

## Draft Genome Sequence of *Paenibacillus* sp. Strain OSY-SE, a Bacterium Producing the Novel Broad-Spectrum Lipopeptide Antibiotic Paenibacterin

## En Huang,<sup>a</sup> Yaoqi Guo,<sup>a</sup> and Ahmed E. Yousef<sup>a,b</sup>

Department of Food Science and Technology, The Ohio State University, Columbus, Ohio, USA,<sup>a</sup> and Department of Microbiology, The Ohio State University, Columbus, Ohio, USA<sup>b</sup>

A strain of *Paenibacillus* sp., OSY-SE, was isolated from soil and found to produce a novel lipopeptide antibiotic. The antibiotic, paenibacterin, is active against Gram-negative and Gram-positive bacterial pathogens. Paenibacterin is biosynthesized by a non-ribosomal peptide synthetase pathway. Here we report the draft genome sequence of *Paenibacillus* sp. OSY-SE.

Resistance of pathogens to antibiotics is a rapidly evolving phe-nomenon with serious public health implications. Methicillin-resistant Staphylococcus aureus (MRSA) exemplifies the emergence of multidrug resistance among Gram-positive bacterial pathogens. Similarly, some Gram-negative pathogens have become resistant to B-lactams, fluoroquinolones, and aminoglycosides, threatening the therapeutic choices for treating these pathogens (2). Antimicrobial lipopeptides may combat antibiotic-resistant pathogens, but some of these compounds have limitations. Polymyxin, for example, has strong activity against Gramnegative bacteria (5, 9), but concerns on nephrotoxicity and neurotoxicity have limited its broad use as a therapeutic agent. Therefore, new and effective antimicrobials are urgently needed to combat these emerging drug-resistant pathogens. Paenibacterin is a newly discovered cyclic lipopeptide produced by Paenibacillus sp. OSY-SE. The lipopeptide comprises 13 amino acids and a  $C_{15}$ fatty acyl moiety (3). Paenibacterin is active against Gram-negative and Gram-positive pathogens, including Escherichia coli O157:H7, Salmonella enterica serovar Typhimurium, Listeria monocytogenes, and methicillin-resistant Staphylococcus aureus. To understand the biosynthesis of paenibacterin, we determined the whole-genome sequence of the producer strain, Paenibacillus sp. OSY-SE.

Genomic DNA of Paenibacillus sp. OSY-SE was isolated using a DNA extraction kit (DNeasy blood and tissue kit; Qiagen, Valencia, CA). RNase-treated genomic DNA in Tris-Cl buffer (pH 8.5) was used for construction of a paired-end library with a TruSeq DNA sample preparation kit (Illumina, San Diego, CA) according to the manufacturer's instruction. The constructed library was sequenced (76-cycle paired-end runs) in a flow cell lane using Illumina Genome Analyzer II. De novo assembly of the short reads with commercial software (CLC Genomics Workbench 4.7.2; CLCBio, Cambridge, MA) yielded 205 contigs (>200 bp each), with a maximum contig size of 359,285 bp. The resulting draft genome of Paenibacillus sp. OSY-SE consists of 6,931,767 bases; the overall GC content of the genome was calculated as 48.66% by the software Artemis (8). Automatic genome annotation was performed using the rapid annotations using subsystems technology (RAST) server (1). Among the 6,475 protein-coding sequences (CDSs), 65.79% have been assigned a putative function by RAST. The chromosome has one rRNA operon and 38 tRNA genes, as predicted by RNAmmer (4) and tRNAscan-SE (6), respectively.

The average nucleotide identities (ANI) between Paenibacillus

sp. OSY-SE and 20 genomes of *Paenibacillus* species that are available in GenBank were determined using the *in silico* DNA-DNA hybridization method implemented in the software JSpecies (7). The results indicated that *Paenibacillus* sp. OSY-SE has the closest genetic relatedness with *Paenibacillus* lactis strain 154 (ANI, 77.21%). The gene cluster responsible for paenibacterin biosynthesis was identified in a 52-kb region encoding three nonribosomal peptide synthetases and two ABC-like transporters.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number ALKF00000000. The version described in this paper is the first version, ALKF01000000.

## ACKNOWLEDGMENT

This research was supported by the Virginia Hutchison Bazler and Frank E. Bazler Designated Professorship in Food Science.

## REFERENCES

- 1. Aziz R, et al. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75.
- Fischbach MA, Walsh CT. 2009. Antibiotics for emerging pathogens. Science 325:1089–1093.
- Guo Y, Huang E, Yuan C, Zhang L, Yousef AE. 2012. Isolation of a strain of *Paenibacillus* sp. and structural elucidation of its broad-spectrum lipopeptide antibiotic. Appl. Environ. Microbiol. 78:3156–3165.
- Lagesen K, et al. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. Nucleic Acids Res. 35:3100–3108.
- Landman D, Georgescu C, Martin DA, Quale J. 2008. Polymyxins revisited. Clin. Microbiol. Rev. 21:449–465.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964.
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc. Natl. Acad. Sci. U. S. A. 106:19126.
- 8. Rutherford K, et al. 2000. Artemis: sequence visualization and annotation. Bioinformatics 16:944–945.
- Zavascki AP, Goldani LZ, Li J, Nation RL. 2007. Polymyxin B for the treatment of multidrug-resistant pathogens: a critical review. J. Antimicrob. Chemother. 60:1206–1215.

Received 17 August 2012 Accepted 29 August 2012 Address correspondence to Ahmed E. Yousef, yousef.1@osu.edu. E.H. and Y.G. contributed equally to this study. Copyright © 2012, American Society for Microbiology. All Rights Reserved. doi:10.1128/JB.01506-12