REPORT

Genetic Variants at 6p21.1 and 7p15.3 Are Associated with Risk of Multiple Cancers in Han Chinese

Guangfu Jin,^{1,2,16} Hongxia Ma,^{1,16} Chen Wu,^{5,16} Juncheng Dai,¹ Ruyang Zhang,¹ Yongyong Shi,⁶ Jiachun Lu,⁷ Xiaoping Miao,⁸ Meilin Wang,³ Yifeng Zhou,⁹ Jiaping Chen,^{1,2} Huizhang Li,¹ Shandong Pan,¹ Minjie Chu,¹ Feng Lu,¹ Dianke Yu,⁵ Yue Jiang,¹ Jing Dong,¹ Lingmin Hu,¹ Yijiang Chen,10 Lin Xu,13 Yongqian Shu,11 Shiyang Pan,12 Wen Tan,5 Baosen Zhou,14 Daru Lu,15 Tangchun Wu,⁸ Zhengdong Zhang,³ Feng Chen,¹ Xinru Wang,^{1,4} Zhibin Hu,^{1,2,4,*} Dongxin Lin,^{5,*} and Hongbing Shen1,2,4

Cancer susceptibility loci identified in reported genome-wide association studies (GWAS) are often tumor-specific; however, evidence of pleiotropy of some genes/loci has also been observed and biologically plausible. We hypothesized that there are important regions in the genome harboring genetic variants associated with risk of multiple types of cancer. In the current study, we attempted to map genetic variants that have consistent effects on risk of multiple cancers using our existing genome-wide scan data of lung cancer, noncardia gastric cancer, and esophageal squamous-cell carcinoma with overall 5,368 cases and 4,006 controls (GWAS stage), followed by a further evaluation in additional 9,001 cases with one of these cancer types and 11,436 controls (replication stage). Five variants satisfying the criteria of pleiotropy with p values from 1.10×10^{-8} to 8.96 \times 10⁻⁶ for genome-wide scans of three cancer types were further evaluated in the replication stage. We found consistent associations of rs2494938 at 6p21.1 and rs2285947 at 7p15.3 with these three cancers in both GWAS and replication stages. In combined samples of GWAS and replication stages, the minor alleles of rs2494938 and rs2285947 were significantly associated with an increased risk of the cancers (odds ratio $[OR] = 1.15$, 95% confidence interval [CI], 1.10–1.19 and OR = 1.17, 95% CI, 1.12–1.21), with the p values being 1.20 \times 10⁻¹² and 1.26 \times 10⁻¹⁶, respectively, which are at a genome-wide significance level. Our findings highlight the potential importance of variants at 6p21.1 and 7p15.3 in the susceptibility to multiple cancers.

Genome-wide association studies (GWAS) have broadened our understanding of genetic variations that confer risk for different types of cancers.^{[1](#page-4-0)} Notably, most of risk-related loci are tumor-specific. However, pleiotropy has been observed for several loci, such as the regions of 8q24, 5p15.33 (TERT [MIM 187270]-CLPTM1L [MIM 612585]), and 9p21.3 (ANRIL [MIM 613149]). Among several loci at 8q24 associated with prostate cancer risk, $2-8$ at least two have also been associated with risk of colorectal cancer, $9-11$ ovarian cancer, 12 or breast cancer.^{[13](#page-5-0)} Preliminary results from functional studies have linked a cancerrelated variant (rs6983267) at 8q24 to an enhancer that may interact with the known oncogene MYC [MIM 190080].^{[14,15](#page-5-0)} The TERT-CLPTM1L locus has initially been implicated in lung cancer risk.^{16,17} In a subsequent study on 17 cancer types, the variant rs401681 allele at the TERT-CLPTM1L locus has been associated with increased

risk of basal cell, lung, urinary bladder, prostate, and cervical cancers, whereas it conferred protection against cutaneous melanoma.[18](#page-5-0) Furthermore, this locus is also associated with risk of cancers of glioma, 19 pancreas, 20 and breast.^{[21](#page-5-0)} The ANRIL locus at 9p21.3 is associated with risk of coronary artery disease,^{[22](#page-5-0)} myocardial infarction,^{[23](#page-5-0)} type 2 diabetes, 24 24 24 and multiple cancers (glioma, 19,25 19,25 19,25 basal cell carcinoma, 26 26 26 melanoma, 27 breast, 28 28 28 and nasopharyngeal carcinoma²⁹). Importantly, mouse models have revealed a pivotal role of Anril in regulation of CDKN2A/B through and in proliferation and senescence. 30 Thus, evidence of pleiotropy might highlight some common etiological pathways in the development of multiple human cancers.

In light of above evidence, we hypothesized that there are other important genomic regions harboring variants that are associated with risk of multiple types of human

¹Department of Epidemiology and Biostatistics and Ministry of Education Key Laboratory for Modern Toxicology, School of Public Health, ²Section of Clinical Epidemiology, Jiangsu Key Lab of Cancer Biomarkers, Prevention and Treatment, Cancer Center, ³Department of Occupational Medicine and Environmental Health, School of Public Health, ⁴State Key Laboratory of Reproductive Medicine, Nanjing Medical University, Nanjing 210029, China; 5 State Key Laboratory of Molecular Oncology and Department of Etiology and Carcinogenesis, Cancer Institute and Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100021, China; ⁶Bio-X Institutes, Key Laboratory for the Genetics of Developmental and Neuropsychiatric Disorders (Ministry of Education), Shanghai Jiao Tong University, Shanghai 200240, China; ⁷The Institute for Chemical Carcinogenesis, State Key Laboratory of Respiratory Disease, Guangzhou Medical College, Guangzhou 510182, China; ⁸Department of Epidemiology and Biostatistics and Ministry of Education Key Lab for Environment and Health, School of Public Health, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, China; ⁹Cyrus Tang Hematology Center, Jiangsu Institute of Hematology, Medical College, Soochow University, Suzhou 215123, China;
¹⁰Departments of Thoracic Surgery, ¹¹Department of Oncology, Jiangsu Key Hospital of Nanjing Medical University, Nanjing 210029, China; 13Department of Thoracic Surgery, Affiliated Cancer Hospital of Nanjing Medical University, Jiangsu Cancer Hospital, Nanjing 210009, China; 14Department of Epidemiology, School of Public Health, China Medical University, Shenyang 110001, China; 15State Key Laboratory of Genetic Engineering, Center for Fudan-VARI Genetic Epidemiology and MOE Key Laboratory of Contemporary Anthropology, School of Life Sciences, Fudan University, Shanghai 200433, China

¹⁶These authors contributed equally to this work

[http://dx.doi.org/10.1016/j.ajhg.2012.09.009.](http://dx.doi.org/10.1016/j.ajhg.2012.09.009) 2012 by The American Society of Human Genetics. All rights reserved.

^{*}Correspondence: zhibin_hu@njmu.edu.cn (Z.H.), lindx72@cicams.ac.cn (D.L.)

cancer. To test this hypothesis, we utilized the existing GWAS data on lung cancer,³¹ noncardia gastric cancer $(NCGC),^{32}$ $(NCGC),^{32}$ $(NCGC),^{32}$ and esophageal squamous-cell carcinoma $(ESCC)^{33}$ $(ESCC)^{33}$ $(ESCC)^{33}$ that have been associated with a common risk factor of cigarette smoking $34-36$ and may have shared susceptibility loci, to map pleiotropic loci contributing to these three types of cancer in Han Chinese populations, followed by further replication in an additionally large case-control set.

Subjects included in the current study and the criteria for their recruitment have been described previously. $31-33$ Briefly, for the GWAS stage, 1,473 cases with lung cancer, 550 cases with NCGC, and their shared 1,962 controls were recruited from central region of China (Jiangsu Province and surrounding areas); 858 cases with lung cancer, 456 cases with NCGC, 2,031 cases with ESCC, and their shared 2,044 controls were recruited from northern region of China (Beijing and surrounding areas). In the replication stage, 1,894 cases with NCGC, 3,006 cases with ESCC, and 6,525 controls were recruited from Jiangsu Province and surrounding areas; 1,534 cases with lung cancer, 1,436 cases with NCGC, and 2,922 controls were recruited from Beijing and surrounding areas. An additional set of 1,131 cases with lung cancer and 1,989 controls was recruited from southern region of China (Guangdong Province). Overall, 5,368 cancer cases and 4,006 controls were included in the GWAS stage, and 9,001 cancer cases (2,665 lung cancer, 3,330 NCGC, and 3,006 ESCC) and 11,436 controls were included in the replication stage. All of these subjects provided informed consent. This study was approved by the Institutional Review Boards of all participating institutions.

All samples in the GWAS stage were scanned using Affymetrix Genome-Wide Human SNP Array 6.0 chips (Affymetrix), as described previously.³¹⁻³³ Standard quality control procedures on the raw genotyping data were used to filter unqualified samples and SNPs. Individuals were removed from subsequent analyses if they satisfy any of the following items: (1) low call rate (overall rat $<$ 95%), (2) ambiguous gender, (3) duplicates or familial relationship (PL_HAT > 0.025), (4) extreme heterozygosity rate ($>$ mean $+$ 6 SD). SNPs were excluded if they (1) were not mapped to autosome chromosomes, (2) had a call rate $<$ 95%, (3) had minor allele frequency (MAF) $<$ 0.05 in controls, (4) had genotype distributions that deviated from those expected as Hardy-Weinberg equilibrium in controls ($p < 1 \times 10^{-5}$), or (5) failed in any of quality control for GWAS of lung cancer, noncardia gastric cancer, and esophageal squamous-cell carcinoma. As a result, 2,331 cases with lung cancer, 1,006 cases with NCGC, 2,031 cases with ESCC and 4,006 controls retained for the association analyses in the GWAS stage.

Association analyses in the GWAS stage were first performed independently in lung cancer, NCGC and ESCC, and in the cohorts from different areas (Central and Northern). The results from different cohorts for NCGC or ESCC were combined by meta-analysis. Pleiotropic loci

were then defined and selected for further confirmation in replication stage according to the following criteria: (1) SNPs had consistent associations ($p < 0.05$) between Central and Northern cohorts for each cancer, (2) consistent associations were shown for all three types of cancer, and (3) $p \leq 1.0 \times 10^{-5}$ for the combined results of three types of cancer in the GWAS stage. As a result, five SNPs were selected for genotyping (see [Table S1](#page-4-0) available online) in the replication stage using TaqMan assays (Applied Biosystems, Foster City, CA). Sample status (case or control) was blinded to laboratory technicians who performed genotyping. Ten percent of random samples were repeated for rs2494938 and rs2285947, and the overall accordance rates were 99.8% and 99.7%, respectively.

We performed logistic regression analyses independently based on additive model with adjustment for first principal component, age, sex, and smoking status on each GWAS. For NCGC and lung cancer studies in the GWAS stage, we first treated the samples from two cohorts as independent studies (Central and Northern), and then combined the results for each cancer by meta-analysis. Because the shared controls were used for Central or Northern studies in GWAS stage, the combined association results of the three cancers for filtered SNPs with consistent association results were analyzed by pooling all cancer cases together as compared with the shared controls. The same analysis approach was used in the replication stage. The results from GWAS and replication stages or from different regions (Central, Northern, and Southern) were combined by either pooling samples together or using meta-analysis. Similar results were observed for these two approaches and the former one was reported as default in this study unless specified. Meta-analysis was performed in a default fixed-effect model with inverse variance weighted and a calculation of Cochran's Q statistics for homogeneity test. A random-effects model (DerSimonian-Laird) was adopted and the corresponding results were reported if there was indication of heterogeneity between groups ($P_Q \leq 0.05$). Otherwise, the fixed effect model was maintained ($P_O > 0.05$). PLINK 1.07 was used for genetic association analysis. 37 Analyses were also performed using SAS version 9.1.3 (SAS Institute, Cary, NC) or Stata version 9.2 (StataCorp LP, TX).

The characteristics of 5,368 cases and 4,006 controls included in the GWAS of three types of cancer are shown in [Table 1.](#page-2-0) We first performed logistic regression analyses based on additive model with adjustment for the first principal component, age, sex, and smoking status on each GWAS independently. For lung cancer and NCGC studies, we treated the samples from two cohorts as independent studies (Central study and Northern study), and then estimated combined effect of each locus by meta-analysis.^{[31,33](#page-5-0)} Pleiotropic loci were defined as having consistent effects among three types of cancer and between Central and Northern studies in GWAS. As a result, 5 SNPs (rs2399395 at 3q13.2, rs2494938 at 6p21.1, rs2285947 at 7p15.3, rs13232645 at 7q22.1, and

Studies	Data Set	Region	No.	Age (Mean \pm SD)	Sex (males, %)	Smoking Status (smokers, %)
Total cases			14,369	59.53 ± 10.68	72.19	50.09
Lung cancer	GWAS	Central	1,473	60.08 ± 10.30	71.76	44.81
	GWAS	Northern	858	60.00 ± 10.23	76.22	59.91
	Replication	Northern	1,534	58.14 ± 9.71	67.08	59.97
	Replication	Southern	1,131	60.27 ± 12.32	70.73	55.88
NCGC	GWAS	Central	550	58.24 ± 11.91	71.27	44.91
	GWAS	Northern	456	56.79 ± 12.42	70.61	28.29
	Replication	Central	1,894	58.71 ± 11.97	70.49	46.73
	Replication	Northern	1,436	57.06 ± 12.11	69.29	25.49
ESCC	GWAS	Northern	2,031	59.84 ± 9.78	80.11	65.24
	Replication	Central	3,006	61.70 ± 8.54	71.92	50.57
Total controls			15,442	59.00 ± 11.73	70.24	43.70
	GWAS	Central	1,962	59.35 ± 9.74	61.88	53.82
	GWAS	Northern	2,044	61.34 ± 8.55	83.46	38.38
	Replication	Central	6,525	58.32 ± 11.92	68.29	28.64
	Replication	Northern	2,922	58.24 ± 12.86	70.12	68.62
	Replication	Southern	1,989	59.59 ± 13.49	71.49	51.33

Table 1. Characteristics of Cases with Lung Cancer, Noncardia Gastric Cancer, or Esophageal Squamous-Cell Carcinoma and Controls ncluded in GWAS and Replication Stage

rs9982863 at 21q22.3) were identified ([Table S1\)](#page-4-0). Because similar frequency and effect on the three types of cancer were observed across studies each of these 5 SNPs, the subjects from different studies were combined directly to test the associations. We observed that the combined p values were 1.10 \times 10⁻⁸ to 8.96 \times 10⁻⁶ for these five SNPs ([Table S1\)](#page-4-0).

We next performed replications of above five promising SNPs in overall 9,001 cases with three types of cancer (2,665 lung cancer, 3,330 NCGC, and 3,006 ESCC) and 11,436 controls. We found that rs2399395 at 3q13.2 and rs13232645 at 7q22.1 were replicated only in ESCC and lung cancer (nominal $p < 0.05$), respectively, whereas rs9982863 at 21q22.3 was not significantly associated with any type of cancer [\(Table S2\)](#page-4-0). Thus, these three SNPs were not further investigated. The remaining two SNPs, rs2494938 at 6p21.1 and rs2285947 at 7p15.3, were both associated with all three types of cancer. In combined sample, the odds ratios (ORs) for rs2494938 and rs2285947 were 1.15 (95% confidence interval [CI], 1.10– 1.19; $p = 1.20 \times 10^{-12}$ and 1.17 (95% CI, 1.12–1.21; $p = 1.26 \times 10^{-16}$, respectively ([Table 2\)](#page-3-0). For both rs2494938 and rs2285947, no significant heterogeneity of associations was observed between the GWAS and replication studies or among the Northern, Central, and Southern cohorts (all $p_O > 0.05$) [\(Figure S1\)](#page-4-0).

To further characterize the loci at 6p21.1 and 7p15.3 associated with risk of multiple cancers, we imputed untyped SNPs in the flanking regions of rs2494938 and

rs2285947 within a 250 kb window for subjects in the GWAS stage using the CHB+JPT data from the 1000 Genomes database (released at June 2010) as reference haplotype using Minimac software. Regional plots were generated by three cancer types according to p value of association for each SNP using LocusZoom.^{[38](#page-5-0)} As shown in [Figure S2](#page-4-0), SNPs with lower p values at both regions were observed for each cancer. After conditioning on lead SNP (rs2494938 or rs2285947) for respective region, none of the associations were found with a $p < 0.001$ for the remaining SNPs at 6p21.1 and 7p15.3 except for association of variants at 7p15.3 with ESCC risk, suggesting that no additional independent loci exist at these two regions for lung cancer and NCGC. However, further studies are warranted to assess the independence of associations of variants at 7p15.3 with ESCC risk.

With an effort to show potential biofeatures of association, the locus at 6p21.1 was annotated based on UCSC genome browser ([Figure S3](#page-4-0)). SNPs in strong LD with rs2494938 at 6p21.1 are located in the initial region (including promoter, exon 1, and intron 1) of LRFN2 [MIM 612808] (encoding leucine-rich repeat and fibronectin type III domain-containing protein 2), a member of synaptic adhesion-like molecules (SALMs) family that interacts with the N-methyl D-aspartate (NMDA) receptor.³⁹ It has been reported that the LRFN2 can subvert hematopoietic differentiation to increase erythropoiesis and lead to erythroblastosis in cooperation with MYC.^{[40](#page-6-0)} NMDA receptors are glutamate receptors, consisting of

^aMajor/minor alleles.
^bMAF: minor allele frequency.

c Odds ratios (ORs), 95% confidence intervals (CIs), and p values were calculated in additive models with adjustment for age, sex, and smoking status.

NR1 and NR2 (2A to 2D) subunits.^{[41,42](#page-6-0)} NMDA receptor 2B (NR2B) is methylated in ESCC and non-small cell lung cancer and exhibit a tumor-suppressive activity. $43,44$ Interestingly, it is recently revealed that NMDA channels may play an important proapoptotic function in gastric surface epithelial cells and regulate cell survival and death pathways during development of gastric cancers associated with H . pylori infection.^{[45](#page-6-0)} However, the exact mechanism of the locus at 6p21.1 is largely unknown though LRFN2 might function as a susceptibility gene for multiple cancers through LRFN2-NMDA receptor pathway.

The locus of 7p15.3 was also annotated in [Figure S4,](#page-4-0) and SNPs in strong LD with rs2285947 are located in the initial parts of both SP4 (Specificity Protein 4 [MIM 600540]) and DNAH11 (Dynein, axonemal, heavy chain 11 [MIM 603339]). Publicly available data of cis-expression quantitative trait loci (eQTLs) indicate that rs7788515, a SNP in strong LD with rs2285947 (r^2 = 0.92), is associated with the expression of *DNAH11* ($p = 5.4 \times 10^{-7}$).^{[46](#page-6-0)} Dyneins are microtubule-associated motor protein complexes. Functional link has been established between

MAPK (mitogen-activated protein kinase) components and dynein motors, which are indispensable for activation of MAPK kinase3/6 and p38 in vivo. 47 The p38-MAPK pathway that is strongly activated by stress plays important roles in the immune and inflammatory $r_{\text{exponents}}^{48,49}$ as well as in the regulation of cell survival, differentiation, and migration. $50,51$ However, the effects of variants in this region could be on other genes or noncoding regulatory elements at a distance, and further functional studies are warranted to understand the potentially functional significance of cancer risk-related locus at 7p15.3.

In the current study, we identified two loci at 6p21.1 and 7p15.3 that may serve as susceptibility loci for multiple cancers. Although the approach searching consistent effects on all three cancer types used in this study may represent a cost-efficient manner to discover loci of pleiotropy, it may be too stringent and miss some real loci associated with multiple cancers even though some loci were inconsistent in one study. Therefore, further studies by removing each study per disease may introduce

more candidate loci for further replication and facilitate to discover additional pleiotropy loci. Moreover, these two loci were not investigated and reported in individual cancer study previously³¹⁻³³ due to relative high p values in GWAS stage ($p > 10^{-5}$). A genome-wide significance was observed after combining GWAS and replication stages for associations of rs2494938 at 6p21.1 with NCGC ($p = 4.91 \times 10^{-9}$) and rs2285947 at 7p15.3 with lung cancer ($p = 1.57 \times 10^{-8}$), suggesting that further comprehensive replication studies for individual cancer GWAS are required to indentify missing susceptibility loci.

In summary, our study indicates variants at 6p21.1 and 7p15.3 as candidate susceptibility loci for multiple types of human cancer, though the biological mechanism remains to be elucidated. Further investigations are warranted to extend these findings to other types of human cancers and to other ethnic populations.

Supplemental Data

Supplemental Data includes four figures, two tables, and Supplemental Experimental Procedures and can be found with this article online at [http://www.cell.com/AJHG/.](http://www.cell.com/AJHG/)

Acknowledgments

We thank all the individuals who kindly participated, as well as the researchers and physicians who recruited them and collected information. This work was supported by the National Outstanding Youth Science Foundation of China (81225053), the Key Project of the National Natural Science Foundation of China (81230067), the National Key Basic Research Program Grant (2011CB503805, 2013CB911400, 2013CB910304), the National Natural Science Foundation of China (81001276, 81270044, 81130022, 81202267, 31000553, and 30901233), the Jiangsu Outstanding Youth Science Foundation (BK2012042), the Jiangsu Natural Science Foundation (BK2012841, BK2012443), the US NIH Grant (U19 CA148127), the China National High-Tech Research and Development Program Grant (2009AA022705, 2009AA022706, and 2009AA022701, and the Priority Academic Program for the Development of Jiangsu Higher Education Institutions (Public Health and Preventive Medicine).

Received: May 21, 2012 Revised: July 14, 2012 Accepted: September 13, 2012 Published online: October 25, 2012

Web Resources

The URLs for data presented herein are as follows:

LocusZoom, <https://statgen.sph.umich.edu/locuszoom/> Minimac, <http://genome.sph.umich.edu/wiki/Minimac/> Online Mendelian Inheritance in Man (OMIM), [http://www.](http://www.omim.org/) [omim.org/](http://www.omim.org/)

Plink, [http://pngu.mgh.harvard.edu/~purcell/plink/](http://pngu.mgh.harvard.edu/%7Epurcell/plink/) UCSC genome browser, <http://genome.ucsc.edu/>

References

- 1. Chung, C.C., and Chanock, S.J. (2011). Current status of genome-wide association studies in cancer. Hum. Genet. 130, 59–78.
- 2. Amundadottir, L.T., Sulem, P., Gudmundsson, J., Helgason, A., Baker, A., Agnarsson, B.A., Sigurdsson, A., Benediktsdottir, K.R., Cazier, J.B., Sainz, J., et al. (2006). A common variant associated with prostate cancer in European and African populations. Nat. Genet. 38, 652–658.
- 3. Gudmundsson, J., Sulem, P., Manolescu, A., Amundadottir, L.T., Gudbjartsson, D., Helgason, A., Rafnar, T., Bergthorsson, J.T., Agnarsson, B.A., Baker, A., et al. (2007). Genome-wide association study identifies a second prostate cancer susceptibility variant at 8q24. Nat. Genet. 39, 631–637.
- 4. Yeager, M., Orr, N., Hayes, R.B., Jacobs, K.B., Kraft, P., Wacholder, S., Minichiello, M.J., Fearnhead, P., Yu, K., Chatterjee, N., et al. (2007). Genome-wide association study of prostate cancer identifies a second risk locus at 8q24. Nat. Genet. 39, 645–649.
- 5. Haiman, C.A., Patterson, N., Freedman, M.L., Myers, S.R., Pike, M.C., Waliszewska, A., Neubauer, J., Tandon, A., Schirmer, C., McDonald, G.J., et al. (2007). Multiple regions within 8q24 independently affect risk for prostate cancer. Nat. Genet. 39, 638–644.
- 6. Yeager, M., Chatterjee, N., Ciampa, J., Jacobs, K.B., Gonzalez-Bosquet, J., Hayes, R.B., Kraft, P., Wacholder, S., Orr, N., Berndt, S., et al. (2009). Identification of a new prostate cancer susceptibility locus on chromosome 8q24. Nat. Genet. 41, 1055–1057.
- 7. Gudmundsson, J., Sulem, P., Gudbjartsson, D.F., Blondal, T., Gylfason, A., Agnarsson, B.A., Benediktsdottir, K.R., Magnusdottir, D.N., Orlygsdottir, G., Jakobsdottir, M., et al. (2009). Genome-wide association and replication studies identify four variants associated with prostate cancer susceptibility. Nat. Genet. 41, 1122–1126.
- 8. Al Olama, A.A., Kote-Jarai, Z., Giles, G.G., Guy, M., Morrison, J., Severi, G., Leongamornlert, D.A., Tymrakiewicz, M., Jhavar, S., Saunders, E., et al.; UK Genetic Prostate Cancer Study Collaborators/British Association of Urological Surgeons' Section of Oncology; UK Prostate testing for cancer and Treatment study (ProtecT Study) Collaborators. (2009). Multiple loci on 8q24 associated with prostate cancer susceptibility. Nat. Genet. 41, 1058–1060.
- 9. Tomlinson, I., Webb, E., Carvajal-Carmona, L., Broderick, P., Kemp, Z., Spain, S., Penegar, S., Chandler, I., Gorman, M., Wood, W., et al.; CORGI Consortium. (2007). A genomewide association scan of tag SNPs identifies a susceptibility variant for colorectal cancer at 8q24.21. Nat. Genet. 39, 984–988.
- 10. Zanke, B.W., Greenwood, C.M., Rangrej, J., Kustra, R., Tenesa, A., Farrington, S.M., Prendergast, J., Olschwang, S., Chiang, T., Crowdy, E., et al. (2007). Genome-wide association scan identifies a colorectal cancer susceptibility locus on chromosome 8q24. Nat. Genet. 39, 989–994.
- 11. Haiman, C.A., Le Marchand, L., Yamamato, J., Stram, D.O., Sheng, X., Kolonel, L.N., Wu, A.H., Reich, D., and Henderson, B.E. (2007). A common genetic risk factor for colorectal and prostate cancer. Nat. Genet. 39, 954–956.
- 12. Ghoussaini, M., Song, H., Koessler, T., Al Olama, A.A., Kote-Jarai, Z., Driver, K.E., Pooley, K.A., Ramus, S.J., Kjaer, S.K., Hogdall, E., et al.; UK Genetic Prostate Cancer Study

Collaborators/British Association of Urological Surgeons' Section of Oncology; UK ProtecT Study Collaborators. (2008). Multiple loci with different cancer specificities within the 8q24 gene desert. J. Natl. Cancer Inst. 100, 962–966.

- 13. Easton, D.F., Pooley, K.A., Dunning, A.M., Pharoah, P.D., Thompson, D., Ballinger, D.G., Struewing, J.P., Morrison, J., Field, H., Luben, R., et al.; SEARCH collaborators; kConFab; AOCS Management Group. (2007). Genome-wide association study identifies novel breast cancer susceptibility loci. Nature 447, 1087–1093.
- 14. Pomerantz, M.M., Ahmadiyeh, N., Jia, L., Herman, P., Verzi, M.P., Doddapaneni, H., Beckwith, C.A., Chan, J.A., Hills, A., Davis, M., et al. (2009). The 8q24 cancer risk variant rs6983267 shows long-range interaction with MYC in colorectal cancer. Nat. Genet. 41, 882–884.
- 15. Tuupanen, S., Turunen, M., Lehtonen, R., Hallikas, O., Vanharanta, S., Kivioja, T., Björklund, M., Wei, G., Yan, J., Niittymäki, I., et al. (2009). The common colorectal cancer predisposition SNP rs6983267 at chromosome 8q24 confers potential to enhanced Wnt signaling. Nat. Genet. 41, 885–890.
- 16. McKay, J.D., Hung, R.J., Gaborieau, V., Boffetta, P., Chabrier, A., Byrnes, G., Zaridze, D., Mukeria, A., Szeszenia-Dabrowska, N., Lissowska, J., et al.; EPIC Study. (2008). Lung cancer susceptibility locus at 5p15.33. Nat. Genet. 40, 1404–1406.
- 17. Wang, Y., Broderick, P., Webb, E., Wu, X., Vijayakrishnan, J., Matakidou, A., Qureshi, M., Dong, Q., Gu, X., Chen, W.V., et al. (2008). Common 5p15.33 and 6p21.33 variants influence lung cancer risk. Nat. Genet. 40, 1407–1409.
- 18. Rafnar, T., Sulem, P., Stacey, S.N., Geller, F., Gudmundsson, J., Sigurdsson, A., Jakobsdottir, M., Helgadottir, H., Thorlacius, S., Aben, K.K., et al. (2009). Sequence variants at the TERT-CLPTM1L locus associate with many cancer types. Nat. Genet. 41, 221–227.
- 19. Shete, S., Hosking, F.J., Robertson, L.B., Dobbins, S.E., Sanson, M., Malmer, B., Simon, M., Marie, Y., Boisselier, B., Delattre, J.Y., et al. (2009). Genome-wide association study identifies five susceptibility loci for glioma. Nat. Genet. 41, 899–904.
- 20. Petersen, G.M., Amundadottir, L., Fuchs, C.S., Kraft, P., Stolzenberg-Solomon, R.Z., Jacobs, K.B., Arslan, A.A., Buenode-Mesquita, H.B., Gallinger, S., Gross, M., et al. (2010). A genome-wide association study identifies pancreatic cancer susceptibility loci on chromosomes 13q22.1, 1q32.1 and 5p15.33. Nat. Genet. 42, 224–228.
- 21. Haiman, C.A., Chen, G.K., Vachon, C.M., Canzian, F., Dunning, A., Millikan, R.C., Wang, X., Ademuyiwa, F., Ahmed, S., Ambrosone, C.B., et al.; Gene Environment Interaction and Breast Cancer in Germany (GENICA) Consortium. (2011). A common variant at the TERT-CLPTM1L locus is associated with estrogen receptor-negative breast cancer. Nat. Genet. 43, 1210–1214.
- 22. Wellcome Trust Case Control Consortium. (2007). Genomewide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature 447, 661–678.
- 23. Helgadottir, A., Thorleifsson, G., Manolescu, A., Gretarsdottir, S., Blondal, T., Jonasdottir, A., Jonasdottir, A., Sigurdsson, A., Baker, A., Palsson, A., et al. (2007). A common variant on chromosome 9p21 affects the risk of myocardial infarction. Science 316, 1491–1493.
- 24. Zeggini, E., Weedon, M.N., Lindgren, C.M., Frayling, T.M., Elliott, K.S., Lango, H., Timpson, N.J., Perry, J.R., Rayner, N.W., Freathy, R.M., et al.; Wellcome Trust Case Control Consortium (WTCCC). (2007). Replication of genome-wide

association signals in UK samples reveals risk loci for type 2 diabetes. Science 316, 1336–1341.

- 25. Wrensch, M., Jenkins, R.B., Chang, J.S., Yeh, R.F., Xiao, Y., Decker, P.A., Ballman, K.V., Berger, M., Buckner, J.C., Chang, S., et al. (2009). Variants in the CDKN2B and RTEL1 regions are associated with high-grade glioma susceptibility. Nat. Genet. 41, 905–908.
- 26. Stacey, S.N., Sulem, P., Masson, G., Gudjonsson, S.A., Thorleifsson, G., Jakobsdottir, M., Sigurdsson, A., Gudbjartsson, D.F., Sigurgeirsson, B., Benediktsdottir, K.R., et al. (2009). New common variants affecting susceptibility to basal cell carcinoma. Nat. Genet. 41, 909–914.
- 27. Bishop, D.T., Demenais, F., Iles, M.M., Harland, M., Taylor, J.C., Corda, E., Randerson-Moor, J., Aitken, J.F., Avril, M.F., Azizi, E., et al. (2009). Genome-wide association study identifies three loci associated with melanoma risk. Nat. Genet. 41, 920–925.
- 28. Turnbull, C., Ahmed, S., Morrison, J., Pernet, D., Renwick, A., Maranian, M., Seal, S., Ghoussaini, M., Hines, S., Healey, C.S., et al.; Breast Cancer Susceptibility Collaboration (UK). (2010). Genome-wide association study identifies five new breast cancer susceptibility loci. Nat. Genet. 42, 504–507.
- 29. Bei, J.X., Li, Y., Jia, W.H., Feng, B.J., Zhou, G., Chen, L.Z., Feng, Q.S., Low, H.Q., Zhang, H., He, F., et al. (2010). A genome-wide association study of nasopharyngeal carcinoma identifies three new susceptibility loci. Nat. Genet. 42, 599–603.
- 30. Pasmant, E., Sabbagh, A., Vidaud, M., and Bièche, I. (2011). ANRIL, a long, noncoding RNA, is an unexpected major hotspot in GWAS. FASEB J. 25, 444–448.
- 31. Hu, Z., Wu, C., Shi, Y., Guo, H., Zhao, X., Yin, Z., Yang, L., Dai, J., Hu, L., Tan, W., et al. (2011). A genome-wide association study identifies two new lung cancer susceptibility loci at 13q12.12 and 22q12.2 in Han Chinese. Nat. Genet. 43, 792–796.
- 32. Shi, Y., Hu, Z., Wu, C., Dai, J., Li, H., Dong, J., Wang, M., Miao, X., Zhou, Y., Lu, F., et al. (2011). A genome-wide association study identifies new susceptibility loci for non-cardia gastric cancer at 3q13.31 and 5p13.1. Nat. Genet. 43, 1215–1218.
- 33. Wu, C., Hu, Z., He, Z., Jia, W., Wang, F., Zhou, Y., Liu, Z., Zhan, Q., Liu, Y., Yu, D., et al. (2011). Genome-wide association study identifies three new susceptibility loci for esophageal squamous-cell carcinoma in Chinese populations. Nat. Genet. 43, 679–684.
- 34. Khuder, S.A. (2001). Effect of cigarette smoking on major histological types of lung cancer: a meta-analysis. Lung Cancer 31, 139–148.
- 35. Ladeiras-Lopes, R., Pereira, A.K., Nogueira, A., Pinheiro-Torres, T., Pinto, I., Santos-Pereira, R., and Lunet, N. (2008). Smoking and gastric cancer: systematic review and meta-analysis of cohort studies. Cancer Causes Control 19, 689–701.
- 36. Tramacere, I., La Vecchia, C., and Negri, E. (2011). Tobacco smoking and esophageal and gastric cardia adenocarcinoma: a meta-analysis. Epidemiology 22, 344–349.
- 37. Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M.A., Bender, D., Maller, J., Sklar, P., de Bakker, P.I., Daly, M.J., and Sham, P.C. (2007). PLINK: a tool set for wholegenome association and population-based linkage analyses. Am. J. Hum. Genet. 81, 559–575.
- 38. Pruim, R.J., Welch, R.P., Sanna, S., Teslovich, T.M., Chines, P.S., Gliedt, T.P., Boehnke, M., Abecasis, G.R., and Willer, C.J. (2010). LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics 26, 2336–2337.
- 39. Wang, C.Y., Chang, K., Petralia, R.S., Wang, Y.X., Seabold, G.K., and Wenthold, R.J. (2006). A novel family of adhesionlike molecules that interacts with the NMDA receptor. J. Neurosci. 26, 2174–2183.
- 40. Castellanos, A., Lang, G., Frampton, J., and Weston, K. (2007). Regulation of erythropoiesis by the neuronal transmembrane protein Lrfn2. Exp. Hematol. 35, 724–734.
- 41. Fernandes, H.B., Baimbridge, K.G., Church, J., Hayden, M.R., and Raymond, L.A. (2007). Mitochondrial sensitivity and altered calcium handling underlie enhanced NMDA-induced apoptosis in YAC128 model of Huntington's disease. J. Neurosci. 27, 13614–13623.
- 42. Skeberdis, V.A., Chevaleyre, V., Lau, C.G., Goldberg, J.H., Pettit, D.L., Suadicani, S.O., Lin, Y., Bennett, M.V., Yuste, R., Castillo, P.E., and Zukin, R.S. (2006). Protein kinase A regulates calcium permeability of NMDA receptors. Nat. Neurosci. 9, 501–510.
- 43. Kim, M.S., Yamashita, K., Baek, J.H., Park, H.L., Carvalho, A.L., Osada, M., Hoque, M.O., Upadhyay, S., Mori, M., Moon, C., and Sidransky, D. (2006). N-methyl-D-aspartate receptor type 2B is epigenetically inactivated and exhibits tumorsuppressive activity in human esophageal cancer. Cancer Res. 66, 3409–3418.
- 44. Tamura, H., Suzuki, M., Moriya, Y., Hoshino, H., Okamoto, T., Yoshida, S., and Yoshino, I. (2011). Aberrant methylation of N-methyl-D-aspartate receptor type 2B (NMDAR2B) in nonsmall cell carcinoma. BMC Cancer 11, 220.
- 45. Seo, J.H., Fox, J.G., Peek, R.M., Jr., and Hagen, S.J. (2011). N-methyl D-aspartate channels link ammonia and epithelial cell death mechanisms in Helicobacter pylori Infection. Gastroenterology 141, 2064–2075.
- 46. Ge, B., Pokholok, D.K., Kwan, T., Grundberg, E., Morcos, L., Verlaan, D.J., Le, J., Koka, V., Lam, K.C., Gagné, V., et al. (2009). Global patterns of cis variation in human cells revealed by high-density allelic expression analysis. Nat. Genet. 41, 1216–1222.
- 47. Cheung, P.Y., Zhang, Y., Long, J., Lin, S., Zhang, M., Jiang, Y., and Wu, Z. (2004). p150(Glued), Dynein, and microtubules are specifically required for activation of MKK3/6 and p38 MAPKs. J. Biol. Chem. 279, 45308–45311.
- 48. Han, J., Lee, J.D., Bibbs, L., and Ulevitch, R.J. (1994). A MAP kinase targeted by endotoxin and hyperosmolarity in mammalian cells. Science 265, 808–811.
- 49. Lee, J.C., Laydon, J.T., McDonnell, P.C., Gallagher, T.F., Kumar, S., Green, D., McNulty, D., Blumenthal, M.J., Heys, J.R., Landvatter, S.W., et al. (1994). A protein kinase involved in the regulation of inflammatory cytokine biosynthesis. Nature 372, 739–746.
- 50. Cuenda, A., and Rousseau, S. (2007). p38 MAP-kinases pathway regulation, function and role in human diseases. Biochim. Biophys. Acta 1773, 1358–1375.
- 51. Cuadrado, A., and Nebreda, A.R. (2010). Mechanisms and functions of p38 MAPK signalling. Biochem. J. 429, 403–417.