

Published in final edited form as:

*Angew Chem Int Ed Engl.* 2011 September 12; 50(38): 8895–8899. doi:10.1002/anie.201103365.

## Combinatorial Catalysis Assisted by a Visible Enzymatic Beacon in Real Time: New Synthetically Versatile (Pseudo)Halometalation/Carbocyclizations

Jacob A. Friest, Sylvain Broussy, Woo Jin Chung, and Prof. David B. Berkowitz

Department of Chemistry, University of Nebraska, Lincoln, NE 68588 (USA), Fax: (+001) 402-472-9402

David B. Berkowitz: dbb@unlserve.unl.edu

### Keywords

stereoselective catalysis; enzyme catalysis; halometalation; carbocyclization; xanthanolides

Combinatorial approaches to catalysis have made an impact in targeted transformation development, including Ag-mediated carbene insertion,<sup>[1]</sup> Sc-pybox-based asymmetric cyclopropanation,<sup>[2]</sup> and Rh/Ir-based asymmetric hydrogenation.<sup>[3]</sup> Useful design elements have emerged from these studies, e.g. the value of ligand self assembly,<sup>[4]</sup> or of the inclusion of peptide-like structural elements<sup>[5],[6],[7]</sup> in building ligand arrays. Efficient screening methods are of paramount importance for such efforts. Methods based on fluorescence,<sup>[8]</sup> REMPI,<sup>[9]</sup> MS,<sup>[10]</sup> NMR,<sup>[11]</sup> and IR thermography<sup>[12]</sup> have appeared. A chromophore may be installed into the substrate<sup>[13]</sup> or product<sup>[14]</sup> of the reaction under study. Alternatively, one can exploit chromophores inherent in proteins<sup>[15]</sup> or enzyme-associated reactions,<sup>[16]</sup> and use these sensors to report back on product formation and composition.

Our group has developed an in situ enzymatic screening (ISES) approach whereby an organometallic reaction under study is coupled to an enzymatic reporting reaction in real time.<sup>[17]</sup> This screening method led to the discovery of the first asymmetric allylic amination with Ni(0)<sup>[18]</sup> and to the identification of novel salen ligands with promise for asymmetric synthesis.<sup>[19]</sup> Those approaches involved dehydrogenase enzymes<sup>[20]</sup> as sensors, utilizing the inherent nicotinamide cofactor to provide a UV signal.

This Communication describes an important new ISES-mode in which the reporting enzymes lead to a visible signal in real time. The advantages of colorimetric approaches have been articulated<sup>[13, 21]</sup> and include the ability to screen a diverse array of catalysts with the naked eye, without employing specialized equipment, increasing convenience and throughput.

Synthetically, this study was directed at developing formal halometalation/carbocyclization transformations. One can envision this providing a rapid entry into the cores of terpenoid natural products featuring exomethylene  $\gamma$ -lactones (Scheme 1). The 6-*exo-trig* substrate **6** would lead into cores of xerophilusin and crassin, a specific modulator of STAT phosphorylation.<sup>[27]</sup> The 5-*exo-trig* substrate **3** is designed to give 5,7-sesquiterpenoid

Correspondence to: David B. Berkowitz, dbb@unlserve.unl.edu.

Homepage: <http://chem.unl.edu/dbb/site/welcome.html>

Supporting information for this article is available on the WWW under <http://www.angewandte.org> or from the author.

lactone cores.<sup>[28]</sup> Key natural products (NPs) here include the guaianolide, ixerin Y (**1**),<sup>[29]</sup> and xanthatin (**2**), which shows anti-MRSA,<sup>[30]</sup> antifungal<sup>[31]</sup> and anti-ulcer<sup>[32]</sup> activity.

Related NP-derived  $\alpha$ -methylene butyrolactone moieties appear to Michael-add Cys-38<sup>[33]</sup> of the transcription factor NF- $\kappa$ B,<sup>[34]</sup> thereby blocking DNA binding.<sup>[35]</sup> Our synthetic approach is particularly attractive in light of such SAR postulates, as it would deliver the NP core with a  $\beta$ -halo- $\alpha,\beta$ -unsaturated lactone moiety, for potential target capture, *in vivo*, and also to tap into cross-coupling chemistry *ex vivo*, for chain extension/library elaboration. The drive toward streamlined methods for the construction of such NP cores is motivated by the effectiveness of NP-core based chemical biology libraries in defining studies by groups such as Schreiber,<sup>[22]</sup> Waldmann,<sup>[23]</sup> Shair,<sup>[24]</sup> Arndt<sup>[25]</sup> and Snapper.<sup>[26]</sup>

While the chemistry envisioned in Scheme 1 remained largely unexplored, there was some precedent from the work of Lu,<sup>[36]</sup> primarily with acetoxy-metalation/carbocyclization, employing Pd(II) catalysis in neat acetic acid<sup>[37]</sup> as solvent. We set out to examine a much broader spectrum of metal, (pseudo)halide and substrate space, combinatorially, using visual colorimetric-ISES for higher throughput.

We demonstrate herein that the combination of alcohol oxidase and peroxidase serves as an effective reporting duo for the title transformation (Figure 1). By utilizing ABTS [2,2-azino-bis(3-ethylbenzothiazoline-6-sulfonate)], as peroxidase cofactor, one achieves particular sensitivity. This is because each molecule of alcohol (by)product emanating from the organic reaction of interest that is oxidized by the alcohol oxidase reporter gives rise to two equivalents of ABTS radical cation, providing for an intense ( $\epsilon_{405-414}$  (2 ABTS<sup>•+</sup>)  $\sim$  70,000 M<sup>-1</sup>cm<sup>-1</sup>)<sup>[38]</sup> colorimetric signal in the visible range (jungle green). This allows for first pass scanning of a large number of potential catalytic combinations with the naked eye (Figure 2). A more quantitative ranking (relative rates) may then be obtained by UV-visible spectrophotometry on first pass hits (Fig. 3).

A broad array of 64 metal catalyst candidates was chosen; subdivided into 4 groups of 16 catalysts each, as detailed in the SI. These were screened against 6 (pseudo)halides (LiF, LiCl, LiBr, LiCN, LiOCN, LiSCN) and 3 candidate substrates (**3**, **7**, **8**), creating a 64x6x3 = 1152 combinatorial array. Figure 2 shows a 96 well tray for the metal set III vs. substrate **3**. These were run in a convenient 300  $\mu$ L format (200  $\mu$ L organic/100  $\mu$ L aq. enz). One sees clear positive readouts for the combination of LiBr with both Rh(II)-perfluorocarboxylates (in contrast to the Rh(II) carboxylate catalyst), as well as with Pd(II)(acac) [but not Ni(II)(acac)] and Cl<sub>2</sub>Pd(NCPh)<sub>2</sub> [but not Cl<sub>2</sub>Pt(NCPh)<sub>2</sub>].

The most interesting hits in the colorimetric tray screen were next “cherry-picked” visually, and then ranked more quantitatively by spectrophotometric analysis in the cuvette (Fig. 3). As can be seen, for Pd(II), the cyclization chemistry proceeds efficiently with (PhCN)<sub>2</sub>PdCl<sub>2</sub> and LiBr for both the 5-*exo-trig*-ester and -ether substrates. Acetic acid clearly is not necessary for these cyclizations. Among other Pd(II) catalysts screened, Pd(acac)<sub>2</sub>, gave the next fastest rates.

However, the most generally effective catalytic combination found was LiBr with the Rh(II)-perfluorocarboxylates, providing efficient formal bromorhodiation/carbocyclization, across all three test substrates, in stark contrast to Rh(II)-acetate dimer, and all Rh(I)- and Rh(III)-complexes examined. This reactivity was verified under standard RB flask conditions, through which product identity, stereochemistry and yield were established (Figure 4). Note that the cyclizations are highly diastereoselective, giving the 1,2-*trans* stereochemistry for the xanthatin core from **3**, and the 1,3-*cis* stereochemistry for the

crassin-type core from **7**. Also of interest, catalyst loading could be lowered to 2.5 mol%, without compromise in yield, by gentle heating or sonication (SI).

This would appear to constitute a new reaction mode for the Rh(II)/LiX combination. Control experiments established that this reactivity is not a function of stray TFA (see SI). The disparate reactivity of Rh(II)-perfluorocarboxylates vs. Rh(II)-carboxylates is reminiscent of observations by Padwa and Doyle, the tendency of only the former to promote electrophilic aromatic substitution over carbene insertion.<sup>[39]</sup> Clearly this unusual and valuable reactivity warrants further exploration.

Perhaps of equal significance, the combination of (PhCN)<sub>2</sub>PdCl<sub>2</sub> with LiSCN yields an unprecedented formal thiocyno-palladation/carbocyclization transformation. As such, this reaction manifold assembles a cyclic NP-core bearing a terminal vinyl thiocyanate in one operation [*Note*: product structure verified both spectroscopically and chemically-SI]. Given the importance of the thiocyanate functionality for elegant vibrational Stark effect studies or active site environment by Boxer,<sup>[40]</sup> this transformation will likely be of real value to chemical biologists.

We next utilized the new Rh(II)-perfluorocarboxylate chemistry to fashion a small xanthatin-core library, based upon stereo-controlled synthesis, followed by tailoring chemistry (Scheme 2). Alpine-Borane-mediated ynone reduction establishes the absolute stereochemistry,<sup>[41]</sup> (see SI), while the bromometallation/carbocyclization sets the relative stereochemistry. Finally, RCM yields the desired xanthanolide core, with the bromomethylene lactone undergoing Pd-mediated chain extension reactions as projected (Scheme 3).

Note that the standard Sonagashira coupling proceeds with double bond migration, yielding bicyclic dienolate in **15**. Use of modified Stille conditions (i.e. stannylated acetylene) prevents this to give ynenolate **16**. All analogues feature more extended Michael acceptors of potential interest, given the mechanistic hypothesis that has been advanced (*vide supra*). In summary, the first application of visible, colorimetric ISES has uncovered both a generalizable Rh(II)-mediated halometallation/carbocyclization and the first formal thiocyanometallation/carbocyclization. Current efforts are to explore the scope of these new tandem bond constructions, and of the colorimetric screen that led to their discovery.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

DBB currently serves as a Program Director at the National Science Foundation. This research was facilitated by the Individual Research & Development (IR/D) program associated with that appointment. The authors acknowledge NSF-CHE-0911732 for research support. We thank the NIH (SIG-1-510-RR-06307) and NSF (CHE-0091975, MRI-0079750) for NMR instrumentation, and the NIH (RR016544) for facilities renovation.

## References

1. Burgess K, Lim HJ, Porte AM, Sulikowski GA. *Angew Chem, Int Ed.* 1996; 35:220–222.
2. Moye-Sherman D, Welch MB, Reibenspies J, Burgess K. *Chem Commun.* 1998:2377–2378.
3. a) Ager DJ, Lefort L, de Vries JG. *ACS Symp Ser.* 2009; 1009:239–250. b) Reetz MT, Bondarev O. *Angew Chem, Int Ed.* 2007; 46:4523–4526. c) de Vries JG, Lefort L. *Chemistry--A European Journal.* 2006; 12:4722–4734.

4. a) Weis M, Waloch C, Seiche W, Breit B. *J Am Chem Soc.* 2006; 128:4188–4189. [PubMed: 16568968] b) Takacs JM, Chaiseeda K, Moteki SA, Reddy DS, Wu D, Chandra K. *Pure Appl Chem.* 2006; 78:501–509.
5. a) Fowler BS, Mikochik PJ, Miller SJ. *J Am Chem Soc.* 2010; 132:2870–2871. [PubMed: 20158213] b) Gustafson JL, Lim D, Miller SJ. *Science.* 2010; 328:1251–1255. [PubMed: 20522769] c) Jordan PA, Kayser-Bricker KJ, Miller SJ. *Proc Natl Acad Sci U S A.* 2010; 107:20620–20624. [PubMed: 20439750] d) Agarkov A, Greenfield S, Xie D, Pawlick R, Starkey G, Gilbertson SR. *Biopolymers.* 2006; 84:48–73. [PubMed: 16235230]
6. Francis MB, Jacobsen EN. *Angew Chem, Int Ed.* 1999; 38:937–941.
7. Wieland LC, Vieira EM, Snapper ML, Hoveyda AH. *J Am Chem Soc.* 2009; 131:570–576. [PubMed: 18980303]
8. a) Biswas R, Maillard N, Kofoed J, Reymond JL. *Chem Commun.* 2010; 46:8746–8748. b) Becker S, Hoebenreich H, Vogel A, Knorr J, Wilhelm S, Rosenau F, Jaeger KE, Reetz MT, Kolmar H. *Angew Chem, Int Ed.* 2008; 47:5085–5088. c) Lewis WG, Magallon FG, Fokin VV, Finn MG. *J Am Chem Soc.* 2004; 126:9152–9153. [PubMed: 15281783] d) Stauffer SR, Hartwig JF. *J Am Chem Soc.* 2003; 125:6977–6985. [PubMed: 12783551] e) Jarvo ER, Evans CA, Copeland GT, Miller SJ. *J Org Chem.* 2001; 66:5522–5527. [PubMed: 11485477]
9. Senkan SM. *Nature.* 1998; 394:350–353.
10. a) Lichter PA, Miller SJ. *ACS Combinatorial Science.* 2011; 13:321–326. [PubMed: 21417485] b) Ebner C, Muller CA, Markert C, Pfaltz A. *J Am Chem Soc.* 2011; 133:4710–4713. [PubMed: 21401128] c) Wassenaar J, Jansen E, van Zeist WJ, Bickelhaupt FM, Siegler MA, Spek AL, Reek JNH. *Nature: Chemistry.* 2010; 2:417–421. d) Mueller CA, Pfaltz A. *Angew Chem, Int Ed.* 2008; 47:3363–3366. e) Chen P. *Angew Chem, Int Ed.* 2003; 42:2832–2847.
11. a) Reetz MT, Tielmann P, Eipper A, Ross A, Schlotterbeck G. *Chem Commun.* 2004:1366–1367. b) Evans MA, Morken JP. *J Am Chem Soc.* 2002; 124:9020–9021. [PubMed: 12148984]
12. a) Reetz MT, Becker MH, Liebl M, Furstner A. *Angew Chem, Int Ed.* 2000; 39:1236–1239. b) Taylor SJ, Morken JP. *Science.* 1998; 280:267–270. [PubMed: 9535652]
13. Loch JA, Crabtree RH. *Pure Appl Chem.* 2001; 73:119–128.
14. a) Lavastre O, Morken JP. *Angew Chem, Int Ed.* 1999; 38:3163–3165. b) Moreira R, Havranek M, Sames D. *J Am Chem Soc.* 2001; 123:3927–3931. [PubMed: 11457142]
15. a) Matsushita H, Yamamoto N, Meijler MM, Wirsching P, Lerner RA, Matsushita M, Janda KD. *Molecular BioSystems.* 2005; 1:303–306. [PubMed: 16880995] b) Matsushita M, Yoshida K, Yamamoto N, Wirsching P, Lerner RA, Janda KD. *Angew Chem, Int Ed.* 2003; 42:5984–5987. c) Taran F, Gauchet C, Mohar B, Meunier S, Valleix A, Renard PY, Creminon C, Grassi J, Wagner A, Mioskowski C. *Angew Chem, Int Ed.* 2002; 41:124–127.
16. a) Hamberg A, Lundgren S, Penhoat M, Moberg C, Hult K. *J Am Chem Soc.* 2006; 128:2234–2235. [PubMed: 16478176] b) Sprout CM, Seto CT. *Org Lett.* 2005; 7:5099–5102. [PubMed: 16235967] c) Abato P, Seto CT. *J Am Chem Soc.* 2001; 123:9206–9207. [PubMed: 11552847]
17. Berkowitz DB, Bose M, Choi S. *Angew Chem, Int Ed.* 2002; 41:1603–1607.
18. a) Berkowitz DB, Shen W, Maiti G. *Tetrahedron: Asymmetry.* 2004; 15:2845–2851. b) Berkowitz DB, Maiti G. *Org Lett.* 2004; 6:2661–2664. [PubMed: 15281738]
19. a) Dey S, Powell DR, Hu C, Berkowitz DB. *Angew Chem, Int Ed.* 2007; 46:7010–7014. b) Dey S, Karukurichi KR, Shen W, Berkowitz DB. *J Am Chem Soc.* 2005; 127:8610–8611. [PubMed: 15954763]
20. For complementary examples of ADHs in asymmetric synthesis, see: Applegate GA, Cheloha RW, Nelson DL, Berkowitz DB. *Chem Commun.* 2011; 47:2420–2422. Friest JA, Maezato Y, Broussy S, Blum P, Berkowitz DB. *J Am Chem Soc.* 2010; 132:5930–5931. [PubMed: 20377222] Broussy S, Cheloha RW, Berkowitz DB. *Org Lett.* 2009; 11:305–308. [PubMed: 19128188]
21. a) Folmer-Andersen JF, Lynch VM, Anslyn EV. *J Am Chem Soc.* 2005; 127:7986–7987. [PubMed: 15926802] b) Kawatsura M, Hartwig JF. *Organometallics.* 2001; 20:1960–1964.
22. a) Schreiber SL. *Nat Chem Biol.* 2007; 3:352. [PubMed: 17576414] b) Burke MD, Berger EM, Schreiber SL. *J Am Chem Soc.* 2004; 126:14095–14104. [PubMed: 15506774]

23. a) Basu S, Ellinger B, Rizzo S, Deraeve C, Schurmann M, Preut H, Arndt HD, Waldmann H. *Proc Natl Acad Sci U S A*. 2011; 108:6805–6810. [PubMed: 21415367] b) Waldmann H. *Nat Chem Biol*. 2009; 5:76–77. [PubMed: 19148173]
24. a) Pelish HE, Peterson JR, Salvarezza SB, Rodriguez-Boulan E, Chen JL, Stamnes M, Macia E, Feng Y, Shair MD, Kirchhausen T. *Nat Chem Biol*. 2006; 2:39–46. [PubMed: 16408091] b) Pelish HE, Westwood NJ, Feng Y, Kirchhausen T, Shair MD. *J Am Chem Soc*. 2001; 123:6740–6741. [PubMed: 11439080]
25. a) Hackenberger CPR, Arndt HD, Schwarzer D. *Chemie in Unserer Zeit*. 2010; 44:198–206. b) Walther T, Renner S, Waldmann H, Arndt HD. *ChemBioChem*. 2009; 10:1153–1162. [PubMed: 19360807]
26. a) Kim BG, Chun TG, Lee HY, Snapper ML. *Bioorg Med Chem*. 2009; 17:6707–6714. [PubMed: 19692248] b) Radeke HS, Digits CA, Bruner SD, Snapper ML. *J Org Chem*. 1997; 62:2823–2831. [PubMed: 11671645]
27. Krutzik PO, Crane JM, Clutter MR, Nolan GP. *Nat Chem Biol*. 2008; 4:132–142. [PubMed: 18157122]
28. a) Kummer DA, Brennehan JB, Martin SF. *Org Lett*. 2005; 7:4621–4623. [PubMed: 16209494] b) Evans MA, Morken JP. *Org Lett*. 2005; 7:3371–3373. [PubMed: 16018663] c) Nosse B, Chhor RB, Jeong WB, Boehm C, Reiser O. *Org Lett*. 2003; 5:941–944. [PubMed: 12633111]
29. Ma J-Y, Wang Z-T, Xu L-S, Xu G-J. *Phytochemistry*. 1999; 50:113–115. [PubMed: 9891935]
30. Sato Y, Oketani H, Yamada T, Singyouchi K-I, Ohtsubo T, Kihara M, Shibata H, Higuti T. *J Pharm Pharmacol*. 1997; 49:1042–1044. [PubMed: 9364417]
31. Pinel B, Landreau A, Seraphin D, Larcher G, Bouchara J-P, Richomme P. *J Enzyme Inhib Med Chem*. 2005; 20:575–579. [PubMed: 16408793]
32. Favier LS, Maria AOM, Wendel GH, Borkowski EJ, Giordano OS, Pelzer L, Tonn CE. *J Ethnopharmacology*. 2005; 100:260–267.
33. Wagner S, Hofmann A, Siedle B, Terfloth L, Merfort I, Gasteiger J. *J Med Chem*. 2006; 49:2241–2252. [PubMed: 16570920]
34. Schmitz ML, Mattioli I, Buss H, Kracht M. *ChemBioChem*. 2004; 5:1348–1358. [PubMed: 15457532]
35. a) Lopez-Anton N, Hermann C, Murillo R, Merfort I, Wanner G, Vollmar AM, Dirsch VM. *Apoptosis*. 2007; 12:41–153. b) Siedle B, Garcia-Pineres AJ, Murillo R, Schulte-Moenting J, Castro V, Ruengeler P, Klaas CA, Da Costa FB, Kisiel W, Merfort I. *J Med Chem*. 2004; 47:6042–6054. [PubMed: 15537359]
36. a) Xie X, Lu X, Liu Y, Xu W. *J Org Chem*. 2001; 66:6545–6550. [PubMed: 11578203] b) Zhu G, Lu X. *Organometallics*. 1995; 14:4899–4904. c) Cook GR, Hayashi R. *Org Lett*. 2006; 8:1045–1048. [PubMed: 16524264]
37. a) Song J, Shen Q, Xu F, Lu X. *Tetrahedron*. 2007; 63:5148–5153. b) Zhang Q, Lu X. *J Am Chem Soc*. 2000; 122:7604–7605.
38. a) Kulys J, Bratkovskaja I. *Talanta*. 2007; 72:526–531. [PubMed: 19071650] b) Solis-Oba M, Ugalde-Saldivar VM, Gonzalez I, Viniegra-Gonzalez G. *J Electroanal Chem*. 2005; 579:59–66. c) Scott SL, Chen WJ, Bakac A, Espenson JH. *J Phys Chem*. 1993; 97:6710–6714.
39. Padwa A, Austin DJ, Price AT, Semones MA, Doyle MP, Protopopova MN, Winchester WR, Tran A. *J Am Chem Soc*. 1993; 115:8669–8680.
40. a) Fafarman AT, Sigala PA, Herschlag D, Boxer SG. *J Am Chem Soc*. 2010; 132:12811–12813. [PubMed: 20806897] b) Sigala PA, Fafarman AT, Bogard PE, Boxer SG, Herschlag D. *J Am Chem Soc*. 2007; 129:12104–12105. [PubMed: 17854190]
41. Midland MM. *Chem Rev*. 1989; 89:1553–1561.

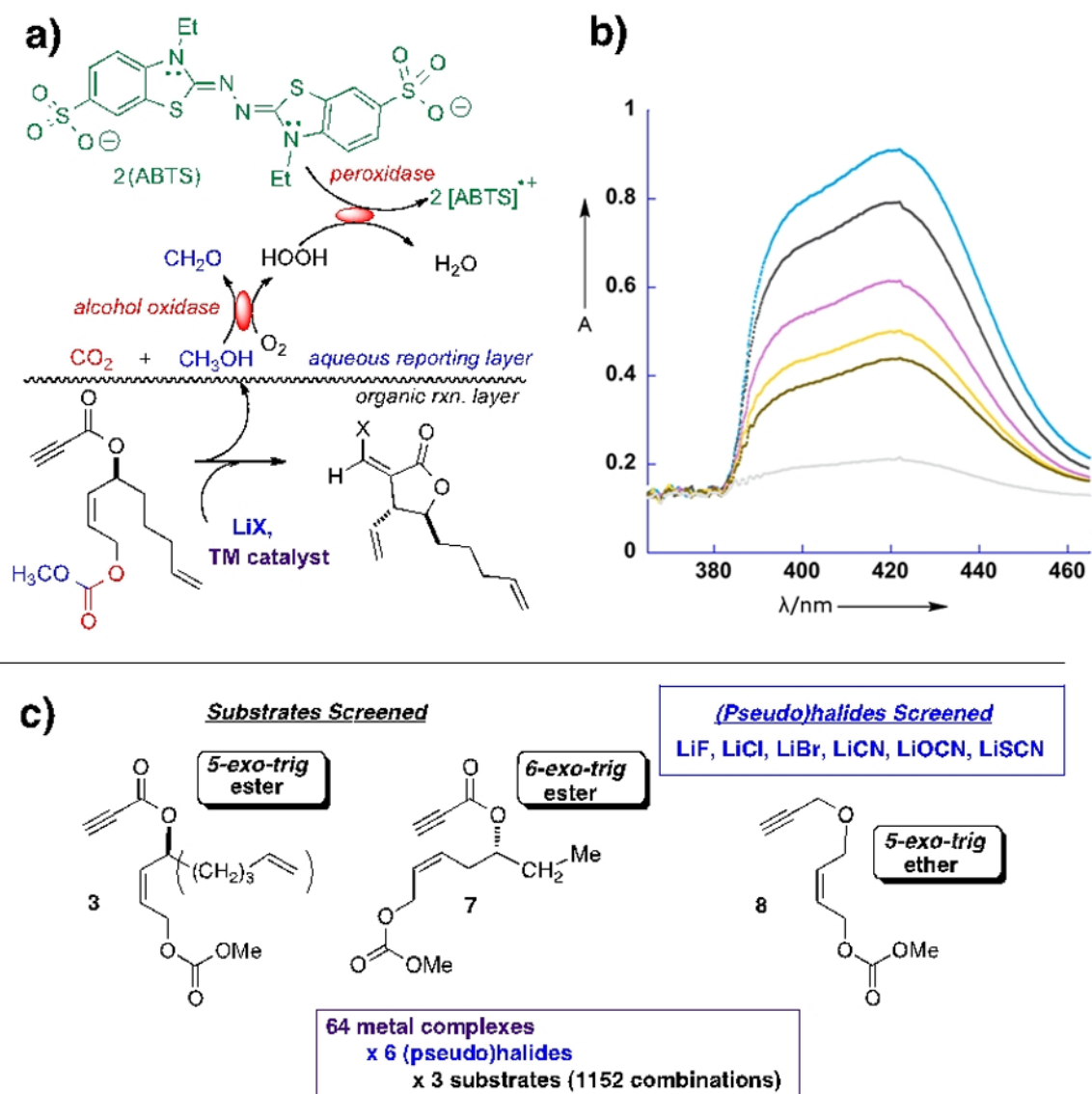
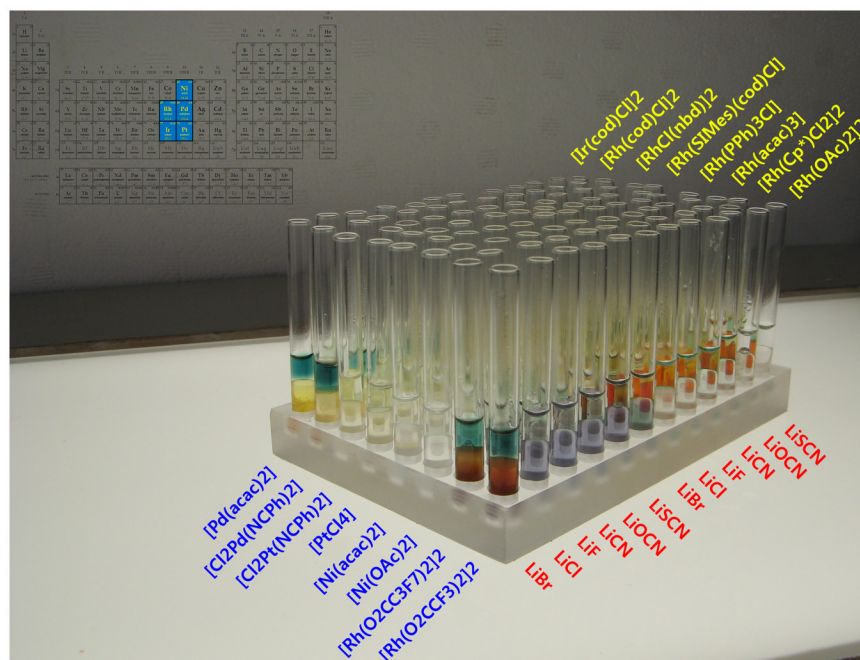
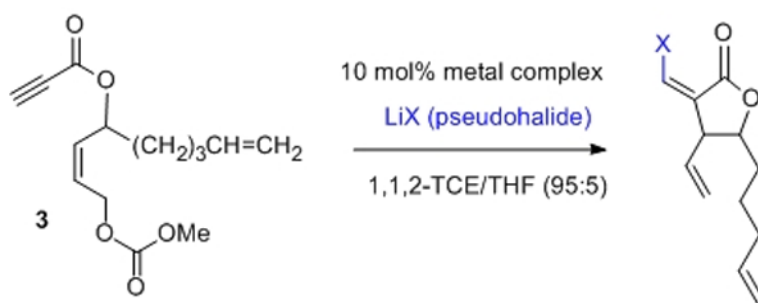
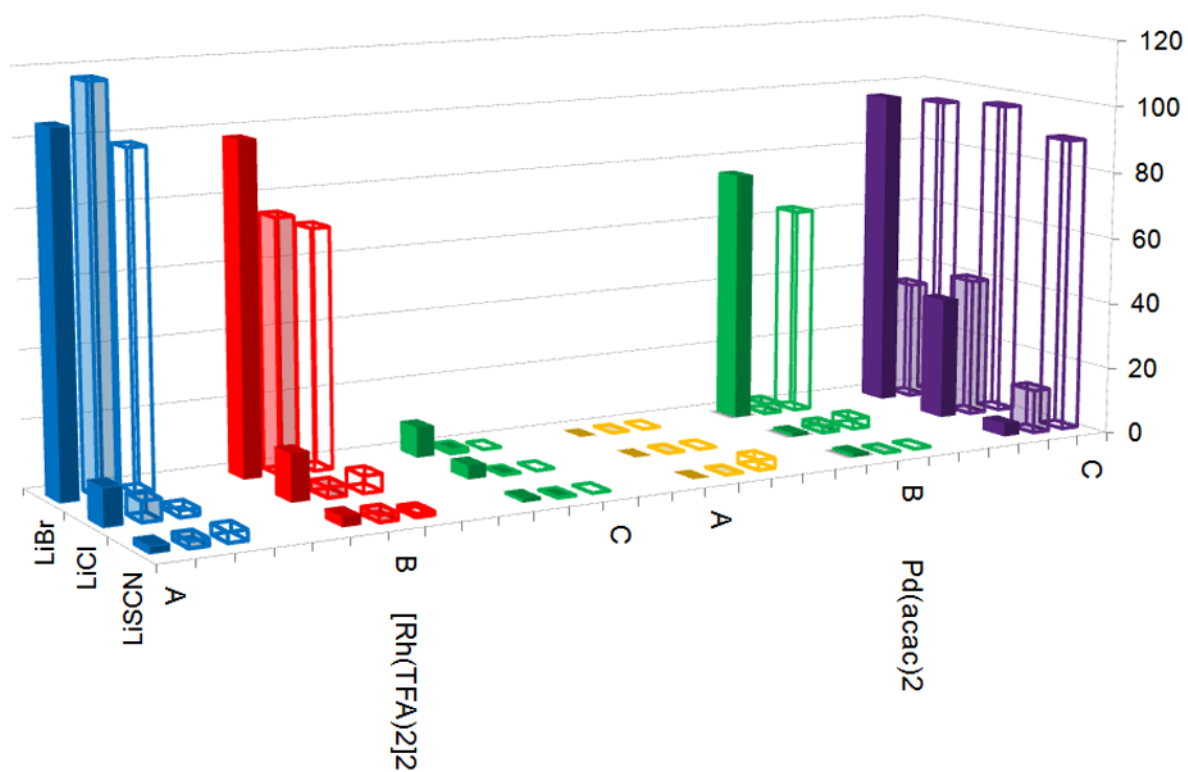


Figure 1.

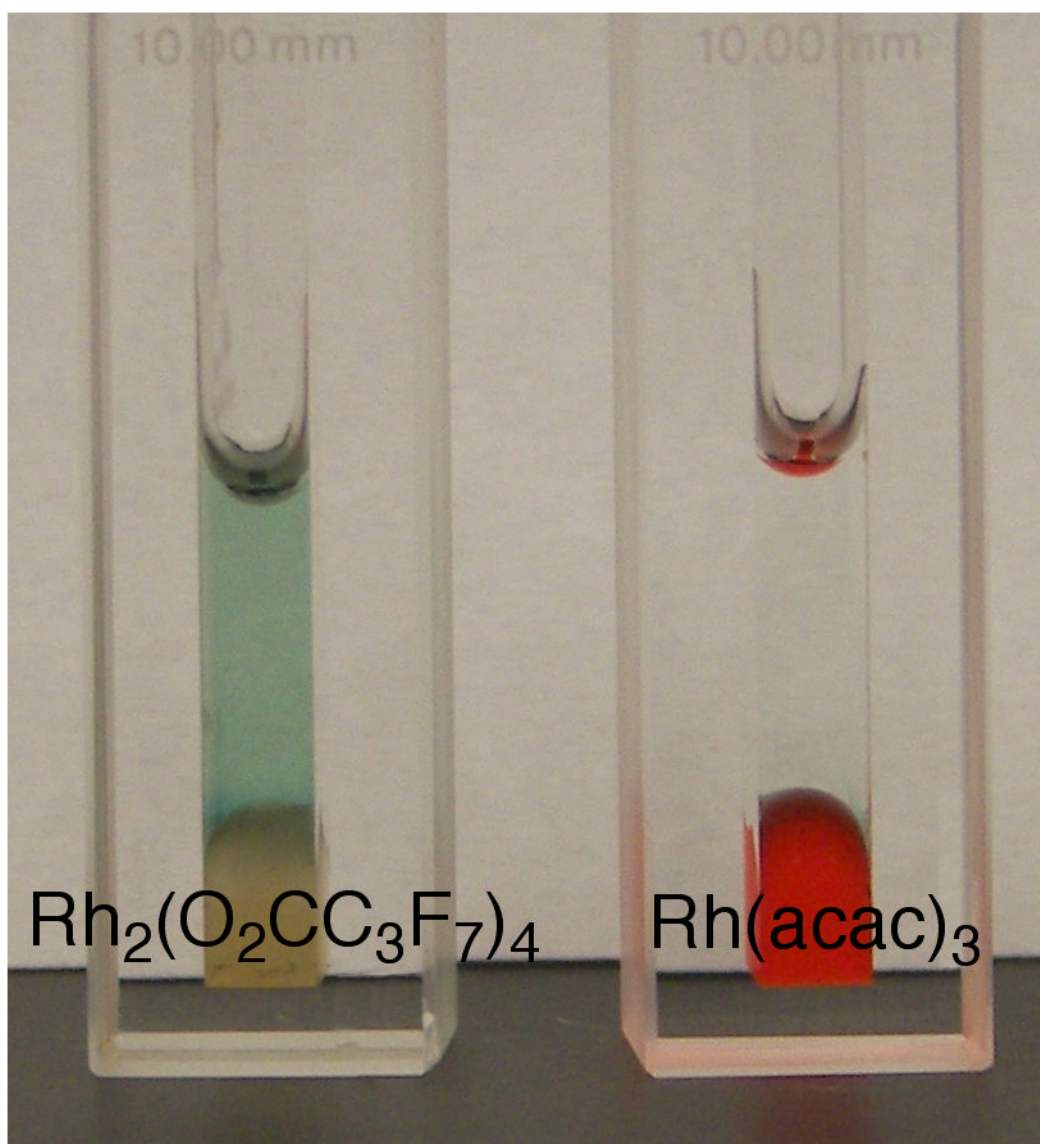
a) - Configuration of this in situ screen; b) - UV spectrum for the formation of ABTS radical cation with time; c) - potential catalytic combinations screened



**Figure 2.** Example of a d9-d10 screening array for substrate **3**—sixteen metal complexes are screened across six (pseudo)halides with propiolate ester **3** (5-*exo*-trig substrate).





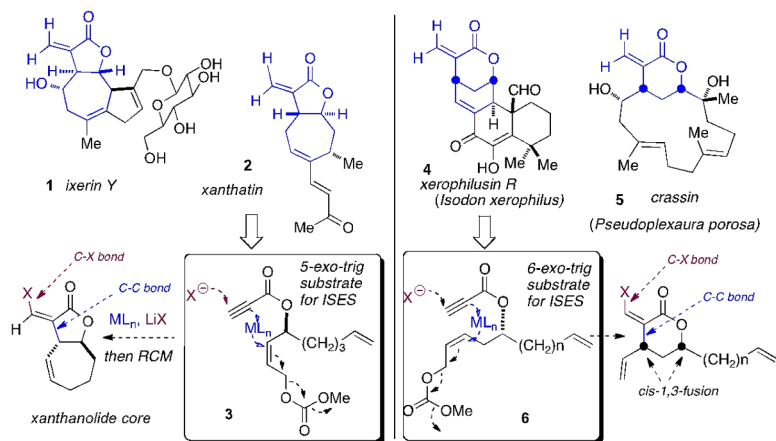


**Figure 3.** Initial hits from visual colorimetric-ISES are ranked spectrophotometrically ( $\text{Abs}_{405}$  in  $\text{mAbs min}^{-1}$ ; A, B and C are respectively substrates **3**, **7** and **8**-see SI for conditions). Right side: example of the cuvette-ISES experiment. *Note:* The ABTS indicator shows turnover with Rh(II)-perfluorobutyrate, while the highly colored Rh(III) catalyst fails.

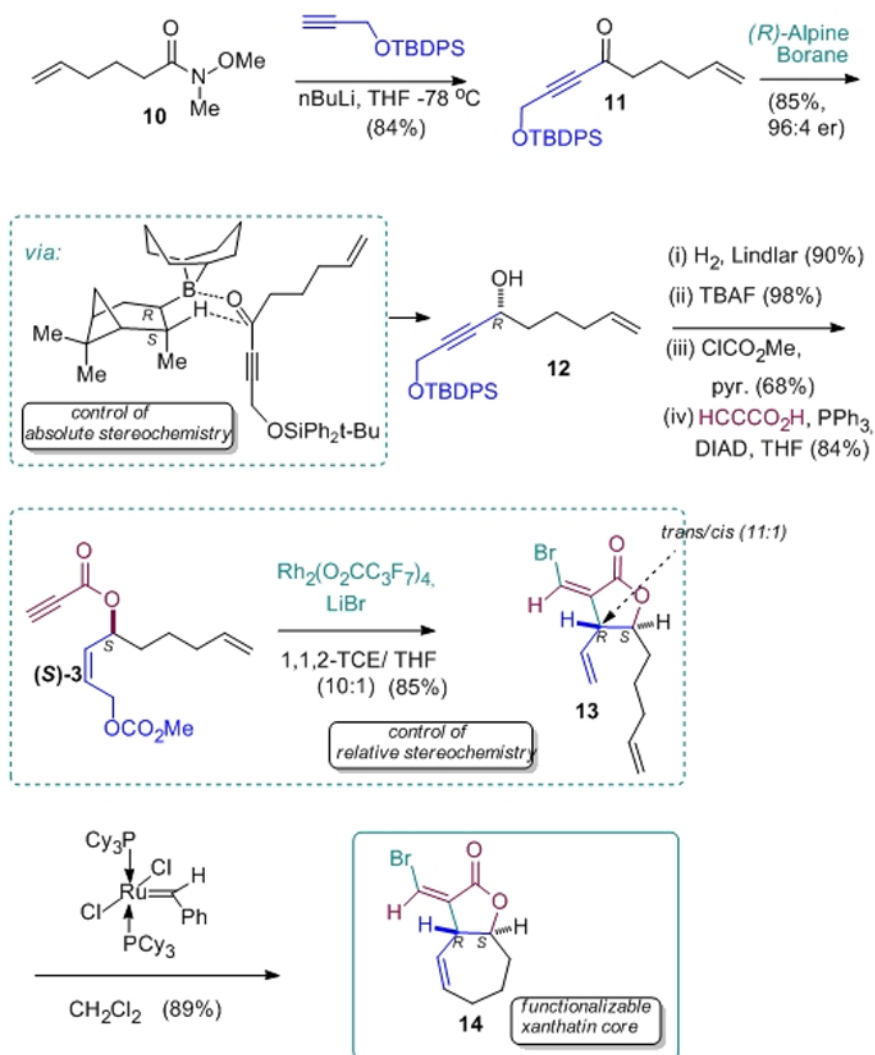
<i>cyclization product</i> <i>catalyst/ nucleophile</i>	(from 3)	(from 9)	(from 7)	(from 8)	(from 8)
[Rh(O <sub>2</sub> CCF <sub>3</sub> ) <sub>2</sub> ] <sub>2</sub> LiBr	<b>80%*</b> (10:1 - trans: cis)	<b>90%*</b>	<b>64%*</b> (>20:1 - cis: trans)	<b>66%**</b> (37%)	—
[Rh(O <sub>2</sub> CC <sub>3</sub> F <sub>7</sub> ) <sub>2</sub> ] <sub>2</sub> LiBr	<b>85%*</b> (11:1 - trans: cis)	<b>90%*</b>	<b>58%*</b> (>20:1 - cis: trans)	<b>62%**</b> (42%)	—
Pd(acac) <sub>2</sub> LiBr or LiSCN	<b>80-89%</b> (~19:1 - trans: cis)	<b>94%</b>	<b>&lt; 15%*</b>	<b>73%**</b>	—
(PhCN) <sub>2</sub> PdCl <sub>2</sub> LiBr or LiSCN	<b>85-92%</b> (~20:1 - trans: cis)	<b>95%</b>	<b>30%*</b>	<b>85%**</b> (48%)	<b>89%</b>

**Figure 4.**

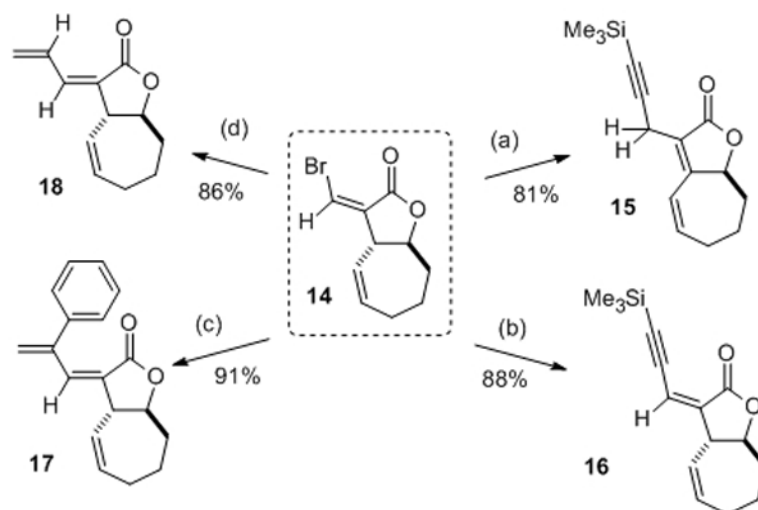
Chart showing success of catalytic metal-(pseudo)halide combinations as a function of substrate. Isolated yields for homogeneous material, following standard RB flask reactions (\*reaction carried out at 60°C; \*\*GC yield)



**Scheme 1.**  
Proposed halometalation/carbocyclization routes into the core structures of terpenoid exomethylene lactone natural products.

**Scheme 2.**

Application of the new halometalation/carbocyclization route to the stereocontrolled synthesis of the xanthatin core.

**Scheme 3.**

Exploitation of the bromomethylene lactone functionality for transition metal-mediated tailoring chemistry upon the bicyclic xanthanolid core: (a) HCC-TMS, CuI, Cl<sub>2</sub>Pd (PPh<sub>3</sub>)<sub>2</sub>, Et<sub>2</sub>NH; (b) Bu<sub>3</sub>SnCC-TMS, cat. Pd<sub>2</sub>dba<sub>3</sub>, Pfu<sub>3</sub>, Δ; (c) BrZnC(Ph)C=CH<sub>2</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>, Δ; (d) Bu<sub>3</sub>SnCH=CH<sub>2</sub>, Pd<sub>2</sub>dba<sub>3</sub>, Pfu<sub>3</sub>, Δ.