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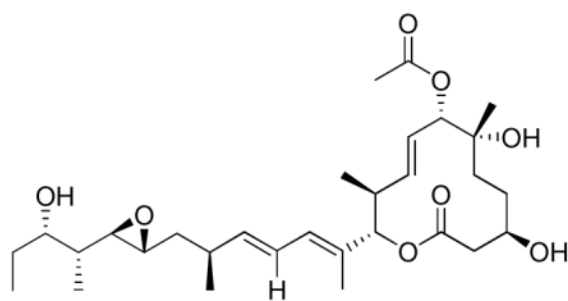
Enantioselective Total Synthesis of Pladienolide B: A Potent Spliceosome Inhibitor

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Abstract



Pladienolide B 1

An enantioselective and convergent total synthesis of pladienolide B (1) is described. Pladienolide B binds to the SF3b complex of a spliceosome and inhibits mRNA splicing activity. The synthesis features an epoxide opening reaction, an asymmetric reduction of a β -keto ester, and a cross metathesis strategy for the side chain synthesis.

In 2004, Sakai et al. reported the isolation and structural characterization of seven novel 12-membered macrocyclic compounds named pladienolide A-G from a culture of an engineered strain of *Streptomyces platensis*, Mer-11107.¹ Initial screening studies showed these compounds were capable of inhibiting the proliferation of human cancer cells with low nanomolar IC₅₀ values.² Interestingly, pladienolides maintained this activity against multiple drug resistant cancer cells. Furthermore, they exhibited a novel mechanism of action by binding to the SF3b subunit of the spliceosome, inhibiting the splicing of pre-mRNA to translatable mRNA. The unspliced pre-mRNA is exported from the nucleus to the cytoplasm resulting in inhibition of cell growth.³ To date, the pladienolides and the structurally distinct FR901464 are the only known molecular scaffolds capable of modulating splicing and generating an antitumor response. Consequently, these compounds have shown tremendous potential to become a new class of anticancer agents. In 2008, clinical trials were initiated using E7107, a pladienolide D analog, for the treatment of various cancers.⁴ Not surprisingly, the chemistry and biology of the pladienolides attracted immense attention. To date, Kotake and co-workers reported the first total synthesis of the most active compounds, pladienolide B and D.⁵ Skaanderup et al. published a synthetic pathway to the macrocyclic core of the enantiomer of pladienolide B, while Burkart and co-

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Supporting Information Available. Experimental procedures and ¹H and ¹³C NMR spectra for all new compounds have been provided. This material is available free of charge via the internet at <http://pubs.acs.org>.

workers have reported their efforts on constructing and modifying the side chain.⁶ Recently, both Maier^{7a} and Webb^{7b} have reported progress in the development of pladienolide based analogs albeit with reduced biological activity. Our interest in pladienolide B arose from its novel mechanism of action and its unique structural features. We sought to develop a convergent route to pladienolide B that could facilitate subsequent structure activity relationship studies and synthesis of structural variants. Herein, we report a concise, enantioselective synthesis of pladienolide B.

As shown in Figure 1, we planned to utilize a late stage Julia-Kocienski olefination⁸ similar to Kotake and co-workers⁵ to append the epoxide containing side chain **2** to the macrocyclic core **3**, providing the *trans-trans* diene system. The hydroxy groups in both intermediates **2** and **3** would be conveniently protected with silyl ether groups thereby allowing a global deprotection at the final stage of the synthesis. Side chain **2** could be efficiently synthesized by cross metathesis of alcohol **4** and mesylate **5**. Shi epoxidation of the resulting olefin would install the desired epoxide moiety.⁹ Both chiral alcohol **4** and mesylate **5** are readily available in optically active form. We planned to construct the macrocyclic core **3** by ring closing metathesis (RCM) of triene **6** in hopes that the free allylic alcohol would produce an activating effect resulting in higher yields and shorter reaction times.¹⁰ Of particular relevance, a related RCM macrocyclization strategy by Kotake and co-workers⁵ with a protected diol provided low yield and significant amount of isomerized olefin. Triene **6** in turn could be synthesized by Yamaguchi esterification of acid **8** and homoallylic alcohol **7**.¹¹ The synthesis of alcohol **7** could be achieved by Brown's crotylation of the corresponding aldehyde.¹² Acid **8** would be synthesized by a ring opening reaction of epoxide **9** with the dianion of *tert*-butyl acetoacetate **10** followed by asymmetric reduction of the resulting β -keto ester. Epoxide **9** could be synthesized using a desymmetrization strategy via Sharpless epoxidation.¹³

The synthesis of the C₁–C₈ fragment is outlined in Scheme 1. Asymmetric desymmetrization of commercially available divinyl carbinol **11** using Sharpless epoxidation followed by PMB protection of the corresponding alcohol provided epoxide **12** in 43% yield as a single isomer according to literature methods.¹⁴ Ring opening of epoxide **12** with the dianion of *tert*-butyl acetoacetate **10** followed by silyl protection of the resulting alcohol with TESCl afforded acyclic β -keto ester.¹⁵ Asymmetric reduction of **13** with NaBH₄-L-tartaric acid complex as prepared by Yatagai and Ohnuki¹⁶ successfully reduced the ketone with excellent stereoselectivity (> 95% dr). The depicted stereochemistry is based on the observed stereochemical outcome of β -keto esters as reported.¹⁶ Silyl protection of the newly formed alcohol provided ester **14** as a single isomer in high yields (61% from **12**). Oxidation of **14** with 3.5 eq. of IBX selectively removed the TES ether and oxidized the alcohol to a ketone without migration of the terminal olefin.¹⁷ The resulting ketone was treated with MeMgBr in THF at –78 °C providing the desired tertiary alcohol **15** as a single isomer (>95% by NMR) in 85% yield (quant. BRSM). The stereochemistry was assigned based on Cram's chelation model.¹⁸ Treatment of alcohol **15** with an excess of TESOTf in the presence of Et₃N in CH₂Cl₂ protected the tertiary alcohol as a TES ether with concomitant formation of the silyl ester which was hydrolyzed upon aqueous workup providing acid **8**. Our asymmetric synthesis of C₁–C₈ segment is quite efficient and provides convenient access to other possible stereoisomers. Previous syntheses of this segment^{5,6a,7a} relied on terpene (nerol and *R*-linalool) modification strategies.

We briefly examined the scope of the NaBH₄/tartaric acid reduction of β -keto ester **13**. As shown in Table 1, reduction of the β -keto ester in the absence of tartaric acid resulted in only a small degree of selectivity (entry 3). With the TES group removed (entry 1) the selectivity was increased suggesting the free hydroxy group had a beneficial directing effect. When the NaBH₄/tartaric acid system was employed, a further improvement in selectivity

was observed (entry 2). However, the best results were obtained with NaBH₄/tartaric acid with the hydroxy group protected as a bulky silyl ether suggesting that steric effects were the dominating factor (entries 5, 7 and 8) for good diastereoselectivity. The size of the ester group also played an important role with the bulky *t*-butyl providing the best selectivity (compare entries 4 and 5). Interestingly, using D-tartaric acid (entry 6) gave a similar degree of selectivity for the other diastereomer suggesting the chirality was transferred from the tartaric acid and not induced from the chirality inherent to the substrate.

Synthesis of optically active homoallylic alcohol **7** is shown in Scheme 2. Prenyl alcohol **16** was protected as a trityl ether with trityl chloride and Et₃N in the presence of a catalytic amount of DMAP. Riley oxidation of the resulting trityl ether provided allylic alcohol **17** in 52% yield along with some aldehyde **18**.¹⁹ The resulting mixture was exposed to MnO₂ oxidation in CH₂Cl₂ to provide aldehyde **18** in good overall yield. Brown's asymmetric crotylation¹² of aldehyde **18** furnished homoallylic alcohol **7** in > 82% *ee* and 64% yield over two steps.

With acid **8** and alcohol **7** in hand we proceeded to form the macrocyclic ring structure. As shown in Scheme 3, esterification of acid **8** with alcohol **7** according to Yamaguchi's protocol¹¹ followed by DDQ oxidative removal of the PMB ether provided allylic alcohol **19**. Ring closing metathesis (RCM) with Grubbs' 2nd generation catalyst (>95% *E* by HPLC) followed by treatment with Ac₂O in pyridine furnished lactone **20**.²⁰ Despite the presence of a free allylic alcohol, the RCM still required elevated temperatures although higher yields were observed compared to Kotake et al.^{5,10} Interestingly, when the allylic alcohol was protected as a PMB ether, RCM was very sluggish. Treatment of **20** with DDQ for 24 h selectively removed the trityl ether while oxidation of the resulting allylic alcohol with IBX afforded aldehyde **3** in 75% yield over two steps.¹⁷ The use of BCl₃ at low temperatures was also effective in selectively removing the trityl ether, however, DDQ provided more robust conditions.²¹

With the synthesis of the macrocycle core complete, we then focused on development of a concise route to the epoxide containing side chain C₁₂–C₂₃. As shown in Scheme 4, cross metathesis between homoallylic alcohol **4** and mesylate **5** proceeded in moderate yield with a high degree of selectivity for the *trans* olefin (5:1 *E:Z*). Subsequent protection of the homoallylic alcohol as a TES ether enabled the separation of olefin **21** from the other undesired cross metathesis products. Displacement of the mesylate with the anion of 1-phenyl-1*H*-tetrazole-5-thiol followed by ammonium molybdate catalyzed hydrogen peroxide oxidation gave sulfone **22** in 71% yield over two steps. Shi epoxidation⁹ of sulfone **22** followed by TES protection of the secondary alcohol afforded the side chain intermediate **2** as a single isomer.

The final assembly of pladienolide B is shown in Scheme 5. Julia-Kocienski olefination⁸ between sulfone **2** and aldehyde **3** provided diene **24** which was subsequently deprotected by the addition of TBAF yielding pladienolide B. The ¹H and ¹³C NMR of our synthetic pladienolide B {[α]_D²³ = +7.3 (*c* 0.26, MeOH)} is identical to the reported spectra for the natural pladienolide B {Lit.₅ [α]_D²⁷ = +7.9 (*c* 1.1, MeOH)} thus confirming the absolute configuration of our synthetic material.

In summary, we have accomplished an enantioselective synthesis of pladienolide B. The synthetic route is convergent and readily scaleable. Overall, our synthesis involved 31 total steps from commercially available material. The longest linear path was 16 steps with an overall yield of 1.4%. This represents a significant improvement over Kotake's route which required 59 total steps (22 longest linear) from commercially available material. The syntheses featured an effective asymmetric reduction of a β-keto ester, a cross metathesis

reaction and Julia-Kocienski olefinations. Other key reactions included a Sharpless asymmetric epoxidation and Brown asymmetric crotylboration reactions. The synthesis will provide convenient access to a variety of derivatives. Further investigations of structural and biological studies are in progress.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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References

1. (a) Sakai T, Sameshima T, Matsufuji M, Kawamura N, Dobashi K, Mizui Y. *J Antibiot.* 2004; 57:173–179. [PubMed: 15152802] (b) Sakai T, Asai N, Okuda A, Kawamura N, Mizui Y. *J Antibiot.* 2004; 57:180–187. [PubMed: 15152803] (c) Asai N, Kotake Y, Niiijima J, Fukuda Y, Uehara T, Sakai T. *J Antibiot.* 2007; 60:364–369. [PubMed: 17617693]
2. Mizui Y, Sakai T, Iwata M, Uenaka T, Okamoto K, Shimizu H, Yamori T, Yoshimatsu K, Asada M. *J Antibiot.* 2004; 57:188–196. [PubMed: 15152804]
3. (a) Kotake Y, Sagane K, Owa T, Mimori-Kiyosue Y, Shimizu H, Uesugi M, Ishihama Y, Iwata M, Mizui Y. *Nat Chem Biol.* 2007; 3:570–5. [PubMed: 17643112] (b) Yokoi A, Kotake Y, Takahashi K, Kadowaki T, Matsumoto Y, Minoshima Y, Sugi NH, Sagane K, Hamaguchi M, Iwata M, Mizui Y. *FEBS J.* 2011; 278:4870–80. [PubMed: 21981285]
4. Butler, MA. *Natural Product-Derived Compounds in Late Stage Clinical Development at the End of 2008.* Buss, A.; Butler, M., editors. The Royal Society of Chemistry; Cambridge, UK: 2009. p. 321–354.
5. Kanada RM, Itoh D, Nagai M, Niiijima J, Asai N, Mizui Y, Abe S, Kotake Y. *Angew Chem Int Ed Engl.* 2007; 46:4350–5. [PubMed: 17437311]
6. (a) Skaanderup PR, Jensen T. *Org Lett.* 2008; 10:2821–2824. [PubMed: 18510332] (b) Mandel AL, Jones BD, La Clair JJ, Burkart MD. *Bioorg Med Chem Lett.* 2007; 17:5159–5164. [PubMed: 17681759]
7. (a) Muller S, Mayer T, Sasse F, Maier ME. *Org Lett.* 2011; 13:3940–3943. [PubMed: 21707025] (b) Gundluru MK, Pourpak A, Cui X, Morris SW, Webb TR. *MedChemComm.* 2011; 2:904–908. [PubMed: 21927710]
8. Blakemore PR, Cole WJ, Kocienski P, Morley A. *Synlett.* 1998; 1:26–28.
9. Wang ZX, Tu Y, Frohn M, Zhang JR, Shi Y. *J Am Chem Soc.* 1997; 119:11224–11235.
10. Hoye T, Zhao H. *Org Lett.* 1999; 1:1123–1125. [PubMed: 10825965]
11. Inanaga J, Hirata K, Saeki H, Katsuki T, Yamaguchi M. *Bull Chem Soc Japan.* 1979; 52:1989–1993.
12. (a) Brown H, Bhat K. *J Am Chem Soc.* 1986; 108:5919–5923. [PubMed: 22175350] (b) Brown H, Bhat K. *J Org Chem.* 1986; 108:293–294.
13. Gao Y, Klunder JM, Hanson RM, Masamune H, Ko SY, Sharpless B. *J Am Chem Soc.* 1987; 109:5765–5780.
14. Schreiber S, Smith D. *J Org Chem.* 1989; 54:9–10.
15. (a) Weiler L. *J Am Chem Soc.* 1970; 92:6702–6704. (b) Lygo B, O'Connor N, Wilson PR. *Tetrahedron.* 1988; 44:6881–6888. (c) Kieczkowski G, Roberts M, Schlessinger R. *J Org Chem.* 1978; 43:788–789.
16. Yotagai M, Ohnuki T. *J Chem Soc, Perkin Trans 1.* 1990; 6:1826–1828.
17. (a) Frigerio M, Santagostino M, Sputore S, Palmisano G. *J Org Chem.* 1995; 60:7272–7276. (b) Wu Y, Huang JH, Shen X, Hu Q, Tang CJ, Li L. *Org Lett.* 2002; 4:2141–2144. [PubMed: 12074652]

18. (a) Cram DJ, Kopecky K. *J Am Chem Soc.* 1959; 81:2748–2755. (b) Reetz M. *Angew Chem Int Ed Engl.* 2003; 23:556–569.
19. Umbriet M, Sharpless K. *J Am Chem Soc.* 1977; 99:5526–5528.
20. (a) Scholl M, Ding S, Lee CW, Grubbs RH. *Org Lett.* 1999; 1:953–956. [PubMed: 10823227] (b) Chatterjee AK, Choi TL, Sanders D, Grubbs R. *J Am Chem Soc.* 2003; 125:11360–11370. [PubMed: 16220959]
21. Jones G, Hynd G, Wright J, Sharma A. *J Org Chem.* 2000; 65:263–265. [PubMed: 10813927]

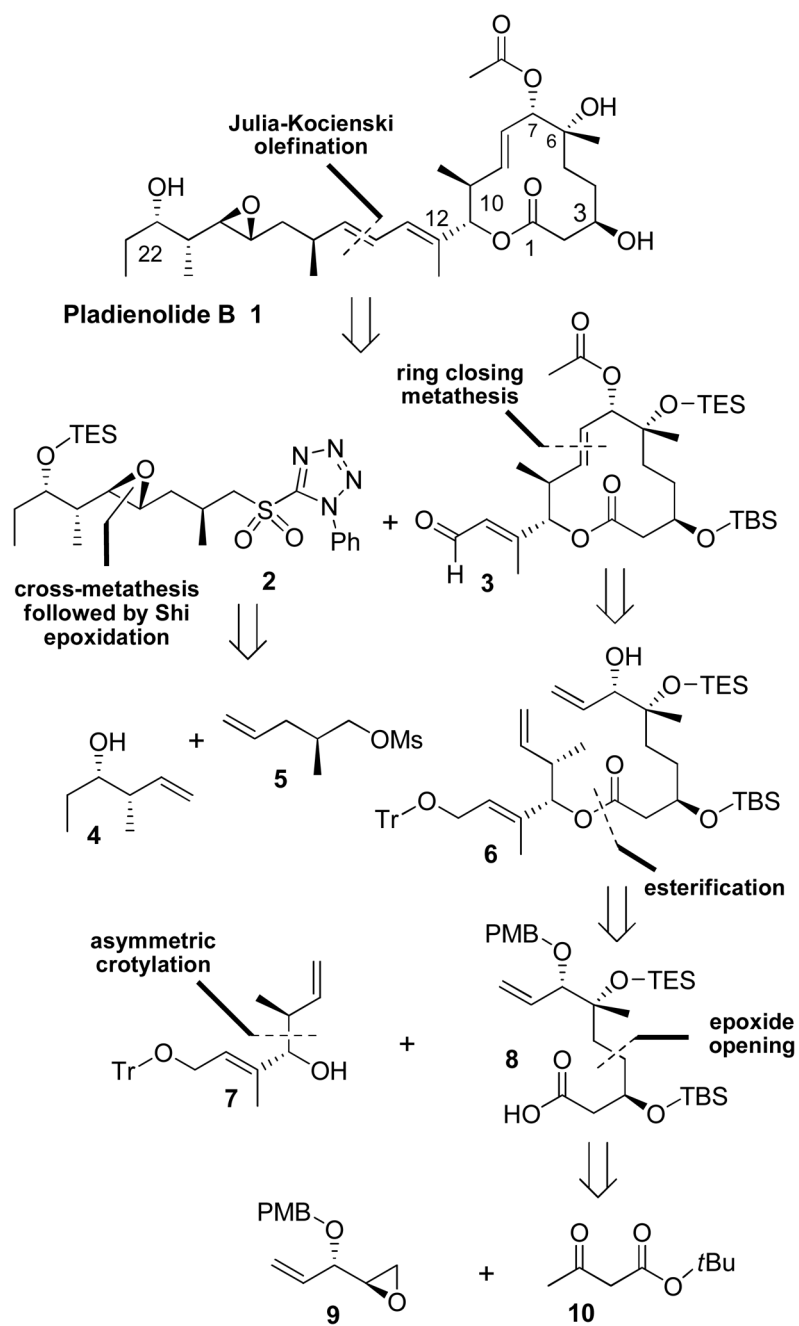
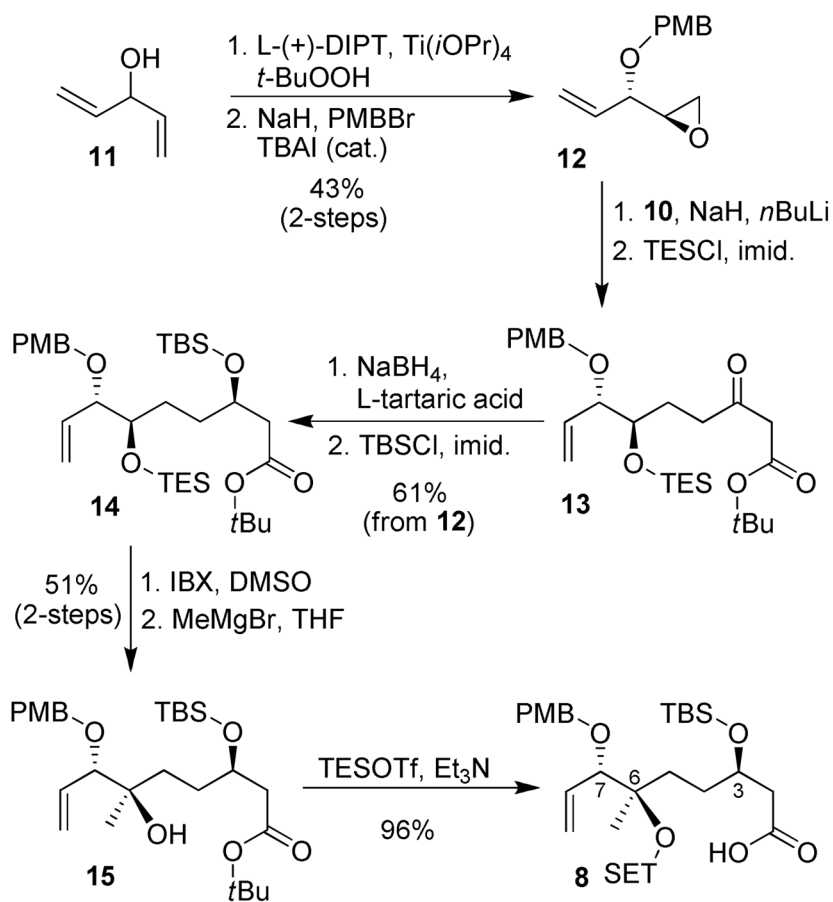
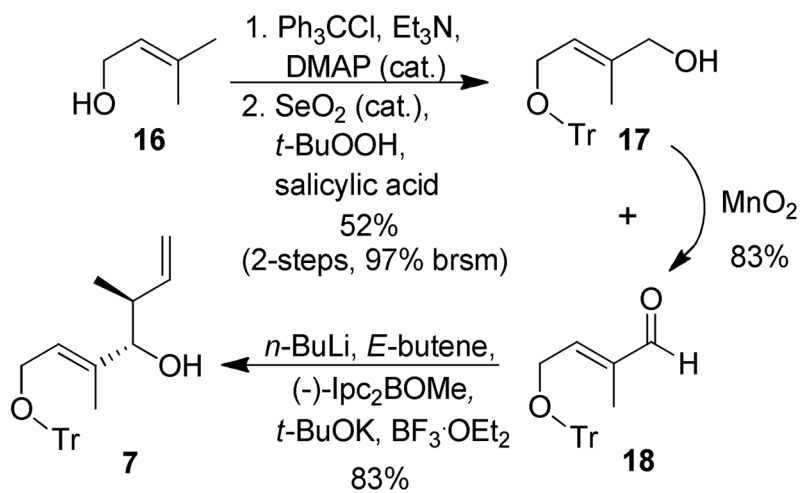


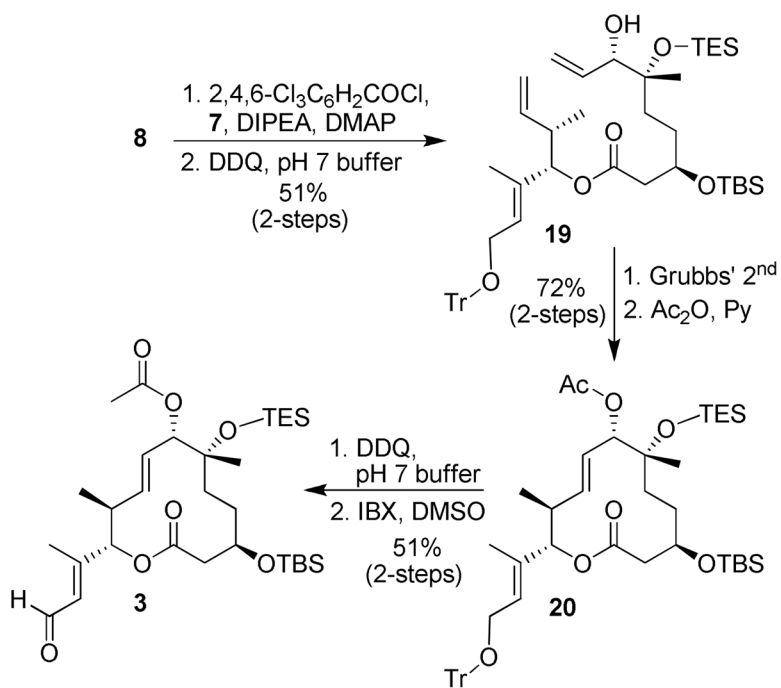
Figure 1.
Retrosynthetic Analysis of Pladienolide B



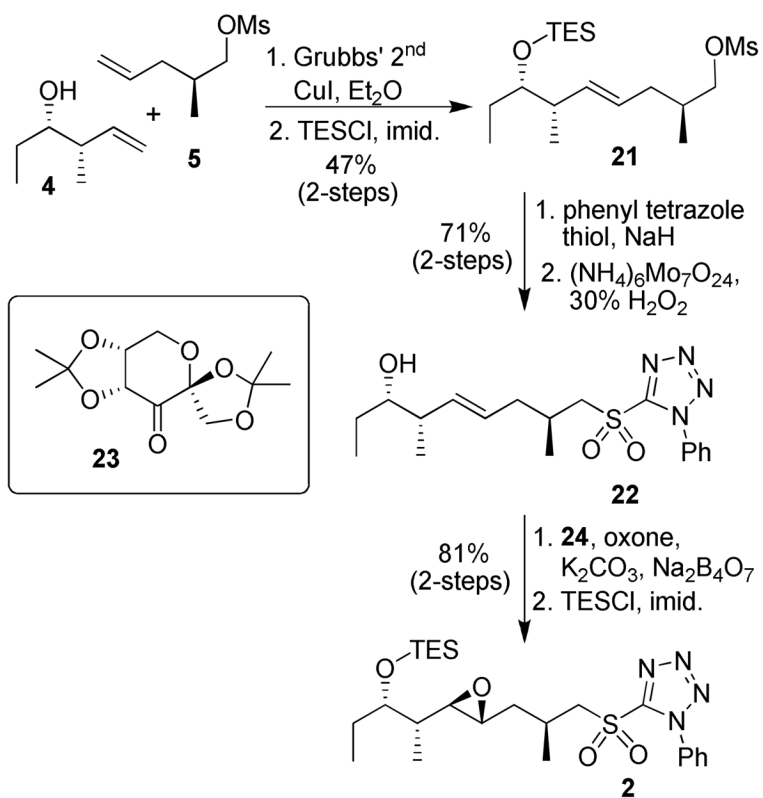
Scheme 1.
Synthesis of C₁-C₈ Intermediate **8**



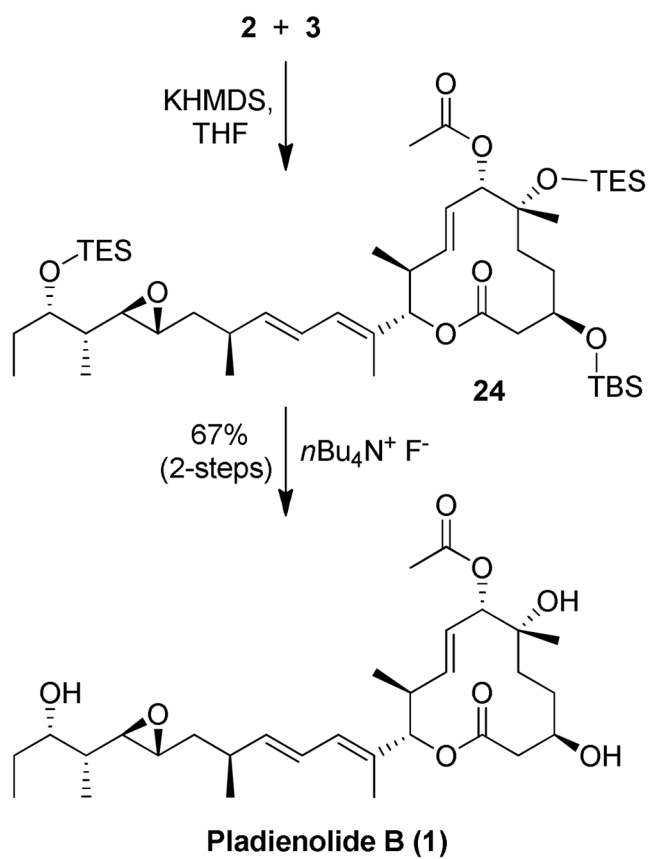
Scheme 2.
Synthesis of $\text{C}_9\text{-C}_{13}$ Intermediate 7



Scheme 3.
Synthesis of Macrocyclic Core 3

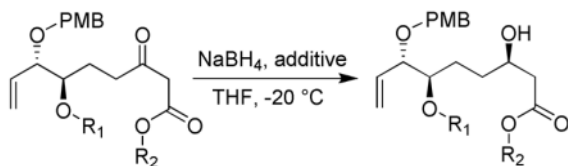


Scheme 4.
Synthesis of Sulfone 2



Scheme 5.
Synthesis of Pladienolide B

Table 1

Asymmetric Reduction of β -keto Ester

entry	R_1	R_2	additive	<i>S</i> : <i>R</i>
1	H	<i>t</i> -butyl	none	37:63
2	H	<i>t</i> -butyl	L-tartaric acid	15:85
3	TES	<i>t</i> -butyl	none	45:55
4	TES	<i>t</i> -butyl	L-tartaric acid	5:95
5	TES	methyl	L-tartaric acid	8:92
6	TES	methyl	D-tartaric acid	94:6
7	TBS	methyl	L-tartaric acid	5:95
8	TPS	methyl	L-tartaric acid	5:95