

# Complete Genome Sequence of a Mammalian Species-Infectious and -Pathogenic H6N5 Avian Influenza Virus without Evidence of Adaptation

Seong-Jun Park,<sup>a</sup> Bong-Kyun Park,<sup>b</sup> Dae-Sub Song,<sup>a,c</sup> and Haryoung Poo<sup>a,c</sup>

Viral Infectious Disease Research Center, Korea Research Institute of Bioscience and Biotechnology, Daejeon, South Korea<sup>a</sup>; Department of Veterinary Medicine Virology Lab, College of Veterinary Medicine and BK21 Program for Veterinary Science, Seoul National University, Gwanak-gu, Seoul, South Korea<sup>b</sup>; and University of Science and Technology, Daejeon, South Korea<sup>c</sup>

**An H6N5 avian influenza virus (AIV) strain, designated A/aquatic bird/Korea/CN5/2009 (H6N5), was isolated from fecal swabs of aquatic birds in 2009, and surprisingly, it showed infectivity and pathogenicity in mammalian species without evidence of adaptation. In this study, we report the first complete genome sequence containing 3' and 5' noncoding regions (NCRs) of a mammalian species-infectious and pathogenic H6N5 AIV, which will help provide important insights into the molecular basis of pathogenesis, transmission, and evolution of AIV.**

Avian influenza virus (AIV) is a segmented, negative-sense, single-stranded RNA virus belonging to the family *Orthomyxoviridae*, genus *Influenzavirus A*. Aquatic birds are the natural reservoirs of influenza A viruses (3, 16). The A/aquatic bird/Korea/CN5/2009 (H6N5) [A/AB/Kor/CN5/09 (H6N5)] strain was isolated from fecal swabs of aquatic birds in 2009. Surprisingly, it showed infectivity and pathogenicity in mammalian species without evidence of adaptation (6). To date, primers targeting the noncoding regions (NCRs) have generally been used to obtain complete genomes of influenza A viruses. Thus, the exact sequences of NCRs have scarcely been determined. Moreover, a complete genome sequence containing 3' and 5' NCRs of a mammalian species-infectious and pathogenic H6N5 AIV with no evidence of adaptation has not been reported despite multifunctions of NCRs in the replication of influenza A viruses (4, 5, 14, 15, 18). For those reasons, it is necessary to analyze the complete genome sequence containing 3' and 5' NCRs of A/AB/Kor/CN5/09 (H6N5) and understand its molecular characteristics.

Viral RNA was isolated from allantoic fluids of embryonated eggs infected with the A/AB/Kor/CN5/09 (H6N5) ( $10^{7.75}$  50% egg infective doses [EID<sub>50</sub>]/ml) by using an RNeasy minikit (Qiagen) and circularized with T4 RNA ligase as described previously (2, 13). The PCR products produced by RNA ligation-mediated reverse transcriptase PCR (RT-PCR) were purified, cloned (9), and sequenced to determine its exact complete genome sequence containing 3' and 5' NCRs on an automated DNA sequencer (ABI system 3700; Applied Biosystems Inc.) by utilizing universal primers (7) with simple modifications and newly designed segment-specific primers.

The complete genome of A/AB/Kor/CN5/09 (H6N5) is 13,607 nucleotides (nt) long; segments 1 (Seg-1) to 8 (Seg-8) are 2,341, 2,341, 2,233, 1,765, 1,565, 1,467, 1,027, and 890 nt, respectively. They encode 12 viral proteins with amino acid lengths as follows: PB2, 759; PB1, 757; N40 (an N-terminally truncated and functionally distinct variant of PB1) (17), 718; PB1-F2, 90; PA, 716; HA, 566; NP, 498; NA, 472; M1, 252; M2, 97; NS1, 230; and NS2 (nuclear export protein [NEP]), 121.

The sizes of NCRs of viral RNA of A/AB/Kor/CN5/09 (H6N5) were variable (17 [Seg-4] to 45 [Seg-5] and 20 [Seg-7] to 58 [Seg-3] nt at the 3' and 5' NCRs, respectively) in the different

genome segments, but the terminal 12 (3'UCGYUUUCGUCC-) and 13 (-GGAACAAAGAUGA5') nt of the 3' and 5' ends, respectively, were highly conserved among all genome segments, which is consistent with results of previous studies (1, 8, 11). Furthermore, a uridine-rich region (5 to 6 U's), which serves as the polyadenylation site (10, 12), was observed from positions 15 through 16 at the 5' end of each segment.

This is the first report of the complete genome sequence containing 3' and 5' NCRs of H6N5 AIV showing infectivity and pathogenicity in mammalian species without evidence of adaptation. We hope that these data will help elucidate the molecular basis of pathogenesis, transmission, and evolution of AIV as well as other influenza A viruses.

**Nucleotide sequence accession numbers.** The complete genome sequence of the A/AB/Kor/CN5/09 (H6N5) has been deposited in GenBank under accession numbers [JX465637](#) to [JX465644](#) for Seg-1 to Seg-8.

## ACKNOWLEDGMENTS

This work was supported by a National Agenda Project grant from the Korea Research Council of Fundamental Science & Technology and the KRIBB Initiative program (KGM3121221).

## REFERENCES

1. Desselberger U, Racaniello VR, Zazra JJ, Palese P. 1980. The 3' and 5'-terminal sequences of influenza A, B and C virus RNA segments are highly conserved and show partial inverted complementarity. *Gene* 8:315–328.
2. de Wit E, et al. 2007. Rapid sequencing of the non-coding regions of influenza A virus. *J. Virol. Methods* 139:85–89.
3. Fouchier RA, et al. 2005. Characterization of a novel influenza A virus hemagglutinin subtype (H16) obtained from black-headed gulls. *J. Virol.* 79:2814–2822.
4. Hagen M, Chung TD, Butcher JA, Krystal M. 1994. Recombinant

Received 25 August 2012 Accepted 27 August 2012

Address correspondence to Dae-Sub Song, [sds1@kribb.re.kr](mailto:sds1@kribb.re.kr), or Haryoung Poo, [haryoung@kribb.re.kr](mailto:haryoung@kribb.re.kr).

Copyright © 2012, American Society for Microbiology. All Rights Reserved.

doi:10.1128/JVI.02301-12

- influenza virus polymerase: requirement of both 5' and 3' viral ends for endonuclease activity. *J. Virol.* **68**:1509–1515.
5. Lee MT, Klumpp K, Digard P, Tiley L. 2003. Activation of influenza virus RNA polymerase by the 5' and 3' terminal duplex of genomic RNA. *Nucleic Acids Res.* **31**:1624–1632.
  6. Nam JH, et al. 2011. Emergence of mammalian species-infectious and -pathogenic avian influenza H6N5 virus with no evidence of adaptation. *J. Virol.* **85**:13271–13277.
  7. Obenauer JC, et al. 2006. Large-scale sequence analysis of avian influenza isolates. *Science* **311**:1576–1580.
  8. Park SJ, et al. 2012. Complete genome sequence of an avian-origin H3N2 canine influenza virus isolated from dogs in South Korea. *J. Virol.* **86**:9548–9549.
  9. Park SJ, Song DS, Ha GW, Park BK. 2007. Cloning and further sequence analysis of the spike gene of attenuated porcine epidemic diarrhea virus DR13. *Virus Genes* **35**:55–64.
  10. Poon LL, Pritlove DC, Fodor E, Brownlee GG. 1999. Direct evidence that the poly(A) tail of influenza A virus mRNA is synthesized by reiterative copying of a U track in the virion RNA template. *J. Virol.* **73**:3473–3476.
  11. Robertson JS. 1979. 5' and 3' terminal nucleotide sequences of the RNA genome segments of influenza virus. *Nucleic Acids Res.* **6**:3745–3757.
  12. Robertson JS, Schubert M, Lazzarini RA. 1981. Polyadenylation sites for influenza virus mRNA. *J. Virol.* **38**:157–163.
  13. Szymkowiak C, et al. 2003. Rapid method for the characterization of 3' and 5' UTRs of influenza viruses. *J. Virol. Methods* **107**:15–20.
  14. Tiley LS, Hagen M, Matthews JT, Krystal M. 1994. Sequence-specific binding of the influenza virus RNA polymerase to sequences located at the 5' ends of the viral RNAs. *J. Virol.* **68**:5108–5116.
  15. Wang L, Lee CW. 2009. Sequencing and mutational analysis of the non-coding regions of influenza A virus. *Vet. Microbiol.* **135**:239–247.
  16. Webster RG, Bean WJ, Gorman OT, Chambers TM, Kawaoka Y. 1992. Evolution and ecology of influenza A viruses. *Microbiol. Rev.* **56**:152–179.
  17. Wise HM, et al. 2009. A complicated message: identification of a novel PB1-related protein translated from influenza A virus segment 2 mRNA. *J. Virol.* **83**:8021–8031.
  18. Zheng H, Palese P, Garcia-Sastre A. 1996. Nonconserved nucleotides at the 3' and 5' ends of an influenza A virus RNA play an important role in viral RNA replication. *Virology* **217**:242–251.