) and coordination (cerebellum) of movement [1]. Furthermore, consistent with the notion that CB₁ has an evolutionarily ancient role in neural pathways that control movement, Valenti *et al.* [97] have reported that the CB₁ receptor antagonist AM 251 (1 µg g⁻¹ body mass) causes a reduction in locomotor activity in the goldfish *Carassius auratus*. Behavioural effects of drugs that bind to the CB₁ receptor have also been investigated in an amphibian species, the rough-skinned newt *T. granulosa*, revealing an inhibitory effect on spontaneous locomotor activity and courtship clasping behaviour [98]. Likewise, the cannabinoid WIN 55,212-2 causes inhibition of locomotor activity in the zebra finch [99].

The brain endocannabinoid system is also involved in regulation of appetite and feeding in mammals [100], and again there is evidence that this role may be evolutionarily ancient. A study by Valenti *et al.* on the goldfish *C. auratus* found that food deprivation was accompanied by a significant increase in anandamide (but not 2-AG) in the telencephalic region of the brain, and intraperitoneal injection of anandamide (1 pg g⁻¹ body mass) caused an increase in food intake within 2 h of administration [97]. Soderstrom *et al.* [101] report that in the zebra finch, a reduction in food availability causes elevation of 2-AG in the caudal telecephalon and a CB₁-mediated reduction in song-stimulated brain expression of the transcription factor ZENK and a CB₁-mediated reduction in singing. Thus, the endocannabinoid system may have a fundamental role in linking behavioural activity with food availability.

The endocannabinoid signalling system is also involved in mechanisms of learning and memory, and studies on rodent models have, for example, provided evidence of roles in mechanisms of synaptic plasticity in brain regions critical for declarative memory (hippocampus) and in neural mechanisms underlying extinction of aversive memories [102]. In this aspect of endocannabinoid signalling, research on a non-mammalian vertebrate, the zebra finch *T. guttata*, has been particularly significant. The zebra finch is an attractive model system for research on neural mechanisms of learning because, in a manner analogous to human language acquisition, male zebra finches learn a song pattern during juvenile development [103]. Soderstrom and co-workers have found that cannabinoid exposure during sensorimotor stages of vocal development alters song patterns produced later during adulthood [104] with distinct sub-periods of sensitivity [105]. Consistent with these findings, the CB₁ receptor is expressed in brain regions involved in song learning [89] and song production [106], with cannabinoid exposure during sensorimotor stages of vocal development leading to alterations in CB₁ expression and 2-AG levels in

the adult brain [107]. Further investigation of mechanisms of action have revealed that cannabinoid exposure during sensorimotor stages of vocal development leads to increased basal expression of the transcription factor FoxP2 in the striatum of adult birds, including the area X song region [108] and increased dendritic spine densities [109].

Analysis of the effects of cannabinoids on adult zebra finches reveals an inhibitory effect on song production [99] and an associated inhibition of expression of the transcription factor ZENK in a brain region that is involved in auditory perception (the caudomedial neostriatum) [110]. Adult exposure to cannabinoids also causes dose-related inhibitory or stimulatory effects on neuronal activity (based on c-Fos expression) in brain regions that control vocal motor output [111].

Thus far, the zebra finch cannabinoid studies have focused primarily on the effects of exogenous cannabinoids (in particular WIN 55,212-2) on song learning and song production. This has provided insights on how developmental exposure to cannabinoids can lead to permanent alterations in brain function and behaviour, which may be highly relevant to an understanding of the risks associated with cannabis use in adolescents [112]. With the recent development of drugs that selectively inhibit degradation of endocannabinoids (e.g. the MAGL inhibitor JZL184 and the FAAH inhibitor PF-3845), it may now be possible to obtain more insights on the physiological roles of the endocannabinoid signalling system in learning using the zebra finch as a model system.

(b) Neurobiology of CB₁/CB₂-type endocannabinoid signalling in invertebrate chordates

As highlighted earlier, the discovery of genes encoding co-orthologues of CB₁ and CB₂ in the urochordate *C. intestinalis* (CiCBR) [76] and in the cephalochordate *B. floridae* (BfCBR) [75] revealed that the evolutionary origin of CB₁/CB₂-type cannabinoid receptors could be traced back beyond the vertebrates to the common ancestor of extant chordates. As of yet, the pharmacological properties of CiCBR and BfCBR have not been determined, and although these receptors are clearly CB₁/CB₂-type receptors based on sequence similarity, it should not be assumed that CiCBR and BfCBR are necessarily activated by the endocannabinoids 2-AG and anandamide *in vivo*. The GPCRs in mammals that are most closely related to CB₁ and CB₂ are activated by other lipid signalling molecules—the lysophospholipids [113]. Therefore, while we cannot assume that CiCBR and BfCBR are activated by the endocannabinoids 2-AG and anandamide, it seems reasonable to assume that these receptors are activated *in vivo* by endocannabinoid/lysophospholipid-like lipid signalling molecules. Thus, determining the identity of endogenous ligands for CiCBR and BfCBR is of great interest because it may shed light on how and when CB₁/CB₂-type receptors acquired their property of binding 2-AG and anandamide.

Although the pharmacological properties of CiCBR and BfCBR are unknown, some insights into the

physiological roles of CiCBR have been obtained by investigation of the distribution CiCBR expression in *C. intestinalis* using specific antibodies that bind to the C-terminal tail of the receptor. These immunocytochemical studies revealed that the approximately 46 kDa CiCBR protein is concentrated in the cerebral ganglion of *C. intestinalis*, which is located between the inhalant and exhalant siphons that confer on this species and on other sea squirts a filter-feeding lifestyle. Furthermore, CiCBR-immunoreactivity is localized in a dense meshwork of neuronal processes in the neuropile of the cerebral ganglion. CiCBR-immunoreactivity is also present in the axons and axon terminals of neurons that project via peripheral nerves over and around the internal surfaces of the inhalant and exhalant siphons [114], a pattern of expression consistent with behavioural effects of cannabinoids on siphon activity in *C. intestinalis* [115].

The axonal targeting of CiCBR in *C. intestinalis* is intriguing because of its similarity to CB₁ receptor localization in mammalian CB₁-expressing neurons. It suggests that CiCBR may have a similar role to CB₁ receptors by acting as an axonal regulator of neurotransmitter release. Furthermore, it implies that the role of CB₁ receptors as presynaptic regulators of neurotransmitter release may be very ancient, preceding the gene duplication that gave rise to CB₁ and CB₂ receptors and dating back at least as far as the common ancestor of vertebrates and urochordates. What is not yet known is the molecular identity of neurotransmitter(s) or neurohormone(s) that are released by CiCBR-expressing neurons in *C. intestinalis*. Is CiCBR expressed in GABAergic and/or glutamatergic neurons, as in mammals, or is CiCBR expressed in other types of neurons such as aminergic or peptidergic neurons? These are questions that need to be addressed if we are to gain an understanding of the physiological roles of CiCBR in *C. intestinalis*. It would also be interesting to determine whether BfCBR is expressed by neurons and targeted to axon terminals in *B. floridae*. If it is, then this would indicate that the axonal targeting of CB₁-type receptors that is seen in vertebrates can be traced back to the common ancestor of all extant chordates.

It is important to note that because CiCBR and BfCBR are co-orthologues of CB₁-type and CB₂-type cannabinoid receptors, then these receptors in invertebrate chordates may have both CB₁-like and CB₂-like functional properties. It is of interest, therefore, that CiCBR is not only expressed in neurons but is also present in haemocytes in *C. intestinalis* [114], which may be indicative of an ancient CB₂-like role in regulation of immunological processes. Thus, we can imagine a scenario where in the invertebrate chordate ancestor of vertebrates a CiCBR/BfCBR-like protein may have had both CB₁-type and CB₂-type functions and, following duplication of the gene encoding a CiCBR/BfCBR-like protein, the duplicated receptors diverged and acquired their more specific CB₁-type and CB₂-type functions. Clearly, this is speculative but it provides a rationale for further investigation of the physiological roles of CiCBR and BfCBR and the physiological roles of CB₁-type and CB₂-type cannabinoid receptors in non-mammalian vertebrates.

(c) *Neurobiology of non-CB₁/CB₂-mediated endocannabinoid signalling in invertebrates*

While CB₁/CB₂-type receptors do not occur in the majority of invertebrates, as highlighted earlier, the biochemical pathways for biosynthesis/inactivation of 2-AG and anandamide occur throughout the animal kingdom. Therefore, it is of interest to review evidence of non-CB₁/CB₂-mediated endocannabinoid signalling in the nervous systems of invertebrates.

(i) *Non-chordate deuterostomes—echinoderms and hemichordates*

Effects of cannabinoids and endocannabinoids on fertilization in the sea urchin *S. purpuratus* [116] and the occurrence of an endocannabinoid-like signalling system in embryonic and larval sea urchins [117] have been reported. Furthermore, opportunities to investigate the existence and functions of endocannabinoid-like signalling systems in echinoderms and hemichordates have been facilitated recently by sequencing of the transcriptomes/genomes of the sea urchin *S. purpuratus* and the hemichordate *S. kowalevskii* [118–120].

(ii) *Lophotrochozoan protostomian invertebrates—annelids*

Investigation of a putative endocannabinoid-like signalling system in annelids has largely focused on the medicinal leech *Hirudo medicinalis*, which is a well-established model system in neurobiology. Stefano *et al.* [121] reported the sequence of a putative leech cDNA encoding a partial (153 amino acids) protein sequence sharing significant similarity with mammalian CB₁ cannabinoid receptors. However, subsequent analysis of the sequence revealed that it was chimaeric, with a central region sharing 98 per cent identity with the bovine adrenocorticotrophic hormone receptor, and outer regions sharing 65–68% identity with mammalian CB₁ receptors [122]. Horizontal transfer of bovine DNA to leeches that feed on bovine blood was offered as a possible explanation for this unusual sequence [122] but perhaps a more likely explanation is that the sequence is an artefact [1]. More recently, the genome of the leech *Helobdella robusta* has been sequenced (<http://genome.jgi-psf.org/Helro1>) and analysis of the genomic sequence data does not reveal the presence of any CB₁-like genes, consistent with analysis of genomic sequence data from other protostomian invertebrates. However, there is evidence that an endocannabinoid-like system may exist in leeches and other annelids.

Detection of binding sites for ³H-anandamide in cell membranes derived from the CNS of *H. medicinalis* suggested the presence of putative receptors for this molecule [121], while binding sites for the cannabinoid ³H-CP55,940 have been detected in the nervous system of another annelid species, the earthworm *Lumbricus terrestris* [78]. Moreover, the detection of both anandamide and 2-AG and associated enzymatic activities in extracts of leech ganglia indicates that the biosynthetic machinery for the synthesis of these molecules exists in annelids [123].

Building upon these biochemical studies are a recent series of papers by Burrell and colleagues that have provided evidence that an endocannabinoid

signalling system modulates synaptic transmission in the leech *H. medicinalis*. Li and Burrell found that in the polysynaptic pathway from touch-sensitive mechanosensory neurons (T) to S interneurons in *Hirudo*, LTD of synaptic transmission is observed following low-frequency electrical stimulation (1 Hz) for 450 or 900 s. LTD elicited by 450 s low-frequency stimulation was blocked by *N*-methyl-D-aspartate (NMDA) receptor antagonists but LTD elicited by 900 s low-frequency stimulation was unaffected by NMDA receptor antagonists. Interestingly, LTD elicited by 900 s low-frequency stimulation was blocked by the cannabinoid receptor antagonist AM251 and by the DAGL inhibitor RHC80267, suggesting the involvement of an endocannabinoid-like signalling mechanism in this particular form of synaptic plasticity. Importantly, application of 2-AG or the cannabinoid receptor agonist CP-55,940 induced LTD of the T-S synaptic pathway, providing further evidence of an endocannabinoid-like mechanism of synaptic plasticity in the leech [124].

Further characterization of this system has revealed that LTD elicited by 900 s low-frequency stimulation requires activation of metabotropic serotonin receptors and is dependent on Ca^{2+} elevation in the S interneuron, mediated by voltage-gated Ca^{2+} channels and intracellular inositol triphosphate receptors. Furthermore, this particular form of LTD also involves stimulation of nitric oxide synthase and a decrease in cAMP signalling [125]. However, because synaptic plasticity is being examined here in the context of a polysynaptic pathway, mechanistic interpretation of these findings is complicated. Nevertheless, given that CB_1/CB_2 -type cannabinoid receptors do not exist in annelids and other protostomian invertebrates, these findings raise intriguing questions concerning the molecular nature of the putative receptors that mediate effects of endogenous or exogenous 2-AG (and other related lipids) in the nervous system of the leech.

Research on mammalian models has provided evidence that TRPV-type receptors are activated by endocannabinoids *in vitro* and mediate *in vivo* effects of endocannabinoids [71,126]. Thus, Burrell and co-workers have investigated TRPV-type receptors as potential mediators of endocannabinoid-dependent LTD in the leech nervous system. In the leech, there are three types of cutaneous mechanosensory neurons: low threshold touch (T), moderate threshold pressure (P) and high threshold nociceptive (N) neurons, all of which synapse onto the longitudinal motor neuron (L cell), which controls contraction during whole-body shortening. Low-frequency stimulation of the T neurons induces heterosynaptic LTD of glutamatergic transmission at the N-to-L synapse and, importantly, Yuan and Burrell found that this was blocked by DAGL inhibitors and the TRPV antagonists capsazepine and SB 366791. Furthermore, application of 2-AG and the TRPV agonist capsaicin mimicked LTD at the N-to-L synapses and these effects of 2-AG and capsaicin were blocked by capsazepine. Pre-treatment with 2-AG or capsaicin occluded subsequent expression of LTD induced by low-frequency stimulation. Finally, presynaptic, but not postsynaptic, intracellular injection of capsazepine

blocked both low-frequency stimulation-induced and 2-AG-induced LTD, indicating that presynaptic TRPV-type receptors mediate LTD at the N-to-L synapse. Collectively, these findings indicate that low-frequency stimulation of T neurons stimulates postsynaptic synthesis of 2-AG or a 2-AG-like molecule in L neurons, which then acts in a retrograde manner to inhibit heterosynaptic neurotransmitter release by N neurons via a TRPV-type-receptor-mediated mechanism [127].

Evidence that presynaptic TRPV-type-receptor-mediated LTD may be a widespread mechanism of synaptic plasticity in the leech nervous system has been obtained in a subsequent study, using the leech T-S synaptic pathway as a model preparation [128]. LTD is induced when a spike train is triggered in the S cell 1–10 s prior to stimulation of the T cell and this is blocked by perfusion of the preparation with the cannabinoid receptor antagonist AM251 or the DAGL inhibitor RHC80267 and by injection of the DAGL inhibitor tetrahydropipstatin into the S cell. Perfusion with the TRPV antagonist capsazepine also blocked LTD induced by a spike train in the S cell 1–10 s prior to stimulation of the T cell. This effect of capsazepine was observed when it was injected into the T cell but not when it was injected into the S cell. Thus, it appears that mechanisms of LTD involving postsynaptic synthesis of 2-AG or 2-AG-like molecules by DAGL and presynaptic activation of TRPV-type receptors occur widely in the leech nervous system. These findings raise interest in determination of the molecular identity of the putative TRPV-type receptors that mediate LTD in the leech nervous system. This would enable investigation of the cellular distribution of these receptors in the leech nervous system and comparison of their molecular properties with mammalian TRPV receptors. Likewise, it would be interesting to investigate the expression of DAGLs in the leech nervous system at a cellular and sub-cellular level to assess DAGLs as potential sources of 2-AG or 2-AG-like molecules that mediate LTD via retrograde synaptic signalling mechanisms.

The discovery of LTD mediated by 2-AG or 2-AG-like molecules and TRPV-type receptors in the leech nervous system suggest that endocannabinoid-mediated retrograde synaptic signalling is an evolutionarily ancient mechanism that predates the origins of CB_1/CB_2 -type cannabinoid receptors in chordates. If this is correct, these mechanisms of synaptic plasticity may also operate in the nervous systems of other invertebrates (see below). Thus, the findings of Burrell and co-workers have paved the way for further investigation of the function of endocannabinoid-type signalling mechanisms in the nervous systems of all animals, extending the scope for research on the comparative neurobiology of endocannabinoid signalling well beyond the phylum Chordata.

(iii) *Lophotrochozoan protostomian invertebrates—molluscs*

There has been relatively little investigation of endocannabinoid-like signalling systems in molluscs. This is perhaps surprising, given the importance of

molluscs as model systems in neurobiology—in particular, the gastropod species *Aplysia californica* and *Lymnaea stagnalis* [129]. The discovery that an endocannabinoid-type signalling system mediates synaptic plasticity in the leech *H. medicinalis*, as highlighted above, may act as a stimulus for researchers to investigate whether similar mechanisms operate in molluscan species.

Importantly, biochemical studies on bivalve molluscan species have revealed the presence of NAEs, including anandamide, putative binding sites for anandamide and a FAAH-like enzymatic activity [130,131]. Furthermore, transcriptomic/genomic sequence data are available for molluscan species, including the gastropod *A. californica* [132] and the bivalve *Crassostrea gigas* [133]. Therefore, identification of genes encoding proteins implicated in endocannabinoid signalling (e.g. DAGLs, MAGL, NAPE-PLD, FAAH) is now feasible for molluscan species, which will facilitate detailed investigation of endocannabinoid-like signalling systems in molluscan species.

(iv) *Ecdysozoan protostomian invertebrates—nematodes*

The nematode *C. elegans* was the first animal species to have its genome sequenced, and analysis of this sequence provided the first evidence that CB₁/CB₂-type cannabinoid receptors do not occur throughout the animal kingdom [1]. A gene encoding a GPCR (C02H7.2) that shares sequence similarity with CB₁/CB₂-type cannabinoid receptors is present in *C. elegans*, but analysis of its sequence indicates that it is not an orthologue [1,77]. Nevertheless, binding sites for the cannabinoid ³H-CP-55,940 have been detected in the nematode *Panagrellus redivivus*, suggesting the presence of other non-CB₁/CB₂-type cannabinoid receptors in nematodes [78].

The presence of the endocannabinoids anandamide and 2-AG has been specifically investigated in nematodes by analysis of three species, *C. elegans*, *Caenorhabditis briggsae* and *Pelodera strongyloides*, and both anandamide and 2-AG were detected in all three species. However, anandamide and 2-AG were not detected in a mutant strain of *C. elegans* (fat-3) that lacks functional activity of the delta-6 desaturase enzyme required for synthesis of long-chain polyunsaturated fatty acids (PUFAs; including arachidonic acid) [134].

Importantly, the physiological roles of anandamide and other NAEs in *C. elegans* have recently been investigated, exploiting the use of techniques to manipulate the expression of genes encoding enzymes involved in NAE metabolism. Suppression of FAAH using RNA interference (RNAi) or FAAH inhibitors (URB597) caused an increase in the levels of anandamide and other NAEs and overexpression of the *faah-1* gene caused a decrease in levels of anandamide and other NAEs, demonstrating the importance of FAAH as a regulator of NAEs in an invertebrate species [135]. Furthermore, *faah-1* overexpression caused a developmental delay that was rescued by *faah-1* RNAi, indicating a role for NAEs in promotion of larval development in *C. elegans*. Peak levels of NAEs are detected during the second larval stage (L2) at which time animals are committed to reproductive growth,

but NAE levels are reduced at L2 in animals committed to an alternative diapause stage (dauer) induced by dietary restriction. This suggested that NAEs may act as signals of an altered metabolic state and, consistent with this notion, exogenous application of the NAE eicosapentaenoyl ethanolamide (EPEA)—and, to a lesser extent, anandamide—was found to rescue dauer formation. Worms overexpressing *faah-1* also exhibited resistance to thermal stress and increased adult lifespan, whereas application of EPEA caused a reduction in thermotolerance and lifespan, most strikingly in animals under dietary restriction. Collectively, the data reported by Lucanic *et al.* indicate that the effect of dietary restriction on lifespan extension is mediated, at least in part, by a reduction in NAE signalling. However, the mechanisms by which NAEs exert effects on lifespan in *C. elegans* are as yet unknown, and therefore identification of NAE receptors in nematodes now represents a fascinating objective for the future.

(v) *Ecdysozoan protostomian invertebrates—arthropods*

Sequencing of the genome of an arthropod species, the insect *Drosophila melanogaster*, provided key evidence that CB₁/CB₂-type cannabinoid receptors do not occur throughout the animal kingdom [1], and this has been supported by sequencing of other arthropod genomes. Nevertheless, binding studies using radio-labelled cannabinoids have revealed the presence of putative binding sites in a variety of arthropod species [78], although identification of membrane proteins that bind cannabinoids has yet to be accomplished for any arthropod species. Evidence that TRPV-type ion channels mediate effects of endocannabinoids in the nervous system of the leech *H. medicinalis* [127] points to these proteins as potential endocannabinoid receptors in arthropods. It is noteworthy that the prototype for the TRP ion channel family was first discovered in *Drosophila* as a result of molecular analysis of the *transient receptor potential (trp)* mutant that has a defective phototransduction mechanism [136]. Interestingly, analysis of phototransduction in *Drosophila* has revealed similarities with mechanisms of 2-AG biosynthesis. Thus, the photon-activated rhodopsin protein is coupled via G-proteins to stimulate activation of PLC, and the DAGs generated by PLC are substrates for a DAGL that is a homologue of the mammalian enzymes DAGL α and DAGL β . The *Drosophila* DAGL mutant (*maE*) is defective in photoreceptor responses to light, and a DAG metabolite is thought to mediate phototransduction by activating TRP and/or TRP-like (TRPL) ion channels [137]. However, the identity of the DAG metabolite that binds to and activates the TRP/TRPL phototransduction channels is not known, although there is evidence that it may be a PUFA [136]. This suggests the existence and activity of a MAGL in the phototransduction pathway, but one has yet to be identified. As highlighted earlier, *Drosophila* and other insects do not have an orthologue of the MAGL enzyme that has such a key role in 2-AG metabolism in mammals. Therefore, other as yet uncharacterized enzymes

may generate endogenous TRP/TRPL ligands from MAG substrates.

Interestingly, DAGL is not only expressed in *Drosophila* photoreceptors but is also expressed more widely in the brain [137]. Therefore, investigation of the physiological roles of DAGL and its metabolites in the nervous system of *Drosophila* might be fruitful in revealing the existence of an endocannabinoid-like signalling system that regulates synaptic activity in insects and in other arthropods.

(vi) Cnidarians

Cnidarians are considered to be the most primitive animals with nervous systems and evidence for the existence of an endocannabinoid-like system in the hydrozoan *Hydra viridis* has been reported [138]. Sequencing the genomes of *Hydra magnipapillata* and the anthozoan *N. vectensis* [139,140] has presented new opportunities for molecular analysis of putative endocannabinoid signalling mechanisms in the simple nervous systems of these animals.

5. CONCLUDING COMMENTS

The value and importance of non-mammalian animals for cannabinoid research has been illustrated here with reference to studies on 'model organisms' ranging from leeches to lampreys and from zebrafish to zebra finches. However, we should not look upon cannabinoid-related studies on non-mammalian animals solely from a utilitarian perspective. Research on the evolution and comparative neurobiology of endocannabinoid signalling is intrinsically interesting and there is great potential to deepen our understanding of this aspect of nervous system function by revealing 'the art of the possible' in the diverse branches of the tree of life.

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