

Galleria mellonella as a model host for human pathogens

Recent studies and new perspectives

Juliana Campos Junqueira

Department of Biosciences and Oral Diagnosis; São José dos Campos Dental School; Universidade Estadual Paulista/UNESP; São Paulo, Brazil

Keywords: *Galleria mellonella*, fungal pathogenesis, bacterial pathogenesis, infection, invertebrate models

The number of studies using *G. mellonella* as a model host for human pathogens has increased significantly in the last few years. Important studies were published from different countries for evaluating the pathogenesis of bacterial and fungal infections and for exploring the host defenses against pathogens. Therefore, standardized conditions for the use of *G. mellonella* larvae need to be established. Recent research showed that the deprivation of *G. mellonella* larvae of food during the experiment caused a reduction in immune responses and an increased susceptibility to infection, suggesting that incubating of larvae in the presence or absence of nutrition may affect the results and comparisons among different laboratories.

Larvae of the greater wax moth *Galleria mellonella* have recently been used as model hosts for studying pathogenic microorganisms as an alternative to vertebrates. A positive correlation between virulence and host response has generally been found in both invertebrate and mammalian host models for a range of microorganisms, such as *Acinetobacter baumannii*,^{1,2} *Francisella tularensis*,³ *Pseudomonas aeruginosa*,^{4,5} *Yersinia pseudotuberculosis*,⁶ *Staphylococcus aureus*,⁷ *Streptococcus pyogenes*,⁸ *Streptococcus mutans*,⁹ *Enterococcus faecalis*,^{10,11} *Candida albicans*¹² and *Cryptococcus neoformans*.¹³

In 2010, Fuchs and colleagues¹⁴ reported in *Virulence* several methods for using *Galleria mellonella* as a model host to study fungal pathogenesis. First, these authors described a number of the benefits of using *G. mellonella* larvae as a model host that are not easily achieved with invertebrate models such as *Caenorhabditis elegans* and *Drosophila melanogaster*. For example, the larvae of *G. mellonella* can be maintained at 37°C. This characteristic is very important, because it allows microorganisms to be studied under the temperature conditions at which they are

pathogenic to human hosts. Another benefit of the *G. mellonella* model is the multiple options for facile delivery of the pathogen, such as topical application, oral delivery and injection. Among these methods, injection offers the benefit that fungi can be injected directly into the larval hemocoel and therefore larvae receive a known amount of pathogens. Moreover, the *G. mellonella* model is not restricted to studies that examine aspects of the pathogenesis of fungal infections but also recommends itself to the study of host defenses against fungal pathogens. *G. mellonella* have an innate immune system comprised of different types of hemocytes, which play a role in fungal-pathogen defense.

Next, Fuchs et al.¹⁴ presented in detail various methods to study fungal virulence and the association of fungal cells with insect hemocytes using *Candida albicans* and *Cryptococcus neoformans* to illustrate the use of this model. These authors showed that *G. mellonella* can be used to monitor fungal pathogenicity by a survival assay. Larvae can also be utilized to observe differences in fungal cell filamentation post-infection. For this experiment, the

fat body and other internal structures of *G. mellonella* can be collected, fixed with formalin, and prepared for histological sectioning. Furthermore, the authors demonstrated how fungal cell-hemocyte associations can be evaluated using fluorescence-activated cell sorting (FACS) analysis. The study by Fuchs et al.¹⁴ has great value to the scientific community, because the protocols presented for the *G. mellonella* infection model can be adapted to the study of other fungal and bacterial pathogens.

In this context, Olsen and colleagues⁸ in 2011 published in *Virulence* the first study to describe *G. mellonella* as model host for group A streptococcus (GAS, *S. pyogenes*). To test the hypothesis that *G. mellonella* is a suitable model host to study GAS pathogenesis, the authors infected larvae with serotype M3 strain MGAS315. The genome of this strain has been sequenced, and it is representative of highly virulent serotype M3 GAS strains that cause severe invasive disease in humans. In addition, strain MGAS315 has been extensively studied in previous experiments using mice and monkeys. All larvae infected with strain MGAS315 had distinct signs of

invasive infection, including melanization, rapid death and formation of a destructive abscess-like lesion at the site of inoculation. These abscesses comprised a dense central core of necrotic tissue and GAS microorganisms surrounded by a well-organized outer band of host hemocytes, coagulated hemolymph and extracellular melanin pigment. According to the authors, these findings are similar to the histopathology that is commonly observed in mouse and monkey models of GAS necrotizing fasciitis and in humans with severe soft tissue infections. Therefore, these results showed that *G. mellonella* larvae are useful host organisms for studying GAS pathogenesis.

In the same year, *Virulence* published another interesting study related to the *G. mellonella* model, in which this insect's immune response to infection was extensively explored by Fallon and colleagues.¹⁵ In this study, the authors demonstrated that prior exposure of *G. mellonella* larvae to non-lethal doses (1×10^4 or 1×10^5) of *Aspergillus fumigatus* conidia increased the larval survival rate when a lethal dose (1×10^7) was administered 24 h later, suggesting that the inoculation of *G. mellonella* with non-lethal doses of *A. fumigatus* conferred a significant protective response against a subsequent lethal inoculum. According to Fallon et al.,¹⁵ insects do not have an immune system that is analogous to the adaptive immune response of mammals in terms of antibody generation, but they do have the capacity to mount an immune response in

anticipation of a subsequent infection that has some elements that are similar to the function of the adaptive immune response in mammals. This study significantly contributes to research exploring *G. mellonella* as a model host, because an understanding of the mechanisms employed by insects to withstand infection is critical to their successful use as models for human pathogens.

Thus, we have observed that *G. mellonella* as a model for the study of infectious diseases has achieved increasing acceptance among scientific researchers, and the use of this invertebrate model in medical research extends to many laboratories around the world. Recently, important studies were published from different countries, such as the US,^{12,16,17} Ireland,^{15,17,18} Canada,¹⁹ the United Kingdom,²⁰ Spain,²¹ Germany,^{22,23} Brazil,²⁴ Tunisia,²⁵ Greece,²⁶ South Korea,²⁷ Poland,²⁸ Italy²⁹ and Norway.³⁰ Therefore, studies need to be developed to determine standardized conditions for the propagation and maintenance of *G. mellonella* larvae.

In this issue of *Virulence*, Banville and colleagues³¹ have published a study to evaluate the effect of nutritional deprivation on the ability of larvae to withstand infection. The objective of this study was to establish standardized conditions for larval treatment for in vivo testing, given that some researchers incubate larvae with a food source during experiments, while others do not. The authors observed that larvae deprived of nutrition for 7 days demonstrated increased susceptibility to

infection with the fungal pathogen *C. albicans*. Starved larvae demonstrated a slight reduction in hemocyte density, but the hemocytes from starved larvae were as effective at killing *C. albicans* cells as those from unstarved larvae. Hemolymph from starved larvae showed reduced expression of a range of antimicrobial peptides and immune proteins. Banville et al.³¹ concluded that the deprivation of *G. mellonella* larvae of food leads to a reduction in cellular and immune responses and an increased susceptibility to infection, indicating that researchers utilizing *G. mellonella* for the study of human pathogens should specify whether food is provided to the larvae to allow valid comparisons between results from different laboratories.

According to the studies cited above, it is evident that the number of studies using *G. mellonella* as a model host has increased significantly in the last few years. In addition, there has been an improvement in the techniques used with this model, which allows further possibilities for the development of other studies. Certainly, the articles published in *Virulence* represent an important scientific contribution for the advancement of research utilizing *G. mellonella* as a model host for human pathogens.

Acknowledgments

The author thanks the São Paulo Council of Research—FAPESP, Brazil (12/02184-9) for supporting the research addressing *G. mellonella* as a model host for fungal pathogens.

References

1. Peleg AY, Jara S, Monga D, Eliopoulos GM, Moellering RC, Jr., Mylonakis E. *Galleria mellonella* as a model system to study *Acinetobacter baumannii* pathogenesis and therapeutics. *Antimicrob Agents Chemother* 2009; 53:2605-9; PMID:19332683; <http://dx.doi.org/10.1128/AAC.01533-08>
2. Gaddy JA, Arivett BA, McConnell MJ, López-Rojas R, Pachón J, Actis LA. Role of acinetobactin-mediated iron acquisition functions in the interaction of *Acinetobacter baumannii* strain ATCC 19606T with human lung epithelial cells, *Galleria mellonella* caterpillars, and mice. *Infect Immun* 2012; 80:1015-24; PMID:22232188; <http://dx.doi.org/10.1128/IAI.06279-11>
3. Aperis G, Fuchs BB, Anderson CA, Warner JE, Calderwood SB, Mylonakis E. *Galleria mellonella* as a model host to study infection by the *Francisella tularensis* live vaccine strain. *Microbes Infect* 2007; 9: 729-34; PMID:17400503; <http://dx.doi.org/10.1016/j.micinf.2007.02.016>
4. Jander G, Rahme LG, Ausubel FM. Positive correlation between virulence of *Pseudomonas aeruginosa* mutants in mice and insects. *J Bacteriol* 2000; 182:3843-5; PMID:10851003; <http://dx.doi.org/10.1128/JB.182.13.3843-3845.2000>
5. Miyata S, Casey M, Frank DW, Ausubel FM, Drenkard E. Use of the *Galleria mellonella* caterpillar as a model host to study the role of the type III secretion system in *Pseudomonas aeruginosa* pathogenesis. *Infect Immun* 2003; 71:2404-13; PMID:12704110; <http://dx.doi.org/10.1128/IAI.71.5.2404-2413.2003>
6. Champion OL, Cooper IA, James SL, Ford D, Karlyshev A, Wren BW, et al. *Galleria mellonella* as an alternative infection model for *Yersinia pseudotuberculosis*. *Microbiology* 2009; 155:1516-22; PMID:19383703; <http://dx.doi.org/10.1099/mic.0.026823-0>
7. Desbois AP, Coote PJ. Wax moth larva (*Galleria mellonella*): an *in vivo* model for assessing the efficacy of antistaphylococcal agents. *J Antimicrob Chemother* 2011; 66:1785-90; PMID:21622972; <http://dx.doi.org/10.1093/jac/dkr198>
8. Olsen RJ, Watkins ME, Cantu CC, Beres SB, Musser JM. Virulence of serotype M3 Group A *Streptococcus* strains in wax worms (*Galleria mellonella* larvae). *Virulence* 2011; 2:111-9; PMID:21258213; <http://dx.doi.org/10.4161/viru.2.2.14338>
9. Abranches J, Miller JH, Martinez AR, Simpson-Haidaris PJ, Burne RA, Lemos JA. The collagen-binding protein Cnm is required for *Streptococcus mutans* adherence to and intracellular invasion of human coronary artery endothelial cells. *Infect Immun* 2011; 79:2277-84; PMID:21422186; <http://dx.doi.org/10.1128/IAI.00767-10>

10. Michaux C, Sanguinetti M, Reffuveille F, Auffray Y, Posteraro B, Gilmore MS, et al. SlyA is a transcriptional regulator involved in the virulence of *Enterococcus faecalis*. *Infect Immun* 2011; 79:2638-45; PMID: 21536798; <http://dx.doi.org/10.1128/IAI.01132-10>
11. Yasmin A, Kenny JG, Shankar J, Darby AC, Hall N, Edwards C, et al. Comparative genomics and transduction potential of *Enterococcus faecalis* temperate bacteriophages. *J Bacteriol* 2010; 192:1122-30; PMID: 20008075; <http://dx.doi.org/10.1128/JB.01293-09>
12. Fuchs BB, Eby J, Nobile CJ, El Khoury JB, Mitchell AP, Mylonakis E. Role of filamentation in *Galleria mellonella* killing by *Candida albicans*. *Microbes Infect* 2010; 12:488-96; PMID:20223293; <http://dx.doi.org/10.1016/j.micinf.2010.03.001>
13. Mylonakis E, Moreno R, El Khoury JB, Idnurm A, Heitman J, Calderwood SB, et al. *Galleria mellonella* as a model system to study *Cryptococcus neoformans* pathogenesis. *Infect Immun* 2005; 73:3842-50; PMID:15972469; <http://dx.doi.org/10.1128/IAI.73.7.3842-3850.2005>
14. Fuchs BB, O'Brien E, Khoury JB, Mylonakis E. Methods for using *Galleria mellonella* as a model host to study fungal pathogenesis. *Virulence* 2010; 1:475-82; PMID:21178491; <http://dx.doi.org/10.4161/viru.1.6.12985>
15. Fallon JP, Troy N, Kavanagh K. Pre-exposure of *Galleria mellonella* larvae to different doses of *Aspergillus fumigatus* conidia causes differential activation of cellular and humoral immune responses. *Virulence* 2011; 2:413-21; PMID:21921688; <http://dx.doi.org/10.4161/viru.2.5.17811>
16. Fuchs BB, Mylonakis E. Using non-mammalian hosts to study fungal virulence and host defense. *Curr Opin Microbiol* 2006; 9:346-51; PMID:16814595; <http://dx.doi.org/10.1016/j.mib.2006.06.004>
17. Lionakis MS. *Drosophila* and *Galleria* insect model hosts: new tools for the study of fungal virulence, pharmacology and immunology. *Virulence* 2011; 2: 521-7; PMID:22186764; <http://dx.doi.org/10.4161/viru.2.6.18520>
18. Cotter G, Doyle S, Kavanagh K. Development of an insect model for the in vivo pathogenicity testing of yeasts. *FEMS Immunol Med Microbiol* 2000; 27:163-9; PMID:10640612; <http://dx.doi.org/10.1111/j.1574-695X.2000.tb01427.x>
19. Leuko S, Raivio TL. Mutations that impact the enteropathogenic *Escherichia coli* Cpx envelope stress response attenuate virulence in *Galleria mellonella*. *Infect Immun* 2012; 80:3077-85; PMID:22710873; <http://dx.doi.org/10.1128/IAI.00081-12>
20. Harding CR, Schroeder GN, Reynolds S, Kosta A, Collins JW, Mousnier A, et al. *Legionella pneumophila* pathogenesis in the *Galleria mellonella* infection model. *Infect Immun* 2012; 80:2780-90; PMID:22645286; <http://dx.doi.org/10.1128/IAI.00510-12>
21. García-Rodas R, Casadevall A, Rodríguez-Tudela JL, Cuenca-Estrella M, Zaragoza O. *Cryptococcus neoformans* capsular enlargement and cellular gigantism during *Galleria mellonella* infection. *PLoS One* 2011; 6:e24485; PMID:21915338; <http://dx.doi.org/10.1371/journal.pone.0024485>
22. Vilcinskas A. Anti-infective therapeutics from the Lepidopteran model host *Galleria mellonella*. *Curr Pharm Des* 2011; 17:1240-5; PMID:21470117; <http://dx.doi.org/10.2174/138161211795703799>
23. Vogel H, Altincicek B, Glöckner G, Vilcinskas A. A comprehensive transcriptome and immune-gene repertoire of the lepidopteran model host *Galleria mellonella*. *BMC Genomics* 2011; 12:308; PMID: 21663692; <http://dx.doi.org/10.1186/1471-2164-12-308>
24. Junqueira JC, Fuchs BB, Muhammed M, Coleman JJ, Suleiman JM, Vilela SF, et al. Oral *Candida albicans* isolates from HIV-positive individuals have similar in vitro biofilm-forming ability and pathogenicity as invasive *Candida* isolates. *BMC Microbiol* 2011; 11: 247; PMID:22053894; <http://dx.doi.org/10.1186/1471-2180-11-247>
25. Fedhila S, Buisson C, Dussurget O, Serron P, Glomski IJ, Lielh P, et al. Comparative analysis of the virulence of invertebrate and mammalian pathogenic bacteria in the oral insect infection model *Galleria mellonella*. *J Invertebr Pathol* 2010; 103:24-9; PMID:19800349; <http://dx.doi.org/10.1016/j.jip.2009.09.005>
26. Schell MA, Lipscomb L, DeShazer D. Comparative genomics and an insect model rapidly identify novel virulence genes of *Burkholderia mallei*. *J Bacteriol* 2008; 190:2306-13; PMID:18223084; <http://dx.doi.org/10.1128/JB.01735-07>
27. Park SY, Kim KM, Lee JH, Seo SJ, Lee IH. Extracellular gelatinase of *Enterococcus faecalis* destroys a defense system in insect hemolymph and human serum. *Infect Immun* 2007; 75:1861-9; PMID: 17261598; <http://dx.doi.org/10.1128/IAI.01473-06>
28. Mak P, Zdybicka-Barabas A, Cytryńska M. A different repertoire of *Galleria mellonella* antimicrobial peptides in larvae challenged with bacteria and fungi. *Dev Comp Immunol* 2010; 34:1129-36; PMID:20558200; <http://dx.doi.org/10.1016/j.dci.2010.06.005>
29. Antunes LC, Imperi F, Minandri F, Visca P. In vitro and in vivo antimicrobial activity of gallium nitrate against multidrug resistant *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 2012; 56:5961-70; PMID:22964249; <http://dx.doi.org/10.1128/AAC.01519-12>
30. Stenfors Arnesen L, Granum PE, Buisson C, Bohlin J, Nielsen-LeRoux C. Using an insect model to assess correlation between temperature and virulence in *Bacillus weihenstephanensis* and *Bacillus cereus*. *FEMS Microbiol Lett* 2011; 317:196-202; PMID:21276046; <http://dx.doi.org/10.1111/j.1574-6968.2011.02229.x>
31. Banville N, Browne N, Kavanagh K. Effect of nutrient deprivation on the susceptibility of *Galleria mellonella* larvae to infection. *Virulence* 2012; 3.