Original Article

Complications

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Correlations between Glucagon Stimulated C-peptide Levels and Microvascular Complications in Type 2 Diabetes Patients

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Background: This study aimed to investigate whether stimulated C-peptide is associated with microvascular complications in type 2 diabetes mellitus (DM).

Methods: A cross-sectional study was conducted in 192 type 2 diabetic patients. Plasma basal C-peptide and stimulated C-peptide were measured before and 6 minutes after intravenous injection of 1 mg glucagon. The relationship between C-peptide and microvascular complications was statistically analyzed.

Results: In patients with retinopathy, basal C-peptide was 1.9 ± 1.2 ng/mL, and stimulated C-peptide was 2.7 ± 1.6 ng/mL; values were significantly lower compared with patients without retinopathy (P=0.031 and P=0.002, respectively). In patients with nephropathy, basal C-peptide was 1.6 ± 0.9 ng/mL, and stimulated C-peptide was 2.8 ± 1.6 ng/mL; values were significantly lower than those recorded in patients without nephropathy (P=0.020 and P=0.026, respectively). Stimulated C-peptide level was associated with increased prevalence of microvascular complications. Age-, DM duration-, and hemoglobin A1c-adjusted odds ratios for retinopathy in stimulated C-peptide value were 4.18 (95% confidence interval [CI], 1.40 to 12.51) and 3.35 (95% CI, 1.09 to 10.25), respectively. The multiple regression analysis between nephropathy and C-peptide showed that stimulated C-peptide was statistically correlated with nephropathy (P=0.03).

Conclusion: In patients with type 2 diabetes, the glucagon stimulation test was a relatively simple method of short duration for stimulating C-peptide response. Stimulated C-peptide values were associated with microvascular complications to a greater extent than basal C-peptides.

Keywords: Basal C-peptide; Diabetes mellitus, type 2; Glucagon stimulation test; Microvascular complications; Stimulated C-peptide

INTRODUCTION

C-peptide and insulin are secreted in equimolar amounts from the β -cells of the pancreas. A large amount of secreted insulin is removed by the liver during passage; in contrast, only a minor portion of serum C-peptide is taken out of the circulation by the liver [1,2]. Therefore, serum C-peptide more accurately

reflects the β -cell function of the pancreas than serum insulin. Furthermore, C-peptide is also less affected in insulin-treated patients and by the presence of insulin-binding antibodies.

Previously, C-peptide was regarded as a biologically inert hormone. According to recent studies, however, C-peptide has been considered an active peptide hormone with an important physiologic function and an intracellular signaling function

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[3]. C-peptide may play a role in microvascular blood flow [4]. C-peptide also stimulates blood flow and glucose transport. A low level of C-peptide is an indicator of the progression of diabetic angiopathies [5]. In addition, C-peptide decreases glomerular hyperfiltration and albuminuria in type 1 diabetes mellitus (DM) [6].

In the Diabetes Control and Complications Trial (DCCT), type 1 DM patients with C-peptide capacity showed a significantly lower prevalence of microvascular complications than patients totally lacking C-peptide secretion by β -cells [7]. In another study, stimulated C-peptide level was found to be associated with a lower risk of microvascular complications in type 1 DM [8].

Methods for stimulating C-peptide response include the mixed-meal tolerance test (MMTT), glucagon stimulation test (GST) and the arginine stimulation test (AST). The GST has the advantage of a shorter duration (6 minutes), simple standardization and no side effects compared to mixed meals and other stimuli [9]. The C-peptide response to glucagon stimulation reflects β -cell function. Moreover, the assessment of stimulated C-peptide levels is recommended rather than that of basal C-peptide levels, due to a lower degree of variation in the former [10].

The purpose of this study was to examine whether stimulated C-peptide levels using GST were significantly associated with microvascular complications in type 2 DM patients.

METHODS

Participants

A cross-sectional study was conducted in diabetic inpatients of Konyang University Hospital from January 2008 to December 2009. We recruited 192 patients with type 2 DM aged 45 years or order. All agreed to participate and gave oral and written consent. The Institutional Review Board of the Clinical Research Center in Konyang University Hospital approved the study protocol. Type 2 DM was diagnosed using the American Diabetes Association criteria as follows: Hemoglobin A1c (HbA1c) \geq 6.5% using an National Glycohemoglobin Standarization Program (NGSP) certified method; plasma glucose \geq 126 mg/dL (7 mmol/L) after an overnight fast; a positive value should be confirmed with a repeat test; symptoms of diabetes (polyuria, polydipsia, fatigue, weight loss) and a random plasma glucose level of >200 mg/dL (11.1 mmol/L); and oral glucose tolerance test that showed a plasma glucose level of

>200 mg/dL (11.1 mmol/L) at 2 hours after ingestion of 75 g of glucose. We included only those patients who were over 30 years of age at diagnosis. Patients with acute inflammation, infection (pneumonia, urinary tract infection and DM foot), hepatitis, malignancy, or type 1 DM were excluded. We also excluded patients who required exogenous insulin and thiazolidinedione treatment, as well as those with renal impairment (glomerular filtration rate [GFR] \leq 60 mL/min/1.73 m²) or plasma basal glucose levels \geq 250 mg/dL, because these factors can affect C-peptide levels. Participants were evaluated for their baseline clinical and laboratory profiles. We assessed systolic and diastolic blood pressures (BP), fasting blood glucose levels, HbA1c, lipid panel, uric acid, homocystein, apolipoprotein (Apo A and B), and laboratory data.

Methods

The duration of diabetes was determined based upon when the patient was first diagnosed with the disorder. Body mass index (BMI) was calculated by dividing weight in kilograms by height in meters squared (kg/m²). We evaluated C-peptide levels in the fasting state because serum glucose levels and the ingestion of drugs known as sulfonylureas (that boost the body's production of insulin) can affect serum C-peptide levels. After an 8-hour fast, venous blood was collected for laboratory analysis. The fasting blood glucose level was determined by a routine hexokinase method. HbA1c and C-peptide levels, lipid panel, uric acid, homocystein, and Apo A and B were estimated. Uric acid, total cholesterol, triglycerides, and low density lipoprotein cholesterol (LDL-C) were measured by enzymatic methods, while high density lipoprotein cholesterol (HDL-C) was assessed via a selective inhibition assay. HbA1c was determined by high-performance liquid chromatography using an automatic glycohemoglobin analyzer (Tosoh HLC-723 7; Tosoh Co., Tokyo, Japan). Plasma C-peptide was analyzed by radioimmunoassay. The intra- and inter-assay coefficients of variations were 4% and 6%, respectively. Systolic and diastolic BPs were measured after the patient had been seated for at least 10 minutes. The homeostasis model assessment of insulin resistance (HOMA-IR) and of β-cell function (HOMA-β) was calculated using the HOMA calculator version 2.2.2 by using the homeostasis model assessment (HOMA2) as a percentage of the values in a normal reference population.

The GST was performed after an overnight fast. Plasma C-peptide levels were measured before and 6 minutes after intravenous injection of 1 mg of glucagon (basal and stimulated C-



peptide, respectively).

Diagnosis of diabetic retinopathy was performed through fundoscopic examination by one ophthalmologist experienced in diabetic retinopathy. We included both patients with nonproliferative diabetic retinopathy and those with proliferative diabetic retinopathy. Retinopathy was defined by the presence of dot and blot hemorrhages, hard exudates, cotton wool spots, laser scars, and a history of vitrectomy as described by the European protocol for diabetic retinopathy. Proliferative retinopathy was classified as the presence of new vessels, which were subsequently confirmed by fluorescein angiography. Urinary albumin excretion was estimated using 24-hour urine collection. Diabetic nephropathy was defined as urinary albumin excretion ≥30 mg/day (microalbuminuria [30 to 300 mg/ day] and macroalbuminuria [>300 mg/day]) in the absence of other reasons for albuminuria. Patients with GFR < 60 mL/min/ 1.73 m² were excluded. GFR was estimated using the Modification of Diet in Renal Disease (MDRD) study equation. Diabetic neuropathy was diagnosed on the basis of abnormal nerve conduction velocity. Furthermore, we included symptoms (decreased sensation, numbness, prickling, and burning) and any positive findings on neurologic examination by the Michigan Diabetic Neuropathy Score in our data.

We created four C-peptide groups that were defined by the relationship between basal C-peptide and Δ C-peptide values. Δ C-peptide was defined as the difference between basal and stimulated C-peptides. Because C-peptide is a continuous variable, we used the mean value of C-peptide measurements (basal C-peptide, Δ C-peptide). The mean basal serum C-peptide value was 1.9 ng/mL. Patients with basal C-peptide values of less than 1.9 ng/mL were classified into group 1 or 2; patients with basal C-peptide values of 1.9 ng/mL or more were placed into group 3 or 4.

The mean value of ΔC was 1.1 ng/mL. Patients with ΔC -peptide values of less than 1.1 ng/mL were classified into either group 1 or 3. Patients with C values of 1.1 ng/mL or more were put into group 2 or 4. Thus, the four groups were defined as follows: Group 1 (basal C-peptide <1.9 ng/mL, ΔC -peptide <1.1 ng/mL), Group 2 (basal C-peptide <1.9 ng/mL, ΔC -peptide \geq 1.1 ng/mL), Group 3 (basal C-peptide \geq 1.9 ng/mL, ΔC -peptide <1.1 ng/mL), and Group 4 (basal C-peptide \geq 1.9 ng/mL, ΔC -peptide \geq 1.1 ng/mL).

First, we investigate the association of C-peptide groups with the presence microvascular complications. Secondly, we evaluated the association of stimulated C-peptide levels with microvascular complications in type 2 DM.

Statistical analysis

All analyses were performed using the SPSS version 18.0 (SPSS Inc., Chicago, IL, USA). Characteristics of C-peptide groups and the relationship between C-peptide groups and microvascular complications were assessed using a one-way analysis of variance (ANOVA) method. Diabetic microvascular complications caused by C-peptides (basal and stimulated C-peptides) were analyzed by independent *t*-tests. Data are presented as mean±standard deviation or percentage, and a *P* value of less than 0.05 was considered statistically significant.

Multiple logistic regression analyses were used to estimate the odds ratios (ORs) for microvascular complications after adjusting for other clinical and biochemical variables such as age, DM duration, and degree of glycemic control (HbA1c). The relationship between nephropathy and C-peptides (basal and stimulated C-peptides) was analyzed by multiple regression analysis. Furthermore, a *P* value of less than 0.05 was considered statistically significant.

RESULTS

Clinical characteristics of the patients in four groups

The clinical characteristics of the patients in the four groups are shown in Table 1. We included 192 patients in the study (98 males [51.0%], 94 females [48.9%]), who were divided into four groups according to the mean value of C-peptide measurements. BMI, HbA1c, total cholesterol, LDL-C, triglycerides, uric acid, Apo B, HOMA-IR and HOMA-β levels were found to be positively associated with the C-peptide groups (Table 1). No differences between groups were found for systolic BP, diastolic BP, fasting blood glucose, or GFR. BMI was significantly higher in Group 4 (high basal C-peptide, high Δ C-peptide) than in Group 1 (low basal C-peptide, low Δ Cpeptide) (P<0.01). HbA1c was significantly lower in Group 4 (high basal C-peptide, high ∆C-peptide) than in Group 1 (low basal C-peptide, low Δ C-peptide) (P<0.01). Total cholesterol, LDL-C and triglyceride were significantly higher in Group 4 than in Group 1 (P=0.01, P=0.02, and P<0.01, respectively); uric acid and Apo B levels were also significantly different between these groups (P=0.04 and P=0.03, respectively). Our results showed that Group 4 experienced increased metabolic components and insulin resistance than Group 1. As insulin resistance increased, the β-cell function compensated; Group



| Table 1. Characteristics of the clinic-b | sed cohort of 192 partic | ipants with type 2 diabetes b | y C-peptide group |
|---|--------------------------|-------------------------------|-------------------|
| | | | |

| Cl | | C-peptide group | | | |
|---------------------------------|------------------|------------------|-------------------|-------------------|---------|
| Characteristic | 1 | 2 | 3 | 4 | P value |
| No. (%) | 57 (29.6) | 40 (20.8) | 41 (21.4) | 54 (28.1) | |
| Age, yr | 59.4±8.3 | 58.9 ± 9.3 | 61.3±9.5 | 58.5 ± 8.3 | 0.47 |
| DM duration, yr | 9.0 ± 7.7 | 10.2 ± 9.4 | 8.6 ± 7.8 | 7.1 ± 6.4 | 0.30 |
| BMI, kg/m ² | 23.6±3.5 | 23.6 ± 2.7 | 25.7 ± 3.8 | 26.7 ± 4.7 | < 0.01 |
| SBP, mm Hg | 121.9 ± 13.0 | 122.5 ± 16.3 | 121.4 ± 16.8 | 122.2 ± 13.1 | 0.99 |
| DBP, mm Hg | 74.4 ± 8.3 | 75.6 ± 9.6 | 74.3 ± 10.0 | 75.2 ± 9.3 | 0.90 |
| FBG, mg/dL | 140.2 ± 54.0 | 147.4 ± 58.1 | 139.7 ± 57.5 | 162.9 ± 61.3 | 0.14 |
| HbA1c, % | 10.0 ± 2.7 | 9.9 ± 2.9 | 8.6 ± 1.9 | 8.5 ± 1.7 | < 0.01 |
| TC, mg/dL | 162.4 ± 44.2 | 185.1 ± 47.0 | 187.3 ± 49.4 | 190.3 ± 53.9 | 0.01 |
| HDL-C, mg/L | 42.2 ± 13.5 | 44.9 ± 10.3 | 42.1 ± 11.4 | 42.3 ± 14.2 | 0.69 |
| LDL-C, mg/dL | 102.5 ± 32.7 | 119.8 ± 34.1 | 118.1 ± 36.7 | 120.9 ± 37.2 | 0.02 |
| Triglyceride, mg/dL | 134.0 ± 61.4 | 158.8 ± 79.9 | 225.6 ± 178.4 | 237.5 ± 213.1 | < 0.01 |
| Homocystein, µmol/L | 11.4 ± 4.5 | 10.4 ± 3.2 | 13.3 ± 5.4 | 17.8 ± 8.5 | 0.06 |
| GFR, mL/min/1.73 m ² | 70.5 ± 23.6 | 74.6 ± 16.2 | 62.4 ± 23.7 | 71.0 ± 21.8 | 0.08 |
| Uric acid, mg/dL | 4.0 ± 1.3 | 4.2 ± 1.2 | 5.4 ± 1.9 | 5.3 ± 5.2 | 0.04 |
| Apo A, mg/dL | 126.1 ± 31.0 | 132.4 ± 23.7 | 129.4 ± 29.3 | 126.2 ± 33.7 | 0.72 |
| Apo B, mg/dL | 82.5 ± 22.8 | 92.9 ± 25.1 | 93.7 ± 30.5 | 98.6 ± 34.6 | 0.03 |
| HOMA- IR | 1.06 ± 0.43 | 1.19 ± 0.36 | 2.70 ± 0.96 | 2.77 ± 1.01 | < 0.01 |
| HOMA-β, % | 50.7 ± 37.9 | 51.8 ± 46.3 | 87.3 ± 45.9 | 78.6 ± 61.1 | 0.02 |

Values are presented as mean \pm standard deviation or number (%). The four groups were defined as follows: Group 1 (basal C-peptide <1.9 ng/mL, Δ C-peptide <1.1 ng/mL), Group 2 (basal C-peptide <1.9 ng/mL, Δ C-peptide ≥1.1 ng/mL), Group 3 (basal C-peptide ≥1.9 ng/mL, Δ C-peptide <1.1 ng/mL), and Group 4 (basal C-peptide ≥1.9 ng/mL, Δ C-peptide ≥1.1 ng/mL).

DM, diabetes mellitus; BMI, body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure; FBG, fasting blood glucose; HbA1c, hemoglobin A1c; TC, total cholesterol; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; GFR, glomerular filtration rate; Apo, apolipoprotein; HOMA-IR, homeostasis model assessment of insulin resistance; HOMA- β , homeostasis model assessment of β -cell function.

Table 2. Diabetic microvascular complications by C-peptide group

| Diabetic micro- | C-peptide group | | | | |
|----------------------------|-----------------|-----------|-----------|-----------|---------|
| vascular com- plication | 1 | 2 | 3 | 4 | P value |
| Retinopathy | 15 (26.3) | 9 (22.5) | 8 (19.5) | 5 (9.3) | 0.14 |
| NPDR/PDR | 1/4 | 8/1 | 7/1 | 5/0 | |
| Nephropathy | 20 (35.1) | 13 (32.5) | 14 (34.1) | 11 (20.4) | 0.32 |
| Micro/Overt | 12/8 | 11/2 | 8/6 | 7/4 | |
| Neuropathy | 30 (52.6) | 24 (60) | 20 (48.8) | 29 (53.7) | 0.79 |

Values are presented as number (%). The four groups were defined as follows: Group 1 (basal C-peptide <1.9 ng/mL, Δ C-peptide <1.1 ng/mL), Group 2 (basal C-peptide <1.9 ng/mL, Δ C-peptide \geq 1.1 ng/mL), Group 3 (basal C-peptide \geq 1.9 ng/mL, Δ C-peptide <1.1 ng/mL), and Group 4 (basal C-peptide \geq 1.9 ng/mL, Δ C-peptide \geq 1.1 ng/mL). NPDR/PDR, non-proliferative/proliferative retinopathy; Micro/Overt, micro albuminuria/overt albuminuria.

4, which had higher β -cell function, showed better controlled blood glucose levels (HbA1c).

Association between C-peptide levels according to patient grouping and microvascular complications

We investigated that the association between C-peptide levels according to patient grouping and microvascular complications (Table 2). Diabetic retinopathy was diagnosed in 26.3% of patients in Group 1, 22.5% in Group 2, 19.5% in Group 3, and 9.3% in Group 4. Diabetic retinopathy showed a higher prevalence in Group 1, but this difference was not statistically significant (P=0.14). The prevalence of diabetic nephropathy was higher in Group 1 than in Group 4, but this difference was not statistically significant, either (P=0.39). The incidence of diabetic neuropathy did not differ significantly between groups (P=0.79). Therefore, we concluded that there was no statisti-



Table 3. Diabetic microvascular complications by basal and stimulated C-peptide

| | Retinopathy | | Nephropathy | | | Neuropathy | | | |
|-----------------------------|---------------|---------------|-------------|---------------|---------------|------------|-------------|---------------|---------|
| | + | - | P value | + | - | P value | + | - | P value |
| Basal C-peptide, ng/mL | 1.9 ± 1.2 | 2.4 ± 1.3 | 0.03 | 1.6 ± 0.9 | 2.2 ± 1.2 | 0.02 | 2.1 ± 1.2 | 2.4 ± 1.5 | 0.12 |
| Stimulated C-peptide, ng/mL | 2.7 ± 1.6 | 3.7 ± 1.9 | < 0.01 | 2.8 ± 1.6 | 3.5 ± 1.8 | 0.03 | 3.3 ± 1.8 | 3.8 ± 2.0 | 0.04 |

cally significant association between microvascular complications and C-peptide groups (basal C-peptide, Δ C-peptide), which was likely because of the small-sized groups involved in our study.

Association between basal and stimulated C-peptide levels and microvascular complications

The association between basal and stimulated C-peptide levels and microvascular complications in our patient groups was evaluated (Table 3). In patients with diabetic retinopathy, the basal C-peptide level was 1.9 ± 1.2 ng/mL and the stimulated C-peptide level was 2.7 ± 1.6 ng/mL. The basal and stimulated C-peptide levels were significantly lower in patients with retinopathy than they were in those without diabetic retinopathy (basal C-peptide: $1.9 \pm 1.2 \text{ ng/mL}$ vs. $2.7 \pm 1.6 \text{ ng/mL}$, P = 0.031; stimulated C-peptide: 2.7 ± 1.6 ng/mL vs. 3.7 ± 1.9 ng/mL, P =0.002, respectively). In patients with diabetic nephropathy, the basal C-peptide level was 1.6±0.9 ng/mL and the stimulated C-peptide level was 2.8 ± 1.6 ng/mL. These values were significantly lower than those in patients without diabetic nephropathy (basal C-peptide: 1.6 ± 0.9 ng/mL vs. 2.2 ± 1.2 ng/mL, P=0.020; stimulated C-peptide: 2.8 ± 1.6 ng/mL vs. 3.5 ± 1.8 ng/mL, P=0.026, respectively). The basal C-peptide level in patients with diabetic neuropathy was lower than that in patients without diabetic neuropathy, but this difference was not statistically significant (2.1 \pm 1.2 ng/mL vs. 2.4 \pm 1.5 ng/mL, P=0.12). The stimulated C-peptide level was 3.3 ± 1.8 ng/mL in patients with diabetic neuropathy, which was significantly lower than that recorded in patients without diabetic neuropathy $(3.3 \pm 1.8 \text{ ng/}$ mL vs. 3.8 ± 2.0 ng/mL, P = 0.044).

Multiple DM risk factors and adjusted ORs for microvascular complications in stimulated and basal C-peptide levels

To evaluate the influence of other clinical factors associated with DM complications, multiple logistic regression analyses were performed. We calculated the DM complication risk factors-adjusted OR and the 95% confidence intervals (95% CI)

Table 4. Multiple risk factors-adjusted ORs and 95% CIs in the relationship between retinopathy and basal C-peptide level

| | OR (95% CI) | P value |
|------------------------|-------------------|---------|
| Age, yr | | |
| Group A (41-59) | 1.55 (0.42-5.65) | 0.51 |
| Group B (59-76) | 1.81 (0.51-6.39) | 0.36 |
| DM duration, yr | | |
| Group A (40-7) | 4.76 (1.98-11.42) | < 0.01 |
| Group B (7-0) | 1.72 (0.32-9.31) | 0.53 |
| Basal C-peptide, ng/mL | | |
| Group A (1.0) | 2.09 (0.80-5.53) | 0.13 |
| Group B (1.0-2.0) | 1.60 (0.38-2.97) | 0.91 |
| HbA1c, % | | |
| Group A (4.9-8.3) | 1.02 (0.27-3.84) | 0.98 |
| Group B (8.3-10.0) | 1.65 (0.45-6.09) | 0.46 |
| Group C (10.0-19.1) | 1.88 (0.46-7.66) | 0.38 |

Groups A, B, and C are composed of serial data (192 patients). OR, odds ratio; CI, confidence interval; DM, diabetes mellitus; HbA1c, hemoglobin A1c.

for the association of microvascular complications with basal and stimulated C-peptide levels (Tables 4-7). Age, DM duration, and HbA1c results were included in the risk factors. Multiple logistic regression analyses of the correlation between stimulated C-peptide and retinopathy prevalence showed that DM duration and stimulated C-peptide were correlated with retinopathy prevalence, which was statistically significant (4.42 [95% CI 1.83 to 10.69]), P<0.01; 1.225, P<0.01; 4.18, P< 0.01; 3.35, P=0.04, respectively). In neuropathy, however, basal and stimulated C-peptide levels did not have statistically significant correlations with neuropathy prevalence when it was adjusted for multiple risk factors (Table 7). The basal Cpeptide-adjusted OR was not statistically significant in retinopathy and neuropathy (Tables 4, 6). We investigated the correlation between nephropathy and DM complication risk factors (such as age, DM duration, stimulated C-peptide, HbA1c, systolic BP, and fasting blood sugar) (Table 8). Multiple regression analysis showed that stimulated C-peptide was correlated



Table 5. Multiple risk factors-adjusted ORs and 95% CIs in the relationship between retinopathy and stimulated C-peptide level

| | OR (95% CI) | P value |
|-----------------------------|-------------------|---------|
| Age, yr | | |
| Group A (41-59) | 1.28 (0.35-4.70) | 0.71 |
| Group B (59-76) | 1.33 (0.37-4.71) | 0.66 |
| DM duration, yr | | |
| Group A (40-7) | 4.42 (1.83-10.69) | < 0.01 |
| Group B (7-0) | 1.23 (0.23-6.57) | 0.81 |
| Stimulated C-peptide, ng/mL | | |
| Group A (1.0) | 4.18 (1.40-12.51) | 0.01 |
| Group B (1.0-2.0) | 3.35 (1.09-10.25) | 0.04 |

Groups A and B are composed of serial data.

OR, odds ratio; CI, confidence interval; DM, diabetes mellitus.

Table 6. Multiple risk factors-adjusted ORs and 95% CIs in the relationship between neuropathy and basal C-peptide level

| | OR (95% CI) | P value |
|------------------------|------------------|---------|
| Age, yr | | |
| Group A (41-59) | 2.08 (0.78-0.53) | 0.15 |
| Group B (59-76) | 1.63 (0.63-4.24) | 0.31 |
| DM duration, yr | | |
| Group A (40-7) | 1.72 (0.78-3.83) | 0.18 |
| Group B (7-0) | 0.80 (0.19-3.43) | 0.76 |
| Basal C-peptide, ng/mL | | |
| Group A (1.0) | 1.25 (0.60-2.65) | 0.56 |
| Group B (1.0-2.0) | 0.73 (0.35-1.55) | 0.42 |
| HbA1c, % | | |
| Group A (4.9-8.3) | 1.77 (0.67-4.71) | 0.25 |
| Group B (8.3-10.0) | 1.62 (0.60-4.38) | 0.35 |
| Group C (10.0-19.1) | 0.10 (0.32-3.06) | 0.10 |

Groups A and B are composed of serial data.

OR, odds ratio; CI, confidence interval; DM, diabetes mellitus; HbA1c, hemoglobin A1c.

with nephropathy prevalence, which was statistically significant (stimulated C-peptide: -2.17, P=0.03) (Table 6).

DISCUSSION

Type 2 DM is characterized by insulin resistance and decreased insulin secretion [11]. Type 2 DM is a progressive disease that is affected by a multi-factorial physiology. In this disorder the

Table 7. Multiple risk factors-adjusted ORs and 95% CIs in the relationship between neuropathy and stimulated C-peptide level

| OR (95% CI) | P value |
|------------------|--|
| | |
| 1.94 (0.74-5.14) | 0.18 |
| 1.56 (0.60-4.04) | 0.36 |
| | |
| 1.73 (0.77-3.85) | 0.18 |
| 0.72 (0.17-3.06) | 0.66 |
| | |
| 1.26 (0.50-3.20) | 0.62 |
| 0.97 (0.48-1.97) | 0.94 |
| | |
| 1.71 (0.65-4.51) | 0.28 |
| 1.58 (0.58-4.31) | 0.37 |
| 0.96 (0.30-3.04) | 0.94 |
| | 1.94 (0.74-5.14) 1.56 (0.60-4.04) 1.73 (0.77-3.85) 0.72 (0.17-3.06) 1.26 (0.50-3.20) 0.97 (0.48-1.97) 1.71 (0.65-4.51) 1.58 (0.58-4.31) |

Groups A and B are composed of serial data.

OR, odds ratio; CI, confidence interval; DM, diabetes mellitus; HbA1c, hemoglobin A1c.

Table 8. Correlation between nephropathy and multiple factors

| | γ^2 | P value |
|-----------------------------|------------|---------|
| Stimulated C-peptide, ng/dL | -2.10 | 0.03 |
| Age, yr | 0.68 | 0.47 |
| DM duration, yr | -0.29 | 0.75 |
| HbA1c, % | -0.64 | 0.54 |
| SBP, mm Hg | 0.15 | 0.09 |
| FBS, mg/dL | 0.11 | 0.30 |

DM, diabetes mellitus; HbA1c, hemoglobin A1c; SBP, systolic blood pressure; FBS, fasting blood sugar.

natural history of β -cell functions has not been clearly identified. Therefore, it is necessary to evaluate the degree to which insulin secretion affects diabetic complications and the clinical course of type 2 DM.

In this cross-sectional study, we evaluated how β -cell functions are associated with clinical manifestations and microvascular complications in patients with type 2 DM. We found that basal C-peptide and secreted C-peptide (Δ C-peptide) levels were associated with clinical characteristics and microvascular complications. HbA1c was significantly higher in Group 1 (low basal C-peptide, low Δ C-peptide) than in Group 4 (high basal



C-peptide, high Δ C-peptide). However, HOMA-IR and HOMA- β scores were significantly higher in Group 4 than in Group1. We can speculate that poor β -cell function is likely related to uncontrolled blood glucose. BMI was significantly higher in Group 4 (high basal C-peptide, high Δ C-peptide) than in Group 1 (low basal C-peptide, low Δ C-peptide). BMI, total cholesterol, LDL-C, triglycerides, uric acid, and Apo B were significantly higher in Group 4 than in Group 1.

The correlation between C-peptide levels and components of the metabolic syndrome is well known [12]. We found that C-peptide levels was associated with dyslipidemia and BMI. When basal C-peptide and secreted C-peptide (ΔC-peptide) level were high, there was an increased likelihood of progression to metabolic syndrome. Kim et al. [13] showed that basal C-peptide level increased significantly in metabolic syndrome patients with diabetes. Therefore, people with a high basal C-peptide level need to decrease it to reduce the risk of coronary heart disease and cerebral vascular disease associated with metabolic syndrome.

The prevalence of diabetic microvascular complications was not significantly different between the groups (basal C-peptide, ΔC-peptide) in this study. The prevalence of diabetic retinopathy was higher in Group 1 than in Group 4. This finding suggests that the higher ΔC -peptide group had a lower prevalence of retinopathy. Kim et al. [13] conducted a larger study than ours, and they reported that ΔC -peptide is more closely associated with microvascular complications than are fasting Cpeptide levels. In their study the prevalence of diabetic retinopathy and nephropathy was higher in Group 1 than in Group 4. This finding suggests that low basal and secreted Cpeptide (ΔC-peptide) levels are associated with a high prevalence of diabetic retinopathy and nephropathy. In this study, however, this observation was not statistically significant. This study had a relatively small number of patients with chronic complications. With a larger sample size, we might have observed a significant association between microvascular complications and C-peptide groups (basal C-peptide, Δ C-peptide), and we could have been able to better clarify the correlation between β-cell function and microvascular complications.

C-peptide and insulin are secreted in equimolar amounts from the β -cells of the pancreas. Secreted insulin passes through the liver into the bloodstream, where about 50% of the insulin is extracted. C-peptide and insulin have different hepatic clearances. Therefore, serum insulin concentration is indicative of post-hepatic insulin levels rather than the value of se-

creted insulin [8]. Accurate estimation of β -cell function in insulin-treated patients is not possible, as measured insulin level cannot be separated into secreted and exogenous insulin levels. As C-peptide has a longer half-life than insulin, C-peptide more accurately reflects β -cell function than serum insulin in insulin-treated patients, and C-peptide levels are less affected by the presence of insulin-binding antibodies.

Stimulated C-peptide reactivity is correlated with β-cell function in patients with type 2 DM [14,15]. Furthermore, the lower degree of variation in the stimulated C-peptide level makes it more preferable than the fasting C-peptide level. Three methods for stimulating C-peptide response have been used in clinical trials. The first method is MMTT, in which a liquid meal is ingested in the fasting state, and C-peptide level is measured over the subsequent 2 to 4 hours. The second method is GST, in which glucagon is injected intravenously and Cpeptide level is measured over the following 6 minutes [10]. In patients with type 2 DM, GST is relatively simple to perform and has a shorter duration than other tests. Adverse effect associated with GST include mild nausea and rarely, vomiting. The third method is the AST, which has a short duration, but its reproducibility has not been assessed. Jones et al. [16] reported that the urine C-peptide creatinine ratio was positively correlated with serum C-peptide and may provide a practical alternative measure. Although they are positively correlated, it is likely that the urine C-peptide creatinine ratio will be less reproducible after individuals consume their own meals at home than are measurements taken after a fixed stimulus. Kim et al. [13] studied these levels using the MMTT. However, GST is more convenient and cost-effective than MMTT and therefore has been regarded as the most suitable test [17]. Therefore, we used GST to obtain stimulated C-peptide levels in this study. This method was the biggest difference between the two studies.

Many studies have evaluated the association between C-peptide level and diabetic complications in patients with type 2 DM. In the DCCT trial, type 1 diabetic patients with C-peptide secretory capacities showed a significant prevalence of microvascular complications than did patients who were deficient in β -cell C-peptide secretion [7]. Low C-peptide levels were associated with the progression of diabetic microangiopathy, such as retinopathy and nephropathy [18]. Shin et al. [19] showed that a lower C-peptide response to oral glucose stimuli was associated with albuminuria. In addition, C-peptide has been shown to be associated with autonomic nerve function in type 2 DM [20]. In another study, low C-peptide immuno-



reactivity was revealed to be associated with diabetic microangiopathy [21]. Recently, Kim et al. [13] found that Δ C-peptide was more closely associated with microvascular complications than was the fasting C-peptide level. In this previous study, stimulated C-peptide was used as an alternative marker of β -cell function, and the association of stimulated C-peptide with microvascular complications was investigated.

However, no association was found between C-peptide level and neuropathy, nephropathy or retinopathy [5]. Klein et al. [22] could not identify any relationship between C-peptide level and retinopathy prevalence, either; however, the serum glucose level was associated with retinopathy. Another study showed no relationship between C-peptide and microangiopathy [23].

In this study, we found that the basal C-peptide level was significantly associated with diabetic retinopathy and nephropathy, but not with neuropathy. The stimulated C-peptide level was associated with all three types of microvascular complications. To adjust for the confounding factor, we performed multiple regression analyses. In retinopathy, DM duration and the stimulated C-peptide level did result in significant attenuation of the relationships. Furthermore, the stimulated C-peptide had statistical significance in the multiple regression analysis with nephropathy and DM complication risk factors. However upon sub-analysis we did not obtain a statistically significant result with neuropathy and multiple DM complication risk factors such as age, DM duration and HbA1c. C-peptide has a dose-dependent effect on Na⁺/K⁺-adenosine triphosphatase activity, endothelial nitrous oxide synthase and endoneurial blood flow [24,25]. Therefore, it seems that diabetic neuropathy was more affected by stimulated C-peptide than by basal C-peptide. If the present study had included a larger sample size, we might have observed a significant association between nephropathy and the stimulated C-peptide level. We showed that a reduction in β -cell function was associated with retinopathy and neuropathy. The purpose of our study was to prove that stimulated C-peptide response to glucagon can be used to assess the β -cell function of the pancreas and predict the prevalence of diabetic microvascular complications. This ability is essential for prevention because early evaluation and therapeutic interventions may help preserve β -cell function in affected patients and could slow the progression of diabetic microvascular complications.

Our study had some limitations. The number of participants was too small to accurately validate the association between

stimulated C-peptide levels and microvascular complications. We also used a cross-sectional study design, which is not suitable for assessing the temporal sequence of the assessed outcome. The stimulated C-peptide response to glucagon has also been shown to be affected by age, glucose level and weight. Furthermore, we excluded patients with GFR <60 mL/min/ 1.73 m², who were associated with diabetic nephropathy. We also excluded patients who required exogenous insulin and thiazolidinedione treatment, but we included sulfonylurea users in our data. However, if the GST were performed after an overnight fast, the effect of sulfonylurea would be minimized. We did not evaluate the association of stimulated C-peptide with the severity of diabetic microvascular complications. We created four groups based upon the mean value of basal Cpeptide and ΔC-peptide because C-peptide is a continuous variable.

In conclusion, in patients with type 2 DM, GST was a relatively simple method with a short duration and few side effects. The stimulated C-peptide values obtained through GST were associated with microvascular complications to a greater extent than the basal C-peptide level in patients with type 2 DM. Prospective and larger studies will be needed to clarify the association between stimulated C-peptide levels and microvascular complications, which may be used to reduce the prevalence of microvascular complications in patients with type 2 DM.

CONFLICTS OF INTEREST

This work did not receive any financial support nor is it affiliated with any organizations. No potential conflict of interest relevant to this article was reported.

REFERENCES

- Faber OK, Hagen C, Binder C, Markussen J, Naithani VK, Blix PM, Kuzuya H, Horwitz DL, Rubenstein AH, Rossing N. Kinetics of human connecting peptide in normal and diabetic subjects. J Clin Invest 1978;62:197-203.
- 2. Kuhl C, Faber OK, Hornnes P, Jensen SL. C-peptide metabolism and the liver. Diabetes 1978;27 Suppl 1:197-200.
- Marx N, Walcher D. C-Peptide and atherogenesis: C-Peptide as a mediator of lesion development in patients with type 2 diabetes mellitus? Exp Diabetes Res 2008;2008:385108.
- 4. Forst T, Kunt T, Wilhelm B, Weber MM, Pfutzner A. Role of



- C-Peptide in the regulation of microvascular blood flow. Exp Diabetes Res 2008;2008:176245.
- Sari R, Balci MK. Relationship between C peptide and chronic complications in type-2 diabetes mellitus. J Natl Med Assoc 2005;97:1113-8.
- Chowta MN, Adhikari PM, Chowta NK, Shenoy AK, D'Souza S. Serum C peptide level and renal function in diabetes mellitus. Indian J Nephrol 2010;20:25-8.
- Steffes MW, Sibley S, Jackson M, Thomas W. Beta-cell function and the development of diabetes-related complications in the diabetes control and complications trial. Diabetes Care 2003; 26:832-6.
- Palmer JP, Fleming GA, Greenbaum CJ, Herold KC, Jansa LD, Kolb H, Lachin JM, Polonsky KS, Pozzilli P, Skyler JS, Steffes MW. C-peptide is the appropriate outcome measure for type 1 diabetes clinical trials to preserve beta-cell function: report of an ADA workshop, 21-22 October 2001. Diabetes 2004;53: 250-64.
- Scheen AJ, Castillo MJ, Lefebvre PJ. Assessment of residual insulin secretion in diabetic patients using the intravenous glucagon stimulatory test: methodological aspects and clinical applications. Diabetes Metab 1996;22:397-406.
- 10. Greenbaum CJ, Mandrup-Poulsen T, McGee PF, Battelino T, Haastert B, Ludvigsson J, Pozzilli P, Lachin JM, Kolb H; Type 1 Diabetes Trial Net Research Group; European C-Peptide Trial Study Group. Mixed-meal tolerance test versus glucagon stimulation test for the assessment of beta-cell function in therapeutic trials in type 1 diabetes. Diabetes Care 2008;31:1966-71.
- 11. Abdullah BB, Patil BS, Thaseen A. Significance of C-peptide in type 2 diabetics: a study in the North Karnataka population of India. Al Ameen J Med Sci 2010;3:65-78.
- Koyama K, Chen G, Lee Y, Unger RH. Tissue triglycerides, insulin resistance, and insulin production: implications for hyperinsulinemia of obesity. Am J Physiol 1997;273(4 Pt 1): E708-13.
- Kim BY, Jung CH, Mok JO, Kang SK, Kim CH. Association between serum C-peptide levels and chronic microvascular complications in Korean type 2 diabetic patients. Acta Diabetol 2012;49:9-15.
- 14. Garcia-Webb P, Bonser A, Welborn TA. Correlation between fasting serum C-peptide and B cell insulin secretory capacity in diabetes mellitus. Diabetologia 1982;22:296.
- 15. Gottsater A, Landin-Olsson M, Fernlund P, Gullberg B, Lernmark A, Sundkvist G. Pancreatic beta-cell function evaluated

- by intravenous glucose and glucagon stimulation: a comparison between insulin and C-peptide to measure insulin secretion. Scand J Clin Lab Invest 1992;52:631-9.
- 16. Jones AG, Besser RE, McDonald TJ, Shields BM, Hope SV, Bowman P, Oram RA, Knight BA, Hattersley AT. Urine C-peptide creatinine ratio is an alternative to stimulated serum Cpeptide measurement in late-onset, insulin-treated diabetes. Diabet Med 2011;28:1034-8.
- 17. Faber OK, Binder C. C-peptide response to glucagon: a test for the residual beta-cell function in diabetes mellitus. Diabetes 1977;26:605-10.
- 18. Inukai T, Matsutomo R, Tayama K, Aso Y, Takemura Y. Relation between the serum level of C-peptide and risk factors for coronary heart disease and diabetic microangiopathy in patients with type-2 diabetes mellitus. Exp Clin Endocrinol Diabetes 1999;107:40-5.
- Shin SJ, Lee YJ, Hsaio PJ, Chen JH, Guh JY, Chen MT, Chen WC, Tsai JH. Relationships between beta-cell function and diabetic duration and albuminuria in type 2 diabetes mellitus. Pancreas 1997;14:192-8.
- 20. Toyry JP, Niskanen LK, Mantysaari MJ, Lansimies EA, Haffner SM, Miettinen HJ, Uusitupa MI. Do high proinsulin and C-peptide levels play a role in autonomic nervous dysfunction? Power spectral analysis in patients with non-insulin-dependent diabetes and nondiabetic subjects. Circulation 1997;96: 1185-91.
- Bo S, Cavallo-Perin P, Gentile L, Repetti E, Pagano G. Relationship of residual beta-cell function, metabolic control and chronic complications in type 2 diabetes mellitus. Acta Diabetol 2000;37:125-9.
- 22. Klein R, Klein BE, Moss SE. The Wisconsin Epidemiologic Study of Diabetic Retinopathy. XVI. The relationship of Cpeptide to the incidence and progression of diabetic retinopathy. Diabetes 1995;44:796-801.
- 23. Sjoberg S, Gunnarsson R, Gjotterberg M, Lefvert AK, Persson A, Ostman J. Residual insulin production, glycaemic control and prevalence of microvascular lesions and polyneuropathy in long-term type 1 (insulin-dependent) diabetes mellitus. Diabetologia 1987;30:208-13.
- 24. Sima AA, Zhang W, Grunberger G. Type 1 diabetic neuropathy and C-peptide. Exp Diabesity Res 2004;5:65-77.
- 25. Kamiya H, Zhang W, Sima AA. The beneficial effects of C-Peptide on diabetic polyneuropathy. Rev Diabet Stud 2009;6:187-202.