

Complete Genome Sequence of Temperate Bacteriophage *AcaML1* from the Extreme Acidophile *Acidithiobacillus caldus* ATCC 51756

Pablo Tapia,^a Francisco Moya Flores,^b Paulo C. Covarrubias,^{b,c} Lillian G. Acuña,^{b,c} David S. Holmes,^{b,c} and Raquel Quatrini^{b,c}

Facultad de Ciencias, Pontificia Universidad Católica de Valparaíso, Valparaíso, Chile^a; Facultad de Ciencias Biológicas, Universidad Andres Bello, Santiago, Chile^b; and Fundación Ciencia & Vida, Santiago, Chile^c

Development of reproducible genetic tools in the industrially important acidithiobacilli is urgently required. Inducible temperate phages which may be modified *in vitro*, propagated in suitable hosts, and used to transduce relevant genetic information to other strains and/or species are potentially valuable tools in this field of research. In order to address these current limitations, the genome sequence of an inducible temperate *Myoviridae*-like bacteriophage from the *Acidithiobacillus caldus* type strain was annotated and analyzed bioinformatically. Here, we announce the genome sequence of *AcaML1* and report major findings from its annotation.

The acidithiobacilli are gammaproteobacteria that are ubiquitous in biomineral biotopes, with several characterized species that play key roles in industrial metal recovery (6). To date, no phage capable of infecting members of this group has been identified.

Several members of this group have recently been sequenced (3, 8–10). Bioinformatic analysis of the genome sequence of the moderately thermophilic, sulfur-oxidizing acidophile *Acidithiobacillus caldus* ATCC 51756 predicted the presence of a 59-kb temperate phage located between *srrA* tmRNA and tRNA^{Met}. Given the absence of reproducible genetic modification tools for the acidithiobacilli (5), identification and genomic characterization of this novel lysogenic, inducible *Myoviridae* bacteriophage, termed *A. caldus* myovirus-like 1 (*AcaML1*), potentially capable of infecting and transducing genetic information into members of this genus, represent a major contribution to the field.

Genomic libraries of 4 kb and 40 kb were constructed and sequenced using the random shotgun strategy (7). Assembly of qualified reads, gene modeling, and annotation were performed as previously described (8–10). Curation of the annotation was carried out using conserved protein motif analyses and motif databases, such as Interproscan (<http://www.ebi.ac.uk/Tools/pfa/iprscan>) and POG (4), and comparative genomic strategies against viral sequence repositories, such as PhAnToMe (<http://www.phantom.org>).

The *AcaML1* genome has 59,353 bp and a GC content of 64.5%, which is higher than the average GC content of its host (61.6%). It is predicted to encode 72 gene products arranged in 10 clusters, 52.8% of which display sequence similarity to known proteins, 34.4% are hypotheticals, and 12.8% present no similarity to any other proteins reported to date. Gene clusters 1 and 2 encode signature proteins implicated in lysogeny establishment and regulation and control of the lysogeny-lytic switch (integrase, excisionase, regulators, primase, RNase endonuclease, methyltransferases). Gene clusters 3 to 9 encode gene products predicted to play a role in viral particle formation and assembly (procapsid shell and maturation protease, baseplate, contractile tail tube, and tail fibers), DNA packaging (terminase, portal protein), and host lysis for viral particle release (holin and endolysin).

The distal gene module in *AcaML1* contains three genes encoding an McrBC DNA restriction system and an accompanying

modification enzyme (cytosine methylase) and two insertion sequences (IS5, IS21). The presence of RM systems in phages is frequent, playing a relevant role in protection of phage DNA from degradation during phage subversion of the host resources and/or the stabilization of the mobile element within the host genome (e.g., reference 2). These gene cassettes are frequent targets of horizontal gene transfer between bacteria, further facilitated by flanking transposases (e.g., reference 1).

Ongoing studies of the genome of *AcaML1* and its life cycle, including its infectivity, latent period, burst size, and host range, will deepen our understanding of its interaction with the *A. caldus* host and provide further insight into its potential as a tool for genetic modification in this group of biotechnologically relevant but genetically recalcitrant microbes.

Nucleotide sequence accession number. The complete genome sequence of *Acidithiobacillus caldus* *Myoviridae*-like phage *AcaML1* is available in GenBank under accession number [JX507079](https://www.ncbi.nlm.nih.gov/nuccore/JX507079).

ACKNOWLEDGMENTS

This work was supported by Fondecyt grant numbers 1100887 and 1090451. P.C.C. and L.A. were the recipients of graduate training CONICYT fellowships.

REFERENCES

1. Kita K, Kawakami H, Tanaka H. 2003. Evidence for horizontal transfer of the EcoT38I restriction-modification gene to chromosomal DNA by the P2 phage and diversity of defective P2 prophages in *Escherichia coli* TH38 strains. *J. Bacteriol.* 185:2296–2305.
2. Kobayashi I. 2001. Behavior of restriction-modification systems as selfish mobile elements and their impact on genome evolution. *Nucleic Acids Res.* 29:3742–3756.
3. Liljeqvist M, Valdes J, Holmes DS, Dopson M. 2011. Draft genome of the psychrotolerant acidophile *Acidithiobacillus ferrivorans* SS3. *J. Bacteriol.* 193:4304–4305.
4. Liu J, Glazko G, Mushegian A. 2006. Protein repertoire of double-stranded bacteriophages. *Virus Res.* 117:68–80.

Received 22 August 2012 Accepted 22 August 2012

Address correspondence to Raquel Quatrini, rquatrini@cienciavida.cl.

Copyright © 2012, American Society for Microbiology. All Rights Reserved.

doi:10.1128/JVI.02261-12

5. Quatrini R, Valdés J, Jedlicki E, Holmes DS. 2007. The use of bioinformatics and genome biology to advance our understanding of bioleaching microorganisms, p 221–239. *In* Donati E, Sand W (ed), *Microbial processing of metal sulfides*. Springer, Dordrecht, The Netherlands.
6. Rawlings DE, Johnson DB. 2007. The microbiology of biomining: development and optimization of mineral-oxidizing microbial consortia. *Microbiology* 153:315–324.
7. Tettelin H, et al. 2001. Complete genome sequence of a virulent isolate of *Streptococcus pneumoniae*. *Science* 293:498–506.
8. Valdés J, et al. 2008. *Acidithiobacillus ferrooxidans* metabolism: from genome sequence to industrial applications. *BMC Genomics* 9:597.
9. Valdés J, et al. 2009. Draft genome sequence of the extremely acidophilic bacterium *Acidithiobacillus caldus* ATCC 51756 reveals metabolic versatility in the *Acidithiobacillus* genus. *J. Bacteriol.* 191:5877–5878.
10. Valdés J, Ossandon F, Quatrini R, Dopson M, Holmes DS. 2011. Draft genome sequence of the extremely acidophilic biomining bacterium *Acidithiobacillus thiooxidans* ATCC 19377 provides insights into the evolution of the *Acidithiobacillus* genus. *J. Bacteriol.* 193:7003–7004.