

NIH Public Access

Author Manuscript

Epilepsy Res. Author manuscript; available in PMC 2013 November 01.

Published in final edited form as:

Epilepsy Res. 2012 November ; 102(1-2): 126–130. doi:10.1016/j.eplepsyres.2012.07.019.

Conditional deletion of TrkC does not modify limbic epileptogenesis

A. Soren Leonard1, **Ram S. Puranam**1,2, **Jeffrey Helgager**1, **Gumei Liu**1, and **James O. McNamara**1,2,3,*

¹Department of Neurobiology, Duke University Medical Center, Durham, NC

²Department of Medicine (Neurology), Duke University Medical Center, Durham, NC

³Department of Pharmacology and Molecular Cancer Biology, Duke University Medical Center, Durham, NC

Abstract

The neurotrophin receptor, tropomyosin-related kinase B (TrkB), is required for epileptogenesis in the kindling model. The role of a closely related neurotrophin receptor, TrkC, in limbic epileptogenesis is unknown. We examined limbic epileptogenesis in the kindling model in TrkC conditional null mice, using a strategy that previously established a critical role of TrkB. Despite elimination of TrkC mRNA, no differences in development of kindling were detected between TrkC conditional null and wild type control mice. These findings reinforce the central role of TrkB as the principal neurotrophin receptor involved in limbic epileptogenesis.

Keywords

Tropomyosin-related kinase C; TrkC; Kindling model; Epileptogenesis; Cre recombinase

INTRODUCTION

Although simultaneous overexpression of exogenous BDNF and FGF2 can limit epileptogenesis in the pilocarpine model (Paradiso et al., 2009), converging lines of evidence demonstrate that excessive activation of the receptor tyrosine kinase, TrkB, by endogenous ligands is critical for induction of limbic epileptogenesis (reviewed by McNamara et al., 2006; Brooks-Kayal et al., 2009). Specifically, the genetic perturbation of brain-derived neurotrophic factor (BDNF) (Kokaia et al., 1995; Croll et al., 1999; He et al., 2004; Barton and Shannon, 2005) or its receptor, tropomyosin-related kinase B (TrkB) (Lähteinen et al., 2002; He et al., 2004, 2010; Kotloski and McNamara, 2010; Heinrich et al., 2011), have convincingly demonstrated that this particular neurotrophin pathway is required for limbic epileptogenesis. In contrast to TrkB, whether TrkC is activated by seizures or is required for epileptogenesis has not been investigated. Nevertheless, this

CONFLICT OF INTEREST

^{© 2012} Elsevier B.V. All rights reserved.

^{*}Corresponding Author: James O. McNamara, Department of Neurobiology, Duke University Medical Center, Durham, NC, 27710. jmc@neuro.duke.edu, Ph: 919 684 4241, Fax: 919 684 8219.

The authors have no conflicts of interest.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

We examined the role of TrkC in limbic epileptogenesis using a genetic strategy identical to that which established a critical role for TrkB in the kindling model (He et al., 2004). Because deletion of TrkC from the germline is lethal shortly after birth (Klein et al., 1994), we used a conditional approach in which mice with floxed alleles of TrkC were mated with mice expressing Cre recombinase driven by a synapsin-1 promoter (Syn-Cre), resulting in mice in which both TrkC alleles were eliminated from a subset of central nervous system neurons. We assessed epileptogenesis by quantifying development of kindling and used both RT-PCR and in situ hybridization to verify deletion of TrkC mRNA.

METHODS

All animal experiments were carried out in accordance with current IACUC guidelines under animal protocol A298-09-03. Detail of the animals used, genotyping, qRT-PCR, insitu hybridization, electrode implantation and kindling procedure are described in the Supplemental Methods section.

Electrode implantation and kindling procedure

Surgical and kindling procedures followed that of He et al., 2010 (See supplemental methods). In brief, the electrographic seizure threshold (EST) was determined by administering a 1 sec train at 50 μ A with additional stimulations increasing by 25 μ A (at 1) min intervals) until an electrographic seizure was detected. Stimulations were subsequently administered twice per day at an intensity of the EST until the animals exhibited 3 consecutive seizures of class 4 or greater. Seizures were classified according to a modified Racine (1972) scale. Kindling data are presented as the mean \pm SEM for each group.

RESULTS

Neuron-specific TrkC conditional knockout mice

To selectively eliminate TrkC expression from CNS neurons, mice in which exon 14 of the TrkC gene was flanked by loxP sites (Chen et al., 2005) were crossed to Syn-Cre transgenic mice. Reduction of TrkC mRNA was evidenced by qRT PCR study of hippocampal homogenates which revealed levels in TrkC^{$-/-$} mutant mice approximating 57.2 ± 0.2% of WT mice (p <0.05, Figure 1D, right and middle). Importantly, the floxed TrkC mice in the absence of Syn-Cre exhibited no decrease in TrkC mRNA levels (Figure 1D, right) relative to non-floxed wildtype controls (Figure 1D, left). *In situ* hybridization revealed striking reductions of TrkC mRNA in the dentate granule and CA3 pyramidal cells with lesser reductions in CA1 pyramidal cells of $TrkC^{-/-}$ mutant compared to WT mice (Figure 1A, B, C), a pattern identical to that found with TrkB mRNA using the same Cre driver line (He et al., 2004). Together these findings demonstrate the efficacy of Cre recombinase in reducing TrkC mRNA expression.

Development and persistence of kindling is equivalent in WT and TrkC−**/**− **mice**

The development of kindling as measured by electrophysiological and behavioral responses to stimulation of amygdala proceeded similarly in WT and $Trk^{-/-}$ mice. No differences were found between $WT(n=7)$ and $TrkC^{-/-}$ (n=7) mice with respect to the following measures: First, the current required to evoke the initial electrographic seizure was similar in WT and TrkC^{-/−} mice (279 ± 42 μ A and 271 ± 50 μ A for WT and TrkC^{-/−}, respectively). Second, the duration of the initial electrographic seizure and the progressive lengthening of

electrographic seizure duration were similar in WT and $TrkC^{-/-}$ mice (Fig. 2A). Third, the development of behavioral seizure intensity progressed similarly in mice of both genotypes (Fig. 2B). No significant differences were found in the number of stimulations required to evoke the first clonic motor seizure (class 4 or greater) or the 3rd consecutive clonic or tonic motor seizure lasting at least 10 sec (Fig 2C).

To determine whether a null mutation of TrkC influenced the persistence of the hyperexcitability following the completion of kindling, $WT(n=7)$ and $TrkC^{-/-}$ mutants $(n=7)$ were stimulated following a stimulation-free period of 2 weeks after the 3rd consecutive class 4 or 5 seizure had been evoked. No significant differences were detected in EST assessed after a 2 week stimulation-free period (average ESTs of $338 \pm 132 \,\mu\text{A}$ and 343 ± 144 µA of WT and TrkC^{-/−} mutants). Moreover, the number of additional stimulations required to evoke a class 4 or 5 seizure did not differ between the WT and TrkC^{$-/-$} mice (1.9 ± 0.4 and 1.2 ± 0.2 stimulations, respectively; Fig 2C, far right column). In addition, there was no significant difference in the duration of electrographic seizure $(23.1 \pm 1.2 \text{ and } 26.0 \pm 1.7 \text{ sec})$ or seizure class evoked in the WT and TrkC^{-/−} mutants. Thus, the persistence of the hyperexcitable state established by kindling was unaffected in the $TrkC^{-/-}$ mutant mice.

DISCUSSION

The objective of this study was to test the hypothesis that a conditional deletion of TrkC inhibits epileptogenesis in the kindling model. Two principal findings emerged: 1) crossing Syn-Cre transgenic mice to floxed-TrkC mutant mice reduced TrkC mRNA content as assessed by two independent methods; 2) this reduction of TrkC content did not affect epileptogenesis as revealed by the development of kindling or persistence of hyperexcitability. We conclude that the partial reduction of TrkC expression in this conditional mutant mouse does not modify limbic epileptogenesis in the kindling model.

To compare the effects of TrkC with TrkB on epileptogenesis in the kindling model, the identical genetic strategy using the same Cre driver line was used. Despite reductions of TrkC in a pattern similar to that observed for TrkB in earlier studies (He et al., 2004), no differences were detected in the development of kindling in $TrkC^{-/-}$ compared to WT control mice. The contrast is striking because the development of kindling was eliminated altogether in the conditional $TrkB^{-/-}$ mice (He et al., 2004). Even more modest reductions of TrkB content in the conditional $TrkB^{+/-}$ heterozygous mice resulted in a 50% increase in the number of stimulations required to induce the development of kindling (He et al., 2004).

The absence of detectable inhibition of the development of kindling in the conditional TrkC^{$-/-$} null mutant mice is particularly surprising in light of previous studies of NT-3 heterozygotes in the kindling model. That is, NT-3, a neurotrophin with high affinity and efficacy for TrkC (Lamballe et al., 1991), is thought to function as the principal neurotrophin agonist of TrkC *in vivo*. Mutant mice carrying just one allele of NT-3 exhibit a 50% increase in the number of stimulations required to induce kindling (Elmer et al., 1997). While the present results were surprising in light of the studies of NT- $3^{+/}$ mice, our findings are consistent with earlier studies of Binder et al. (1999) that examined the effects on kindling development of intraventricular (ICV) infusion of recombinant proteins in which the ligand recognition domain of TrkA or B or C was fused in frame with the Fc portion of human IgG1, proteins that bind to and scavenge the endogenous neurotrophins. Whereas ICV infusion of TrkB-Fc markedly inhibited the development of kindling, infusion of either TrkA- or TrkC-Fc had no effect (Binder et al., 1999). The failure of ICV infused TrkC-Fc to inhibit kindling development is consistent with the similarity of kindling development in WT and $TrkC^{-/-}$ conditional mutant mice in the present study. The possibility that

endogenous NT-3 promotes the development of kindling by activation of TrkB seems unlikely because ICV TrkC-Fc would be expected to scavenge NT-3 and partially inhibit development of kindling, yet it did not (Binder et al., 1999).

The present findings underscore several unanswered questions arising with respect to NT-3 in epileptogenesis. What is the cellular and molecular mechanism by which the development of kindling is inhibited in NT-3+/− mice? What is the functional consequence of the seizureinduced reduction of NT-3 mRNA described in multiple animal models (Gall, 1993; Kokaia et al., 1996; Elmer et al., 1997; Ferencz et al., 1997)? Does TrkC undergo activation following seizures, a possibility suggested by the small increases of pTrk immunoreactivity detected in Western blot analyses of TrkB mutant mice (He et al., 2010)? Addressing this last question is hampered by the lack of antibodies that selectively detect TrkC in western blot or by immunoprecipitation. The answers to these questions notwithstanding, the present results demonstrate that TrkC exerts neither a detectable pro- nor anti-epileptogeneic action in the kindling model. These findings underscore the specificity of TrkB among the neurotrophin tyrosine kinase receptors regulating epileptogenesis in animal models.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We thank Dr. David Ginty for the TrkC^{-/−} mice, and Ms. Claudia Clarke for technical support. This work was supported by a grant from the National Institutes of Health (5R01NS056217) to J.O.M. and by a Ruth L. Kirschstein National Research Service Award (F32 NS051064-01) to A.S.L.

References

- Barton ME, Shannon HE. The seizure-related phenotype of brain-derived neurotrophic factor knockdown mice. Neuroscience. 2005; 136:563–569. [PubMed: 16198489]
- Binder DK, Routbort MJ, Ryan TE, Yancopoulos GD, McNamara JO. Selective inhibition of kindling development by intraventricular administration of TrkB receptor body. J Neurosci. 1999; 19:1424– 1436. [PubMed: 9952419]
- Brooks-Kayal AR, Raol YH, Russek SJ. Alteration of epileptogenesis genes. Neurotherapeutics. 2009; 6:312–318. [PubMed: 19332325]
- Chen X, Ye H, Kuruvilla R, Ramanan N, Scangos KW, Zhang C, Johnson NM, England PM, Shokat KM, Ginty DD. A chemical-genetic approach to studying neurotrophin signaling. Neuron. 2005; 46:13–21. [PubMed: 15820690]
- Croll SD, Suri C, Compton DL, Simmons MV, Yancopoulos GD, Lindsay RM, Wiegand SJ, Rudge YS, Scharfman HE. Brain-derived neurotrophic factor transgenic mice exhibit passive avoidance deficits, increased seizure severity and in vitro hyperexcitability in the hippocampus and entorhinal cortex. Neuroscience. 1999; 93:1491–1506. [PubMed: 10501474]
- Elmér E, Kokaia M, Ernfors P, Ferencz I, Kokaia Z, Lindvall O. Suppressed kindling epileptogenesis and perturbed BDNF and TrkB gene regulation in NT-3 mutant mice. Exp Neurol. 1997; 145:93– 103. [PubMed: 9184113]
- Ferencz I, Kokaia M, Keep M, Elmér E, Metsis M, Kokaia Z, Lindvall O. Effects of cholinergic denervation on seizure development and neurotrophin messenger RNA regulation in rapid hippocampal kindling. Neuroscience. 1997; 80:389–399. [PubMed: 9284342]
- Gall CM. Seizure-induced changes in neurotrophin expression: implications for epilepsy. Exp Neurol. 1993; 124:150–166. [PubMed: 8282072]
- He XP, Pan E, Sciarretta C, Minichiello L, McNamara JO. Disruption of TrkB-mediated phospholipase Cgamma signaling inhibits limbic epileptogenesis. J Neurosci. 2010; 30:6188–6196. [PubMed: 20445044]

- He XP, Kotloski R, Nef S, Luikart BW, Parada LF, McNamara JO. Conditional deletion of TrkB but not BDNF prevents epileptogenesis in the kindling model. Neuron. 2004; 43:31–42. [PubMed: 15233915]
- Heinrich C, Lähteinen S, Suzuki F, Anne-Marie L, Huber S, Häussler U, Haas C, Larmet Y, Castren E, Depaulis A. Increase in BDNF-mediated TrkB signaling promotes epileptogenesis in a mouse model of mesial temporal lobe epilepsy. Neurobiol Dis. 2011; 42:35–47. [PubMed: 21220014]
- Kaplan DR, Miller FD. Neurotrophin signal transduction in the nervous system. Curr Opin Neurobiol. 2000; 10:381–391. [PubMed: 10851172]
- Klein R, Silos-Santiago I, Smeyne RJ, Lira SA, Brambilla R, Bryant S, Zhang L, Snider WD, Barbacid M. Disruption of the neurotrophin-3 receptor gene trkC eliminates la muscle afferents and results in abnormal movements. Nature. 1994; 368:249–251. [PubMed: 8145824]
- Kokaia M, Ernfors P, Kokaia Z, Elmér E, Jaenisch R, Lindvall O. Suppressed epileptogenesis in BDNF mutant mice. Exp Neurol. 1995; 133:215–224. [PubMed: 7649227]
- Kokaia Z, Kelly ME, Elmer E, Kokaia M, McIntyre DC, Lindvall O. Seizure-induced differential expression of messenger RNAs for neurotrophins and their receptors in genetically fast and slow kindling rats. Neuroscience. 1996; 75:197–207. [PubMed: 8923534]
- Kotloski R, McNamara JO. Reduction of TrkB expression de novo in the adult mouse impairs epileptogenesis in the kindling model. Hippocampus. 2010; 20:713–723. [PubMed: 19603519]
- Lamballe F, Klein R, Barbacid M. trkC, a new member of the trk family of tyrosine protein kinases, is a receptor for neurotrophin-3. Cell. 1991; 66:967–979. [PubMed: 1653651]
- Lähteinen S, Pitkänen A, Saarelainen T, Nissinen J, Koponen E, Castrén E. Decreased BDNF signalling in transgenic mice reduces epileptogenesis. Eur J Neurosci. 2002; 15:721–734. [PubMed: 11886452]
- McNamara JO, Huang YZ, Leonard AS. Molecular signaling mechanisms underlying epileptogenesis. Sci STKE. 2006; 356:re12. [PubMed: 17033045]
- Paradiso B, Marconi P, Zucchini S, Berto E, Binaschi A, Bozac A, Buzzi A, Mazzuferi M, Magri E, Navarro Mora G, Rodi D, Su T, Volpi I, Zanetti L, Marzola A, Manservigi R, Fabene PF, Simonato M. Localized delivery of fibroblast growth factor-2 and brain-derived neurotrophic factor reduces spontaneous seizures in an epilepsy model. Proc Natl Acad Sci U S A. 2009; 106(17):7191–7196. [PubMed: 19366663]
- Racine RJ. Modification of seizure activity by electrical stimulation. II. Motor seizure, Electroencephalogr. Clin Neurophysiol. 1972; 32:281–294.
- Zhu Y, Romero MI, Ghosh P, Ye Z, Charnay P, Rushing EJ, Marth JD, Parada LF. Ablation of NF1 function in neurons induces abnormal development of cerebral cortex and reactive gliosis in the brain. Genes Dev. 2001; 15:859–876. [PubMed: 11297510]

Figure 1.

In situ hybridization and qRT PCR of TrkC mRNA in the hippocampus of WT and TrkC^{-/−} mutant mice. **A–C)** The granule cell layer (GCL) of the dentate gyrus adjacent to the hilus (H) and molecular layer (ML) exhibited a clear decrease in TrkC mRNA in the $TrkC^{-/-}$ mutant mice (A, right panel) relative to WT controls (A, left panel). The CA3 pyramidal layer (P) adjacent to the stratum oriens (SO) and stratum lucidem (SL) exhibited a clear decrease in TrkC mRNA in the $TrkC^{-/-}$ mutant mice (B, right panel) relative to WT controls (B, left panel). The TrkC mRNA levels in the CA1 pyramidal layer (P) adjacent to the SO and stratum radiatum (SR) exhibited a modest reduction in the $TrkC^{-/-}$ mutant mice (C, right panel) compared to WT controls (C, left panel). Scale bar = 50 μ m. **D**) qRT PCR analysis demonstrating $TrkC^{-/-}$ (n=7; Cre⁺, floxed/floxed) mutant mice exhibit reduced

Leonard et al. Page 7

hippocampal mRNA content (†, p<0.05) relative to WT (n=3; Cre−, floxed/floxed) and nonfloxed $WT^*(n=4)$ controls.

\$watermark-text \$watermark-text

Figure 2.

Kindling development is equivalent in $WT(n=7;$ open squares) and $TrkC^{-/-}$ (n=7; closed squares) mutant mice. **A&B)** Kindling development is presented as electrographic seizure duration (A) and behavioral seizure class (B). Stimulation number (x axis) refers to the number of stimulations that evoked an electrographic seizure with duration of least 5 sec. **C)** Number of stimulations to reach different seizure classes (y axis) in WT and Trk $C^{-/-}$ mice. Left to right: First class 1, class 2 and class 4/5 behaviors; Fully kindled refers to the third consecutive class 4/5 behavioral seizures. The maintenance stimulation was to assess persistence of the kindled state.