

Published in final edited form as:

Tetrahedron. 2012 December 2; 68(48): 10049–10058. doi:10.1016/j.tet.2012.08.044.

Straightforward protocol for the efficient synthesis of varied *N*-acylated (aza)indole 2-/3-alkanoic acids and esters: optimization and scale-up

Andy J. Liedtke^a, Kwangho Kim^b, Donald F. Stec^b, Gary A. Sulikowski^{b,*}, and Lawrence J. Marnett^{a,*}

^aA.B. Hancock Jr. Memorial Laboratory for Cancer Research, Departments of Biochemistry, Chemistry and Pharmacology, Vanderbilt Institute of Chemical Biology, Center in Molecular Toxicology and Vanderbilt-Ingram Cancer Center, Vanderbilt University School of Medicine, Nashville, TN 37232, USA

^bDepartments of Chemistry and Biochemistry, Vanderbilt University, Vanderbilt Institute of Chemical Biology, Nashville, TN 37235 (USA)

Abstract

A library of approximately 40 *N*¹-acylated (aza)indole alkanolic esters and acids was prepared employing a microwave-assisted approach. The optimized synthetic route allows for parallel synthesis, variation of the indole substitution pattern and high overall yield. Additionally, the procedure has been scaled up to yield multi-gram amounts of preferred indole compounds, e.g.: 2'-*des*-methyl indomethacin **2**. The reported compounds were designed as biomedical tools for primary and secondary *in vitro* and *in vivo* studies at relevant molecular targets.

Keywords

Heterocycles; Indoles; (2'-*Des*-methyl) indomethacin; Microwave synthesis; Synthesis scale-up

1. Introduction

N-Acylated indole acetic acids are drawing continued attention due to their wide-ranging pharmacological activities.^{1–5} For example, indomethacin (INDO, **1**, Fig. 1), a traditional non-steroidal anti-inflammatory drug (NSAID), is a potent, time-dependent inhibitor of cyclooxygenases (COX-1 and COX-2) with a long history of clinical use in human subjects.⁶ It promotes anti-inflammatory, anti-pyretic and analgesic activity, but also contributes to severe stomach irritation and ulcers due to secondary pharmacodynamic effects.^{7,8} Recently, **1** and some of its derivatives have been shown to hold interesting and potentially useful 'off-target' activities, e.g. regulation of Th2-mediated asthma and other

© 2012 Elsevier Ltd. All rights reserved.

*Corresponding author 1. Tel.: +1-615-343-7329; fax: +1-615-343-7534; larry.marnett@vanderbilt.edu. *Corresponding author 2. Tel.: +1-615-343-4155; fax: +1-615-343-6361; gary.a.sulikowski@vanderbilt.edu.

Supplementary Material

Detailed synthetic procedures, routine spectroscopic and spectrometric data and HPLC data, as well as exemplified ¹H and ¹³C NMR spectra of intermediates and final compounds are available.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

allergic diseases or chemoprevention of tumors by addressing targets different from COX.^{9–14} Of these INDO derivatives, those that do not (strongly) interfere with arachidonic acid (AA) metabolism, have greater potential to be valuable drug candidates because of their presumably lower gastrointestinal toxicity. Very recently we have probed 2'-*des*-methyl indomethacin (2'-DM-INDO, **2**, Fig. 1),¹⁵ a structurally edited version of **1**, which exhibits drastically reduced inhibitory potency against both COX enzymes and reduced gastric toxicity relative to **1** in C57BL6 mice.^{16,10} To further probe the effects of indoles related to **1** and **2**, we required rapid production of iterative series of *N*-acyl indole alkanonic acids and esters in good overall yield.

In the present manuscript we report a very efficient indolization/acylation approach toward *N*-substituted (aza)indole compounds and we describe the successful scale-up of the synthesis of two prototype indole analogs.

2. Results and Discussion

2.1. Small-scale approach

We accomplished a robust synthetic routine and generated a series of novel INDO (**1**) analogues **18–31** (Tables 2 and 3) with individual compounds emerging in milligram or multi-gram quantities and good overall yield (>55%). The general reaction sequence leading to indoles **18** through **31**, followed three simple steps: (a) Fischer indolization of substituted phenylhydrazine hydrochlorides condensed with selected ketoacids/esters, (b) base-catalyzed indole *N*¹-acylation reactions (*S*_N2 substitutions) with different acid chlorides, and (c) ester hydrolysis of the intermediate methyl or cleavage of allyl *N*-acyl indole alkanooates in the presence of a metal organic catalyst. Structural modifications with respect to **1** (and **2**) were made at the C², C³, C⁵, COOH and *N*¹-position of the indole ring. Previous procedures using similar synthetic strategies have not been reported on large scale and oftentimes proceed in low overall yield.

Several creative methods for assembly of *N*¹-substituted indole compounds have been reported. These include one-pot Stille couplings of *N*-acyl-2-iodoanilines,¹⁷ catalytic dehydrogenative *N*¹-couplings of indoles with primary alcohols,¹⁸ distinct indolization reactions with cyclic enol ethers and enol lactones,¹⁹ and variations of the Fischer Indole synthesis.²⁰

In the refined standard procedure of Yamamoto for compound **1** (Scheme 1),^{21,22} an arylhydrazone precursor **4** is benzoylated at the *N*¹ position to give the *N*¹,*N*¹-disubstituted hydrazine **5**. Typically, commercial hydrazine hydrochloride starting materials **3** need to be deprotonated prior to reaction. The intermediate **5** is then hydrolyzed with hydrogen chloride to give the hydrazine hydrochloride salt **6**, which is cyclized in the final reaction step with levulinic acid to afford **1**.²³ The synthesis of 2'-*des*-methyl INDO **2** has been conducted accordingly with the exception that succinic semialdehyde was used for the ring-closure.

Isolated product yields are often unsatisfactory using this literature procedure due to multiple reaction steps, the formation of by-products, and the requirement for extensive purification of intermediates.¹⁹ The early insertion of the *N*¹-acyl moiety in the reaction sequence is another limitation of this entry, which involves the addition and subsequent displacement of an ethylidene protection group (*N*²). Therefore, in order to vary the *N*-acyl substituent for a series of indole analogues **18–21** (Table 2), each derivative needed to be constructed individually. To overcome this limitation and enhance the flexibility toward the introduction of multiple substituents in the indole scaffold, we pursued a strategy (Scheme 2) that has not been intensively investigated for the serial production of substituted indoles because it has proven to be rather uneconomical, especially for larger scales. This strategy

employs indolization first followed by late-stage N^1 -substitution and release of the carboxylic acid under optimized reaction conditions. Cyclization reactions of arylhydrazines with different ketoacids/esters were conducted in AcOH or refluxing methanol/ H_2SO_4 . We used methyl or allyl esters for our small-scale and large-scale reactions, respectively, because of their stability to the reaction conditions and ease of removal. Utilization of *t*-BuONa in THF or DMAP/ Et_3N in DCM as reaction medium facilitated indole *N*-acylation with select aryl acid chlorides. Final ester cleavage was done in 1,2-DCE with trimethyltin hydroxide²⁴ (methyl esters) or in THF using Pd(PPh_3)₄/morpholine as catalyst complex (allyl esters). Concurrent *N*-deacylation was not observed with these expedient procedures, which clearly privileges them over standard saponification methods with metal alkaline hydroxide/water. We also investigated the synthesis under microwave irradiation for numerous compounds with a view to speed up reaction rates and increase isolable yields.

2.1. 1. Preparation of different indole methyl-alkanoates—Initially, we generated a set of 10 different $C^2/C^3/C^5$ -substituted (aza)indole intermediates. The structures, specific conditions and yields are summarized in Table 1. Detailed experimental data can be also found in Supporting Information.

Methyl esters **9a**, and **9b** of commercial 5-methoxy-2-methylindole-3-acetic acid **7a**, and 5-methoxyindole-3-acetic acid **7b** were generated overnight at room temperature in yields >85% using BOP-Cl/triethylamine and methanol (Scheme 2).²⁵

The preparation of different C^2/C^3 -substituted indole compounds **9c-e**, **14** and **15** is summarized in Scheme 3. Substituted aryl hydrazine hydrochlorides **3** were directly subjected to reaction with selected ketoacids or esters **11** through **13**. Particularly, indole compound **15a** was generated in 90% yield under standard reaction conditions (AcOH, 80 °C, 3h) using 4-methoxyphenyl hydrazine hydrochloride **3a** and methyl 4-ketocyclohexanecarboxylate. Next, in order to allow investigation of the significance of a fluoro substitution at the indole 5-position, we pursued Fischer indole synthesis of C^2/C^3 -substituted indole analogues **9c-e**, **14** and **15b**. Whereas *para*-methoxylated phenylhydrazine hydrochlorides **3a** (e.g. for compound **15a**) readily dissolved in AcOH upon heating, this was not the case for fluorinated starting materials **3b** (leaving unreacted starting materials). Therefore, AcOH was replaced by methanol/ H_2SO_4 as solvent mixture for better solubility of the fluorinated precursors. In this case, selected 'ketoacids' rather than their low alkyl esters were employed in the reaction, as clean COOH-esterification usually occurred alongside the 'indole' ring-closure (compare experimental section for **9c** vs. **9e**). Reactions were accelerated by microwave irradiation and ran approx. 50 times faster (5 to 10 min instead 3h to 6h) than with conventional heating affording consistently high and reproducible yields (~80%). In many cases products precipitated upon pouring the reaction mixture on crushed ice or could be extracted from the resulting aqueous mixture, washed and obtained in acceptable quality after removal of solvent *in vacuo* and trituration with hexanes.

The same results were achieved when using keto acids whose carboxylic acid group had been esterified beforehand (see experimental details for **9c**). (Partial) re-esterification to the corresponding methyl esters **9** even occurred when starting for example with ethyl esters of **11** (data not shown). For compound **9d** bearing an extended exocyclic alkyl chain on position C^3 of the indole, 4-F-phenylhydrazine hydrochloride was converted with 5-oxohexanoic acid under the conditions described. Finally a 2-propionic acid derivative was realized with indole **14**, using 4-oxohexanoic acid as cyclization reagent. In this case, however, the desired product could be isolated only in 46% yield.

Not only did this reaction eliminate the need for an *N*-protection, it also allowed the variation of all relevant substituents R^1 thru R^3 as well as the length and position of the

carboxyalkyl chain at the indole framework within only one reaction step. Generally, linear ketones **11** or **12** were of importance for the later configuration of the carboxy alkyl substituted indoles. Certain 'keto' alkanolic acids/esters provided high regioselectivity in the 3,3-sigmatropic rearrangement in the course of the Fischer indole synthesis. In the case of **11** (acetyl or formyl tail end), the alkyl acid functionality was solely directed to position 3 of the indole product, whereas reagents **12** (ethylcarbonyl tail end) typically produce the alkyl acid functionality at position 2 (isolated products).²⁶ Structure examples are given in Scheme 2 and Table 1.²⁶ Elongation of the alkyl chain in between the two functional groups with respect to reagents **11** was tolerated and didn't affect the selectivity (e.g. **9d**). On the contrary, the inclusion of one or more extra methylene units into the terminal alkyl chain (beyond carbonyl) and/or extension of the alkyl spacer (between CO and COOH) of **12** continuously yielded isomeric mixtures (C²/C³). Such compounds are not part of the current work.

Poor regioselectivity is a general problem of Fischer indole reactions as evidenced in the use of unsymmetrical cyclic ketones. Interestingly, there have been a few examples for the selective cyclization of phenylhydrazones with e.g. 3-ketocyclo-hexanecarboxylic acid or 5-aryldihydro-3(2*H*)-thiophenones.^{27,28} To simplify matters in our present study we used symmetrical 4-ketocyclohexane-carboxylic acid to produce indole derivatives **15a** (6'-methoxy, see above) and **15b** (6'-fluoro) and to demonstrate the later availability for use of similar intermediates in subsequent *N*-acylation/ester cleavage reactions. In practice, indole derivative **15b** could be isolated quantitatively after short reaction times using microwave-aided synthesis.

In order to demonstrate the feasibility of additional heteroatoms and to modify the hydrophilicity of the core skeletons, we also produced a 2-aza (**10**) and a 4-aza indole-3-acetic acid methyl ester analogue (**17**) (Schemes 2 and 4). The reaction for **10** was completed in 77% yield according to a patent method starting from 2-(1*H*-indazol-3-yl)acetic acid (14h reflux in methanol/H₂SO₄).²⁹ In this case the originally attempted esterification with BOP-Cl/methanol did not succeed. Another literature method allowed for the synthesis of the 4-aza indole derivative **17**.^{30,31} In contrast to the published procedure, we adopted the reaction under microwave irradiation with the respective methyl ester. Customized one-pot catalytic reduction/cyclization of a 3-cyano-3-(nitropyridin-2-yl)-methyl propionate precursor **16** in methanol using Pd/C and cyclohexadiene as hydrogen source (10 min at 120 °C) readily afforded **17** in 60% yield.

2.1. 2. Substitution at nitrogen (N¹)—*N*-substitution of indoles, **9** and **15**, yielded a series of *N*-acyl-indole methyl alkanooates, **18** and **19**, respectively (Table 2). Different benzoyl/aryl acid chlorides were chosen as acylating reagents. However, direct *N*-acylation of an indole nucleus is demanding and necessitates deprotonation of the N-H center, as neutral indoles are non nucleophilic at the nitrogen atom. Due to the low acidity of the indole-N¹, full deprotonation to the reactive anion can be only realized under strong base catalysis in (polar) aprotic solvents.

For the small-scale one-pot reactions on indole **9** we first tried sodium hydride (powdered material) in DMF, though, with less satisfying results due to several handling issues (e.g. with compounds **18g**, **18k** or **18l**). Then we switched to sodium *tert*-butoxide and used a 2M commercial solution in THF to produce a larger series of compounds **18–21** having different benzoyl (*p*-Cl, *p*-CH₂Cl, *p*-F, *p*-Me, *p*-OMe, *m/p*-CF₃) and other acyl moieties (Scheme 5, first step). Not only did we observe higher yields and reproducibility with this base (yields ranging from 20–60% for numerous analogues), but also total reaction processing and workup were cleaner than with NaH/DMF (pipetting volumes instead of weighing moisture-sensitive solids; phase separation after water quench). Reactions were run on a small scale

(80–100 mg each) under argon on a parallel synthesis apparatus. *N*-Acylation with different acid chlorides occurred best at temperatures between 0 °C and 25 °C. Therefore, in a typical experiment, we pipetted the base to an ice-cold stirred solution of the indole in THF and after 20 min we added the acid chloride and let the mixture react at ambient temperature until the starting material was consumed or no further product formation could be detected (usually overnight). Progress of the reactions was monitored by LC-MS and TLC. In most cases, one major product was detected. By-products were predominantly unreacted or *N*-deacylated/hydrolyzed indole starting materials and the carboxyl counterparts of the applied benzoyl chlorides.

Isolated amounts of products with varied *meta*- and *para*-substituted *N*-benzoyl moieties appeared to be better (24–57%), when a 2-methyl group was present on the indole ring (**18a-f** and **18m-o**) and were lower (13–29%) among the 2-*des*-methyl indole analogues (**18g-k**), except for **18q** (42%). In compounds **18k** and **18p** the acyl substituent was ‘extended’ by an additional methylene unit, which further reduced the percent yields [**18k**: 13% (impure) and **18q**: 4%, respectively]. A simple aliphatic *N*-propanoyl substituent was realized with indole acetic acid ester analogue **18l**. C²/C³-cyclized indole intermediates, **12a** and **12b** could be substituted on their nitrogens with an *m*-CF₃- and *p*-Cl-benzoyl substituent in 24 and 14% yield, respectively (**19a**, **19b**). Acylation of an N¹, C², C³-unsubstituted 5-methoxy indole precursor using sodium *tert*-butoxide and *p*-Cl-benzoyl chloride afforded a mixture of three compounds, two of which were isomers with the benzoyl moiety on either position C³ (faint, not isolated) or N¹ (**20a**, 25%). The other isolated product **20b** (8%) was analyzed to be an N¹/C³-dibenzoylated indole derivative. These reactions reflect the enhanced reactivity of the unsubstituted indole 3-position upon hydrogen abstraction at the nitrogen *via* the conjugated double-bond system (conjugate-like substitution) and in turn the facilitated N¹-deprotonation reaction by the 3-acyl ring substituent.

The initial attempt to acylate a 2-propionic acid ester indole derivative (e.g., **11**) with 4-chlorobenzoyl chloride using similar conditions failed and considerable amounts of starting material were detected beside some undefined products by LCMS. Nevertheless, *N*-substitution of a comparable indole derivative was successfully followed up later under larger scale and with optimized reaction conditions (compound **30**, see below).

Lastly, compound **21** exemplifies an N¹-benzoylated 2-aza-indole acetic acid methyl ester emerging in 37% yield from the parallel routine described above.

2.1.3. Methyl Ester Cleavage—In order to obtain the free acid derivatives **2**, **22**, **23**, and **24** (Table 3) of the *N*-acylated indole compounds **18**, **19**, and **21**, the methyl ester groups had to be hydrolyzed. Different standard ‘saponification’ methods exist to achieve this task, however common drawbacks include elimination reactions induced by the aqueous-basic conditions.³² This circumstance became particularly significant, as the *N*-acyl bonds of our indole derivatives were perceptibly fragile toward basic (and strong acidic) treatment and easily hydrolyzed to liberate *N*-unsubstituted indole alcanoic acids. This was true for numerous 2-*des*-methyl indole analogues and also for the sterically more demanding 2-propionic acid derivatives (see below), notwithstanding a limited number of literature instances for successful ester saponification toward *N*-acyl 2-methyl indole acetic acids.²⁰

We attempted a recently reported non-aqueous metal catalytic method for the mild and selective hydrolysis of esters with trimethyltin hydroxide.²⁴ It had been illustrated for many (complex) aromatic and aliphatic systems that methyl, allyl and benzyl esters are hydrolyzed in quantitative yields using this reagent. For our current study, we once more transferred the stated settings to microwave conditions (for detailed parameters see experimental part) and consistently applied 5 equivalents of Me₃SnOH beside our starting methyl esters in 1,2-

dichloroethane for the expedited ester hydrolysis. Reactions normally ran to completion within 20 to 30 min (IPC by TLC and LC-MS) and workup was quite simple, as products could be extracted into dichloromethane after addition of 50% acetic acid. Residual Me₃SnOH was removed with water or brine; the organic layer was dried over Na₂SO₄, filtered and concentrated. Flash chromatography on silica gel afforded the pure *N*-acyl indole alkanolic acids **2**, **22** and **23** quantitatively. As shown in Table 3, this technique proved truly valuable in getting the selective and high-yielding (76–99%) hydrolysis of methyl esters within the sensitive substrates **18** and **19**. Unfortunately, this procedure failed with the 2-aza derivative **21** to obtain the corresponding free acid derivative **24**. The latter compound repeatedly decomposed into several unidentified products, regardless of whether the reaction was conducted with microwaves or under conventional heating conditions (60–80 °C).

To extend this strategy to the large-scale synthesis of individual *N*-acyl indole derivatives, we further optimized the reaction conditions as described below, particularly with a view to improving capacities of the *N*-acylation step.

2.2. Large-scale synthesis

New synthetic methodology was adopted for the scale-up to multi-gram scale. Beside the efforts with *N*-substitution, the major problematic step in the synthesis remained the final hydrolysis of the ester in the presence of the *N*-acyl functional group. As mentioned above, the *N*-benzoyl bond is labile under basic conditions, so it was hydrolyzed easily at normal hydrolysis condition for the ester hydrolysis step.³² Therefore we considered using allyl,^{33,34} benzyl,^{35,36} and *tert*-butyl ester³⁷ to overcome the side reaction and allyl ester **26** was evaluated.

A large quantity of **2** was prepared from the commercially available 2-(5-methoxy-1*H*-indol-3-yl)acetic acid **7b** in three steps starting with esterification with allyl bromide to afford allylester **25** in 75% yield (Scheme 6). Efficient acylation of **25** followed by treatment with triethylamine and DMAP then gave *N*-benzoate **26** (92% yield). Subsequently, cleavage of the allyl ester was examined. Optimal conditions employed tetrakis(triphenylphosphine) palladium in combination with morpholine in THF to quantitatively provide **2** without unwanted hydrolysis of the *N*-acyl group.³⁸

From the result of the synthesis for **2**, allyl ester was projected as an advanced intermediate in our synthesis so it was applied to another large-scale synthesis of a 2-propionic acid derivative **30** *en route* to a more complex sulfonimide derivative (COOH-bioisostere) **31**.

Indole **28** was not commercially available and therefore should be prepared from succinyl chloride using Boger's synthesis.³⁹ Succinyl chloride was treated with ethyldichloroaluminum and allyl alcohol in dichloromethane to provide allyl-4-oxohexanoate **27**. Fischer indolization of **27** with 4-methoxyphenyl hydrazine followed by treatment with acetic acid then gave indole compound **28** in 71%. Acylation (87%) and ester cleavage (79%) produced carboxylic acid, **30**.¹⁵ Finally compound **31** was obtained by amide coupling between methylsulfonylamine and acid **30** using CDI coupling reagent.

3. Conclusions

In summary, we have found that different INDO analogues, including 2'-*des*-methyl INDO **2** are efficiently prepared in a parallel approach *via* their *N*-unsubstituted indole methyl-/allylalkanoate intermediates starting from commercially available arylhydrazine hydrochlorides and a variety of ketoacids/esters. Substituted phenylhydrazine precursors were subjected to Fischer cyclization in a one-pot procedure to provide respective indole

products. The *N*-unsubstituted indole alcanoic acid esters (optimally methyl or allyl esters) were deprotonated and further elaborated by *N*-acylation followed by aqueous-free ester cleavage in the presence of different metal-based catalyst systems. Concomitant *N*-deacylation was not observed with the applied technique. The synthetic strategy enabled assembly and variation of all relevant substituents within only three reaction steps. Target compounds were readily formed under optimized settings and could be isolated in good to high overall yields and in acceptable analytical quality. The procedure also allowed for scale-up to afford large amounts of selected compounds of interest for further derivatization reactions and/or direct use in *in vitro* and *in vivo* experiments. The range of indole analogs will be evaluated for activity against peroxisome proliferator activated receptor γ , induction of tumor cell apoptosis, and inhibition of aldo-keto reductase enzymes.^{10,40}

4. Experimental part

All commercial reagents, solvents and other materials were used as received without further purification. Flash chromatography was conducted on a Biotage SP1 automated flash chromatography system equipped with a fixed wavelength UV detector ($\lambda = 254$ nm) using prefabricated 'Flash KP-SIL' columns (size according to requirements). Thin-layer chromatography was performed on precoated fluorescent silica gel 60 F₂₅₄ plates (250 μ m) from Whatman (Partisil® LK6D, Cat. No. 4865-821). Spots were visualized under natural light, and UV illumination at $\lambda = 254$ and 365 nm. Microwave reactions were performed in an automated Biotage Initiator Eight Synthesizer. NMR spectra were recorded with a Bruker AV-400 instrument with sample changer (BACS 60) (400 MHz for ¹H, 100 MHz for ¹³C) or a Bruker AV-300 system (282 MHz for ¹⁹F) and calibrated with DMSO-*d*₆ as solvent and TMS as internal standard signal. Chemical shifts (δ) are reported in parts per million (ppm). Coupling constants (*J*) are given in Hz. Low resolution mass spectra (LCMS) were obtained on an Agilent 1200 LCMS system with electrospray ionization. High resolution mass spectra (HRMS) were recorded on a Waters QToF-API-US plus Acquity system with ES as the ion source. Analytical high pressure liquid chromatography (HPLC) was performed on an Agilent 1200 analytical LCMS with UV detection at 214 nm and 254 nm along with ELSD detection. General methods are specified in the Supporting Information. Compounds **9a**, **9b**, **10** and **17** were prepared according to literature procedures.^{25,31,41,30}

Synthesis of exemplified intermediates and target compounds:

4.1. Methyl 2-(5-fluoro-1H-indol-3-yl)acetate **9e**

According to general procedure B_variant2 (conventional heating), (4-fluorophenyl)hydrazine hydrochloride (100 mg, 0.62 mmol), 4-oxobutanoic acid (79 mg, 0.68 mmol) and 160 μ L H₂SO₄ in 2 mL methanol were refluxed for 3 h under argon to afford 110 mg (86%) of **9e** as brownish viscous mass. C₁₁H₁₀FNO₂, *M_r* = 207.20; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 3.60 (s, 3H), 3.72 (s, 2H), 6.91 (td, *J*=2.6/9.2 Hz, 1H), 7.22 (dd, *J*=2.4/10.0 Hz, 1H), 7.31–7.35 (m, 2H); ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ : -123.52 (5' -F); LCMS (ESI) *t_R*: 2.16 min (>99%, ELSD), *m/z*: 208.2 [M+H]⁺; HRMS (TOF, ES+) C₁₁H₁₀FNO₂ [M+H]⁺ calc. mass 208.0774, found 208.0772.

4.2. Methyl 2-(1H-indazol-3-yl)acetate **10**

According to general procedure A_variant2, 2-(1*H*-indazol-3-yl)acetic acid (300 mg, 1.70 mmol) was refluxed in methanol (15 mL) for 14 h. The reaction mixture was worked up as described to afford 250 mg (77%) of **10**. C₁₀H₁₀N₂O₂, *M_r* = 190.20; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 3.62 (s, 3H), 4.01 (s, 2H), 7.09 (td, *J*=0.8/7.4 Hz, 1H), 7.33 (td, *J*=1.0/7.7 Hz, 1H), 7.48 (d, *J*=8.4 Hz, 1H), 7.69 (d, *J*=8.4 Hz, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 33.04 (s, -CH₂-), 52.17 (s, -OCH₃), 110.49 (s, C7'), 120.29 (s), 120.43 (s), 122.20 (s, C4a'),

126.40 (s, C6'), 138.83 (s, C7a'), 141.20 (s, C3'), 171.03 (s, >C=O); LCMS (ESI) *t*R: 1.65 min (>99%, UV254), *m/z*: 191.2 [M+H]⁺; HRMS (TOF, ES+) C₁₀H₁₀N₂O₂ [M+H]⁺ calc. mass 191.0821, found 191.0821.

4.3. Methyl 3-(5-fluoro-3-methyl-1H-indol-2-yl)propanoate 14

According to general procedure B_variant2, the title compound was obtained from (4-fluorophenyl)hydrazine hydrochloride (100 mg, 0.62 mmol), 4-oxohexanoic acid (88.8 g, 0.68 mmol) and 80 μL H₂SO₄ in 3 mL methanol after 10 min at 120 °C in 46% yield (67 mg) as brownish viscous mass. C₁₃H₁₄FNO₂, M_r = 235.25; ¹H NMR (400 MHz, DMSO-*d*₆) δ: 2.12 (s, 3H), 2.66 (t, *J*=8.0 Hz, 2H), 2.93 (t, *J*=7.6 Hz, 2H), 3.58 (s, 3H), 6.80 (td, *J*=2.8/9.2 Hz, 1H), 7.10 (dd, *J*=2.4/10.2 Hz, 1H), 7.20 (dd, *J*=4.6/8.6 Hz, 1H), 10.74 (bs, 1H); ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ: -124.03 (d, 5'-F); LCMS (ESI) *t*R: 2.58 min (>99%, UV220), *m/z*: 236.2 [M+H]⁺; HRMS (TOF, ES+) C₁₃H₁₄FNO₂ [M+H]⁺ calc. mass 236.1087, found 236.1085.

4.4. Methyl 6-methoxy-2,3,4,9-tetrahydro-1H-carbazole-3-carboxylate 15a

According to general procedure B_variant1, the title compound was obtained from (4-Methoxyphenyl)hydrazine hydrochloride (80.0 mg, 0.46 mmol) and methyl 4-oxocyclohexanecarboxylate (85.9 mg, 0.55 mmol) in 0.5 mL glacial acetic acid after 3 h at 80 °C in 90% yield (107 mg) as an off-white solid. C₁₅H₁₇NO₃, M_r = 259.30; ¹H NMR (400 MHz, DMSO-*d*₆) δ: 1.81–1.91 (m, 1H), 2.15–2.19 (m, 1H), 2.66–2.84 (m, 4H), 2.90 (dd, *J*=4.6/14.2 Hz, 1H), 3.65 (s, 3H), 3.72 (s, 3H), 6.61 (dd, *J*=2.4/8.8 Hz, 1H), 6.86 (d, *J*=2.4 Hz, 1H), 7.11 (d, *J*=8.8 Hz, 1H), 10.51 (s, 1H); LCMS (ESI) *t*R: 2.27 min (>99%, ELSD), *m/z*: 260.2 [M+H]⁺; HRMS (TOF, ES+) C₁₅H₁₇NO₃ [M+H]⁺ calc. mass 260.1287, found 260.1285.

4.5. Methyl 2-(5-methoxy-1H-pyrrolo[3,2-b]pyridin-3-yl)acetate 17

A 2 mL microwave process vial with a stir bar was charged with crude methyl 3-cyano-3-(6-methoxy-3-nitropyridin-2-yl)propanoate **16** (50 mg, 0.19 mmol), 10% Pd/C (5 Mol%, 20 mg, 0.01 mmol), and methanol (1.5 mL). An excess of 1,4-cyclohexadiene (91 mg, 1.13 mmol) was added and the vessel flooded with argon, capped and heated under microwave conditions at 120 °C for 5 min. The reaction was filtered through Celite® and the solvent was evaporated *in vacuo*. The crude material was purified by flash chromatography (SiO₂, ethyl acetate/hexane gradient) to yield the product as greenish oil (25 mg, 60%). C₁₁H₁₂N₂O₃, M_r = 220.22; ¹H NMR (400 MHz, DMSO-*d*₆) δ: 3.61 (s, 3H), 3.72 (s, 2H), 3.83 (s, 3H), 6.53 (d, *J*=8.8 Hz, 1H), 7.38 (d, *J*=2.8 Hz, 1H), 7.66 (d, *J*=8.4 Hz, 1H), 11.01 (bs, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 29.41 (s, -CH₂-), 51.86 (s, -C(O)OCH₃), 52.83 (s, -OCH₃), 104.86 (s, C6'), 107.19 (s, C3'), 122.78 (s, C2'), 124.60 (s, C7a'), 126.80 (s, C7'), 141.29 (s, C3a'), 159.16 (s, C5'), 172.47 (s, >C=O); LCMS (ESI) *t*R: 0.69 min (>97%, UV220, ELSD), *m/z*: 221.2 [M+H]⁺; HRMS (TOF, ES+) C₁₁H₁₂N₂O₃ [M+H]⁺ calc. mass 221.0926, found 221.0926.

4.6. Methyl 2-(5-methoxy-2-methyl-1-(4-methylbenzoyl)-1H-indol-3-yl)acetate 18d

According to general procedure C_variant2, the title compound was obtained from from methyl 2-(5-methoxy-2-methyl-1H-indol-3-yl)acetate (80 mg, 0.34 mmol), 4-methylbenzoyl chloride (63.6 mg, 0.41 mmol) and ^tBuONa (2 M in THF, 206 μL, 0.41 mmol) in anhydrous THF (2.5 mL). The crude residue was subjected to flash chromatography (SiO₂, ethyl acetate/hexane gradient) to afford the pure title compound in 57% yield (68.5 mg). C₂₁H₂₁NO₄, M_r = 351.40; ¹H NMR (400 MHz, DMSO-*d*₆) δ: 2.22 (s, 3H), 2.42 (s, 3H), 3.62 (s, 3H), 3.74 (s, 3H), 3.77 (s, 2H), 6.68 (dd, *J*=2.6/9.0 Hz, 1H), 6.85 (d, *J*=8.8 Hz, 1H), 7.01 (d, *J*=2.4 Hz, 1H), 7.37 (d, *J*=8.0 Hz, 2H), 7.54 (pseudo-d, *J*=8.4 Hz, 2H); LCMS (ESI)

t_R : 2.74 min (>95%, ELSD), m/z : 352.2 [M+H]⁺; HRMS (TOF, ES+) C₂₁H₂₁NO₄ [M+H]⁺ calc. mass 352.1549, found 352.1548.

4.7. Methyl 3-(1-(4-chlorobenzoyl)-5-fluoro-2-methyl-1H-indol-3-yl)propanoate 18o

According to general procedure C_variant2, the title compound was obtained from methyl 3-(5-fluoro-2-methyl-1H-indol-3-yl)propanoate (80 mg, 0.34 mmol), 4-chlorobenzoyl chloride (71.4 mg, 0.41 mmol) and ^tBuONa (2 M in THF, 204 μ L, 0.41 mmol) in anhydrous THF (2.5 mL). The crude residue was subjected to flash chromatography (SiO₂, ethyl acetate/hexane gradient) to afford the pure title compound as yellow oil in 49% yield (62 mg). C₂₀H₁₇ClFNO₃, M_r = 373.81; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 2.19 (s, 3H), 2.59 (t, J =7.4 Hz, 2H), 2.92 (t, J =7.6 Hz, 2H), 3.56 (s, 3H), 6.94 (td, J =2.4/9.2 Hz, 1H), 7.08 (dd, J =4.6/9.0 Hz, 1H), 7.39 (dd, J =2.4/9.2 Hz, 1H), 7.63–7.68 (m, 4H); LCMS (ESI) t_R : 3.21 min (>95%, ELSD), m/z : 374.3 [M+H]⁺; HRMS (TOF, ES+) C₂₀H₁₇ClFNO₃ [M+H]⁺ calc. mass 374.0959, found 374.0962.

4.8. Methyl 6-methoxy-9-(3-(trifluoromethyl)benzoyl)-2,3,4,9-tetrahydro-1H-carbazole-3-carboxylate 19a

According to general procedure C_variant2, the title compound was obtained from methyl 6-methoxy-2,3,4,9-tetrahydro-1H-carbazole-3-carboxylate (80 mg, 0.31 mmol), 3-(trifluoromethyl) benzoyl chloride (77.2 mg, 0.37 mmol) and ^tBuONa (2 M in THF, 185 μ L, 0.37 mmol) in anhydrous THF (2.5 mL). The crude residue was subjected to flash chromatography (SiO₂, ethyl acetate/hexane gradient) to afford the pure title compound in 24% yield (32 mg). C₂₃H₂₀F₃NO₄, M_r = 431.40; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 1.70–1.80 (m, 1H), 2.03–2.07 (m, 1H), 2.47 (m, 2H, partly overlaid by DMSO signal), 2.73–2.79 (m, 1H), 2.82–2.89 (m, 1H), 2.94 (dd, J =5.0/15.8 Hz, 1H), 3.65 (s, 3H), 3.77 (s, 3H), 6.73 (dd, J =2.6/9.0 Hz, 1H), 7.04 (d, J =2.4 Hz, 1H), 7.08 (d, J =9.2 Hz, 1H), 7.79 (t, J =8.0 Hz, 1H), 7.93 (d, J =8.0 Hz, 1H), 7.99 (s, 1H), 8.04 (d, J =8.0 Hz, 1H); ¹⁹F NMR (282 MHz, DMSO-*d*₆) δ : –59.41 (*m*-CF₃); LCMS (ESI) t_R : 0.95 min (>95%, UV254), m/z : 432.0 [M+H]⁺; HRMS (TOF, ES+) C₂₃H₂₀F₃NO₄ [M+H]⁺ calc. mass 432.1423, found 432.1422.

4.9. Methyl 2-(1-(4-chlorobenzoyl)-1H-indazol-3-yl)acetate 21

According to general procedure C_variant2, methyl 2-(1H-indazol-3-yl)acetate (80 mg, 0.42 mmol) was subjected to reaction with 4-chlorobenzoyl chloride (88.3 mg, 0.50 mmol) (252 μ L ^tBuONa). The crude product was purified by flash chromatography (SiO₂, ethyl acetate/hexane gradient) to afford 13.5 mg (52%) of the title compound as bright yellow solid. The compound permanently crystallized upon drying at high vacuum and storage at –20 °C. Yield: 51 mg (37%). C₁₇H₁₃ClN₂O₃, M_r = 328.75; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 3.65 (s, 3H), 4.16 (s, 2H), 7.50 (td, J =0.8/7.6 Hz, 1H), 7.63–7.66 (m, 2H), 7.71 (td, J =1.0/7.6 Hz, 1H), 7.91 (d, J =8.0 Hz, 1H), 8.00–8.03 (m, 2H), 8.41 (d, J =8.4 Hz, 1H); LCMS (ESI) t_R : 2.80 min (>99%, ELSD), m/z : 329.0 [M+H]⁺; HRMS (TOF, ES+) C₁₇H₁₃ClN₂O₃ [M+H]⁺ calc. mass 329.0693, found 329.0693.

4.10. 2-(1-(4-(chloromethyl)benzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)acetic acid 22e

According to general procedure D, methyl 2-(1-(4-(chloromethyl)benzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)acetate (15 mg, 0.039 mmol) was subjected to reaction with trimethyltin hydroxide (35 mg, 0.194 mmol) in 1,2-DCE (1 mL). The crude product was chromatographed on a short silica gel column using 1:1 ethyl acetate/hexane (0.5% AcOH) mixtures of increasing polarity as eluants to afford the title compound quantitatively as off-white solid. Yield: 14.1 mg (98%). C₂₀H₁₈ClNO₄, M_r = 371.81; ¹H NMR (400 MHz, DMSO-*d*₆) δ : 2.19 (s, 3H), 3.65 (bs, 2H), 3.75 (s, 3H), 4.88 (s, 2H), 6.68 (dd, J =2.4/9.2 Hz, 1H), 6.90 (d, J =8.8 Hz, 1H), 7.02 (d, J =2.4 Hz, 1H), 7.61–7.67 (m, 4H); ¹³C NMR (125

MHz, DMSO- d_6) δ : 13.34 (s), 29.93 (s), 45.23 (s), 55.72 (s), 101.14 (s), 111.68 (s), 115.08 (s), 128.79 (s, 2C), 130.14 (s, 2C), 130.41 (s), 130.85 (s), 135.38 (s), 136.27 (s), 142.26 (s), 156.02 (s), 168.79 (s), 176.3 (s), one signal invisible; LCMS (ESI) t_R : 2.48 min (>99%, UV220, UV254), m/z : 372.0 [M+H]⁺; HRMS (TOF, ES+) C₂₀H₁₈ClNO₄ [M+H]⁺ calc. mass 372.1003, found 372.1003.

4.11. 2-(1-(4-chlorobenzoyl)-5-fluoro-1H-indol-3-yl)acetic acid 22n

According to general procedure D, methyl 2-(1-(4-chlorobenzoyl)-5-fluoro-1H-indol-3-yl)acetate (15 mg, 0.043 mmol) was subjected to reaction with trimethyltin hydroxide (39 mg, 0.216 mmol) in 1,2-DCE (1 mL). The crude product was chromatographed on a short silica gel column using 1:1 ethyl acetate/hexane (0.5% AcOH) mixtures of increasing polarity as eluants to afford the title compound quantitatively as off-white solid. Yield: 11.2 mg (78%). C₁₇H₁₁ClFNO₃, M_r = 331.73; ¹H NMR (400 MHz, DMSO- d_6) δ : 3.66 (bs, 2H), 7.23 (td, J =2.6/9.2 Hz, 1H), 7.40–7.43 (m, 2H), 7.65–7.69 (m, 2H), 7.75–7.79 (m, 2H), 8.28 (dd, J =4.8/9.2 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ : 30.62 (s), 104.99 (d, J =24.3 Hz, 1C), 113.34 (d, J =24.9 Hz, 1C), 113.92 (s), 117.76 (d, J =8.9 Hz, 1C), 127.35 (s), 129.08 (s, 2C), 130.57 (s, 2C), 131.38 (d, J =9.7 Hz, 1C), 132.37 (s), 132.47 (s), 138.58 (s), 159.96 (d, J =24.2 Hz, 1C), 167.09 (s), 175.85 (s); ¹⁹F NMR (282 MHz, DMSO- d_6) δ : -116.72 (s, -F); LCMS (ESI) t_R : 2.54 min (>99%, UV215, UV254, ELSD), m/z : 332.0 [M+H]⁺; HRMS (TOF, ES+) C₁₇H₁₁ClFNO₃ [M+H]⁺ calc. mass 332.0490, found 332.0490.

4.12. 6-methoxy-9-(3-(trifluoromethyl)benzoyl)-2,3,4,9-tetrahydro-1H-carbazole-3-carboxylic acid 23

According to general procedure D, methyl 6-methoxy-9-(3-(trifluoromethyl)benzoyl)-2,3,4,9-tetrahydro-1H-carbazole-3-carboxylate (15 mg, 0.035 mmol; ~60% pure) was subjected to reaction with trimethyltin hydroxide (31 mg, 0.174 mmol) in 1,2-DCE (1 mL). The crude product was chromatographed on a short silica gel column using 1:1 ethyl acetate/hexane (0.5% AcOH) mixtures of increasing polarity as eluants to afford the title compound quantitatively as bright yellow solid. Yield: 8.5 mg (98%). C₂₂H₁₈F₃NO₄, M_r = 417.38; ¹H NMR (400 MHz, DMSO- d_6) δ : 1.69–1.75 (m, 1H), 2.02–2.06 (m, 1H), ~2.47 (m, 2H, partially overlaid by DMSO signal), 2.71–2.74 (m, 2H), 2.89–2.92 (m, 1H), 3.78 (s, 3H), 6.73 (dd, J =2.6/9.0 Hz, 1H), 7.03 (d, J =2.4 Hz, 1H), 7.10 (d, J =9.2 Hz, 1H), 7.78 (t, J =7.8 Hz, 1H), 7.93 (d, J =7.6 Hz, 1H), 7.99 (s, 1H), 8.03 (d, J =7.6 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ : 23.51 (s), 24.73 (s), 25.81 (s), 38.74 (s), 55.68 (s), 101.09 (s), 111.96 (s), 115.49 (s), 116.78 (s), 123.48 (d, J =272 Hz, 1C), 126.18 (s), 128.81 (s), 129.34 (s), 130.63 (s), 130.95 (s), 131.37 (d, J =29.0 Hz, 1C), 132.42 (s), 135.55 (s), 136.53 (s), 156.35 (s), 167.29 (s), 179.92 (s); ¹⁹F NMR (282 MHz, DMSO- d_6) δ : -59.38 (*m*-CF₃); LCMS (ESI) t_R : 2.74 min (>99%, UV254, ELSD), m/z : 418.0 [M+H]⁺; HRMS (TOF, ES+) C₂₂H₁₈F₃NO₄ [M+H]⁺ calc. mass 418.1266, found 418.1267.

4.13. Allyl 2-(5-methoxy-1H-indol-3-yl)acetate 25

To a solution of 2-(5-methoxy-1H-indol-3-yl)acetic acid **7b** (15.25 g, 74.3 mmol) in 250 mL of acetone was added 29 g of cesium carbonate (89.2 mmol) and allyl bromide (7.1 mL, 81.74 mmol). The reaction mixture was stirred at room temperature for 12 h. The residual cesium carbonate was removed by filtration and solvent was concentrated *in vacuo*. The residue was purified by column chromatography using Hex/EtOAc (gradient: 0 to 30% EtOAc) to afford allyl 2-(5-methoxy-1H-indol-3-yl)acetate 25 (13.6 g, 75 %). C₁₄H₁₅NO₃, M_r = 245.27; ¹H NMR (400 MHz, CDCl₃) δ 3.8 (s, 2H), 3.89 (s, 3H), 4.64 (dt, J =1.2, 5.7, 2H), 5.2 (dd, J =1.2, 10.4, 1H), 5.32 (dd, J =1.5, 17.2, 1H), 6.0–5.90 (m, 1H), 6.89 (dd, J =2.4, 8.8 Hz, 1H), 7.09 (d, J =2.4 Hz, 1H), 7.17 (s, 1H), 7.26 (d, J =8.8 Hz, 1H).

4.14. Allyl 2-(1-(4-chlorobenzoyl)-5-methoxy-1H-indol-3-yl)acetate 26

To a solution of allyl 2-(5-methoxy-1H-indol-3-yl)acetate 25 (10 g, 40.8 mmol) in dichloromethane (150 mL) was added triethylamine (17.1 mL, 122.4 mmol) and DMAP (4.9 g, 40.8 mmol) at 0 °C. After stirring 30 min, 4-chlorobenzoyl chloride (7.9 mL, 61.2 mmol) was added to the reaction mixture, which was allowed to warm to room temperature and stirred for 12 h. The reaction mixture was then quenched with saturated ammonium chloride (200 mL), extracted with dichloromethane (3 × 150 mL), dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography using Hex/EtOAc (gradient: 0 to 30% EtOAc) to afford a yellow solid of allyl 2-(1-(4-chlorobenzoyl)-5-methoxy-1H-indol-3-yl)acetate **26** (14.3 g, 92 %). C₂₁H₁₈ClNO₄, M_r = 383.82; ¹H NMR (400 MHz, CDCl₃) δ 3.69 (s, 2H), 3.88 (s, 3H), 4.62 (d, *J*=6.0 Hz, 2H), 5.23 (dd, *J*=1.2, 10.4 Hz, 1H), 5.29 (dd, *J*=1.2, 17.2 Hz, 1H), 5.95–5.84 (m, 1H), 7.01 (d, *J*=7.6 Hz, 2H), 7.25 (s, 1H), 7.50 (d, *J*=8.4 Hz, 2H), 7.67 (d, *J*=8.4 Hz, 2H), 8.28 (d, *J*=10.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ: 30.97 (s), 55.70 (s), 65.74 (s), 101.93 (s), 113.76 (s), 114.68 (s), 117.39 (s), 118.69 (s), 126.31 (s), 128.92 (s, 2C), 130.50 (s, 2C), 130.73 (s), 131.43 (s), 131.78 (s), 132.85 (s), 138.15 (s), 156.89 (s), 166.96 (s), 170.25 (s); LCMS (ESI), single peak, *t*_R: 0.92 min (>99%, UV, ELSD), *m/z*, 384.0 [M+H]⁺; HRMS (TOF, ES⁺) C₂₁H₁₈ClNO₄ [M+H]⁺ calc. mass 384.1003, found 384.1003.

4.15. 2-(1-(4-chlorobenzoyl)-5-methoxy-1H-indol-3-yl)acetic acid 2

To a solution of allyl 2-(1-(4-chlorobenzoyl)-5-methoxy-1H-indol-3-yl)acetate 3 (14.3 g, 37.3 mmol) in THF (200 mL) was added morpholine (32.5 mL, 373 mmol) and Pd(PPh₃)₄ (2.3 g, 1.9 mmol) under argon at room temperature. Stirring was continued until LCMS analysis indicated starting material has disappeared. The mixture was filtered and concentrated *in vacuo*. The residue was redissolved in dichloromethane and acidified with 2N HCl (50 mL) to afford white solid **2** (10 g, 79 %). C₁₈H₁₄ClNO₄, M_r = 343.76; ¹H NMR (400 MHz, DMSO-*d*₆) δ: 3.67 (s, 2H), 3.81 (s, 3H), 6.99 (dd, *J*=2.4/9.2 Hz, 1H), 7.12 (d, *J*=2.4 Hz, 1H), 7.32 (s, 1H), 7.65–7.77 (m, 4H), 8.17 (d, *J*=9.2 Hz, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 30.15 (s), 55.47 (s), 102.70 (s), 113.05 (s), 115.70 (s), 116.70 (s), 126.84 (s), 128.82 (s, 2C), 129.99 (s), 130.69 (s, 2C), 131.87 (s), 133.00 (s), 136.57 (s), 156.30 (s), 166.60 (s), 171.97 (s); LCMS (ESI) *t*_R: 2.60 min (>99%, ELSD), *m/z*: 344.0 [M+H]⁺; HRMS (TOF, ES⁺) C₁₈H₁₄ClNO₄ [M+H]⁺ calc. mass 344.0690, found 344.0687.

4.16. Allyl 4-oxohexanoate 27

Succinyl dichloride 4 (4.6 mL, 41.6 mmol) in dichloromethane (200 mL) was cooled to –40 °C and ethylaluminum dichloride (50 mL, 50 mmol) was added at –40 °C. The reaction mixture was stirred at –40 °C for 3.5 h and quenched with the addition of anhydrous allyl alcohol (100 mL). The reaction mixture was concentrated to dryness and redissolved in dichloromethane (100 mL). The organic layer was washed with aqueous sodium potassium tartrate (2 × 50 mL), water (50 mL) and saturated aqueous NaCl (50 mL). The organic layer was dried over MgSO₄, filtered and concentrated to provide pure allyl 4-oxohexanoate **27** (5.2 g, 73 %). C₉H₁₄O₃, M_r = 170.21; ¹H NMR (400 MHz, CDCl₃) δ 1.07 (t, *J*=7.2 Hz, 3H), 2.47 (q, *J*=7.2 Hz, 2H), 2.62 (t, *J*=6.4 Hz, 2H), 2.73 (t, *J*=6.4 Hz, 2H), 4.57 (d, *J*=5.6 Hz, 2H), 5.23 (dd, *J*=1.2, 10.4 Hz, 1H), 5.31 (dd, *J*=1.2, 18.0 Hz, 1H), 5.96–5.84 (m, 1H).

4.17. Allyl 3-(5-methoxy-3-methyl-1H-indol-2-yl)propanoate 28

To a mixture of (4-methoxyphenyl)hydrazine (10.4 g, 59.7 mmol) and allyl 4-oxohexanoate 5 (10.2 g, 59.7 mmol) in 1,4-dioxane (60 mL) was added acetic acid (30 mL) at room temperature. The reaction was heated at 80 °C for 6 h then quenched with saturated NaHCO₃ (150 mL), extracted with EtOAc (3 × 60 mL), and dried over MgSO₄. The residue was purified by column chromatography using Hex/EtOAc (gradient: 0 to 40 % EtOAc) to

afford brown oil allyl 3-(5-methoxy-3-methyl-1*H*-indol-2-yl)propanoate **28** (11.6 g, 71 %). $C_{16}H_{19}NO_3$, $M_r = 273.33$; 1H NMR (400 MHz, $CDCl_3$) δ 2.21 (s, 3H), 2.68 (t, $J=6.4$ Hz, 2H), 3.03 (t, $J=6.4$ Hz, 2H), 3.86 (s, 3H), 4.60 (d, $J=7.2$ Hz, 2H), 5.23 (dd, $J=1.2, 10.4$ Hz, 1H), 5.29 (dd, $J=1.2, 17.2$ Hz, 1H), 5.95-5.83 (m, 1H), 6.78 (dd, $J=2.4, 8.8$ Hz, 1H), 6.93 (d, $J=2.4$ Hz, 1H), 7.16 (d, $J=8.8$ Hz, 1H); ^{13}C NMR (100 MHz, $CDCl_3$) δ : 8.43 (s), 20.91 (s), 34.09 (s), 55.95 (s), 65.47 (s), 100.50 (s), 106.91 (s), 111.04 (s), 111.08 (s), 118.54 (s), 129.32 (s), 130.39 (s), 131.85 (s), 134.45 (s), 153.73 (s), 173.45 (s); LCMS (ESI), single peak, t_R : 0.79 min, m/z : 274.0 $[M+H]^+$; HRMS (TOF, ES+) $C_{16}H_{19}NO_3$ $[M+H]^+$ calc. mass 274.1443, found 274.1441.

4.18. Allyl 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1*H*-indol-2-yl)propanoate **29**

To a solution of allyl 3-(5-methoxy-3-methyl-1*H*-indol-2-yl)propanoate **28** (16 g, 61.8 mmol) in 1,2-dichloroethane (200 mL) was added triethylamine (34.5 mL, 247.2 mmol) and DMAP (7.55 g, 61.8 mmol) at 0°C. After stirring for 30 min, 4-chlorobenzoyl chloride (19.7 mL, 154.6 mmol) was added to the reaction mixture, which was allowed to stir for 12 h under reflux condition. The reaction mixture was cooled down, quenched with saturated ammonium chloride (200 mL), extracted with dichloromethane (3 \times 150 mL), dried over $MgSO_4$, and concentrated *in vacuo*. The residue was purified by column chromatography using Hex/EtOAc (gradient: 0 to 30% EtOAc) to afford brown oil of allyl 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1*H*-indol-2-yl)propanoate **29** (22.2 g, 87 %). $C_{23}H_{22}ClNO_4$, $M_r = 411.88$; 1H NMR (400 MHz, $CDCl_3$) δ 2.24 (s, 3H), 2.69 (t, $J=7.6$ Hz, 2H), 3.29 (t, $J=7.6$ Hz, 2H), 3.83 (s, 3H), 4.53 (d, $J=6.0$ Hz, 2H), 5.18 (dd, $J=1.2, 10.4$ Hz, 1H), 5.25 (dd, $J=1.2, 17.2$ Hz, 1H), 5.93-5.78 (m, 1H), 6.44 (d, $J=9.2$ Hz, 1H), 6.59 (dd, $J=2.8, 8.4$ Hz, 1H), 6.89 (d, $J=2.8$ Hz, 1H), 7.46 (d, $J=8.4$ Hz, 2H), 7.65 (d, $J=8.4$ Hz, 2H); ^{13}C NMR (100 MHz, $CDCl_3$) δ : 8.67 (s), 21.67 (s), 34.14 (s), 55.65 (s), 65.18 (s), 101.38 (s), 111.48 (s), 114.78 (s), 116.38 (s), 118.28 (s), 129.10 (s, 2C), 129.33 (s), 130.76 (s), 131.15 (s, 2C), 131.66 (s), 131.84 (s), 132.06 (s), 133.88 (s), 136.71 (s), 139.15 (s), 155.77 (s), 168.13 (s), 172.23 (s); LCMS (ESI), single peak, t_R : 0.99 min, m/z : 412.0 $[M+H]^+$; HRMS (TOF, ES+) $C_{23}H_{22}ClNO_4$ $[M+H]^+$ calc. mass 412.1316, found 412.1320.

4.19. 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1*H*-indol-2-yl)propanoic acid **30**

To a solution of allyl 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1*H*-indol-2-yl)propanoate **29** (11.1 g, 26.9 mmol) in THF (200 mL) was added morpholine (23.0 mL, 269 mmol) and $Pd(PPh_3)_4$ (1.67 g, 1.35 mmol) under argon at room temperature. Stirring was continued until LCMS analysis indicated starting material has disappeared. The mixture was filtered and concentrated *in vacuo*. The residue was redissolved in dichloromethane, acidified with 2N HCl (50 mL), concentrated *in vacuo*, and purified by column chromatography using Hex/EtOAc (gradient: 0 to 100 % EtOAc) to afford yellow solid of 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1*H*-indol-2-yl)propanoic acid **30** (7.9 g, 79%). $C_{20}H_{18}ClNO_4$, $M_r = 371.81$; 1H NMR (400 MHz, $CDCl_3$) δ 2.24 (s, 3H), 2.72 (t, $J=7.6$ Hz, 2H), 3.29 (t, $J=7.6$ Hz, 2H), 3.83 (s, 3H), 6.42 (d, $J=9.2$ Hz, 1H), 6.59 (dd, $J=2.8, 9.2$ Hz, 1H), 6.90 (d, $J=2.8$ Hz, 1H), 7.45 (d, $J=8.4$ Hz, 2H), 7.64 (d, $J=8.4$ Hz, 2H); ^{13}C NMR (100 MHz, $DMSO-d_6$) δ : 8.40 (s), 21.24 (s), 33.74 (s), 55.42 (s), 101.55 (s), 111.58 (s), 114.38 (s), 115.74 (s), 129.15 (s, 2C), 130.26 (s), 131.24 (s, 2C), 134.04 (s), 136.72 (s), 137.78 (s), 155.46 (s), 167.77 (s), 173.40 (s), one signal invisible; LCMS (ESI), single peak, t_R : 0.82 min, m/z : 372.0 $[M+H]^+$; HRMS (TOF, ES+) $C_{20}H_{18}ClNO_4$ $[M+H]^+$ calc. mass 372.1003, found 372.1005.

4.20. 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1H-indol-2-yl)-N-(methylsulfonyl)propanamide **31**

The reaction mixture of 3-(1-(4-chlorobenzoyl)-5-methoxy-3-methyl-1H-indol-2-yl)propanoic acid **30** (7 g, 18.8 mmol) and methanesulfonamide (1.97 g, 20.7 mmol) in dichloromethane (94 mL) was treated with 1,1'-carbonyldiimidazole (3.05 g, 18.8 mmol) and DBU (3.4 mL, 22.6 mmol) at 0°C. The reaction mixture was stirred at room temperature for 4h then quenched with acetic acid (5 mL) and washed with brine (3 × 40 mL). The organic layer was then dried over MgSO₄ and the solvent was removed *in vacuo*. The yellow solid **31** (5.6 g, 61 %) was obtained after column chromatography purification using Hex/EtOAc (gradient: 0 to 100 % EtOAc with 0.5 % of acetic acid). C₂₁H₂₁ClN₂O₅S, M_r = 170.21; ¹H NMR (400 MHz, CDCl₃) δ 2.20 (s, 3H), 2.54 (dd, *J*=7.2, 7.6 Hz, 2H), 3.10 (dd, *J*=7.2, 7.6 Hz, 2H), 3.14 (s, 3H), 3.75 (s, 3H), 6.42 (d, *J*=9.2 Hz, 1H), 7.70-7.64 (m, 4H), 6.63 (dd, *J*=2.4, 9.2 Hz, 1H), 7.01 (d, *J*=2.4 Hz, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 8.42 (s), 20.57 (s), 35.30 (s), 40.96 (s), 55.42 (s), 101.59 (s), 111.66 (s), 114.45 (s), 116.01 (s), 129.13 (s, 2C), 130.25 (s), 131.20 (s), 131.27 (s, 2C), 134.00 (s), 136.31 (s), 137.77 (s), 155.48 (s), 167.74 (s), 171.42 (s); LCMS (ESI), single peak, 0.81 min, m/z, 449.08 [M+H]⁺; HRMS (TOF, ES+) C₂₁H₂₁ClN₂O₅S [M+H]⁺ calc. mass 449.0938, found 449.0941.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

The authors are grateful to the National Institutes of Health (CA89450) and the National Foundation for Cancer Research for financial support. We thank Dr. Matthew Mulder (Craig Lindsley Group) for recording all high-resolution mass spectra and Christian Wallen (2011 Vanderbilt REU Trainee) for the assistance with the parallel syntheses of the fluorinated indole precursors. A.J.L. also thanks the German Research Foundation (DFG) for obtaining a Research Stipend (LI2019/1-1, Einzelprojekt).

References and notes

- Bartolini, W.; Cali, B.M.; Chen, B.; Chien, Y.-T.; Currie, M.G.; Milne, G.T.; Pearson, J.P.; Talley, J.J.; Zimmerman, C. Application: US US. USA: 2005. p. 118Cont -in-part of U S Ser No 883, 900
- Munoz, B.; Prasit, P.; Stock, N.S. Application: WO WO. Merck & Co., Inc; USA: 2004. p. 40
- Torisu K, Kobayashi K, Iwahashi M, Egashira H, Nakai Y, Okada Y, Nanbu F, Ohuchida S, Nakai H, Toda M. Bioorganic & Medicinal Chemistry Letters. 2004; 14:4557–4562. [PubMed: 15357992]
- Torisu K, Kobayashi K, Iwahashi M, Egashira H, Nakai Y, Okada Y, Nanbu F, Ohuchida S, Nakai H, Toda M. European Journal of Medicinal Chemistry. 2005; 40:505–519. [PubMed: 15893024]
- Wey SJ, Augustyniak ME, Cochran ED, Ellis JL, Fang X, Garvey DS, Janero DR, Letts LG, Martino AM, Melim TL, Murty MG, Richardson SK, Schroeder JD, Selig WM, Trocha AM, Wexler RS, Young DV, Zemtseva IS, Zifcak BM. Journal of Medicinal Chemistry. 2007; 50:6367–6382. [PubMed: 17994684]
- Blobaum AL, Marnett LJ. Journal of Medicinal Chemistry. 2007; 50:1425–1441. [PubMed: 17341061]
- Rainsford KD. Sub-cellular biochemistry. 2007; 42:3–27. [PubMed: 17612044]
- Wallace JL, McKnight W, Reuter BK, Vergnolle N. Gastroenterology. 2000; 119:706–714. [PubMed: 10982765]
- Sandham DA, Adcock C, Bala K, Barker L, Brown Z, Dubois G, Budd D, Cox B, Fairhurst RA, Furegati M, Leblanc C, Manini J, Profit R, Reilly J, Stringer R, Schmidt A, Turner KL, Watson SJ, Willis J, Williams G, Wilson C. Bioorg Med Chem Lett. 2009; 19:4794–8. [PubMed: 19592244]
- Felts AS, Ji C, Stafford JB, Crews BC, Kingsley PJ, Rouzer CA, Washington MK, Subbaramaiah K, Siegel BS, Young SM, Dannenberg AJ, Marnett LJ. ACS Chemical Biology. 2007; 2:479–483. [PubMed: 17602619]

11. Byrns MC, Steckelbroeck S, Penning TM. *Biochemical Pharmacology*. 2008; 75:484–493. [PubMed: 17950253]
12. Piazza GA, Keeton AB, Tinsley HN, Whitt JD, Gary BD, Mathew B, Singh R, Grizzle WE, Reynolds RC. *Pharmaceuticals*. 2010; 3:1652–1667.
13. Schuligoi R, Sturm E, Luschnig P, Konya V, Philipose S, Sedej M, Waldhoer M, Peskar BA, Heinemann A. *Pharmacology*. 2010; 85:372–82. [PubMed: 20559016]
14. Hata AN, Lybrand TP, Marnett LJ, Breyer RM. *Mol Pharmacol*. 2005; 67:640–7. [PubMed: 15563582]
15. Marnett, LJ.; Prusakiewicz, JJ.; Felts, AS.; Ji, C. Application: US US. Vanderbilt University; USA: 2005. p. 63
16. Prusakiewicz JJ, Felts AS, Mackenzie BS, Marnett LJ. *Biochemistry*. 2004; 43:15439–15445. [PubMed: 15581355]
17. Mukai C, Takahashi Y. *Organic Letters*. 2005; 7:5793–5796. [PubMed: 16354068]
18. Maki BE, Scheidt KA. *Organic Letters*. 2009; 11:1651–1654. [PubMed: 19320508]
19. Campos KR, Woo JCS, Lee S, Tillyer RD. *Organic Letters*. 2004; 6:79–82. [PubMed: 14703355]
20. Takahashi H, Wakamatsu S, Tabata H, Oshitari T, Harada A, Inoue K, Natsugari H. *Organic Letters*. 2011; 13:760–763. [PubMed: 21250752]
21. Yamamoto H. *Journal of Organic Chemistry*. 1967; 32:3693–5.
22. Yamamoto H. *Chem Pharm Bull (Tokyo)*. 1968; 16:17–9. [PubMed: 5677227]
23. Maguire AR, Plunkett SJ, Papot S, Clynes M, O'Connor R, Touhey S. *Bioorganic & Medicinal Chemistry*. 2001; 9:745–762. [PubMed: 11310610]
24. Nicolaou KC, Estrada AA, Zak M, Lee SH, Safina BS. *Angewandte Chemie, International Edition*. 2005; 44:1378–1382.
25. Kalgutkar AS, Crews BC, Saleh S, Prudhomme D, Marnett LJ. *Bioorganic & Medicinal Chemistry*. 2005; 13:6810–6822. [PubMed: 16169736]
26. Conn RSE, Douglas AW, Karady S, Corley EG, Lovell AV, Shinkai I. *Journal of Organic Chemistry*. 1990; 55:2908–13.
27. Allen GR Jr. *J Heterocycl Chem*. 1970; 7:239–41.
28. Karthikeyan SV, Perumal S, Shetty KA, Yogeewari P, Sriram D. *Bioorganic & Medicinal Chemistry Letters*. 2009; 19:3006–3009. [PubMed: 19403307]
29. Vasudevan A, Souers AJ, Freeman JC, Verzal MK, Gao J, Mulhern MM, Wodka D, Lynch JK, Engstrom KM, Wagaw SH, Brodjian S, Dayton B, Falls DH, Bush E, Brune M, Shapiro RD, Marsh KC, Hernandez LE, Collins CA, Kym PR. *Bioorganic & Medicinal Chemistry Letters*. 2005; 15:5293–5297. [PubMed: 16203136]
30. Jeanty M, Suzenet F, Guillaumet G. *Journal of Organic Chemistry*. 2008; 73:7390–7393. [PubMed: 18729406]
31. Mylari, BL.; Zembrowski, WJ. Application: EP EP. Pfizer Inc; USA: 1989. p. 15
32. Honba K, Yamada T, Yoneda M, Ohkawa Y, Nagai H. *Iyakuin Kenkyu*. 1982; 13:323–33.
33. Guibe F. *Tetrahedron*. 1998; 54:2967–3042.
34. Tsuji J, Mandai T. *Synthesis*. 1996:1–24.
35. Goossen LJ, Doehring A. *Synlett*. 2004:263–266.
36. Lee JC, Oh YS, Cho SH, Lee J II. *Org Prep Proced Int*. 1996; 28:480–483.
37. Anderson GW, Callahan FM. *J Am Chem Soc*. 1960; 82:3359–63.
38. Kunz H, Waldmann H. *Helv Chim Acta*. 1985; 68:618–22.
39. Choi Y, Ishikawa H, Velcicky J, Elliott GI, Miller MM, Boger DL. *Organic Letters*. 2005; 7:4539–4542. [PubMed: 16178578]
40. Adeniji AO, Twenter BM, Byrns MC, Jin Y, Chen M, Winkler JD, Penning TM. *J Med Chem*. 2012; 55:2311–23. [PubMed: 22263837]
41. Mylari, BL.; Zembrowski, WJ. Application: US US. Pfizer Inc; USA: 1993. p. 8Cont of U S Ser No 499, 279, abandoned

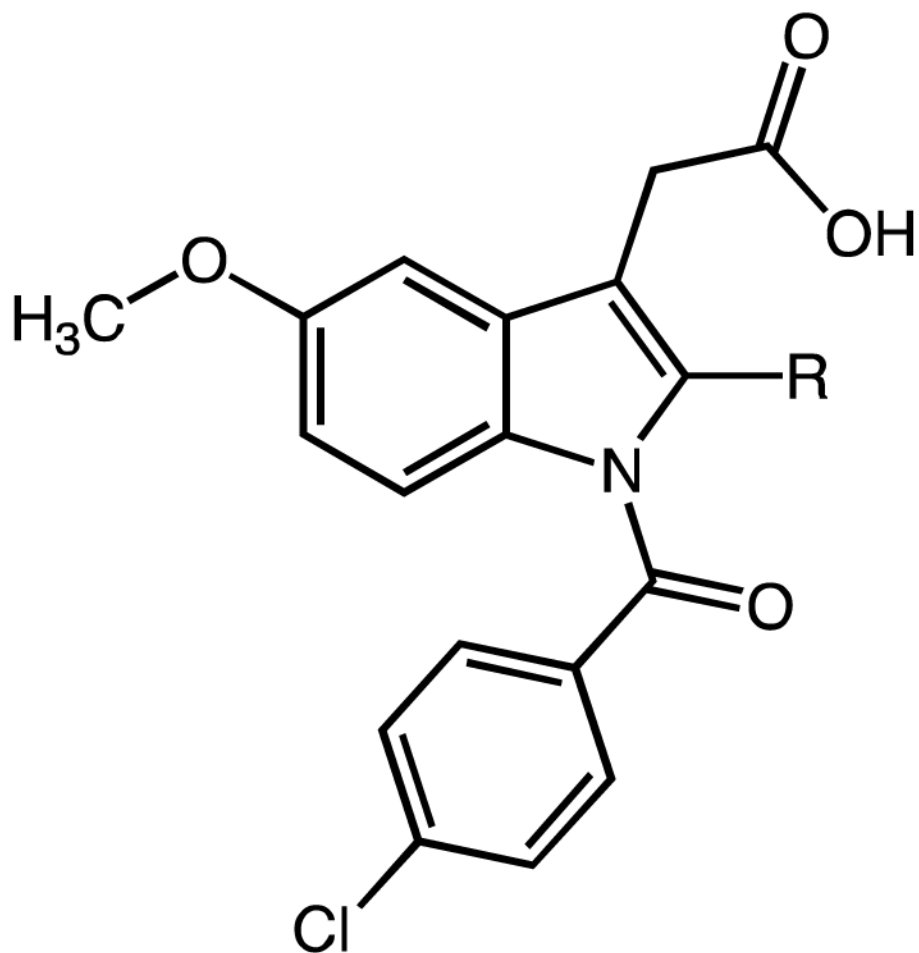
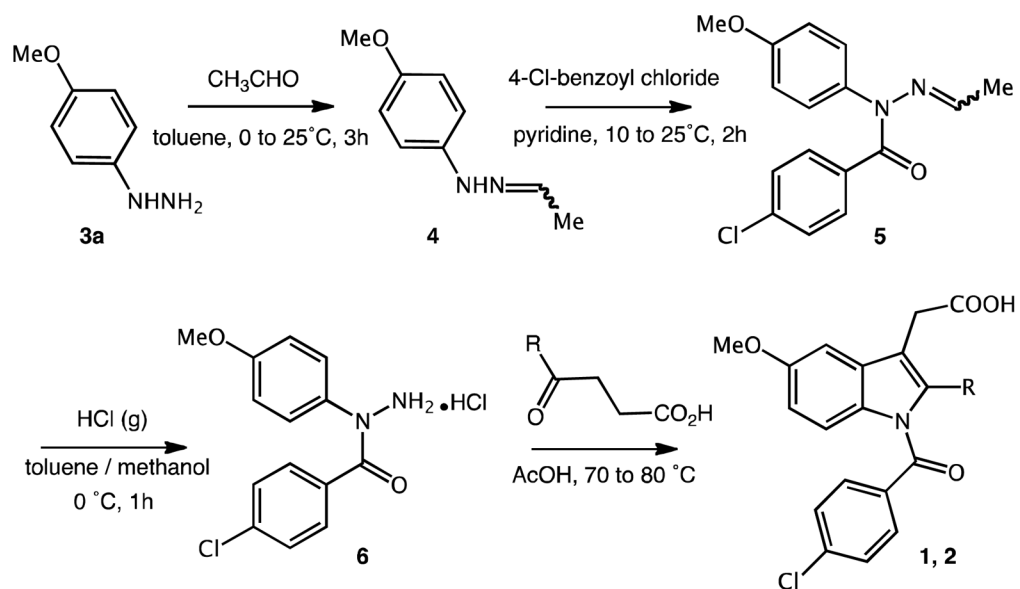
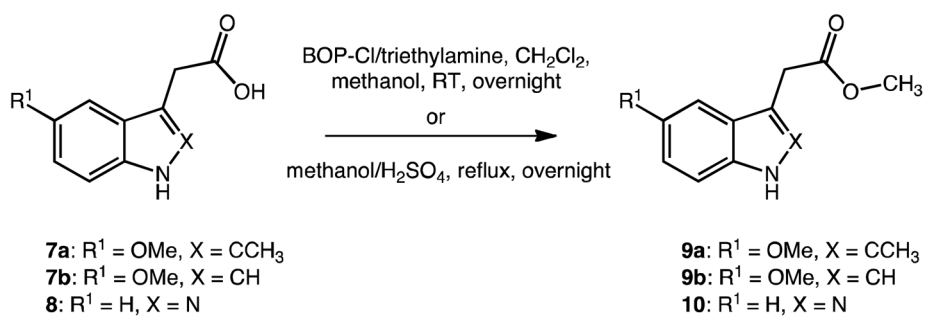


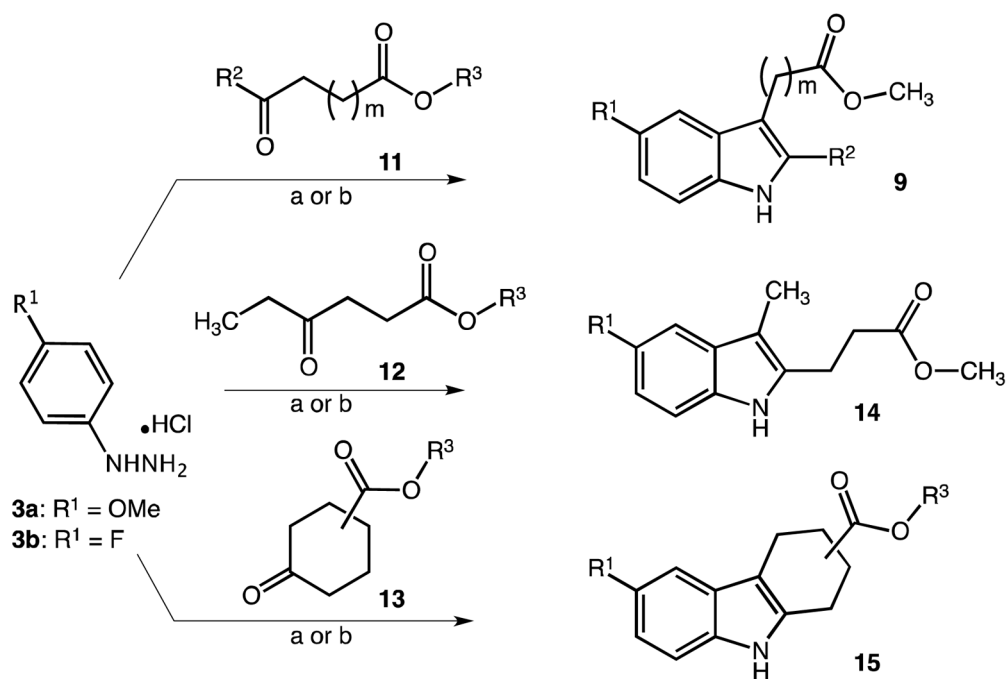
Fig. 1.

**Scheme 1.**

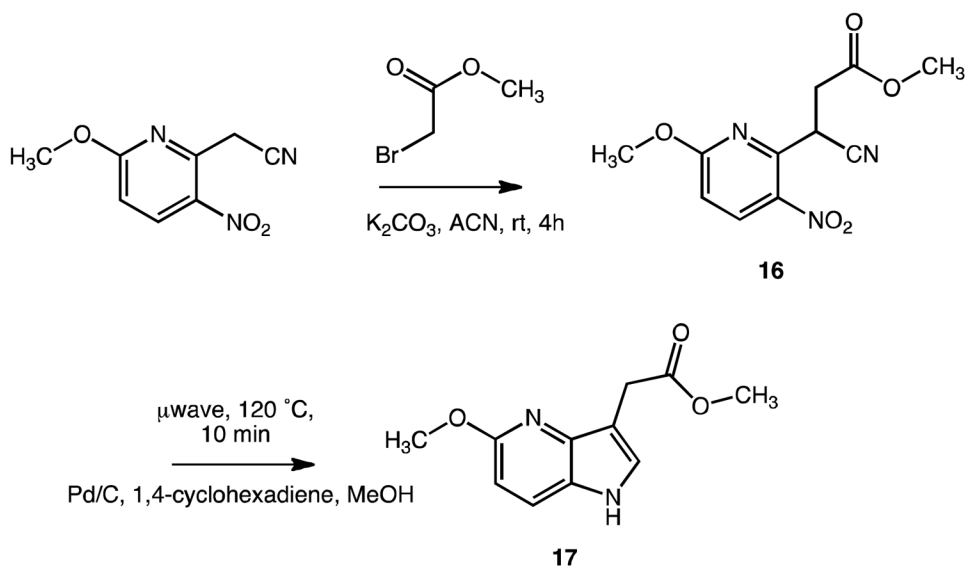
Yamamoto's procedure for the synthesis of (2'-des-methyl) indomethacin; R = CH₃ or H

**Scheme 2.**

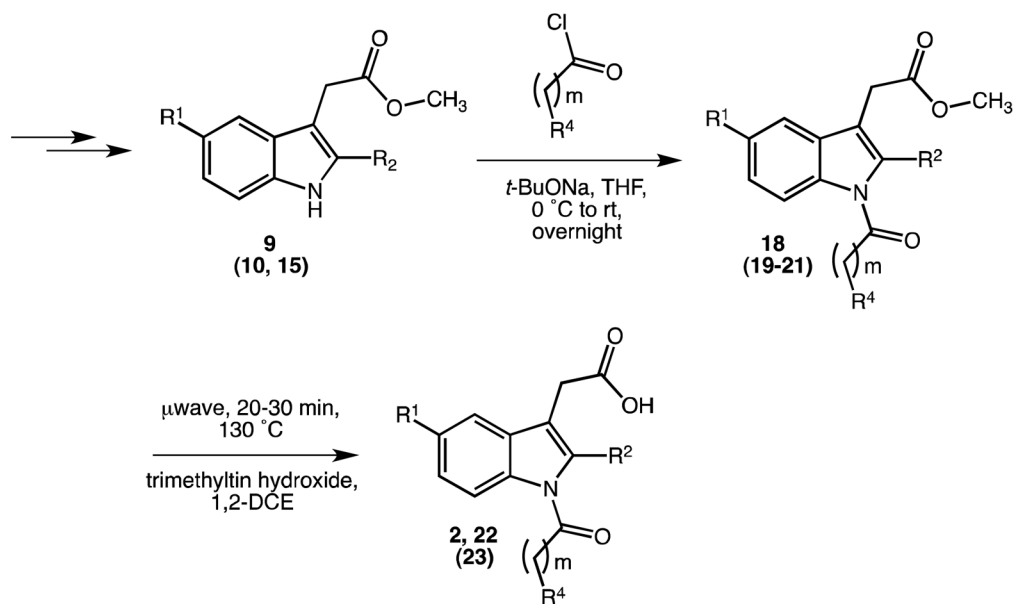
Esterification of (2'-aza-/2'-des-methyl) indole-3'-acetic acids

**Scheme 3.**

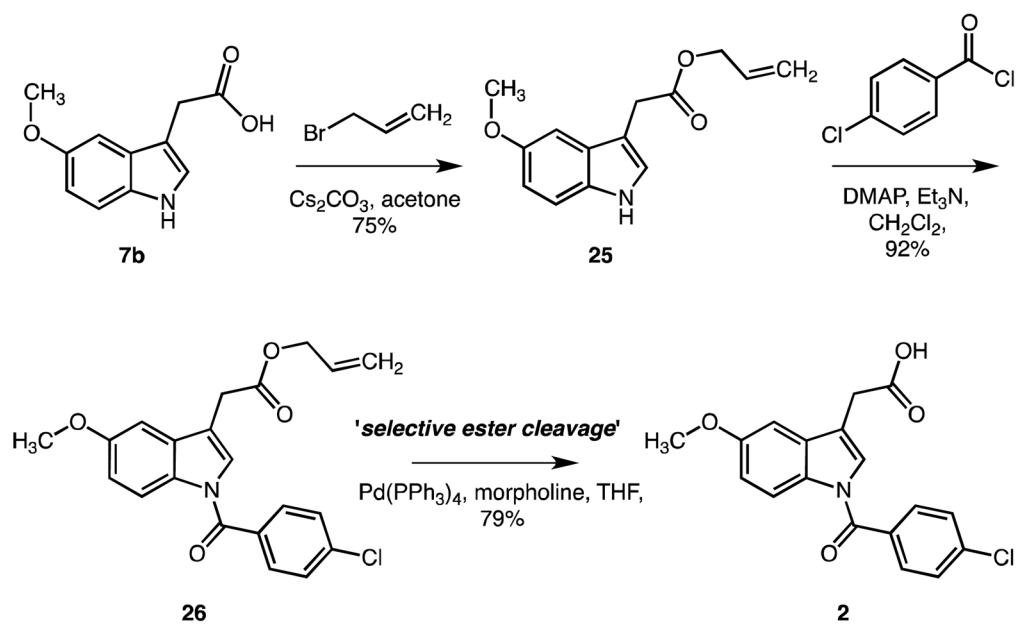
Fischer indolization reactions toward different 2'-/3'-alkylcarboxylic acid esters. Reagents and conditions: (a) AcOH, 80 °C, 3h; (b) MeOH/H₂SO₄, 5 min (μwave) to 3h (conventional heating)



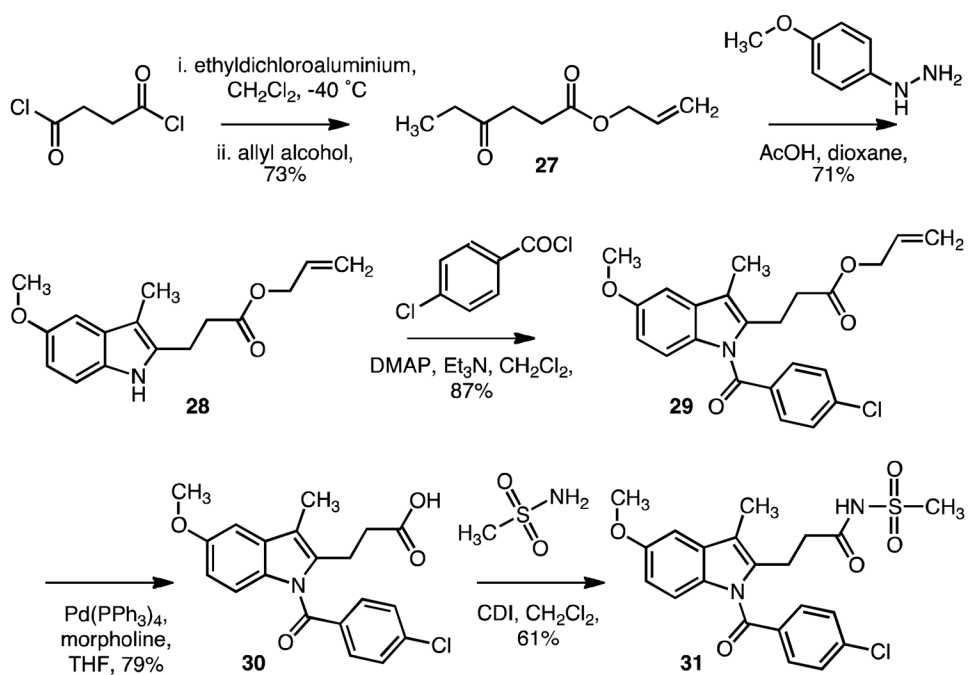
Scheme 4.
Synthesis of a 4-aza-indole-3-acetic acid ester analogue



Scheme 5.
N-Acylation and ester cleavage reactions.



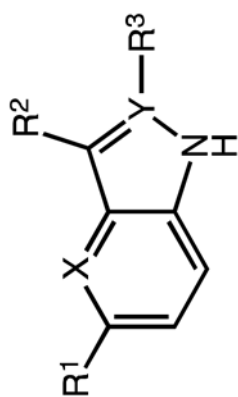
Scheme 6.
Large-scale synthesis of compound 2.


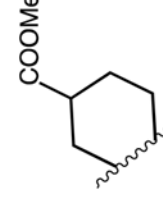


Scheme 7.
Large-scale synthesis of compounds **30** (free acid) and **31** (COOH bioisostere).

Table 1

Preparation of different (aza)indole alkanolic acid ester precursors



#	X	Y	R ¹	R ²	R ³	method	yield (%) ^a
9a	C	C	OMe	CH ₂ C(O)OMe	Me	A1	>99
9b	C	C	OMe	CH ₂ C(O)OMe	H	A1	85
9c	C	C	F	CH ₂ C(O)OMe	Me	B2	84
9d	C	C	F	CH ₂ CH ₂ C(O)OMe	Me	B2	76
9e	C	C	F	CH ₂ C(O)OMe	H	B ₂ ^b	86
14	C	C	F	Me	CH ₂ CH ₂ C(O)OMe	B2	46
15a	C	C	OMe			B1	90
15b	C	C	F			B2	99
10	C	N	H	CH ₂ C(O)OMe	---	A2	77
17	N	C	OMe	CH ₂ C(O)OMe	H	other	60

\$watermark-text

\$watermark-text

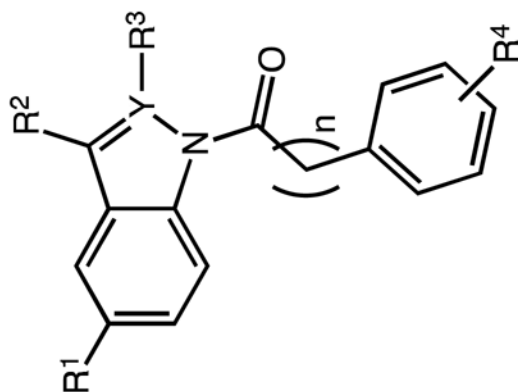
\$watermark-text

Reagents and Conditions: [method A1]: BOP-Cl, triethylamine, CH₂Cl₂, MeOH, rt, overnight; [method A2]: MeOH, H₂SO₄, reflux, overnight; [method B1]: arylhydrazine HCl, ketoacid ester, AcOH, 80 °C, 3h; [method B2]: arylhydrazine HCl, ketoacid or ketoacid ester, MeOH, H₂SO₄, microwave, 120 °C, 10 min.

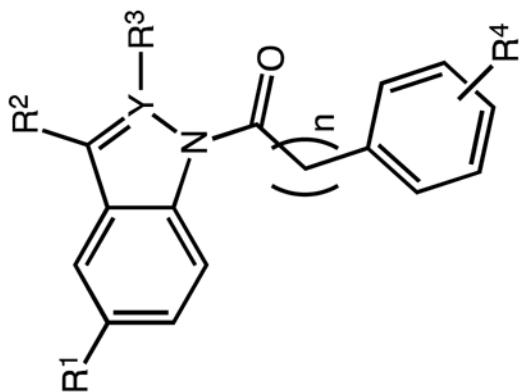
^a isolated yield;

^b conventional heating, 3h instead of microwave

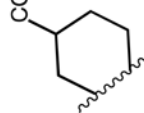
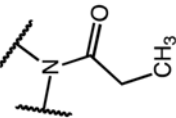
Table 2

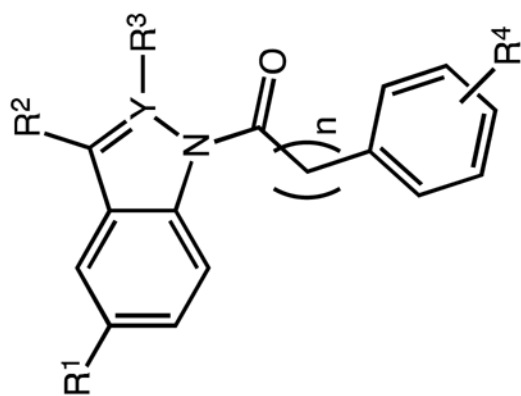
Synthesis of *N*-acyl methyl (aza)indole(cyclo)alkanoates

#	Y	R ¹	R ²	R ³	n	R ⁴	method	yield (%) ^a
18a	C	OMe	CH ₂ C(O)OMe	Me	0	<i>p</i> -F	C2	25
18b	C	OMe	CH ₂ C(O)OMe	Me	0	<i>m</i> -CF ₃	C2	32
18c	C	OMe	CH ₂ C(O)OMe	Me	0	<i>p</i> -CF ₃	C2	33
18d	C	OMe	CH ₂ C(O)OMe	Me	0	<i>p</i> -Me	C2	57
18e	C	OMe	CH ₂ C(O)OMe	Me	0	<i>p</i> -CH ₂ Cl	C2	42
18f	C	OMe	CH ₂ C(O)OMe	Me	0	<i>p</i> -OMe	C2	32
18g	C	OMe	CH ₂ C(O)OMe	H	0	<i>p</i> -Cl	C1	18
18h	C	OMe	CH ₂ C(O)OMe	H	0	<i>p</i> -F	C2	22
18i	C	OMe	CH ₂ C(O)OMe	H	0	<i>m</i> -CF ₃	C2	13
18j	C	OMe	CH ₂ C(O)OMe	H	0	<i>p</i> -CF ₃	C2	29
18k	C	OMe	CH ₂ C(O)OMe	H	1	<i>p</i> -OMe	C1	(13) [*]



#	Y	R ¹	R ²	R ³	n	R ⁴	method	yield (%) ^a
18l	C	OMe	CH ₂ C(O)OMe	H	0		C1	21
18m	C	F	CH ₂ C(O)OMe	Me	0	<i>p</i> -Cl	C2	42
18n	C	F	CH ₂ C(O)OMe	Me	0	<i>m</i> -CF ₃	C2	33
18o	C	F	CH ₂ CH ₂ C(O)OMe	Me	0	<i>p</i> -Cl	C2	49
18p	C	F	CH ₂ C(O)OMe	Me	1	<i>p</i> -Cl	C2	4
18q	C	F	CH ₂ C(O)OMe	H	0	<i>p</i> -Cl	C2	42
19a	C	OMe	COOMe		0	<i>m</i> -CF ₃	C2	24





#	Y	R ¹	R ²	R ³	n	R ⁴	method	yield (%) ^a
19b	C	F	COOMe		0	<i>p</i> -Cl	C2	14
20a	C	OMe	H	Me	0	<i>p</i> -Cl	C2	25
20b	C	OMe		Me	0	<i>p</i> -Cl	C2	8 ^b
21	N	OMe	CH ₂ C(O)OMe	...	0	<i>p</i> -Cl	C2	37

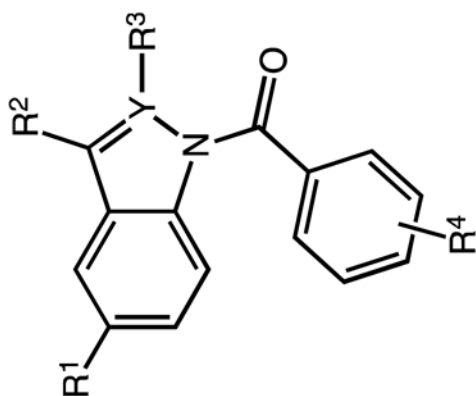
Reagents and Conditions: [method C1]: NaH, DMF, acyl chloride, 0 °C to rt, overnight; [method C2]: *t*-BuONa, THF, acyl chloride, 0 °C to rt, overnight.

^a isolated yield;

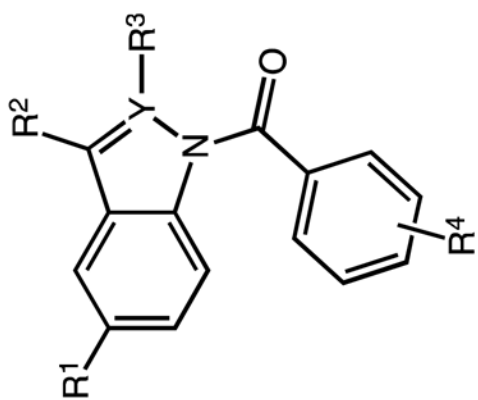
^b **20b** emerged as one side product in the synthesis of **20a**;

* compound after repeated chromatography only about 50% pure

Table 3

Synthesis of *N*-acyl indole(cyclo)alkanoic acids

#	Y	R ¹	R ²	R ³	R ⁴	method	yield (%) ^a
2	C	OMe	CH ₂ C(O)OH	H	<i>p</i> -Cl	D	81
22a	C	OMe	CH ₂ C(O)OH	Me	<i>p</i> -F	D	90
22b	C	OMe	CH ₂ C(O)OH	Me	<i>m</i> -CF ₃	D	97
22c	C	OMe	CH ₂ C(O)OH	Me	<i>p</i> -CF ₃	D	90
22d	C	OMe	CH ₂ C(O)OH	Me	<i>p</i> -Me	D	82
22e	C	OMe	CH ₂ C(O)OH	Me	<i>p</i> -CH ₂ Cl	D	98
22f	C	OMe	CH ₂ C(O)OH	Me	<i>p</i> -OMe	D	93 [*]
22g	C	OMe	CH ₂ C(O)OH	H	<i>p</i> -F	D	76
22h	C	OMe	CH ₂ C(O)OH	H	<i>m</i> -CF ₃	D	96
22i	C	OMe	CH ₂ C(O)OH	H	<i>p</i> -CF ₃	D	99



#	Y	R ¹	R ²	R ³	R ⁴	method	yield (%) ^d
22j	C	OMe	CH ₂ C(O)OH	H		D	94
22k	C	F	CH ₂ C(O)OH	Me	<i>p</i> -Cl	D	87
22l	C	F	CH ₂ C(O)OH	Me	<i>m</i> -CF ₃	D	95
22m	C	F	CH ₂ CH ₂ C(O)OH	Me	<i>p</i> -Cl	D	95
22n	C	F	CH ₂ C(O)OH	H	<i>p</i> -Cl	D	78
23	C	OMe			<i>m</i> -CF ₃	D	98 [*]
24	N	OMe	CH ₂ C(O)OH	---	Cl	D	No product (decomposition)

\$watermark-text

\$watermark-text

\$watermark-text

Reagents and Conditions: [method D]: trimethyltin hydroxide, 1,2-DCE, microwave, 130 °C, 30 min.

^d isolated yield;

* crude educts used; impurity of the starting material taken into account for the calculation of the percent yield (see Supporting Information!)