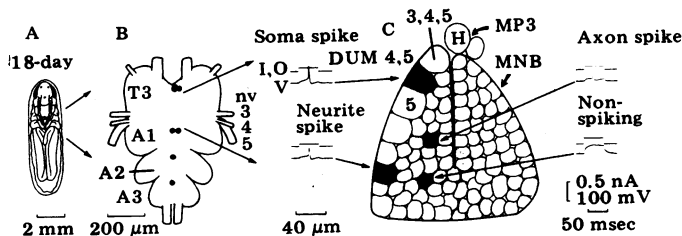


**Correction.** In the article "Cloning of the active thymidine kinase gene of herpes simplex virus type I in *Escherichia coli* K-12" by Florence Colbere-Garapin, Suzanne Chousterman, Florian Horodniceanu, Philippe Kourilsky, and Axel-Claude Garapin, which appeared in the August 1979 issue of *Proc. Natl. Acad. Sci. USA* (76, 3755-3759), the authors request that the following correction be noted. The reference cited in lines 2-4 of the Introduction should be: Kit, S. & Dubbs, D. R. (1963) *Biochem. Biophys. Research Commun.* 11, 55-59.

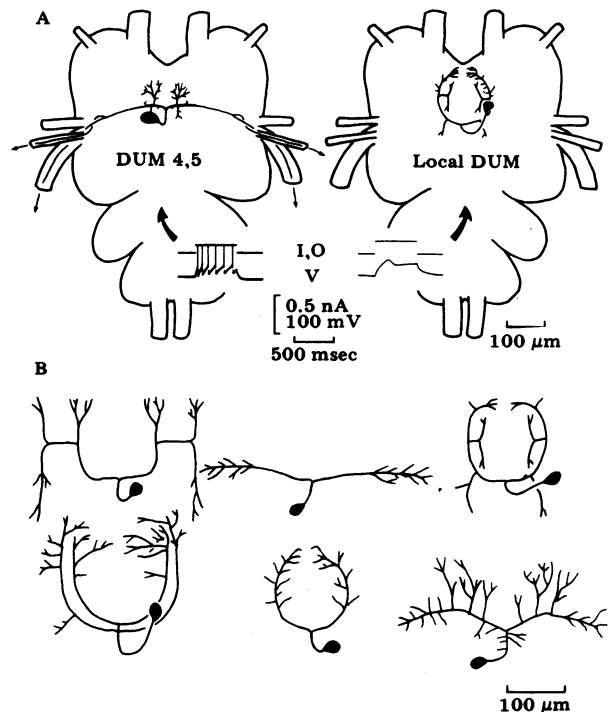
**Correction.** In the article "Inheritance of acquired immunological tolerance to foreign histocompatibility antigens in mice" by R. M. Goczynski and E. J. Steele, which appeared in the May 1980 issue of *Proc. Natl. Acad. Sci. USA* (77, 2871-2875), there was an error in Table 1. Under "Outcross" the entry should be: "Normal CBA♀ × δ<sub>1</sub> (1st gen.<sup>tn</sup>)."

**Correction.** In the article "Large and persistent electrical currents enter the transected lamprey spinal cord" by Richard B. Borgens, Lionel F. Jaffe, and Melvin J. Cohen, which appeared in the February 1980 issue of *Proc. Natl. Acad. Sci. USA* (77, 1209-1213), there was an error in the affiliation line. The work actually was done in the Biology Department at Purdue University. Two of the authors (R.B.B. and M.J.C.) were in the Biology Department at Yale University and came to Purdue to carry out this study in L.F.J.'s laboratory.

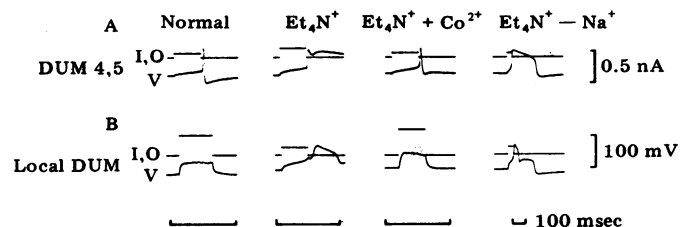
**Correction.** In the article "Electrical excitability: A spectrum of properties in the progeny of a single embryonic neuroblast" by Corey S. Goodman, Keir G. Pearson, and Nicholas C. Spitzer, which appeared in the March 1980 issue of *Proc. Natl. Acad. Sci. USA* (77, 1676-1680), details of some of the figures were not printed well or were retouched incorrectly by the printer. Corrected Figs. 1, 2, and 4 are printed here.



**FIG. 1.** Grasshopper embryo at day 18; diagrams and camera lucida drawings of living specimens (hatching occurs on day 20 at 35°C). (A) Embryo at day 18 viewed within the egg case. (B) Dorsal outline of metathoracic ganglion (T3) fused with first three abdominal ganglia (A1-A3), showing the location of progeny produced by the DUM neuroblast (or median neuroblast, MNB) in each segment (stippled areas) and the one or two remaining progeny of the single cell division of midline precursor 3 (MP3) in each segment (black dots). Lateral nerves (nv) 3-5 are indicated. (C) Camera lucida drawing of the packet of ≈100 progeny of the DUM (MNB) neuroblast and two progeny of MP3. Four of the large DUM neurons are individually identified in the drawing (see text). The DUM neuron somata in a packet appear under Nomarski optics to be encased in a glial sheath. The glial sheath has a median boundary that divides the packet into left and right portions. The progeny of the DUM neuroblast show the complete spectrum of electrical excitability, from soma spikes, to neurite spikes, to axon spikes, to nonspiking, I,0 indicate injected current and zero reference voltage; V, the voltage of the intracellular microelectrode. This range of electrical properties is recorded in cells that appear by other criteria to have reached their mature phenotypes.



**FIG. 2.** (A) Comparison of the morphology and physiology of two progeny of the DUM neuroblast in the grasshopper: DUM 4,5 and a local DUM neuron. DUM 4,5 has a large-diameter soma and peripheral axons in lateral nerves 4 and 5, and generates overshooting soma spikes. The local DUM neuron has a small-diameter soma, is intraganglionic, and is incapable of generating action potentials in normal saline (it shows delayed rectification). Drawings are based on injections of the dye Lucifer yellow in an 18-day embryo. (B) Six other examples of the intraganglionic morphology of small nonspiking DUM neurons in an 18-day embryo.



**FIG. 4.** Comparison of electrical excitability of two progeny of the DUM neuroblast in the grasshopper: DUM 4,5 and a local DUM neuron in an 18-day embryo. (A) The soma of DUM 4,5 generates action potentials that depend on Na<sup>+</sup> and Ca<sup>2+</sup> for their inward current (1, 10, 11). Addition of 30 mM Et<sub>4</sub>N<sup>+</sup> converts the brief action potential into a long and more complex response. The long plateau is abolished by the addition of 10 mM Co<sup>2+</sup>, whereas the initial spike is nearly completely eliminated by removal of Na<sup>+</sup>. (B) The local DUM neuron does not produce action potentials in normal saline. Addition of 30 mM Et<sub>4</sub>N<sup>+</sup> induces the cell to produce long-duration Ca<sup>2+</sup> action potentials that are blocked by the addition of 10 mM Co<sup>2+</sup> and are unaffected by the removal of Na<sup>+</sup>. See text for discussion of different plateau amplitudes.