

NIH Public Access

Author Manuscript

Mol Pharm. Author manuscript; available in PMC 2013 June 04

Published in final edited form as:

Mol Pharm. 2012 June 4; 9(6): 1847–1851. doi:10.1021/mp300140h.

The Fraction Dose Absorbed, in Humans, and High Jejunal Human Permeability Relationship

Arik Dahan^{1,*}, Hans Lennernäs², and Gordon L. Amidon³

¹ Department of Clinical Pharmacology, School of Pharmacy, Faculty of Health Sciences, Ben-Gurion University of the Negev, Beer-Sheva 84105, Israel

² Department of Pharmacy, Biopharmaceutics Research Group, Uppsala University, S-75123 Uppsala, Sweden

³ Department of Pharmaceutical Sciences, College of Pharmacy, University of Michigan, Ann Arbor, MI 48109, USA

Abstract

The drug intestinal permeability ($P_{\rm eff}$) measure has been widely used as one of the main factors governing both the rate and/or extent of drug absorption (F_{abs}) in humans following oral administration. In this communication we emphasize the complexity behind and the care that must be taken with this in-vivo $P_{\rm eff}$ measurement. Intestinal permeability, considering the whole of the human intestine, is more complex than generally recognized, and this can lead to misjudgment regarding $F_{\rm abs}$ and $P_{\rm eff}$ in various settings, e.g. drug discovery, formulation design, drug development and regulation. Setting the adequate standard for the low/high permeability class boundary, the different experimental methods for the permeability measurement, and segmentaldependent permeability throughout the human intestine due to different mechanisms, are some of the main points that are discussed. Overall, the use of jejunal $P_{\rm eff}$ as a surrogate for extent of absorption is sound and scientifically justified; a compound with high jejunal $P_{\rm eff}$ will have high $F_{\rm abs}$, eliminating the risk for misclassification as a BCS Class I drug. Much more care should be taken, however, when jejunal Peff does not support a high-permeability classification; a thorough examination may reveal high-permeability after all, attributable to e.g. segmental-dependent permeability due to degree of ionization or transporter expression. In this situation, the use of multiple permeability experimental methods, including the use of metabolism, which except of luminal degradation requires absorption, is prudent and encouraged.

Keywords

intestinal permeability; fraction dose absorbed; biopharmaceutics classification system; oral absorption

The drug intestinal permeability measure has been widely used as one of the main factors governing both the rate and/or extent of drug absorption (F_{abs}) in humans following oral administration¹⁻³. Extensive scientific research has established that a good correlation exists between the human jejunal permeability (P_{eff}) measured using single-pass perfusion techniques and the fraction of dose absorbed from an immediate-release, rapidly dissolving, dosage form^{4, 5}. The best practice to determine F_{abs} is from pharmacokinetic/mass-balance studies in humans^{3, 4, 6-8}. The complexity and cost of carrying out these studies are very

^{*} Corresponding Author. Department of Clinical Pharmacology, School of Pharmacy, Faculty of Health Sciences, Ben-Gurion University of the Negev, P.O. Box 653, Beer-Sheva 84105, Israel. Tel: +972-8-6479483. Fax: +972-8-6479303. arikd@bgu.ac.il..

high, as they require radiolabelled API for the validation of high drug and metabolites recovery ^{9, 10}. Since the FDA uses F_{abs} as the basis for high-permeability BCS classification², an alternative channel of using P_{eff} as a surrogate for F_{abs} was suggested by the FDA BCS guidance². In this communication we emphasize the complexity behind and the care that must be taken with this in-vivo permeability measurement. Human intestinal permeability (HIP), considering the whole of the human intestine, is more complex than generally recognized, and this can lead to misjudgment regarding F_{abs} and P_{eff} in various settings, e.g. drug discovery, formulation design, drug development and regulation.

The fundamental basis of the biopharmaceutics classification system (BCS) is the application of Fick's first law to a membrane transport (regardless of mechanism), revealing that the drug flux (mass/area/time) through the intestinal wall is equal to the permeability times the concentration of the drug at the intestinal membrane surface. This is a boundary condition on the fundamental mass balance equation for drug absorption⁷. This is a local mass balance requirement, pertaining to each point along the intestinal membrane; the permeability, in general, must be considered as *position* and *time* dependent. This dependence may be due to drug concentration changes (e.g. in carrier-mediated transport), changes in luminal fluid content, physiological/biochemical variations, water absorption, cellular differentiation, up/down regulation of drug transporters, membrane structure/ composition variations along the intestine, etc. However, theoretically reliable prediction of extent of absorption can be made based on this mass-balance, resulting in the equation:

 $F_{abs} = (Dose^{-1}) \int_0^{t_{res}} \iint P_{eff} C_w dA dt$ Eq 1

as was developed in the first presentation of the BCS¹. Absorption, as defined in equation 1, is taken to be the transport of drug, via diffusion and/or carrier mediated transport ^{11, 12}, across the intestinal membrane and/or through the tight-junctions, and thus, 'in the body'.

However, jejunal permeability (alone) may not always adequately predict F_{abs} . As noted above, the effective permeability (P_{eff}) in Equation 1 is, in general, position and time dependent (Figure 1), the (surface) area integration is over the entire intestine, and the time integration is from zero (time of dosing) until t_{res} , the residence time of the (whole) intestine. Equation 1, based on Gauss' Theorem (Divergence theorem), relates drug lost from the intestinal lumen (luminal volume) to the drug transported through the intestinal wall. This, of course, excludes luminal and brush border degradation in the 'lumen' prior to absorption. The experimental studies in humans determine the jejunal $P_{\rm eff}$, a 10 cm segment of the intestine approximately 40 cm distal to the gastro-duodenal junction, and represent the average $P_{\rm eff}$ of that 10 cm of proximal jejunum. The $P_{\rm eff}$ of this segment, which is the best human measurement available today, may not represent the $P_{\rm eff}$ of the whole of the human intestine and thus may not always adequately predict F_{abs} as required by Equation 1. This view was explicitly discussed in the BCS presentation paper¹, and was later efficiently implemented in physiologically-based pharmacokinetic (PBPK) modeling software e.g. GastroPlus®, that divide the intestine into segments to account for regional-dependent permeability and metabolism^{13, 14}. While for nonionizable and transcellular passively absorbed compounds the regional differences in the permeability along the intestine may be small, for drugs that are ionizable and/or their absorption is carrier-mediated, complex position-dependent permeability can be expected, as will be discussed below¹⁵. The FDA definition of F_{abs} >90% for high-permeability classification, i.e. the low/high permeability class boundary for classification of drugs, was initially conservative based on the need to consider these more complex permeability concerns. Notably, the European Medicines Agency, EMA, that initially followed the $F_{abs}>90\%$ definition for high-permeability classification, has recently decreased the permeability class boundary to 85% fraction of dose absorbed¹⁶.

Dahan et al.

The β -blocker metoprolol has been widely used as a marker for the low/high permeability class boundary, that is, if a compound shows higher P_{eff} than metoprolol then it is considered to be high-permeability, and *vice versa*. The corresponding FDA's F_{abs} based high-permeability classification states that a drug substance is classified as high-permeability when the fraction of dose absorbed in humans is 90% or higher². Examination of the human intestinal absorption of metoprolol reveals that it is in fact completely (100%) absorbed¹⁷, as evidenced by radiolabel mass-balance studies in five human subjects, thus making metoprolol a conservative reference drug for the low/high permeability class boundary. Indeed, the angiotensin II blocker losartan has been shown to have a lower human P_{eff} than that of metoprolol yet a high fraction dose absorbed based on mass-balance. Similarly, isotretinoin was reported to have 90% fraction dose absorbed yet a human P_{eff} lower than that of metoprolol (Table 1). The original choice of metoprolol as a reference drug was intentionally conservative in the initial guidance, to minimize the chances for exceptions to the dissolution standard that was going to be recommended for 'Biowaivers'.

The results of more recent studies show that not only does metoprolol represent an overly conservative boundary reference drug, it carries an additional complexity. We have shown that metoprolol's intestinal permeability in the rat is segmental-dependent; as a basic drug $(pK_a=9.5)$ with Log P value ~2, the pH changes along the small intestine were significant enough to lead to 5-fold higher permeability in the ileum (pH 7.5) than in the jejunum (pH 6.5). This raises the question of choice of $P_{\rm eff}$ reference value even for metoprolol. An IR oral dose of metoprolol has been shown to be completely absorbed in the upper small intestine in both humans and animal models; Jobin et al have used the intubation technique in humans to show that 60% of a metoprolol oral dose is absorbed from the duodenum, and an additional 20% is absorbed from the following 30 cm of the jejunum, leading to absorption of 80% of the dose from the upper 50 cm of the small intestine¹⁸. Masaoka et al have shown that in rats, 90% of an oral metoprolol dose is absorbed prior to the jejunum, and the remaining 10% is absorbed in the upper jejunum¹⁹. It follows, hence, that metoprolol's P_{eff} value at pH 7.5 (ileum) is not likely to be physiologically relevant, in terms of mass of drug absorbed, for an IR dosage form; rather, the permeability at pH 6.5, the average pH of the human jejunum (and the pH used in human jejunal permeability studies), would govern metoprolol's *in-vivo* intestinal absorption from an IR dosage form, and this permeability value allows the complete absorption of metoprolol. This analysis points out a possible extension to the regulatory high-permeability criterion: taking metoprolol's jejunal $P_{\rm eff}$ at pH 6.5 as the benchmark for high-permeability, it is suggested that if a compound matches/exceeds this threshold anywhere in the intestine, and not necessarily in the jejunum, it is a high-permeability compound with complete absorption (F_{abs} >90%). This illustrates the issue of segmental-dependent permeability, in this case due to luminal degree of ionization and its effect on the (complex) intestinal membrane permeability, a very relevant consideration as will be discussed below.

We have recently addressed the distinctive absorption characteristics of the β -blocker sotalol, for which low apparent permeability across Caco-2 cell monolayers has been reported, but also a high fraction dose absorbed ($F_{abs}>90\%$) in humans has been reported ^{20, 21}. We have shown that, similarly to metoprolol, sotalol's intestinal permeability in the rat was segmental-dependent, with higher P_{eff} at distal small intestinal regions (with higher average pH) than in proximal segments. At any given small intestinal segment and pH, sotalol's permeability was lower than that of metoprolol; however, most significanly, sotalol's permeability in the ileum at pH 7.5 exceeded that of metoprolol in the jejunum at pH 6.5 and matched metoprolol's P_{eff} in the middle small intestinal length, an oral dose of sotalol would have an apparent high-permeability (i.e. greater than that of metoprolol in the jejunum) throughout a significant portion of the small intestinal residence time (~2 hours),

resulting in its high fraction dose absorbed²². Significantly, the human t_{max} values of sotalol and metoprolol from an IR product support this analysis; metoprolol reaches C_{max} within ~1 hr, indicating rapid absorption from the proximal small intestine (following gastric emptying) consistent with the independent studies noted above, while sotalol's C_{max} is reached only after ~4 hr, strongly suggesting higher rate and extent of absorption from the distal parts of the small intestine. This analysis illustrates the limitations, as well as the care that must be used, in making a case for high/low permeability classification based merely on jejunal $P_{\rm eff}$ values and comparison to metoprolol; sotalol would be falsely classified as lowpermeability based on jejunal P_{eff} value determined at pH 6.5, but when more thoroughly evaluated, the permeability basis for sotalol's nearly complete oral absorption is apparent, and the correct high-permeability classification was made. This case also stresses the care that must be taken when relying on (solely) Caco-2 studies for permeability characterization, and the limitations of this experimental method in representing the intestinal membrane permeability process in all of its complexity. Yet, very notably, published results thus far indicate that there are no known false-positives in Caco-2 permeability studies, indicating the significance/strength of this experimental method 20, 23, 24. That is, if a compound shows high-permeability across Caco-2 cell monolayer in a well validated and conducted study, it will have high F_{abs} .

Equation 1 requires one to consider the permeability, in all of its complexity, and in order to be valid requires one to consider the whole of the intestine, under the luminal conditions of dosing and is valid based on mass balance if one uses or determines the correct (complex) permeability. Further, metoprolol is absorbed, based on mass-balance studies, to 100%, while a high-permeability drug is defined at $F_{abs}>90\%$. Thus the permeability value for high-permeability classification should be adjusted down to reflect this difference. It was suggested that the β -blocker labetalol, which exhibits lower P_{eff} than metoprolol yet 95% F_{abs} , could serve as a more reflective marker for the low/high permeability class boundary.

When carrier-mediated intestinal permeability, both influx and efflux, contribute to the drug absorption process, the expression levels of the relevant transporters along the intestine may lead to a segmental dependent permeability phenomenon. For instance, we have shown that the expression level of the efflux transporter P-glycoprotein (P-gp) along the rat small intestine is regional-dependent and follows a gradient, increasing from the proximal to the distal segments^{25, 26}. The same trend, although with higher variability, has been shown in humans as well^{27, 28}. We have also shown that the permeability of the H_2 blockers and P-gp substrates cimetidine and famotidine varies along the intestine, and is inversely correlated with the P-gp expression levels in the different segments^{25, 29}. The same segmentaldependent permeability, at the same magnitude (approximately 2-fold) was reported for cimetidine intestinal absorption in humans³⁰, showing the correlation between human and rat segmental-dependent permeability, also when carrier-mediated transport is involved. In the case of (solely) P-gp substrates, the jejunal $P_{\rm eff}$ may still adequately correlate with $F_{\rm abs}$ in spite of the potential segmental-dependent permeability and the decreased $P_{\rm eff}$ downstream, because high-permeability in the jejunum will likely allow the compound a high extent of absorption from an IR drug product regardless of the subsequent permeability in the more distal intestinal segments, similarly to the analysis of metoprolol's absorption. On the other hand, we have shown that the expression of the efflux transporter MRP2 throughout the rat small intestine follows an opposite gradient, decreasing from the proximal to the distal segments²⁶. In this case, a (solely) MRP2 substrate may show low jejunal P_{eff} , but sufficiently high ileal permeability to enable high F_{abs} . This may lead to misclassification when judging merely by the jejunal permeability. Additional complexity may rise when multiple transporters are involved in the intestinal absorption of the drug^{26, 31, 32}. It should be noted that this analysis is relevant only for low/borderline

permeability compounds; intestinal transporters will have no or little apparent effect on the intestinal absorption of drugs with significantly high permeability³²⁻³⁴.

The segmental-dependent intestinal permeability of drugs along the human intestine, and its relevance to the overall absorption, is (at present) largely unknown for most drugs. Regional infusions of the human intestine have been performed and support the segmental dependence of permeability, but due to the unknown- and time-dependent surface area, a permeability value cannot be calculated. Given the lack of direct experimental data in humans, especially human ileal permeability, the best we can do today is to evaluate segmental dependence in animal models that can provide a strong indication of the expected situation in humans³⁵⁻³⁹. Indeed, it was recognized in the FDA guidance on BCS that animal data, particularly rat small intestinal perfusion, can be correlated very well with human jejunal permeability². Further it was recognized that tissue culture methods, e.g. the commonly used Caco-2 cell monolayers, could reduce the need for human and animal experiments but carried much more uncertainty since the biochemical and cellular expression of proteins and other cellular components and developmental differences (compared to human intestine) are difficult to quantify or unknown. Thus, a stronger scientific case, in the absence of human data, would need to be made for a convincing highpermeability classification. The requirement for 20 reference or validation compounds was made for these (among other) reasons, so these could be used to validate a case for highpermeability determination ²⁴. Overall, human data, particularly human fraction absorbed would be the 'gold' standard in the determination of high-permeability; however permeability can be estimated, with varying degrees of confidence, much more easily than mass balance studies can be performed.

Notwithstanding the complexity behind the permeability value, human jejunal $P_{\rm eff}$ values of 42 compounds with various absorption mechanisms determined principally at Uppsala University and at the University of Michigan over a period of 18 years have been shown to have an excellent correlation with the fraction dose absorbed in humans^{4, 6, 8}. Overall, therefore, the use of jejunal $P_{\rm eff}$ as a surrogate for extent of absorption is sound and scientifically justified; a compound with high jejunal permeability (exceeding that of metoporolol's jejunal permeability) will certainly have a high fraction dose absorbed, eliminating the risk for misclassification as a BCS Class I drug. Much more care should be taken, however, when jejunal $P_{\rm eff}$ does not support a high-permeability classification; a thorough examination may reveal high-permeability after all, attributable to e.g. segmental-dependent permeability due to degree of ionization or transporter expression (whatever the underlying biochemical mechanism for this dependence). In this situation, the use of multiple permeability experimental methods, including the use of metabolism, which except of luminal degradation requires absorption, as suggested by the BDDCS^{40, 41}, is prudent and encouraged ^{3, 20, 23}.

References

- Amidon GL, Lennernäs H, Shah VP, Crison JR. A theoretical basis for a biopharmaceutic drug classification: the correlation of in vitro drug product dissolution and in vivo bioavailability. Pharmaceutical Research. 1995; 12(3):413–420. [PubMed: 7617530]
- CDER/FDA. Guidance for industry: Waiver of in vivo bioavailability and bioequivalence studies for immediate release dosage forms based on a biopharmaceutical slassification system. Center for Drug Evaluation and Research; 2000.
- Dahan A, Miller JM, Amidon GL. Prediction of solubility and permeability class membership: provisional BCS classification of the world's top oral drugs. AAPS Journal. 2009; 11(4):740–746. [PubMed: 19876745]

- Lennernäs H. Human jejunal effective permeability and its correlation with preclinical drug absorption models. Journal of Pharmacy and Pharmacology. 1997; 49(7):627–638. [PubMed: 9255703]
- Lennernäs H. Human intestinal permeability. Journal of Pharmaceutical Scinces. 1998; 87(4):403– 410.
- Lennernäs H. Modeling gastrointestinal drug absorption requires more in vivo biopharmaceutical data: experience from in vivo dissolution and permeability studies in humans. Current Drug Metabolism. 2007; 8(7):645–57. [PubMed: 17979653]
- Sun D, Lennernäs H, Welage LS, Barnett JL, Landowski CP, Foster D, Fleisher D, Lee K-D, Amidon GL. Comparison of human duodenum and caco-2 gene expression profiles for 12,000 gene sequences tags and correlation with permeability of 26 drugs. Pharmaceutical Research. 2002; 19(10):1400–1416. [PubMed: 12425456]
- Cook JA, Davit BM, Polli JE. Impact of biopharmaceutics classification system-based biowaivers. Molecular Pharmaceutics. 2010; 7(5):1539–1544.
- Roffey S, Obach R, Gedge J, Smith D. What is the objective of the mass balance study? A retrospective analysis of data in animal and human excretion studies employing radiolabeled drugs. Drug Metabolism Reviews. 2007; 39(1):17–43. [PubMed: 17364879]
- 11. Dobson PD, Kell DB. Carrier-mediated cellular uptake of pharmaceutical drugs: an exception or the rule? Nature Reviews Drug Discovery. 2008; 7(3):205–220.
- Sugano K, Kansy M, Artursson P, Avdeef A, Bendels S, Di L, Ecker GF, Faller B, Fischer H, Gerebtzoff G, Lennernäs H, Senner F. Coexistence of passive and carrier-mediated processes in drug transport. Nature Reviews Drug Discovery. 2010; 9(8):597–614.
- Yu LX, Amidon GL. A compartmental absorption and transit model for estimating oral drug absorption. International Journal of Pharmaceutics. 1999; 186(2):119. [PubMed: 10486429]
- Yu LX, Lipka E, Crison JR, Amidon GL. Transport approaches to the biopharmaceutical design of oral drug delivery systems: prediction of intestinal absorption. Advanced Drug Delivery Reviews. 1996; 19(3):359–376. [PubMed: 11540095]
- Tannergren C, Bergendal A, Lennernäs H, Abrahamsson B. Toward an increased understanding of the barriers to colonic drug absorption in humans: implications for early controlled release candidate assessment. Molecular Pharmaceutics. 2009; 6(1):60–73. [PubMed: 19183105]
- 16. EMA. Guideline on the investigation of bioequivalence. European Medicines Agency; 2010.
- Regardh C, Borg K, Johansson R, Johnsson G, Palmer L. Pharmacokinetic studies on the selective beta1-receptor antagonist metoprolol in man. Journal of Pharmacokinetics and Biopharmaceutics. 1974; 2(4):347–64. [PubMed: 4155762]
- Jobin G, Cortot A, Godbillon J, Duval M, Schoeller J, Hirtz J, Bernier J. Investigation of drug absorption from the gastrointestinal tract of man. I. Metoprolol in the stomach, duodenum and jejunum. British Journal of Clinical Pharmacology. 1985; 19(Suppl 2):97S–105S. [PubMed: 4005135]
- Masaoka Y, Tanaka Y, Kataoka M, Sakuma S, Yamashita S. Site of drug absorption after oral administration: Assessment of membrane permeability and luminal concentration of drugs in each segment of gastrointestinal tract. European Journal of Pharmaceutical Scinces. 2006; 29(3-4):240– 250.
- Chen M-L, Yu L. The use of drug metabolism for prediction of intestinal permeability. Molecular Pharmaceutics. 2009; 6(1):74–81. [PubMed: 19132929]
- Yang Y, Faustino PJ, Volpe DA, Ellison CD, Lyon RC, Yu LX. Biopharmaceutics classification of selected beta-blockers: solubility and permeability class membership. Molecular Pharmaceutics. 2007; 4(4):608–614. [PubMed: 17637014]
- Dahan A, Miller JM, Hilfinger JM, Yamashita S, Yu LX, Lennernäs H, Amidon GL. Highpermeability criterion for BCS classification: Segmental/pH dependent permeability considerations. Molecular Pharmaceutics. 20107(5):1827–1834.

Dahan et al.

- Amidon KS, Langguth P, Lennernäs H, Yu L, Amidon GL. Bioequivalence of oral products and the biopharmaceutics classification system: science, regulation, and public policy. Clinical Pharmacology and Therapeutics. 2011; 90(3):467–470. [PubMed: 21775984]
- Thiel-Demby VE, Humphreys JE, St. John Williams LA, Ellens HM, Shah N, Ayrton AD, Polli JW. Biopharmaceutics classification system: Validation and learnings of an in vitro permeability assay. Molecular Pharmaceutics. 2009; 6(1):11–18. [PubMed: 19248229]
- 25. Dahan A, Amidon GL. Segmental dependent transport of low permeability compounds along the small intestine due to P-glycoprotein: The role of efflux transport in the oral absorption of BCS class III drugs. Molecular Pharmaceutics. 2009; 6(1):19–28. [PubMed: 19248230]
- 26. Dahan A, Sabit H, Amidon GL. Multiple efflux pumps are involved in the transpithelial transport of colchicine: Combined effect of P-gp and MRP2 leads to decreased intestinal absorption throughout the entire small intestine. Drug Metabolism and Disposition. 2009; 37(10):2028–2036. [PubMed: 19589874]
- Mouly S, Paine M. P-glycoprotein increases from proximal to distal regions of human small intestine. Pharmaceutical Research. 2003; 20(10):1595–9. [PubMed: 14620513]
- Thorn M, Finnstrom N, Lundgren S, Rane A, Loof L. Cytochromes P450 and MDR1 mRNA expression along the human gastrointestinal tract. British Journal of Clinical Pharmacology. 2005; 60(1):54–60. [PubMed: 15963094]
- 29. Dahan A, West BT, Amidon GL. Segmental-dependent membrane permeability along the intestine following oral drug administration: Evaluation of a triple single-pass intestinal perfusion (TSPIP) approach in the rat. European Journal of Pharmaceutical Scinces. 2009; 36(2–3):320–329.
- 30. Sutcliffe F, Riley S, Kaser-Liard B, Turnberg L, Rowland M. Absorption of drugs from human jejunum and ileum. British Journal of Clinical Pharmacology. 1988; 26:206P–207P.
- Dahan A, Amidon GL. Small intestinal efflux mediated by MRP2 and BCRP shifts sulfasalazine intestinal permeability from high to low, enabling its colonic targeting. American Journal of Physiology - Gastrointestinal & Liver Physiology. 2009; 297(2):G371–G377. [PubMed: 19541926]
- 32. Giacomini K, Huang S, Tweedie D, Benet L, Brouwer K, Chu X, Dahlin A, Evers R, Fischer V, Hillgren K, Hoffmaster K, Ishikawa T, Keppler D, Kim R, Lee C, Niemi M, Polli J, Sugiyama Y, Swaan P, Ware J, Wright S, Yee S, Zamek-Gliszczynski M, Zhang L. Membrane transporters in drug development. Nature Reviews Drug Discovery. 2010; 9(3):215–236.
- 33. Cao X, Yu LX, Barbaciru C, Landowski CP, Shin H-C, Gibbs S, Miller HA, Amidon GL, Sun D. Permeability dominates in vivo intestinal absorption of P-gp substrate with high solubility and high permeability. Molecular Pharmaceutics. 2005; 2(4):329–340. [PubMed: 16053336]
- Custodio JM, Wu CY, Benet LZ. Predicting drug disposition, absorption/elimination/transporter interplay and the role of food on drug absorption. Advanced Drug Delivery Reviews. 2008; 60(6): 717. [PubMed: 18199522]
- 35. Cao X, Gibbs S, Fang L, Miller H, Landowski C, Shin H, Lennernäs H, Zhong Y, Amidon G, Yu L, Sun D. Why is it challenging to predict intestinal drug absorption and oral bioavailability in human using rat model. Pharmaceutical Research. 2006; 23(8):1675–86. [PubMed: 16841194]
- 36. Chiou W, Barve A. Linear correlation of the fraction of oral dose absorbed of 64 drugs between humans and rats. Pharmaceutical Research. 1998; 15(11):1792–5. [PubMed: 9834005]
- Dahan A, Amidon GL. Grapefruit juice and its constituents augment colchicine intestinal absorption: Potential hazardous interaction and the role of P-glycoprotein. Pharmaceutical Research. 2009; 26(4):883–892. [PubMed: 19048359]
- Kim JS, Mitchell S, Kijek P, Tsume Y, Hilfinger J, Amidon GL. The suitability of an in situ perfusion model for permeability determinations: Utility for BCS class I biowaiver requests. Molecular Pharmaceutics. 2006; 3(6):686–694. [PubMed: 17140256]
- Lennernäs H. Animal data: the contributions of the Ussing chamber and perfusion systems to predicting human oral drug delivery in vivo. Advanced Drug Delivery Reviews. 2007; 59(11): 1103–1120. [PubMed: 17900749]
- Benet L, Amidon GL, Barends D, Lennernäs H, Polli J, Shah V, Stavchansky S, Yu L. The use of BDDCS in classifying the permeability of marketed drugs. Pharmaceutical Research. 2008; 25(3): 483–488. [PubMed: 18236138]

41. Wu C-Y, Benet LZ. Predicting drug disposition via application of BCS: Transport/absorption/ elimination interplay and development of a biopharmaceutics drug disposition classification system. Pharmaceutical Research. 2005; 22(1):11–23. [PubMed: 15771225] Dahan et al.



Figure 1.

Schematic illustration of (some of) the complexity behind absorption processes and the permeability measure, considering the whole of the human intestine, as discussed in this communication.

Table 1

Literature human jejunal permeability (P_{eff}) and fraction dose absorbed (F_{abs}) values for metoprolol, losartan and isotretinoin. Metoprolol is a conservative reference drug for the low/high permeability class boundary, as losartan and isotretinoin show lower P_{eff} yet high F_{abs} .

Drug	Human jejunal P _{eff} (×10 ⁴ cm/sec)	$F_{\rm abs}(\%)$
Metoprolol	1.34 ⁴	100 17
Losartan	1.15 7	100 4
Isotretinoin	0.99 ⁷	90 ⁴

Table 2

Effective permeability values (×10⁵ cm/sec) obtained for sotalol and metoprolol after *in-situ* single-pass perfusion to the rat proximal jejunum at pH 6.5, mid-small intestine at pH 7.0, and to the distal ileum at pH 7.5 ²². Mean (S.D.); *n*=6. Sotalol's ileal permeability exceeds/matches metoprolol's jejunal/mid small intestinal $P_{\rm eff}$ respectively, resulting in its high $F_{\rm abs}$.

	Proximal Jejunum, pH 6.5	Mid-Small Intestine, pH 7.0	Distal Ileum, pH 7.5
Metoprolol	3.7 (0.4)	5.6 (0.6)	20 (4.5)
Sotalol	0.4 (0.05)	1.7 (0.6)	5.1 (0.6)