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Beta-Adrenergic Receptor Polymorphisms and Cardiac Graft Function in Potential Organ Donors

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Abstract

Prior studies have demonstrated associations between β -adrenergic receptor polymorphisms and left ventricular dysfunction—an important cause of allograft non-utilization for transplantation. We hypothesized that β AR polymorphisms predispose donor hearts to LV dysfunction after brain death. 1,043 organ donors managed from 2001-2006 were initially studied. The following β AR single nucleotide polymorphisms were genotyped: β 1AR 1165C/G (Arg389Gly), β 1AR 145A/G (Ser49Gly), β 2AR 46G/A (Gly16Arg), and β 2AR 79C/G (Gln27Glu). In multivariable regression analyses, the β 2AR46 SNP was significantly associated with LV systolic dysfunction, with each minor allele additively decreasing the odds for LV ejection fraction <50%. The β 1AR1165 and β 2AR46 SNPs were associated with higher dopamine requirement during the donor management period: donors with the GG and AA genotypes had ORs of 2.64 (95% CI 1.52-4.57) and 2.70 (1.07-2.74) respectively for requiring >10 mcg/kg/min of dopamine compared to those with the CC and GG genotypes. However, no significant associations were found between β AR SNPs and cardiac dysfunction in 364 donors managed from 2007-2008, perhaps due to changes in donor management, lack of power in this validation cohort, or the absence of a true association. β AR polymorphisms may be associated with cardiac dysfunction after brain death, but these relationships require further study in independent donor cohorts.

Keywords

allograft function; brain death; cardiac allograft; cardiac transplant; cohort study; donor evaluation; donor management; donor outcomes; genetic polymorphism; genotyping

INTRODUCTION

Left ventricular (LV) systolic dysfunction has been well-described in the setting of brain death,(1) and is likely a multi-factorial process resulting from activation of the sympathetic nervous system, diffuse loss of vasomotor tone, endothelial dysfunction, and hormone depletion.(2, 3) Classic baboon studies have demonstrated that the initial Cushing reaction that accompanies brainstem herniation results in direct myocardial injury. Within minutes after brain death, an “autonomic storm” occurs(4) in which serum epinephrine levels increase by 1,100%, norepinephrine by 300%, and dopamine by 200%.(5, 6) Endomyocardial biopsy specimens have shown contraction band necrosis—histologic evidence of microinfarction secondary to catecholamine-mediated calcium overload. This initial period of heightened sympathetic activity may then be followed by a period of parasympathetic dominance characterized by reduced catecholamine sensitivity.(7)

It is unknown why only a subgroup of people develops LV dysfunction after brain death. This phenomenon, however, appears to affect acceptance of allografts for cardiac transplantation, leading to refusal of donor organs in approximately 25% of cases,(8) even for organs from young, otherwise healthy donors.(4) The beta-adrenergic receptors (β AR) are major mediators of catecholamine effects on the heart, and β AR polymorphism genotypes are associated with adverse cardiovascular outcomes in the general population. (9-14) We hypothesized that specific β AR polymorphism genotypes increase cardiac sensitivity to circulating catecholamines in the setting of brain death, and are therefore associated with a higher prevalence of left ventricular dysfunction in potential organ donors for cardiac transplantation.

MATERIALS and METHODS

Subjects

This is a retrospective genetics study utilizing a cohort of 2,048 consecutive organ donors managed by the California Transplant Donor Network (CTDN, Oakland, CA) from 2001-2008. The inclusion criteria included availability of stored DNA samples from brain dead organ donors and consent for at least one organ to be donated (n=1624). Our discovery cohort was comprised of 1,043 donors managed from 2001-2006; we then studied an additional 364 donors managed from 2007-2008 as a validation cohort, recognizing that changes in donor management protocols had been implemented after 2006. CTDN is the largest organ procurement agency in Northern California and supplies donor organs mainly to transplant centers in northern and central California and occasionally to neighboring states. Potential brain dead organ donors were identified by treating physicians at hospitals throughout the region, and consent for organ donation was obtained from family members or next-of-kin. Management of the organ donor was subsequently assumed by CTDN staff, and consent was obtained from the donor's family to collect patient data and biological samples. This study was approved by CTDN, the Committee on Human Research at the University of California San Francisco, and by the Research Compliance Office at Stanford University.

Measurement of Phenotypic and Echocardiographic Parameters

Upon assumption of donor management, comprehensive data on donor-level variables were recorded by CTDN staff in a standardized fashion, including demographic variables, cause of death, health-related behaviors, and past medical history. Data on comprehensive laboratory testing were also recorded, including serologies; hematologic, hepatic, and renal function indices; and cardiac troponin assays. Standard testing for potential donors who were not immediately ruled-out for cardiac graft donation (due to known coronary artery disease treated with percutaneous stents or bypass surgery, prior cardiac valve surgery, age > 65 years, lack of consent for donation, or coroner exclusion) included an electrocardiogram; one or more echocardiograms; and a coronary angiogram for male donors over 40 years and female donors over 45 years. All cardiac testing was performed and interpreted at the donor hospital and results were recorded by CTDN personnel. Data on vital signs, invasive hemodynamics, and medications were also recorded.

Donor data were extracted from the medical records and were entered into the study database by three CTDN study investigators (VH, RLM, and JN) who were blinded to outcomes. A subsequent quality-assessment review of 5% of the medical records was performed (KK, JZ, RLM), reviewing 177 fields per donor chart, and demonstrated 97% accuracy of data collection.

Polymorphism Selection and Genotyping

Four β -adrenergic receptor polymorphisms previously reported to be functionally significant were chosen for analysis (Table 1). The uncommon β 2AR 491C/T (Thr164Ile) SNP was also genotyped, but was not chosen for analysis due to the very low frequency of the T allele (1.7%) at this locus. Genomic DNA was isolated from the blood or spleen of deceased donors using the salting out method or commercial product (QIAmp DNA kits, Valencia, CA) at the Immunogenetics and Transplantation Laboratory at the University of California, San Francisco. Polymorphisms were genotyped by template-directed dye-terminator incorporation assay with fluorescence polarization detection (FP-TDI), using the AcycloPrime-FP kit (Perkin-Elmer),⁽¹⁵⁾ as previously described.⁽¹⁴⁾ β 1AR 1165C/G (Arg389Gly) was genotyped using the TaqMan SNP Genotyping Assay (Applied Biosystems, Assay ID: C_8898494_10) per the manufacturer's protocol. Positive (duplicates) and negative controls were included in all plates. The genotyping call rates were

as follows: β 1AR 1165=98.7%, β 1AR 145=93.7%, β 2AR 46=95.3%, β 2AR 79=96.9%. Genotyping was performed by investigators blinded to clinical variables.

Statistical Analysis

Statistical analyses were performed with R (version 2.13). The primary outcome variable in this study was donor left ventricular ejection fraction (LVEF), which was treated as a dichotomous variable (LVEF < or = 50%). Secondary outcome variables included any left ventricular regional wall motion abnormalities (RWMA) and maximum dopamine requirement during the donor management period (< or > 10 mcg/kg/min). The threshold values for dichotomization of LVEF and dopamine requirement were determined a priori, based on the collective clinical experience of the study investigators with respect to donor management and organ allocation.(8, 16, 17) Each polymorphism was tested for Hardy-Weinberg equilibrium (HWE) by χ^2 test within each of the four main racial/ethnic groups (Caucasian, African-American, Hispanic, and Asian). A racial group was dropped from analysis for that SNP if the HWE p-value was less than 0.001.

Multivariable logistic regression models were fit to estimate the association between the polymorphisms in each gene and the outcome of interest. Genotypes were coded as additive effects (0-1-2) with each level indicating the presence of an additional minor allele. Models were adjusted for donor age, sex, race (treated as a categorical variable, with Caucasians as the reference group), and cause of death (dichotomized into death due to intracranial hemorrhage/stroke versus all other causes). Interactions between SNPs in the same gene were also explored. Individuals with missing covariates or genotypes were dropped from the analysis. Sensitivity analyses were then performed, restricting the cohort to the largest racial-ethnic subgroup (Caucasians), to investigate potential population stratification effects.

For those genes and outcomes that passed the nominal significance threshold in the additive model ($p < .05$), we also present results from a co-dominant model treating genotypes as a categorical variable to allow for non-linear effects and to further explore the patterns of association (Supplementary Table 1). This strategy is analogous to performing an ANOVA and then follow-up t-tests only if the overall association is significant. No correction for multiple testing was performed.

The above-described models were initially fit for donors managed by CTDN from 2001 to 2006. We then refit the models for donors managed from 2007 to 2008 as a validation cohort. Finally, we calculated our power to replicate associations in the validation cohort empirically, assuming an alpha level of 0.05 and 10,000 simulations using the effect estimates, along with the upper and lower boundaries of their 95% confidence intervals.

RESULTS

Subjects

A total of 1407 of the 2048 potential organ donors managed by CTDN from 2001 and 2008 had stored DNA and complete genotyping available for analysis and defined the two study cohorts: a discovery cohort of 1043 donors from 2001-2006 and a validation cohort of 364 donors managed in 2007 and 2008. Eight hundred ninety four donors (75%) in the discovery cohort and 288 (67%) in the validation cohort had an echocardiogram. Donor characteristics, by cohort, are summarized in Table 2. Median values and the associated inter-quartile ranges (IQR) are shown for continuous variables and percentages for dichotomous variables. P-values were calculated by Wilcoxon rank-sum and chi-square tests respectively. Differences in donor management protocols between the two study cohorts included use of higher doses of phenylephrine and corticosteroids, and greater use of thyroid hormone in the latter cohort.

The clinical characteristics of the genetics study cohorts did not differ significantly from the entire CTDN donor cohort (data not shown).

Genotype frequencies of the four adrenergic receptor polymorphisms, by cohort and racial group, are shown in Table 3. Genotype distributions differed across the racial groups but were consistent with Hardy-Weinberg equilibrium within racial groups, except for the β 2AR79 SNP in African-Americans in the validation cohort (2007-2008 donors, $p < 0.0003$); this group was dropped from further analysis. Within each racial group, genotype frequencies were similar across the two study cohorts.

Association of Adrenergic Receptor Genotypes with Outcomes

Table 4 summarizes associations between specific β AR genotypes and cardiac outcomes, by study cohort. No significant interactions were identified between SNPs in the same gene or across genes.

Discovery cohort: 2001-2006 donors

Left Ventricular Ejection Fraction: Seven hundred seventy eight donors had an echocardiogram that included measurement of LVEF and complete covariate information. The median LVEF was 65%, with 10% of donors having an LVEF less than 50%. Figures 1 and 2 show boxplots for the β 1AR and β 2AR genotypes. Donors who were homozygous for the minor G allele for both β 1AR SNPs had lower LVEF compared to heterozygotes and major allele homozygous groups. Similarly, for the β 2AR46 SNP, the AA genotype group had a lower median LVEF than heterozygotes or major allele homozygous groups. Conversely, for the β 2AR79 SNP, the CC genotype group had a lower median LVEF than heterozygotes or minor allele homozygous groups.

In multivariable analysis, the β 2AR46 SNP was significantly associated with donor left ventricular systolic dysfunction, defined as having LVEF $< 50\%$, after adjusting for the effects of age, sex, race and cause of death. Assuming an additive model, each additional minor allele A of β 2AR46 (Gly16Arg) decreased the odds of LVEF $< 50\%$ by 40% ($p = 0.012$, Table 4). Figure 3a shows the results of the co-dominant model for each of the β 2AR SNPs (without the presence of the other). Compared to the β 2AR46 GG genotype group, the odds ratio (OR) for LVEF $< 50\%$ was 0.45 (95% CI 0.25-0.80) for the AG genotype and 0.34 (95% CI 0.17-0.67) for the AA genotype group, consistent with an additive genetic model. The β 2AR79 (Gln27Glu) SNP also suggests an additive relationship with LV dysfunction, with ORs for the CG and GG alleles being 1.42 (95% CI 0.85-2.35) and 2.07 (95% CI 0.88-4.89), though this association is not significant in the presence of the β 2AR46 SNP (Table 4). The β 1AR SNPs did not show a significant association with LVEF. Restricting the cohort to Caucasians yielded similar results (Supplementary Table 2).

Left Ventricular Regional Wall Motion Abnormalities: Seven hundred seventy six donors had echocardiograms with RWMA data as well as complete covariate information. Twenty percent of donors exhibited left ventricular RWMA. The β 1AR1165 SNP (p -value < 0.05) showed a marginal association with each minor allele associated with a 28% decrease in the odds of having RWMA ($p = 0.046$, Table 4). Figure 3b shows that the association of β 1AR1165 genotypes and RWMA follows an additive model whereas a dominant model might be a better fit for the β 2AR79 SNP, as one additional minor allele has the same protective effect as having both alleles.

Peak Dopamine Requirement: Dopamine dosing and complete covariate information was available for 977 donors. The median maximum dopamine dose required during the donor management period was 5.2 (IQR: 3.0, 10.0) mcg/kg/min. A donor dopamine requirement of

>10 mcg/kg/min is often considered a contraindication for cardiac allograft acceptance for transplantation, and 18% of donors managed from 2001-2006 exhibited this high dopamine requirement. SNPs from both β 1AR and β 2AR genes showed significant association with peak dopamine dose (Table 4). Compared to the β 1AR 1165CC genotype, the GG genotype group shows a significant association with peak dopamine dose >10 mcg/kg/min [OR = 2.63 (95% CI 1.51-4.57)], while the CG genotype was not associated [OR = 1.12 (95% CI 0.78-1.62)], suggestive a recessive model. The β 2AR46 SNP also shows a recessive effect, with the AA (but not GA) genotype significantly associated with peak dopamine requirement [OR = 2.63 (95% CI 1.51-4.57)] compared to the GG genotype group.

Validation cohort: 2007-2008 donors—After 2006, significant staffing changes occurred at CTDN, including the hiring of many new coordinators responsible for donor management. Several major changes in donor management strategies ensued. Donors in this latter cohort had a higher incidence of left ventricular dysfunction, higher peak central venous pressure, worse renal and hepatic function, and lower rates of cardiac allograft acceptance for transplantation (Table 2).

No significant associations were found between β -adrenergic receptor polymorphisms and cardiac allograft function in the 2007-2008 cohort (Table 4). A full model including donors from all years showed a significant interaction between the cohort and the SNPs significantly associated with outcome in the 2001-2006 cohort. In order to investigate whether specific changes in donor management strategies may have accounted for the lack of replication found in the latter cohort, we constructed additional multivariable regression models adjusting for inotrope/vasopressor doses, doses of steroid and thyroid hormone, and hemodynamic differences, among other relevant covariates. We also tested for interactions between the individual β AR polymorphisms and medications administered. Neither strategy accounted for the lack of replication in this validation cohort. Post-hoc power estimates for the replication of the five significant associations observed in the discovery cohort ranged from 20%-36%, below the standard 80% threshold for type II error (Supplementary Table 3).

DISCUSSION

In our discovery cohort of 1,043 organ donors managed between 2001 and 2006 we identified several significant associations between β -adrenergic receptor polymorphisms and the presence of left ventricular dysfunction after brain death. Specifically, genotypes known to be associated with increased sensitivity to circulating catecholamines were associated with a higher risk of LV dysfunction during the donor management period.

The β -adrenergic receptors are located at the cell membrane of cardiomyocytes and mediate the effects of circulating catecholamines.(18) The β 1-adrenergic receptor is the predominant β -adrenergic receptor expressed on the cardiomyocyte and is responsive to circulating epinephrine and to local norepinephrine derived from cardiac sympathetic nerves.(13) In rodents, transgenic cardiac overexpression of β 1-adrenergic receptors causes progressive cardiomyopathy and heart failure.(19, 20) We therefore considered the β 1AR 1165C/G (Arg389Gly) and 145A/G (Ser49Gly) polymorphisms as potential risk factors for donor LV dysfunction, as they affect the sensitivity and response of β 1-adrenergic receptors to circulating catecholamines,(21, 22) which are present at very high levels early after brain death.(5, 23) Similarly, the β 2-adrenergic receptors are also present in human myocardium, as well as in vascular smooth muscle beds. Stimulation of β 2-adrenergic receptors mediates cardiac inotropic and chronotropic effects,(24) induces cardiomyocyte apoptosis,(25) and causes vascular smooth muscle relaxation and vasodilation in response to sympathetic tone.(26) We therefore evaluated the associations between the β 2-adrenergic receptor

polymorphisms 46A/G (Gly16Arg) and 79C/G (Gln27Glu) and cardiac injury after brain death. These polymorphisms in the β 1- and β 2-adrenergic receptors are well-studied and have previously been associated with altered response to sympathetic stimulation,(27) resting heart rate,(22) risk of coronary events,(10) arrhythmias,(12) vascular reactivity,(9) and survival in patients with heart failure.(28)

The initial “catecholamine storm”(23) that occurs after brain death is often followed by a period of functional denervation, during which there is a dominance of vagal parasympathetic or inhibitory effects.(7) This theory is supported by the observed excessive activity of the inhibitory G protein $G_{i\alpha}$ in brain-dead organ donors with LV dysfunction. (29) These dramatic physiologic changes after brain death may mediate the relationship between the “catecholamine insensitive” β 1AR 1165GG genotype and high dopamine requirements during the subsequent donor management period.

Our initial provocative findings were then studied in a second cohort of organ donors managed between 2007 and 2008. The original study findings did not replicate in the latter cohort, and there are several potential explanations for this discrepancy. First, as discussed previously, changes in staffing and donor management strategies may have masked the influence of β AR genetic variation on allograft function. For example, significantly fewer donors in the 2007-2008 cohort were treated with dopamine (a β -receptor agonist), and higher doses of phenylephrine (an α -receptor agonist) were used. This change in inotrope/vasopressor support during donor management may have overshadowed the role of β AR signaling on cardiac function. In addition, there was a dramatic increase in the use of thyroid hormone supplementation between the 2001-2006 cohort (14%) and the 2007-2008 cohort (54%). As thyroid hormone can increase cardiac contractility, this strategic change in donor management could have also masked the relationship seen between β AR genotype and cardiac function in our discovery cohort. Second, there were notable differences in laboratory values, hemodynamics, and allograft function, suggesting that the 2007-2008 cohort was comprised of a “sicker” donor population. This observation may account for the decrease in allograft acceptance for transplantation in the latter cohort. Third, it is possible that unrecognized or unmeasured differences between study cohorts may have accounted for lack of replication. Finally, the smaller sample size in the second cohort (364 versus 1,043 donors) impacted the power to replicate our initial findings (20-36%). Thus, it is possible that our initial findings may have been false positive results, or may represent true associations, but our study was underpowered to replicate.

Understanding the pathophysiology of LV dysfunction after brain death plays a crucial role in the graft selection process for heart transplantation. Currently, approximately 60% of available cardiac grafts are discarded due to stringent acceptance criteria,(17) leading to a great discrepancy between the number of critically-ill patients on the waiting list compared to the number of available grafts for transplantation.(30) While non-utilization of donor hearts is a multi-factorial problem, encompassing diverse donor characteristics and logistical issues, LV dysfunction is the most frequently cited reason for non-utilization.(8, 31) Left ventricular dysfunction in a cardiac donor raises the specter of irreversible cardiac injury which may lead to clinically significant graft dysfunction and graft failure in the transplant recipient. However, animal and human studies now support the hypothesis that catecholamine toxicity plays a central role in reducing myocardial contractility after brain death(29, 32) and that cardiac dysfunction is often reversible in organ donors.(16) Supporting this hypothesis are our discovery cohort findings of significant associations between β AR polymorphisms that mediate myocardial catecholamine sensitivity and LV dysfunction after brain death. Similarly, many transplant centers consider an allograft to be unsuitable if inotrope requirements are high during the donor management period. Our

results suggest that high inotrope requirements may be associated with β AR genotypes that confer insensitivity to circulating catecholamines.

This study has significant strengths and limitations that deserve discussion. First and foremost, this represents the largest existing research database of detailed, rigorously adjudicated clinical and genetic data on over 1,400 potential organ donors managed in the current era. Second, this study represents a unique approach to the study of genetic influences in organ transplantation. We chose to study candidate gene polymorphisms in organ *donors*, and their influences on allograft function. Most genetic studies to-date in organ transplantation have examined associations between *recipient* genetic variation and post-transplant outcomes. Finally, we studied functional β AR polymorphisms that were previously shown to be associated with adverse cardiac outcomes in the general population as a means to study the pathogenesis of cardiac dysfunction after brain death, utilizing a very unique organ donor population. Limitations of this study include non-replication of initial findings in the validation cohort, which we were unable to account for by adjusting for recognized (and measurable) differences in donor management strategies during the study period. This is further exacerbated by the fact that donors were managed at a variety of local hospitals that may have had different medical management strategies prior to assumption of donor management by CTDN staff. Furthermore, characteristics and outcomes of donors managed by CTDN may not be equivalent to donor outcomes in other regions of the country, due to nationwide variations in donor management strategies. We also recognize the possible influence of uncontrolled confounding or population admixture on our genetic analyses. Although we did see consistent results when repeating our analyses in the sub-population of Caucasian donors, subtle population substructure may still be present within this racial group. Finally, complete phenotypic data could not be obtained for every donor, due to the retrospective nature of the data collection.

In conclusion, β -adrenergic receptor polymorphisms may contribute to the development of cardiac dysfunction after brain death. While we initially identified several compelling associations between the β AR SNPs of interest and cardiac function, our findings did not replicate in the validation cohort and there are several potential explanations for these discrepant results, as described above. Additional studies are therefore needed to examine the influence of donor genetic variants on post-transplant outcomes, and to assess for interactions between donor and recipient genetic modifiers in organ transplantation.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Abbreviations

LV	Left ventricular
LVEF	Left ventricular ejection fraction
CTDN	California Transplant Donor Network
RWMA	Regional Wall Motion Abnormalities

βAR	Beta Adrenergic Receptor
SNP	Single Nucleotide Polymorphism
HWE	Hardy Weinberg Equilibrium

REFERENCES

1. Goel R, Johnson F, Mehra MR. Brain injury and ventricular dysfunction: insights into reversible heart failure. *Congest Heart Fail.* 2005; 11(2):99–101. [PubMed: 15860979]
2. Allman FD, Herold W, Bosso FJ, Pilati CF. Time-dependent changes in norepinephrine-induced left ventricular dysfunction and histopathologic condition. *J Heart Lung Transplant.* 1998; 17(10):991–997. [PubMed: 9811407]
3. Lang SA, Maron MB, Bosso FJ, Pilati CF. Temporal changes in left ventricular function after massive sympathetic nervous system activation. *Can J Physiol Pharmacol.* 1994; 72(6):693–700. [PubMed: 7954102]
4. Audibert G, Charpentier C, Seguin-Devaux C, Charretier PA, Gregoire H, Devaux Y, et al. Improvement of donor myocardial function after treatment of autonomic storm during brain death. *Transplantation.* 2006; 82(8):1031–1036. [PubMed: 17060850]
5. Novitzky D, Horak A, Cooper DK, Rose AG. Electrocardiographic and histopathologic changes developing during experimental brain death in the baboon. *Transplant Proc.* 1989; 21(1 Pt 3):2567–2569. [PubMed: 2705268]
6. Novitzky D, Wicomb WN, Cooper DK, Rose AG, Reichart B. Prevention of myocardial injury during brain death by total cardiac sympathectomy in the Chacma baboon. *Ann Thorac Surg.* 1986; 41(5):520–524. [PubMed: 3707246]
7. Samuels MA. The brain-heart connection. *Circulation.* 2007; 116(1):77–84. [PubMed: 17606855]
8. Zaroff JG, Babcock WD, Shiboski SC. The impact of left ventricular dysfunction on cardiac donor transplant rates. *J Heart Lung Transplant.* 2003; 22(3):334–337. [PubMed: 12633701]
9. Cockcroft JR, Gazis AG, Cross DJ, Wheatley A, Dewar J, Hall IP, et al. Beta(2)-adrenoceptor polymorphism determines vascular reactivity in humans. *Hypertension.* 2000; 36(3):371–375. [PubMed: 10988267]
10. Heckbert SR, Hindorff LA, Edwards KL, Psaty BM, Lumley T, Siscovick DS, et al. Beta2-adrenergic receptor polymorphisms and risk of incident cardiovascular events in the elderly. *Circulation.* 2003; 107(15):2021–2024. [PubMed: 12682000]
11. Iwai C, Akita H, Kanazawa K, Shiga N, Terashima M, Matsuda Y, et al. Arg389Gly polymorphism of the human beta1-adrenergic receptor in patients with nonfatal acute myocardial infarction. *Am Heart J.* 2003; 146(1):106–109. [PubMed: 12851615]
12. Kanki H, Yang P, Xie HG, Kim RB, George AL Jr, Roden DM. Polymorphisms in beta-adrenergic receptor genes in the acquired long QT syndrome. *J Cardiovasc Electrophysiol.* 2002; 13(3):252–256. [PubMed: 11942593]
13. Small KM, Wagoner LE, Levin AM, Kardia SL, Liggett SB. Synergistic polymorphisms of beta1- and alpha2C-adrenergic receptors and the risk of congestive heart failure. *N Engl J Med.* 2002; 347(15):1135–1142. [PubMed: 12374873]
14. Zaroff JG, Pawlikowska L, Miss JC, Yarlagadda S, Ha C, Achrol A, et al. Adrenoceptor polymorphisms and the risk of cardiac injury and dysfunction after subarachnoid hemorrhage. *Stroke.* 2006; 37(7):1680–1685. [PubMed: 16728691]
15. Hsu TM, Kwok PY. Homogeneous primer extension assay with fluorescence polarization detection. *Methods Mol Biol.* 2003; 212:177–187. [PubMed: 12491910]
16. Zaroff JG, Babcock WD, Shiboski SC, Solinger LL, Rosengard BR. Temporal changes in left ventricular systolic function in heart donors: results of serial echocardiography. *J Heart Lung Transplant.* 2003; 22(4):383–388. [PubMed: 12681416]
17. Zaroff, JG.; Rosengard, BR.; Armstrong, WF.; Babcock, WD.; D'Alessandro, A.; Dec, GW., et al. *Circulation*; Consensus conference report: maximizing use of organs recovered from the cadaver donor: cardiac recommendations; Crystal City, Va. March 28-29, 2001; 2002. p. 836-841.

18. Green SA, Turki J, Innis M, Liggett SB. Amino-terminal polymorphisms of the human beta 2-adrenergic receptor impart distinct agonist-promoted regulatory properties. *Biochemistry*. 1994; 33(32):9414–9419. [PubMed: 7915137]
19. Bisognano JD, Weinberger HD, Bohlmeyer TJ, Pende A, Reynolds MV, Sastravaha A, et al. Myocardial-directed overexpression of the human beta(1)-adrenergic receptor in transgenic mice. *J Mol Cell Cardiol*. 2000; 32(5):817–830. [PubMed: 10775486]
20. Engelhardt S, Hein L, Wiesmann F, Lohse MJ. Progressive hypertrophy and heart failure in beta1-adrenergic receptor transgenic mice. *Proc Natl Acad Sci U S A*. 1999; 96(12):7059–7064. [PubMed: 10359838]
21. Mason DA, Moore JD, Green SA, Liggett SB. A gain-of-function polymorphism in a G-protein coupling domain of the human beta1-adrenergic receptor. *J Biol Chem*. 1999; 274(18):12670–12674. [PubMed: 10212248]
22. Ranade K, Jorgenson E, Sheu WH, Pei D, Hsiung CA, Chiang FT, et al. A polymorphism in the beta1 adrenergic receptor is associated with resting heart rate. *Am J Hum Genet*. 2002; 70(4):935–942. [PubMed: 11854867]
23. Hachinski VC, Smith KE, Silver MD, Gibson CJ, Ciriello J. Acute myocardial and plasma catecholamine changes in experimental stroke. *Stroke*. 1986; 17(3):387–390. [PubMed: 3715933]
24. Brodde OE, Michel MC. Adrenergic and muscarinic receptors in the human heart. *Pharmacol Rev*. 1999; 51(4):651–690. [PubMed: 10581327]
25. Menon B, Singh M, Ross RS, Johnson JN, Singh K. beta-Adrenergic receptor-stimulated apoptosis in adult cardiac myocytes involves MMP-2-mediated disruption of beta1 integrin signaling and mitochondrial pathway. *Am J Physiol Cell Physiol*. 2006; 290(1):C254–261. [PubMed: 16148033]
26. Feldman RD. Beta-adrenergic receptor alterations in hypertension--physiological and molecular correlates. *Can J Physiol Pharmacol*. 1987; 65(8):1666–1672. [PubMed: 2825942]
27. Dishy V, Sofowora GG, Xie HG, Kim RB, Byrne DW, Stein CM, et al. The effect of common polymorphisms of the beta2-adrenergic receptor on agonist-mediated vascular desensitization. *N Engl J Med*. 2001; 345(14):1030–1035. [PubMed: 11586955]
28. Borjesson M, Magnusson Y, Hjalmarson A, Andersson B. A novel polymorphism in the gene coding for the beta(1)-adrenergic receptor associated with survival in patients with heart failure. *Eur Heart J*. 2000; 21(22):1853–1858. [PubMed: 11052857]
29. Owen VJ, Burton PB, Michel MC, Zolk O, Bohm M, Pepper JR, et al. Myocardial dysfunction in donor hearts. A possible etiology. *Circulation*. 1999; 99(19):2565–2570. [PubMed: 10330389]
30. Gridelli B, Remuzzi G. Strategies for making more organs available for transplantation. *N Engl J Med*. 2000; 343(6):404–410. [PubMed: 10933740]
31. Hornby K, Ross H, Keshavjee S, Rao V, Shemie SD. Non-utilization of hearts and lungs after consent for donation: a Canadian multicentre study. *Can J Anaesth*. 2006; 53(8):831–837. [PubMed: 16873351]
32. White M, Wiechmann RJ, Roden RL, Hagan MB, Wollmering MM, Port JD, et al. Cardiac beta-adrenergic neuroeffector systems in acute myocardial dysfunction related to brain injury. Evidence for catecholamine-mediated myocardial damage. *Circulation*. 1995; 92(8):2183–2189. [PubMed: 7554200]

LV Ejection Fraction By β 1-AR Genotypes

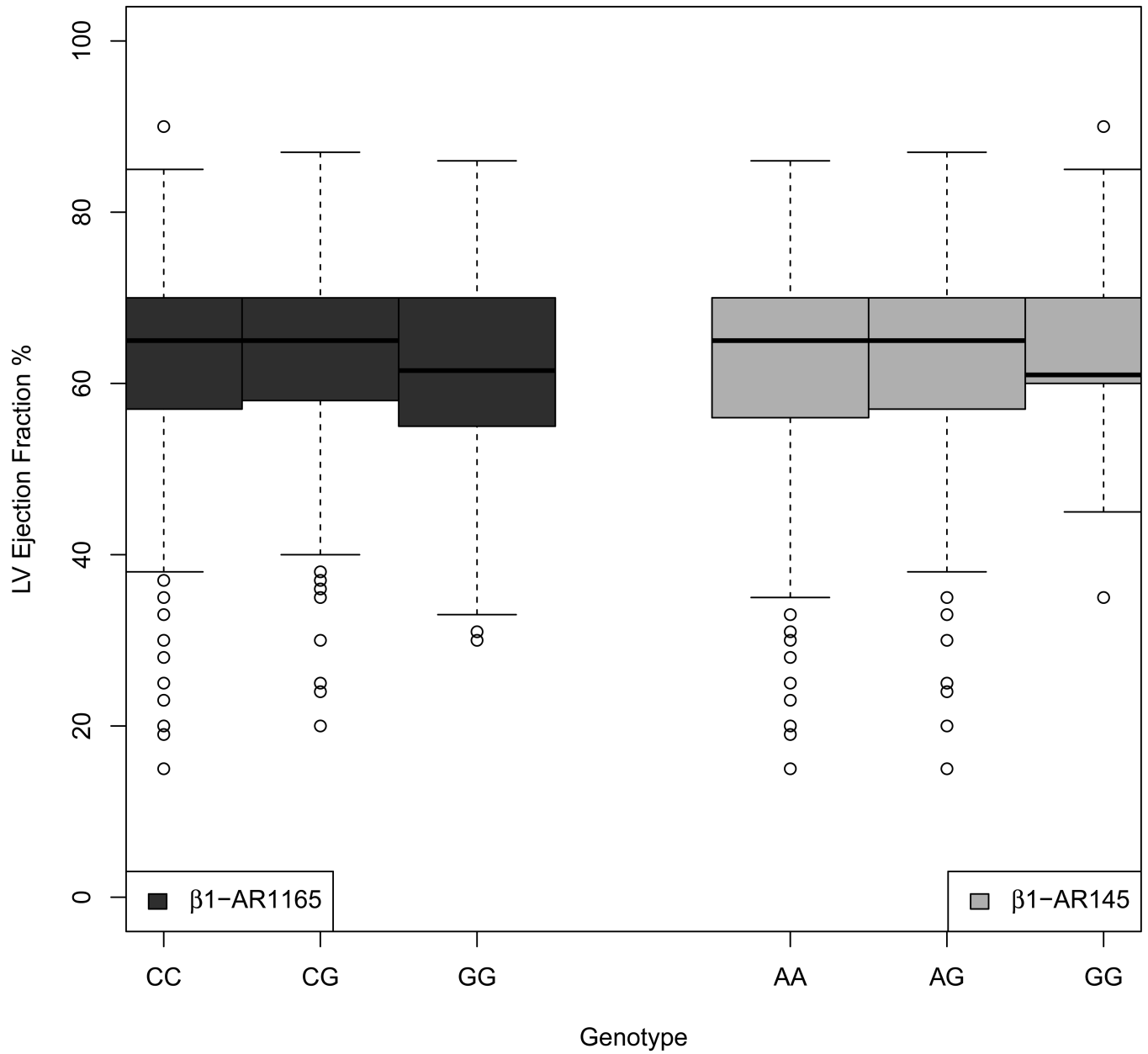


Figure 1. LV ejection fraction, by β 1-adrenergic receptor genotypes, in CTDN donors (2001-2006 cohort)

LV Ejection Fraction By β 2-AR Genotypes

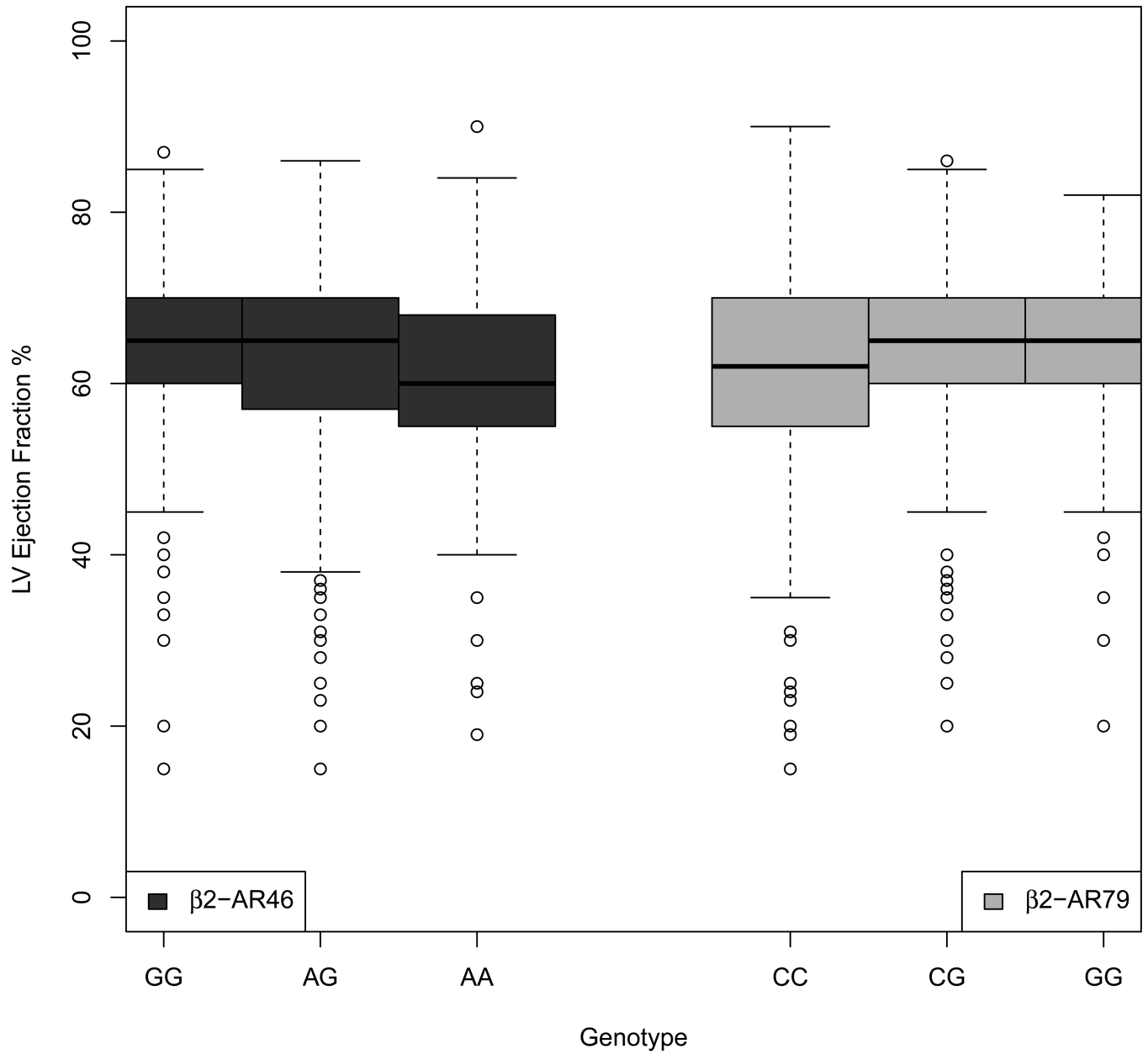


Figure 2. LV ejection fraction, by β 2-adrenergic receptor genotypes, in CTDN donors (2001-2006 cohort)

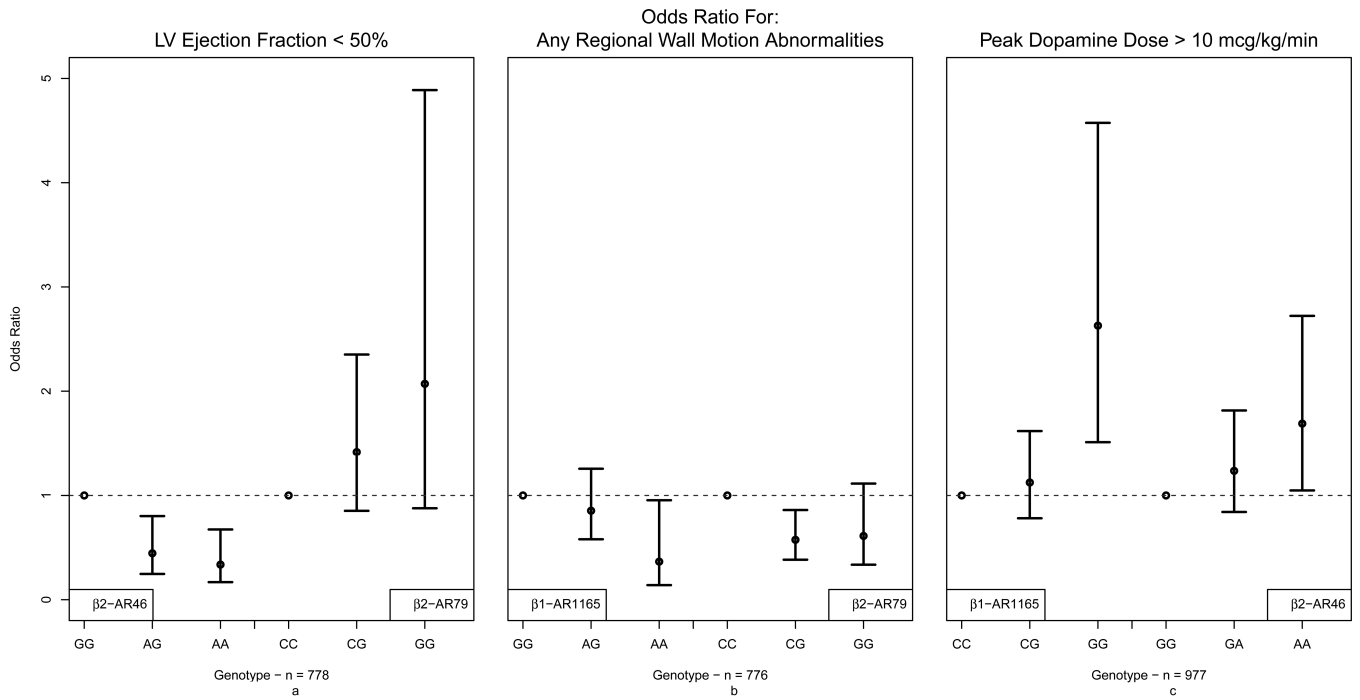


Figure 3. Odds ratio for LV ejection fraction < 50%, regional wall motion abnormalities, and peak dopamine requirement > 10 mcg/kg/min, by donor β -adrenergic receptor in CTDN donors (2001-2006 cohort)

Table 1

Rationale for Polymorphism Selection

Gene	Variant	dbSNP ID	Functional Rationale	Clinical Rationale	References
β 1AR (ADRB1)	1165 C>G (Arg 389Gly)	rs 1801252	C allele: \uparrow response to β -agonist stimulation	CC genotype associated with \uparrow risk of AMI CC genotype associated with \uparrow risk of HF	11, 13, 21
	145 A>G (Ser49Gly)	rs1801253	G allele: \downarrow response to β -agonist stimulation	GG or AG genotype associated with improved survival in HF	22, 28
β 2AR (ARDB2)	46 G>A (Gly16Arg)	rs1042713	G allele: alters agonist-promoted downregulation	GG genotype associated with \uparrow left ventricular ejection fraction	9, 10, 12
	79 C>G Gln27Glu	rs1042714	G allele: alters agonist-promoted downregulation	CC genotype associated with \uparrow risk of CAD events	9, 10, 12, 18

AMI: acute myocardial infarction; HF: heart failure; CAD: coronary artery disease

Table 2

Donor characteristics, by study cohort

	Discovery cohort 2001-2006	Validation cohort 2007-2008	p-value*
Donors, n	1043	364	
Age (years)	42 (24, 52)	41 (25.5, 51)	0.85
Female	41%	37%	0.13
Body mass index (kg/m ²)	25.5 (22.2, 29.4)	26.4 (23.0, 30.7)	0.0025
Race/Ethnicity			0.9300
Caucasian	55%	53%	
African-American	10%	11%	
Hispanic	25%	26%	
Asian	8%	8%	
Other	2%	2%	
Cause of death			<0.0001
Anoxia	11%	19%	
Cerebrovascular Accident/Stroke	48%	42%	
Head Trauma	40%	38%	
CNS	0.3%	0.5%	
Other	0.6%	0.5%	
Medical history			
Hypertension	28%	30%	0.16
Diabetes	7%	11%	0.01
Smoking	51%	52%	0.35
History of coronary artery disease	4%	6%	0.32
Drug use (cocaine, methamphetamines, inhaled drugs)	41%	38%	0.41
Hemodynamics			
Peak heart rate (bpm)	124 (110, 137)	122 (110, 136)	0.12
Final heart rate (bpm)	100 (90, 111)	100 (88, 110)	0.11
Peak systolic blood pressure (mmHg)	157 (144, 178)	161 (149, 180)	0.01
Final systolic blood pressure (mmHg)	124 (110, 139)	125 (114, 139)	0.10
Peak diastolic blood pressure (mmHg)	90 (78, 100)	89 (76, 100)	0.97
Final diastolic blood pressure (mmHg)	68 (60, 77)	69 (60, 76)	0.52
Peak central venous pressure (mmHg)	11 (9, 13)	12 (10, 15)	< 0.001
Final central venous pressure (mmHg)	7 (6, 9)	9 (7, 10)	< 0.001
Laboratory values			
Peak sodium (mmol/L)	153 (148, 160)	155 (149, 161)	0.0007
Peak creatinine (mg/dL)	1.3 (1.0, 1.6)	1.4 (1.1, 2.0)	< 0.001
Peak ALT (U/L)	44 (28, 84)	59 (36, 133.5)	< 0.001
Peak AST (U/L)	66 (40, 128)	82 (46, 204.5)	< 0.001
Lowest hemoglobin (g/dL)	9.8 (8.4, 11.6)	9.1 (8.1, 10.7)	< 0.001
Peak troponin (ng/mL)	0.38 (0.10, 1.84)	0.29 (0.05, 1.69)	0.04
Peak CPK-MB (ng/mL)	1.6 (0.8, 3.1)	0.07 (0.04, 1.3)	0.12

	Discovery cohort 2001-2006	Validation cohort 2007-2008	p-value*
Echocardiogram			
Echocardiogram performed, %	76%	71.0%	0.06
Median LVEF, %	65 (57, 70)	64 (55, 70)	0.07
EF<50%, %	11%	15%	0.05
Regional wall motion abnormalities, %	20%	20%	0.96
Left ventricular hypertrophy [†] , %	11%	9%	0.35
Vasoactive Medications			
Dopamine (%)	83%	65%	<0.001
Peak dopamine dose, mcg/kg/min	5.2 (3.0, 10.0)	5.0 (3.9, 8.8)	0.87
Peak dopamine>10 mcg/kg/min, %	18%	14%	0.18
Phenylephrine (%)	74%	78%	0.13
Peak phenylephrine dose, mcg/min	67 (8, 140)	109 (50, 200)	<0.0001
Epinephrine (%)	6%	7%	0.57
Esmolol (%)	17%	17%	0.99
Hormone replacement			
Steroids	98%	95%	0.0012
Solmedrol dose over 24 hrs (grams)	2.0 (1.0, 2.0)	3.0 (2.0, 3.0)	<0.0001
Thyroid hormone (T4), %	14%	54%	<0.0001
Cardiac allograft acceptance for transplantation	47%	39%	0.0062

* Wilcoxon Test for continuous variables (median with interquartile range), chi2 test for categorical variables

[†] defined as septal or posterior wall thickness > 1.1 cm; ALT, alanine aminotransferase; AST, aspartate aminotransferase; CPK-MB, creatine phosphokinase, myocardial fraction; LVEF, left ventricular ejection fraction; EF, ejection fraction

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Table 3
Beta Adrenergic Receptor Polymorphism Genotype Frequency, by Study Cohort and Race

βAR Receptor Polymorphism	Discovery cohort 2001-2006					Validation cohort 2007-2008				
	Overall	Caucasian	Hispanic	African-American	Asian	Overall	Caucasian	Hispanic	African-American	Asian
BIAR 1165 (Arg389Gly)										
CC	59%	56%	70%	44%	61%	63%	63%	73%	39%	61%
CG	34%	37%	25%	42%	32%	32%	33%	24%	41%	36%
GG	7%	7%	5%	14%	7%	6%	5%	3%	20%	3%
BIAR 145 (Ser49Gly)										
AA	71%	78%	56%	63%	74%	67%	77%	46%	64%	71%
AG	25%	20%	35%	29%	21%	28%	22%	45%	26%	21%
GG	4%	1%	9%	8%	5%	5%	1%	10%	10%	9%
B2AR 46 (Gly16Arg)										
GG	36%	39%	36%	25%	27%	40%	40%	37%	43%	27%
AG	46%	45%	47%	54%	45%	38%	39%	40%	36%	39%
AA	18%	15%	18%	21%	28%	22%	21%	24%	21%	33%
B2AR 79 (Gln27Glu)										
CC	50%	37%	63%	66%	69%	50%	35%	65%	64%	73%
CG	38%	43%	32%	29%	29%	37%	48%	27%	18%	20%
GG	13%	20%	5%	5%	2%	14%	17%	8%	18%	7%

Table 4

Associations Between Beta Adrenergic Receptor Polymorphisms and Cardiac Allograft Dysfunction, by Study Cohort

	Discovery cohort 2001-2006		Validation cohort 2007-2008	
	OR (95% CI)	p-value	OR (95% CI)	p-value
LV Ejection Fraction < 50%				
	N=778		N=249	
B1AR 1165G	1.14 (0.77 – 1.71)	0.51	1.04 (0.55 – 1.95)	0.91
β1AR 145G	1.04 (0.65 – 1.66)	0.86	2.00 (0.86 – 4.70)	0.11
β2AR 46A	0.60 (0.40, 0.89)	0.012	1.68 (0.89 – 3.17)	0.11
β2AR 79G	1.04 (0.67 – 1.63)	0.86	1.17 (0.63 – 2.19)	0.62
LV Regional Wall Motion Abnormalities				
	N=776		N=261	
B1AR 1165G	0.72 (0.53 – 0.99)	0.046	0.86 (0.48 – 1.51)	0.59
β1AR 145G	0.92 (0.65 – 1.30)	0.64	0.75 (0.39 – 1.44)	0.38
β2AR 46A	1.06 (0.78 – 1.43)	0.71	1.03 (0.60 – 1.77)	0.90
β2AR 79G	0.73 (0.53 – 1.02)	0.06	1.22 (0.71 – 2.10)	0.48
Peak Dopamine > 10 mcg/kg/min				
	N=977		N=234	
B1AR 1165G	1.37 (1.05 – 1.78)	0.020	0.45 (0.19 – 1.07)	0.07
β1AR 145G	0.80 (0.57 – 1.14)	0.23	0.87 (0.42 – 1.82)	0.72
β2AR 46A	1.39 (1.04 – 1.85)	0.026	1.03 (0.56 – 1.89)	0.92
β2AR 79G	1.15 (0.85 – 1.57)	0.36	1.21 (0.63 – 2.33)	0.57

Multivariable regression models assuming additivity in minor allele adjusted for donor age, sex, race, cause of death, and the other polymorphism (SNP) within the same gene.

The point estimate is the odds ratio for each additional minor allele.

LV: left ventricular