

Published in final edited form as:

Chem Commun (Camb). 2012 December 21; 48(98): . doi:10.1039/c2cc37221e.

Reversible Binding of Nitric Oxide to an Fe(III) Complex of a Tetra-amido Macrocyclic

Michael D. Pluth^b and Stephen J. Lippard^a

Stephen J. Lippard: lippard@mit.edu

^aDepartment of Chemistry, Massachusetts Institute of Technology (MIT), Cambridge, MA, 02139 USA

Abstract

Nitric oxide binds reversibly to the Fe(III) complex of a well-developed tetra-amido macrocyclic ligand. Reaction with NO results in formation of a species consistent with an S=1 {Fe-NO}⁶ ground state as characterized by UV-vis, IR, EPR, and Mössbauer spectroscopy. The resultant nitrosyl is labile and dissociates readily upon purging with N₂, thus providing a rare example of reversible NO binding to non-heme iron.

Nitric oxide plays an important role in biological signaling, with numerous pathways involving interactions with transition metal centers.¹ Accordingly, bioinorganic chemists have been inspired to investigate the interactions of NO with a variety of transition metal centers.² Our interest in transition metal-based nitrosyl chemistry, which includes the design and implementation of intra- and extracellular sensors of NO, prompted our investigation of non-heme iron(III) complexes as a platform for binding NO *reversibly*. Current small-molecule-based methods for biological NO detection rely on irreversible NO reactions, thereby only allowing for observation of cumulative NO production, rather than real-time concentration changes. A system that could react with NO reversibly would allow for direct detection of both increases and decreases in intracellular NO concentrations; a currently unmet need in NO sensing.

Transition metals offer an attractive platform for reversible NO binding. For this study, we focused on non-heme {M-NO}⁶ centers, where *n* in {M-NO}^{*n*} refers to the total the total number electrons from the metal-*d* and NO- * orbitals,³ particularly Fe(III) and Ru(III) nitrosyls stabilized by poly-anionic ligands.⁴ Although many of these compounds release NO upon photolysis,⁵ release of NO without an external stimulus remains difficult. A major challenge in developing systems that bind NO reversibly is to ensure that NO is released as ·NO rather than NO⁻ or NO⁺, for the latter events are accompanied by metal oxidation or reduction, respectively. We envisioned that an Fe(III) center housed in a ligand that stabilizes this oxidation state might promote NO binding and release without formal reduction of iron. We therefore explored the reaction of NO with an Fe(III) complex of the well-studied tetra-amido macrocycle (TAML) scaffold⁶ depicted in Scheme 1. Although this and related systems have previously supported chemistry of high-valent metal complexes,⁷ the reactivity with NO was previously unexplored.

© The Royal Society of Chemistry

Correspondence to: Stephen J. Lippard, lippard@mit.edu.

^bCurrent Address: Department of Chemistry, University of Oregon, Eugene OR, 97403

[†]Electronic Supplementary Information (ESI) available: Experimental details, spectroscopic methods, and details of DFT calculations. See DOI: 10.1039/b000000x/

Anaerobic treatment of $[\text{Fe}(\text{TAML})]^-$ with NO in a 1:100 MeOH:MeCN solution⁸ resulted in immediate conversion of the initial orange solution to deep purple with concomitant appearance of new bands at 533 ($\epsilon = 1600 \text{ M}^{-1} \text{ cm}^{-1}$) and 888 ($\epsilon = 3500 \text{ M}^{-1} \text{ cm}^{-1}$) nm in the UV-vis spectrum (Figure 1). Purging the solution with dry nitrogen either under ambient light or in the dark regenerated the optical spectrum of $[\text{Fe}(\text{TAML})]^-$. Further addition of NO restored the purple solution. These results suggest that bound NO is in equilibrium with free NO in solution, and that purging promotes complete release of the gas, subsequent reintroduction of which restores the NO adduct. This process was repeated numerous times without apparent decomposition of $[\text{Fe}(\text{TAML})]^-$.

$[\text{Fe}(\text{TAML})]^-$ has two available axial coordination sites, and reaction with NO could potentially afford both mono- and/or dinitrosyl adducts. To probe the stoichiometry of NO binding, a Job's plot was prepared by varying the molar ratios of $[\text{Fe}(\text{TAML})]^-$ to NO. Monitoring the absorbance at 888 nm as a function of the altered molar ratio of reactants revealed a clear break at 0.5, which is characteristic of 1:1 adduct formation (Figure 2a). This result is consistent with formation of a mononitrosyl, in which iron could either be a five-coordinate or octahedral six-coordinate with solvent coordinated at an axial site trans to the nitrosyl ligand.

To further investigate NO binding, variable temperature UV-vis spectroscopic experiments were performed. The strong visible and near-IR absorptions of $[\text{Fe}(\text{TAML})\text{NO}]^-$ at 533 and 888 nm, respectively, are qualitatively similar to absorptions at 565 and 970 nm recently reported for the $S=1$ $\{\text{Fe}-\text{NO}\}^6$ adduct of a tetra-anionic N_2S_2 ligand scaffold from which NO could be released upon heating.⁹ In the N_2S_2 system, the near-IR absorptions were assigned to electronic transitions in the $S=1$ manifold, but cooling the nitrosyl to 0 °C bleached these bands owing to spin crossover from the $S=1$ to the $S=0$ spin state. To perform similar studies with $[\text{Fe}(\text{TAML})\text{NO}]^-$, variable temperature UV-vis experiments from -30 to 80 °C in 100:1 $\text{CH}_3\text{CN}:\text{MeOH}$ or 0 to -78 °C in MeOH were performed (Figure 3b,c). In both cases, the intensities of the near-IR absorptions changed but the wavelength of the absorbance maxima was not appreciably altered. These results suggest that the change in UV-vis intensity reflects the solvent dependence of the NO binding thermodynamics rather than $[\text{Fe}(\text{TAML})\text{NO}]^-$ spin state interconversion. The solvent dependence of NO binding can further be observed by addition of H_2O to a solution of $[\text{Fe}(\text{TAML})\text{NO}]^-$, which leads to partial displacement of coordinated NO (Figure S1).

To provide direct evidence of iron nitrosyl bond formation, solution IR studies were performed, monitoring the growth and disappearance of a nitrosyl stretch as a function of NO addition and N_2 purging. Addition of 5 equiv of NO to a 20 mM solution of $[\text{Fe}(\text{TAML})]^-$ in 5:1 $\text{CH}_3\text{CN}:\text{MeOD}$ resulted in the appearance of a new band at 1799 cm^{-1} , consistent with formation of an Fe(III) nitrosyl (Figure 3). Purging the headspace of the reaction vessel with N_2 to remove the NO completely bleached the nitrosyl stretch. Readdition of NO regenerated the 1799 cm^{-1} stretch with an identical intensity as the initial cycle (Figure 3). Isotopic labeling with ^{15}NO resulted in an observed NO stretch at 1761 cm^{-1} , which agrees well with the predicted isotopic shift of 1767 cm^{-1} based on the simple harmonic oscillator model.

To further explore the electronic structure of the resultant iron nitrosyl, EPR and NMR spectroscopy were employed. Samples of 10 mM $[\text{Fe}(\text{TAML})]^-$ in MeOH were treated with 10 equiv of NO and frozen at 20 K. Upon reaction with NO, the characteristic $S=3/2$ EPR signal of $[\text{Fe}(\text{TAML})]^-$ bleached completely.¹⁰ These results are consistent with formation of an EPR-silent, integer spin iron product (Figure S2).

NMR spectroscopy was used to further probe the spin state of the nitrosyl product. Although $[\text{Fe}(\text{TAML})]^-$ is paramagnetic, the ^1H NMR spectrum shows characteristic resonances that can be monitored upon reaction with NO. If the metal nitrosyl product is diamagnetic, as is the case with many $\{\text{Fe}-\text{NO}\}^6$ complexes, the ^1H NMR spectrum should sharpen significantly and show resonances consistent with a typical diamagnetic compound. Alternatively, if the product is a less-common paramagnetic $S=1$ or $S=2$ $\{\text{Fe}-\text{NO}\}^6$ species, the NMR spectrum may show new characteristic features of the $\{\text{Fe}-\text{NO}\}^6$. To test these possible outcomes, a 10 mM solution of $[\text{Fe}(\text{TAML})]^-$ in either 9:1 $\text{CD}_3\text{CN}:\text{CD}_3\text{OD}$ or neat CD_3OD was titrated with different amounts of NO. Diamagnetic sharpening was not observed in either solvent system, but new paramagnetic resonances were observed. For example, in CD_3OD , the resonance at -13 ppm in the NMR spectrum of $[\text{Fe}(\text{TAML})]^-$ disappeared upon treatment with NO, and the shape of the spectrum from 0 to -5 ppm changed upon NO addition (Figure S3). These results are consistent with formation of a paramagnetic $[\text{Fe}(\text{TAML})\text{NO}]^-$ species.¹¹

We further characterized the nitrosyl product by Mössbauer spectroscopy. Isolating the nitrosyl adduct in solid form proved problematic owing to the ease by which the coordinated NO was displaced. Solid material containing $[\text{Fe}(\text{TAML})\text{NO}]^-$ was isolated by bubbling NO through a solution of $[\text{Fe}(\text{TAML})]^-$ in methanol at a rate sufficient to evaporate the solvent. The IR spectrum of the resultant black solid revealed a nitrosyl stretch at 1797 cm^{-1} (KBr pellet) consistent with the solution IR measurements. Mössbauer spectroscopy at 80 K was performed, and the resultant data were fit to a three-site model corresponding to the $[\text{Fe}(\text{TAML})]^-$ starting material (64%), the nitrosyl complex (36%), and a minor uncharacterized impurity (0.07%) (Figure 4).¹² The parameters corresponding to the nitrosyl adduct were $\delta = -0.20\text{ mm/s}$, $\delta_{\text{eq}} = 3.29\text{ mm/s}$, $\delta_{\text{L,R}} = 0.31$. The observed negative isomer shift is similar to those of several Fe(IV) compounds coordinated by similar tetra-amido ligands,^{7e,7j} suggesting the formation of an $\text{Fe}^{\text{IV}}-\text{NO}^-$ core.

In addition to the solution and solid state measurements of the structure and properties of $[\text{Fe}(\text{TAML})\text{NO}]^-$, we performed DFT calculations to probe the electronic configuration of the complex at the B3LYP/6-311+g(d,p) level of theory. Previous calculations of the coordinatively-unsaturated $[\text{Fe}(\text{TAML})]^-$ complex showed high spin density in the dz^2 orbital, poised to react with NO.^{7g} Our calculations confirmed this prior result and suggest that the $S=1$ nitrosyl complex is 4.0 kcal/mol more stable than the analogous $S=0$ complex.¹³ This energetic difference is consistent with the solution data, which suggest an $S=1$ ground state electronic configuration for $[\text{Fe}(\text{TAML})\text{NO}]^-$. Time dependent DFT (TDDFT) calculations were performed using PB1PBE/6-311+g(d,p) to calculate the UV-vis spectra of the optimized $S=0$ and $S=1$ $[\text{Fe}(\text{TAML})\text{NO}]^-$ structures. The calculated UV-spectrum of the $S=1$ complex (Figure S4, S5) qualitatively reproduces the experimentally observed near-IR absorbance, whereas the calculated $S=0$ spectrum was featureless above 680 nm. Taken together with the spectroscopic evidence, the energy and TDDFT calculations support an $S=1$ spin state for $[\text{Fe}(\text{TAML})\text{NO}]^-$.

Conclusions

A rare example of reversible binding of NO to a synthetic non-heme Fe(III) complex is presented. Upon addition of NO, the $S=3/2$ $[\text{Fe}(\text{TAML})]^-$ complex binds NO with a 1:1 stoichiometry and spectroscopic parameters consistent with formation of an $S=1$ $\{\text{Fe}-\text{NO}\}^6$ complex. Upon purging with N_2 , the coordinated NO is released to regenerate the parent Fe(III) complex.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

The work was supported by the National Science Foundation (CHE-061194 to SJL) and National Institutes of Health (K99GM092970 to MDP). The authors thank Dr. Loi Do and Mr. Justin Wilson for helpful discussions in addition to assistance with Mössbauer spectroscopy and DFT calculations, respectively.

references

1. a.) Tennyson AG, Lippard SJ. *Chem. Biol.* 2011; 18:1211–1220. [PubMed: 22035790] b.) Thomas DD, Ridnour LA, Isenberg JS, Flores-Santana W, Switzer CH, Donzelli S, Hussain P, Vecoli C, Paolucci N, Ambs S, Colton CA, Harris CC, Roberts DD, Wink DA. *Free Radical Biol. Med.* 2008; 45:18–31. [PubMed: 18439435] c.) Goodrich LE, Paulat F, Praneeth VKK, Lehnert N. *Inorg. Chem.* 2010; 49:6293–6316. [PubMed: 20666388] d.) Roncaroli F, Videla M, Slep LD, Olabe JA. *Coord. Chem. Rev.* 2007; 251:1903–1930.
2. a.) Schopfer MP, Wang J, Karlin KD. *Inorg. Chem.* 2010; 49:6267–6282. [PubMed: 20666386] b.) McCleverty JA. *Chem. Rev.* 2004; 104:403–418. [PubMed: 14871130] c.) Hayton TW, Legzdins P, Sharp WB. *Chem. Rev.* 2002; 102:935–991. [PubMed: 11942784]
3. Enemark JH, Feltham RD. *Coord. Chem. Rev.* 1974; 13:339–406.
4. For selected examples, see: Schweitzer D, Ellison JJ, Shoner SC, Lovell S, Kovacs JA. *J. Am. Chem. Soc.* 1998; 120:10996–10997. Patra AK, Afshar R, Olmstead MM, Mascharak PK. *Angew. Chem. Int. Ed.* 2002; 41:2512–2515. Sellmann D, Blum N, Heinemann FW, Hess BA. *Chem.-Eur. J.* 2001; 7:1874–1880. [PubMed: 11405465] Serres RG, Grapperhaus CA, Bothe E, Bill E, Weyhermuller T, Neese F, Wieghardt K. *J. Am. Chem. Soc.* 2004; 126:5138–5153. [PubMed: 15099097]
5. For selected examples, see: Merkle AC, McQuarters AB, Lehnert N. *Dalton Trans.* 2012; 41:8047–8059. [PubMed: 22653336] Fry NL, Mascharak PK. *Acc. Chem. Res.* 2011; 44:289–298. [PubMed: 21361269] Fry NL, Wei J, Mascharak PK. *Inorg. Chem.* 2011; 50:9045–9052. [PubMed: 21815610] Ghosh K, Kumar S, Kumar R, Singh UP, Goel N. *Inorg. Chem.* 2010; 49:7235–7237. [PubMed: 20690733] De P, Sarkar B, Maji S, Das AK, Bulak E, Mobin SM, Kaim W, Lahiri GK. *Eur. J. Inorg. Chem.* 2009:2702–2710. Rose MJ, Mascharak PK. *Inorg. Chem.* 2009; 48:6904–6917. [PubMed: 19480433] Halpenny GM, Mascharak PK. *Inorg. Chem.* 2009; 48:1490–1497. [PubMed: 20560617] Rose MJ, Mascharak PK. *Coord. Chem. Rev.* 2008; 252:2093–2114. [PubMed: 21052477]
6. a.) Collins TJ, Powell RD, Slebodnick C, Uffelman ES. *J. Am. Chem. Soc.* 1991; 113:8419–8425. b.) Bartos MJ, Gordon-Wylie SW, Fox BG, Wright LJ, Weintraub ST, Kauffmann KE, Münck E, Kostka KL, Uffelman ES, Rickard CEF, Noon KR, Collins TJ. *Coord. Chem. Rev.* 1998; 174:361–390.
7. a.) Popescu DL, Vrabel M, Brausam A, Madsen P, Lente G, Fabian I, Ryabov AD, van Eldik R, Collins TJ. *Inorg. Chem.* 2010; 49:11439–11448. [PubMed: 21086984] b.) Ellis WC, McDaniel ND, Bernhard S, Collins TJ. *J. Am. Chem. Soc.* 2010; 132:10990–10991. [PubMed: 20698652] c.) Ellis WC, Tran CT, Denardo MA, Fischer A, Ryabov AD, Collins TJ. *J. Am. Chem. Soc.* 2009; 131:18052–+; d.) Ghosh A, Mitchell DA, Chanda A, Ryabov AD, Popescu DL, Upham EC, Collins GJ, Collins TJ. *J. Am. Chem. Soc.* 2008; 130:15116–15126. [PubMed: 18928252] e.) Chanda A, Shan XP, Chakrabarti M, Ellis WC, Popescu DL, de Oliveira FT, Wang D, Que L, Collins TJ, Münck E, Bominaar EL. *Inorg. Chem.* 2008; 47:3669–3678. [PubMed: 18380453] f.) de Oliveira FT, Chanda A, Banerjee D, Shan XP, Mondal S, Que L, Bominaar EL, Münck E, Collins TJ. *Science.* 2007; 315:835–838. [PubMed: 17185561] g.) Chanda A, Popescu DL, de Oliveira FT, Bominaar EL, Ryabov AD, Münck E, Collins TJ. *J. Inorg. Biochem.* 2006; 100:606–619. [PubMed: 16464502] h.) Collins TJ. *Acc. Chem. Res.* 2002; 35:782–790. [PubMed: 12234208] i.) Kostka KL, Fox BG, Hendrich MP, Collins TJ, Rickard CEF, Wright LJ, Münck E. *J. Am. Chem. Soc.* 1993; 115:6746–6757. j.) Collins TJ, Fox BG, Hu ZG, Kostka KL, Münck E, Rickard CEF, Wright LJ. *J. Am. Chem. Soc.* 1992; 114:8724–8725.

8. MeOH was added to help solubilize the Fe(III) complex
9. Rose MJ, Betterley NM, Oliver AG, Mascharak PK. *Inorg. Chem.* 2010; 49:1854–1864. [PubMed: 20067276]
10. A minor signal was observed at $g = 2.01$ which we attribute to either a minor impurity, or potential formation of a small amount of an $S=1/2$ six-coordinate trans-dinitrosyl. The overall spin of this species accounts for less than 5% of the total sample by spin integration.
11. Attempts to measure the spin multiplicity directly using the Evans method proved unsuccessful, probably due to the excess free NO required to drive the equilibrium to the nitrosyl product
12. Mossbauer parameters for the starting material were $a = 0.15$ mm/s, $a_{eq} = 4.02$ mm/s, $L_R = 0.68$, 0.74 , and for the minor impurity were $a = 0.12$ mm/s, $a_{eq} = 0.57$ mm/s, $L_R = 0.25$.
13. Attempts to optimize the geometry of the $S=2$ nitrosyl from numerous initial geometries always resulted in dissociation of the NO ligand, suggesting that the $S=2$ spin state was energetically favorable.

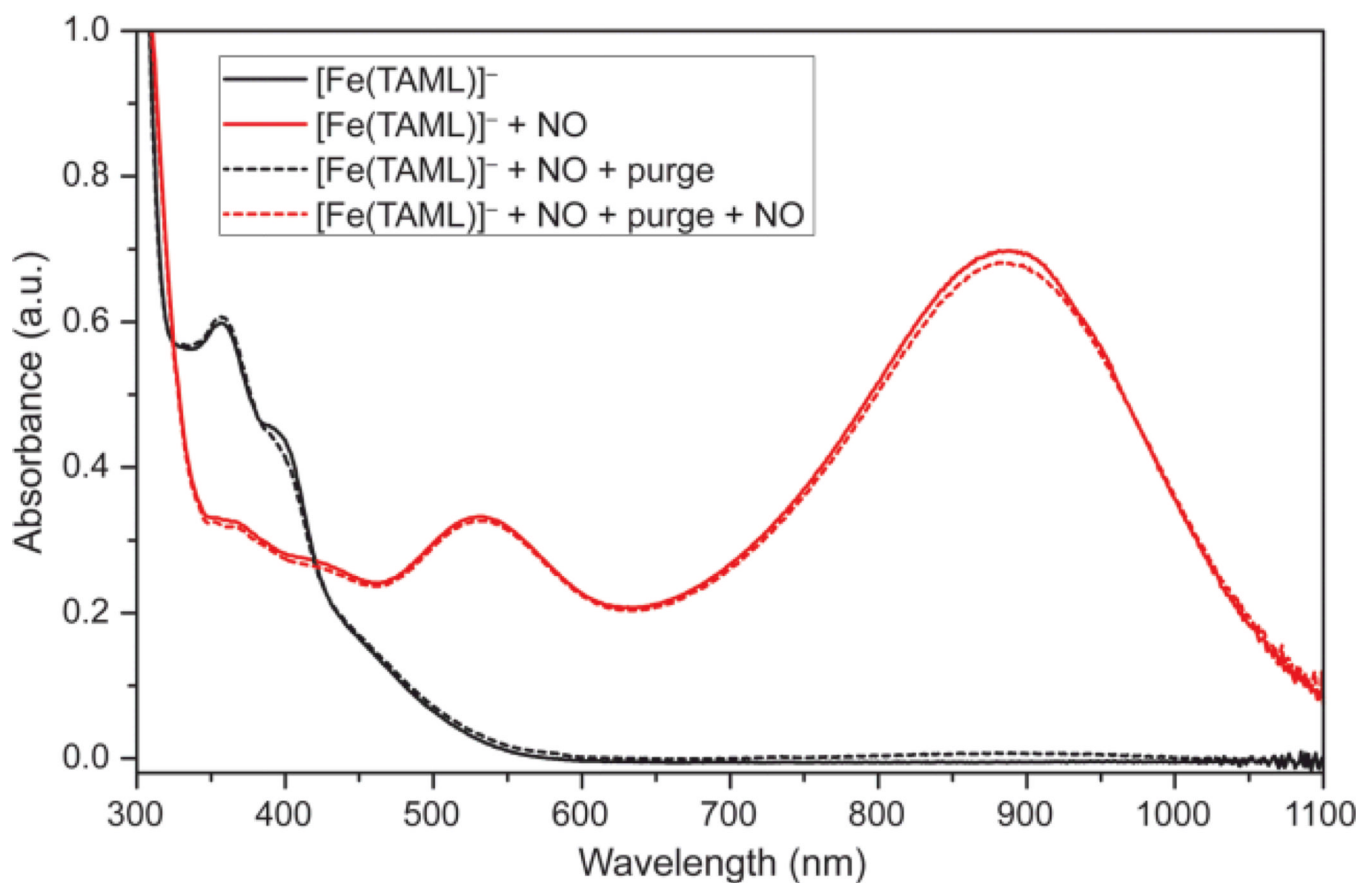


Figure 1.

UV-vis spectra of [Fe(TAML)]⁻ before (black) and after (red) addition of NO. Purging the sample 60 s with N₂ (dotted black) regenerates the starting spectrum. Further addition of NO (dotted red) regenerates the NO adduct.

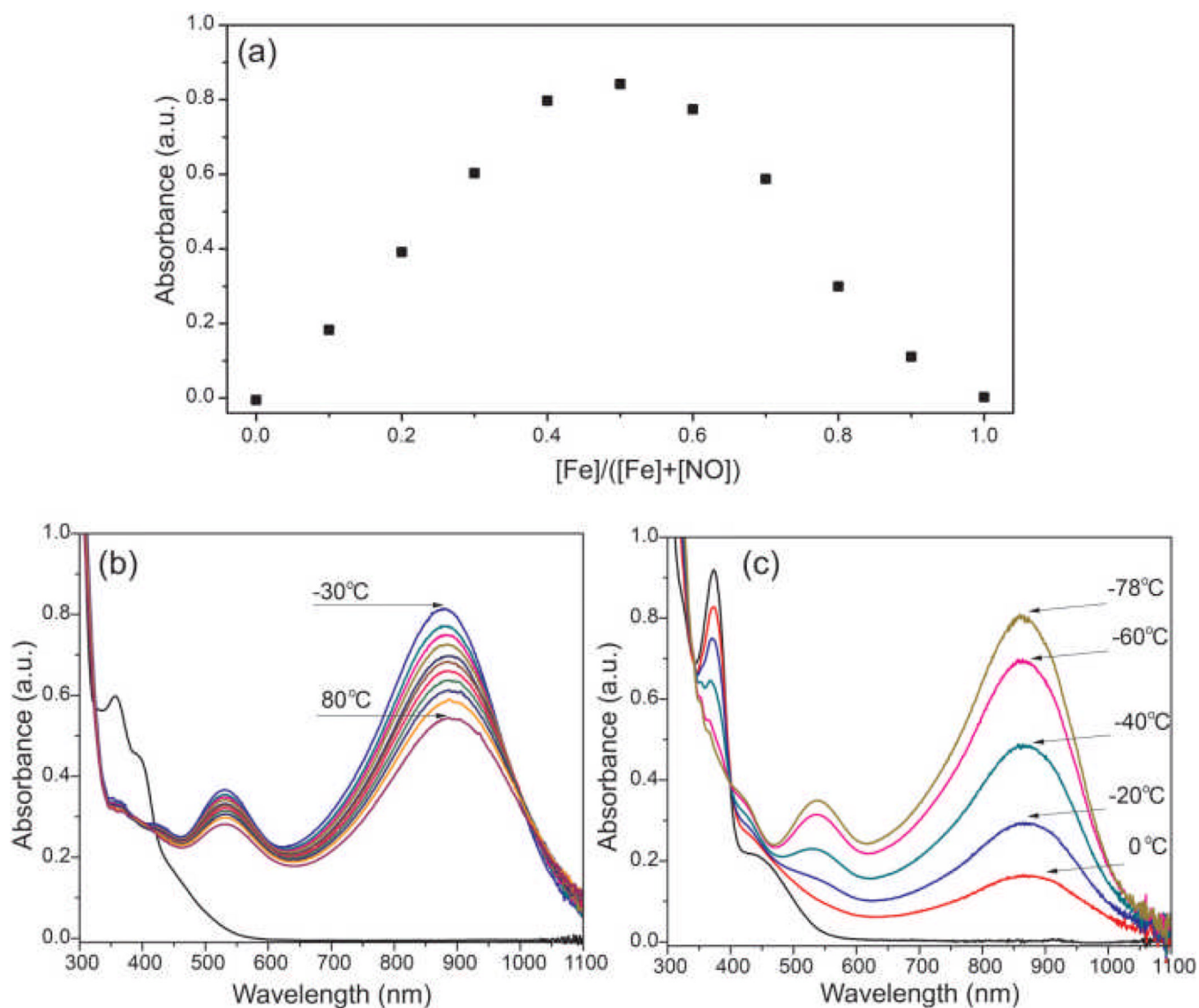


Figure 2.

(a) Job's plot of NO binding to $[\text{Fe}(\text{TAML})]^-$ confirming a 1:1 binding stoichiometry. (b) Variable temperature UV-vis spectrum of 200 μM $[\text{Fe}(\text{TAML})]^-$ and 20 mM NO in 100:1 $\text{CH}_3\text{CN}:\text{MeOH}$. (c) Variable temperature UV-vis spectrum of 200 μM $[\text{Fe}(\text{TAML})]^-$ and 20 mM NO in MeOH.

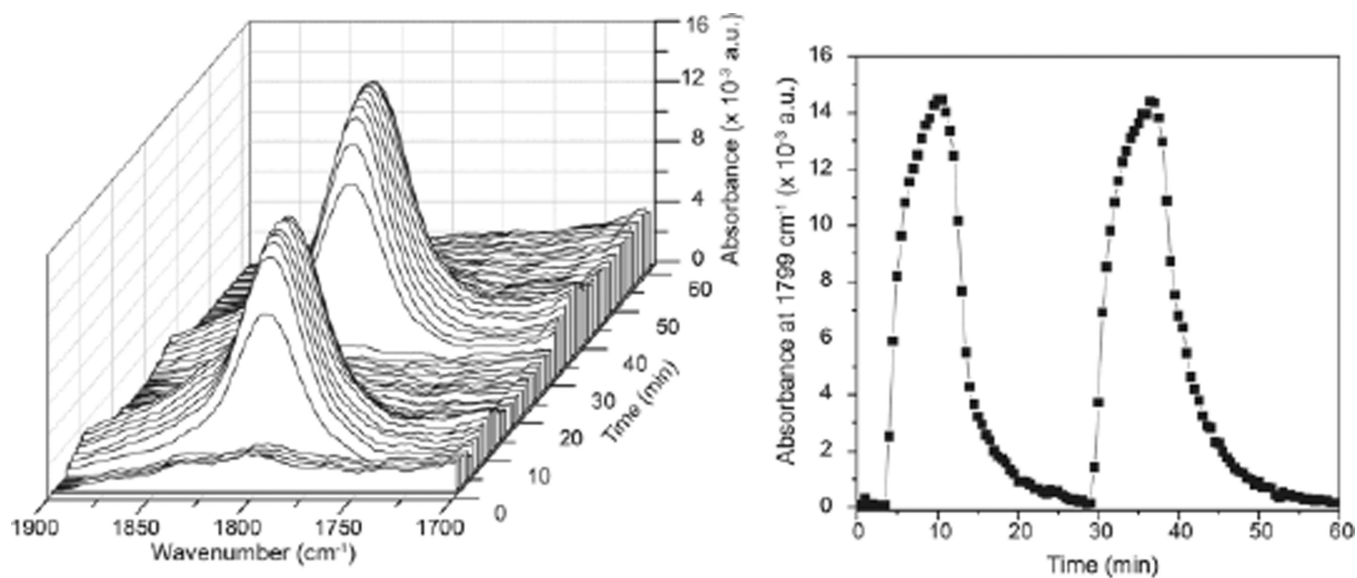


Figure 3.

Left: React-IR trace of the reaction of 20 mM [Fe(TAML)]⁻ with 5 equiv of NO. Two cycles of NO addition and subsequent purging with N₂ are shown. Right: Intensity of the 1799 cm⁻¹ (ν_{NO}) stretch as a function of time. Two cycles of NO/N₂ cycling are depicted.

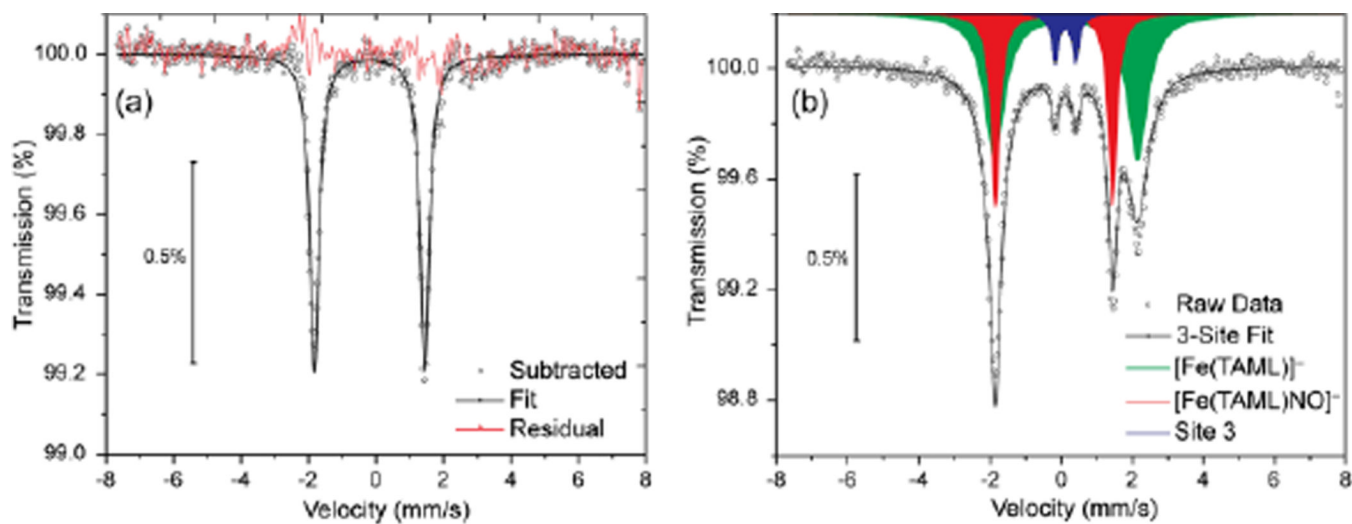
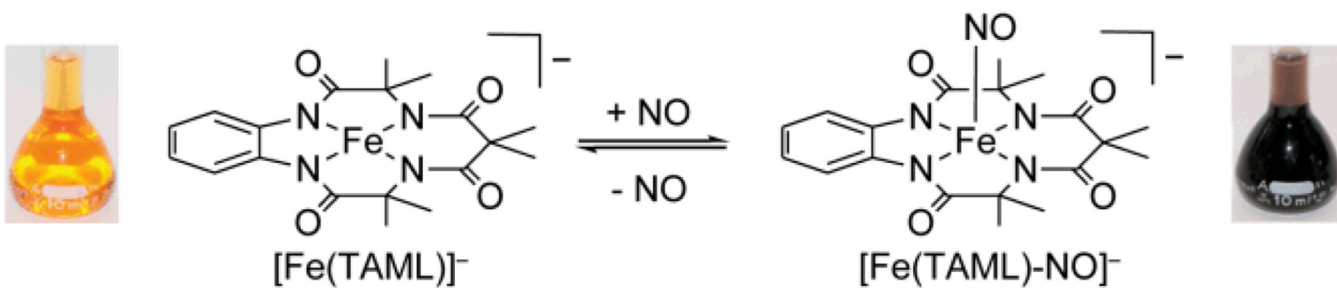


Figure 4.

(a) Isolated spectrum of $[\text{Fe}(\text{TAML})\text{NO}]^-$ after subtraction of the signal from $[\text{Fe}(\text{TAML})]^-$ and a minor impurity. (b) Three site fit of the raw data.



Scheme 5.
Reversible binding of NO to Fe(III) complexes.