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Radiother Oncol. Author manuscript; available in PMC 2014 January 01.

Published in final edited form as:

Radiother Oncol. 2013 January; 106(1): 101-105. doi:10.1016/j.radonc.2012.08.007.

Spinal cord tolerance to single-session uniform irradiation in pigs: implications for a dose-volume effect

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Abstract

Background and Purpose—This study was performed to test the hypothesis that spinal cord radiosensitivity is significantly modified by uniform versus laterally non-uniform dose distributions.

Materials and Methods—A uniform dose distribution was delivered to a 4.5–7.0cm length of cervical spinal cord in 22 mature Yucatan minipigs for comparison with a companion study in which a laterally non-uniform dose was given[1]. Pigs were allocated into four dose groups with mean maximum spinal cord doses of 17.5 ± 0.1 Gy(n=7), 19.5 ± 0.2 Gy(n=6), 22.0 ± 0.1 Gy(n=5), and 24.1 ± 0.2 Gy(n=4). The study endpoint was motor neurologic deficit determined by a change in gait within one year. Spinal cord sections were stained with a Luxol fast blue/periodic acid Schiff combination.

Results—Dose-response curves for uniform versus non-uniform spinal cord irradiation were nearly identical with ED_{50} 's (95% confidence interval) of 20.2 Gy(19.1–25.8) and 20.0 Gy(18.3– 21.7), respectively. No neurologic change was observed for either dose distribution when the maximum spinal cord dose was 17.8 Gy while all animals experienced deficits at doses 21.8Gy.

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Conflicts of Interest Notification Paul Medin and Timothy Solberg teach in radiosurgery courses sponsored by BrainLAB AG. Ryan Foster and Timothy Solberg have received research funding from Elekta, Ltd. All remaining authors have no conflicts to report.

Conclusion—No dose-volume effect was observed in pigs for the dose distributions studied and the endpoint of motor neurologic deficit; however, partial spinal cord irradiation resulted in less debilitating neurologic morbidity and histopathology.

Keywords

dose-volume effect; spinal cord tolerance; normal tissue tolerance; stereotactic spinal radiosurgery; swine

Introduction

Dose-volume effects are of great significance in radiation therapy and have been summarized for many organs by the Quantitative Analysis of Normal Tissue Effect in the Clinic (QUANTEC) collaboration[2]. Early efforts to investigate dose-volume effects in the spinal cord were limited to characterizing the influence of irradiated length on response[3–5]. Pioneering work in frame-based spinal radiosurgery at the University of Arizona[6], followed by the development of image-guidance and dose-shaping technologies, provided tools to localize and irradiate lesions of the spine while minimizing dose to the spinal cord but the effect of the resulting complex dose distributions on spinal cord tolerance was unknown. Since 2001, many studies performed in rats have demonstrated that spinal cord tolerance is modified by non-uniform dose distributions[7] including steep lateral dose gradients[8–10], longitudinal dose inhomogeneity[3, 11], and selective regional irradiation[9]. These studies were never repeated in a large animal. A general perception that dose-volume effects play a role in human spinal cord tolerance permeates the radiosurgery literature[12–15].

In this study, pigs received de novo single-session irradiation using a uniform dose distribution for comparison to a previous study in which the same cervical spinal cord segments received a steep lateral dose gradient[1]. Spinal cord tolerance to uniform irradiation has not previously been studied in a large animal under conditions relevant to clinical stereotactic body radiotherapy and the lateral dose-volume effect has not previously been studied in a model with spinal cord dimensions equivalent to humans. The inclusion of human clinical treatment parameters in this study may be responsible for the difference between these results and those of previous studies and may serve to refine models of normal tissue complication probability[16].

Methods and Materials

This study conformed to all national and local regulations regarding the use of animals for research and was approved by the Institutional Animal Care and Use Committee. A total of 50 female Yucatan minipigs were enrolled to study dose-volume effects in the cervical spinal cord. Pigs were randomly assigned to receive either uniform or partial-volume single-session irradiation to the same spinal cord segments. Twenty-two pigs received uniform irradiation and are described here. Twenty-six pigs were assigned to receive partial-volume irradiation and have been reported previously[1]. Two pigs served as unirradiated controls.

Animals that received uniform spinal cord irradiation were 42–102 weeks old and weighed approximately 35–60 kg when irradiated. Treatment parameters for all individual animals are presented in Table 1 including: a) prescription dose group, b) irradiated spinal cord length, c) irradiated spinal cord level, d) image-guidance/irradiation platform, e) maximum spinal cord dose, f) age, g) followup period, h) latency to response, and i) overall treatment time. All animals received a treatment planning CT scan with 0.75–1.5 mm thick slices and a 300–500 cm field of view. Treatment planning calculations were performed using either

Brainscan 5.31 software (BrainLAB, AG, Feldkirchen) or Pinnacle³ 8.0m (Philips Electronics N.V., Eindhoven). Radiation was delivered in a single session to a cylindrical target volume 4.5–7.0 cm in length and 5 cm in diameter that was centered on the spinal cord. In the rostral/caudal direction, the target volume was centered at the level of the sixth cervical vertebral body. Dose distributions in the axial and sagittal planes are shown in Figure 1. Treatment plans consisted of a series of 4-6 dynamically-shaped arcs or 12 conformal fields arranged with the goal of creating a uniform dose distribution through the spinal cord. The spinal cord volume was defined on CT images by contracting the thecal sac contour by 1.5mm in the axial plane. This method was based on CT/MRI fusion of two animals and is consistent with the method used in a companion study[1]. The spinal cord was contoured 5.5-6.5 mm beyond the irradiated volume in the rostral and caudal directions and the dose calculation grid resolution through the spinal cord ranged from 1.5-1.8 mm. Dose distributions were normalized to the global plan maximum and dose was prescribed to the 90% isodose line. Animals were stratified into 4 prescription dose groups as follows: 22 Gy (4), 20 Gy (5), 18 Gy (6), and 16 Gy (7). Treatment planning dose-volume histogram statistics for the spinal cord are summarized in Table 2 including: a) mean maximum point dose, b) mean percentage volume to receive 10 Gy, c) mean volume to receive 14 Gy, and d) mean maximum dose to 1 cc volume. For all procedures, animals were anesthetized with a mixture of *Telazol*® and xylazine and maintained on isoflurane. Animals were positioned supine in a body-length, vacuum-molded immobilization cushion that was individually molded for each pig and kept unchanged between simulation and irradiation. Image-guided localization was performed either using stereoscopic kilovoltage x-rays (Novalis Body Xray 6D, BrainLAB AG) or conebeam computed tomography (CBCT) (XVI, Elekta AB, Stockholm). For stereoscopic image guidance, a pair of digital radiographs was exposed and automatically fused with digitally reconstructed radiographs (DRR's) generated from the pre-treatment CT scan to determine if any positioning adjustments were necessary. After visual evaluation of the fusion results, the treatment table was shifted until the actual position and required treatment position differed by less than 1 mm in the three primary axes. The image-guidance process was repeated following table adjustments to ensure that shifts were made correctly. The same general positioning and verification processes were followed for CBCT image guidance.

Irradiation was performed with either 6 MV (*Novalis*, BrainLAB AG) or 10 MV (*Synergy S*, Elekta AB) image-guided linear accelerators. Radiation from the *Novalis* was delivered through dynamically-shaped arcs at a rate of 800 monitor units per minute that equated to an approximate instantaneous dose rate of 5.0–6.3 Gy/min at the spinal cord, varying with the depth of overlying tissue. Radiation from the *Synergy S* was delivered through conformal fields at a rate of 500 monitor units per minute that equated to an approximate instantaneous dose rate of 3.8–4.4 Gy/min at the spinal cord. Overall delivery time (first beam on to last beam off) varied with prescription dose but the 20 Gy prescription was delivered in 10–20 minutes on the *Novalis* while the 22 Gy prescription was delivered in 20–26 minutes on the *Synergy S*.

After irradiation, animals were followed for 51–56 weeks or until a neurologic response was observed. The general health of animals was observed daily with attention toward unusual restlessness, vocalizing, loss of mobility, licking, biting, or guarding of a painful area, failure to groom, unkempt appearance, open sores, skin lesions, loss of appetite, and weight loss. Gait was observed approximately weekly with the animal walking freely in a large space. Response was defined as any study-related change in gait. Animals recognized to have a change in gait were evaluated by a veterinarian for symptoms indicative of pain and humanely killed. The cervical spinal cords of all study animals (irradiated plus controls) were removed and fixed in formalin before being sectioned and processed for embedding in paraffin. Five uniformly distributed axial sections through the irradiated volume of each

animal were cut and stained with a Luxol fast blue/periodic acid Schiff combination and were examined for histology. The dose-related incidence of motor deficit was used as an endpoint to obtain quantal data that was then analyzed by probit analysis[4, 8] to establish a dose-response curve and to calculate the probability of a deficit at increasing dose levels with the associated 95% confidence bounds.

Results

Parameters such as prescription dose, maximum spinal cord dose, length of follow-up and latency until response are shown for individual animals in Table 1. Ten of twenty-two pigs developed motor neurologic changes while the neurologic status of twelve pigs remained normal throughout the study. Bilateral deficits were observed in the hind limbs either prior to or in coincidence with the forelimbs in nine of ten pigs. Only forelimb deficits were noted in pig #19 but this animal was humanely killed one day after changes were first noted. Motor deficits presented initially as general weakness and uncoordinated gait in all responders but the progression of deficits was variable. Pig #8 (19.2 Gy maximum spinal cord dose) experienced hind limb weakness 8 weeks after irradiation but fully recovered without intervention by 10 weeks only to relapse at 25 weeks. The response of pigs in the 20 Gy prescription group varied from transient weakness at 10 weeks after irradiation, followed by a complete recovery (pig #14), to rapid onset of weakness and immobility (pig #16) without recovery. All pigs in the 22 Gy prescription group experienced rapid onset of weakness and loss of coordination without recovery. The latent period for initial onset of motor deficit ranged from 8–13 weeks. No animal exhibited behavior indicative of pain throughout this study.

A clear dose-response relationship was observed for uniformly irradiated spinal cords, with no response when the maximum spinal cord dose was 17.8 Gy and 100% rate of motor neurologic deficits when the maximum spinal cord dose was 21.8 Gy. Calculated estimates of the maximum spinal cord doses resulting in 1, 5, 10, 50 and 90% probability of paralysis along with the associated 95% confidence bounds are shown in Supplementary File, Table 3. The dose-response curve for uniformly irradiated spinal cords has been overlayed with the dose-response curve from a previous study of non-uniformly irradiated spinal cords[1] and is presented in Figure 2. Observed response data points are superimposed on the dose-response curves and annotated to show the number of responders per number of animals at each dose point. The associated upper and lower 95% confidence bounds are also plotted (dashed lines) for each curve. The test for parallelism yielded a p-value of 0.577 indicating no significant difference between the two dose-response curves. The dose associated with a 50% incidence of paralysis (ED₅₀) and 95% confidence interval (CI) due to uniform spinal cord irradiation was 20.2 Gy(19.1–25.8). A Pearson goodness-of-fit chi-square indicated (p=0.982) that the probit model provides an excellent fit.

General agreement was observed between the occurrence of neurologic change and the presence of histological lesions. All uniformly irradiated animals with motor deficits showed some degree of demyelination and focal white matter necrosis, seemingly randomly distributed in lateral, ventral and dorsal white matter tracts with sparing of the gray matter. Although the distribution of histologic damage differed from previously described non-uniformly irradiated spinal cords[1], the type of damage was the same. The only responder in the 18 Gy group (pig #8) with transient neurological changes in the hindlimbs was histologically characterized by extensive demyelination without frank white matter necrosis. In a few cases near the threshold dose for myelopathy, histology showed only some very small foci of demyelination. In addition to diffuse demyelination of the white matter, foci of white matter necrosis were observed increasingly with dose.

Discussion

This study was performed to test the hypothesis that spinal cord radiosensitivity is significantly modified by uniform versus laterally non-uniform dose distributions. Results of uniform spinal cord irradiation are reported here for comparison with a companion study in which a laterally non-uniform dose was delivered[1]. Dose-volume characteristics for pigs from the companion study are presented in Supplementary File, Table 4 for comparison with characteristics from the present study (Table 2). Despite vast differences in 10 Gy and 14 Gy volumes between studies, the dose-response curves for motor neurologic deficit are nearly identical suggesting that the maximum spinal cord point dose is the best predictor of response. The resulting ED_{50} 's (95% CI) are 20.2 Gy(19.1–25.8) and 20.0 Gy(18.3–21.7) for uniform and non-uniform irradiation, respectively.

Overall radiation delivery times varied among individual animals in every dose group of this study and the previous study that incorporated non-uniform spinal cord irradiation[1]. Overall delivery times are presented for all animals from both studies in Tables 1 and Supplementary File, Table 5, respectively. Pop et al.[17] investigated the impact of temporal dose distribution on the radiation tolerance of the rat spinal cord. An 192-Iridium brachytherapy source was stepped through catheters implanted lateral to the vertebral bodies to irradiate the spinal cord with varying overall treatment times and average dose rates. ED_{50} values were observed to increase by 2.1 Gy as the overall treatment time was increased from 4–8 to 28–37 minutes. Direct comparison of this rat data with the current pig study is confounded by considerable differences in study design and methods but the ED_{50} values derived from this pig study may have been influenced by varying degrees of sublethal damage repair that occurred due to differences in overall treatment time. The influence of sublethal damage repair is not readily apparent in this study.

Dose-response results from the present study are consistent with previous studies in mice[18], rats[3, 11] and guinea pigs[19] that also incorporated uniform spinal cord irradiation. All of these preclinical studies have reported steep dose-response curves with ED_{50} values ranging from 18.9–21.5 Gy. The only previous study in pigs reported a 37% greater ED_{50} value of 27.7(±0.6) Gy but our data suggests that the greater ED_{50} is almost certainly due to the low dose rate (0.2–0.3 Gy/min) used in that study[4].

van Luijk et. al., [8] investigated the influence of laterally non-uniform dose distributions on spinal cord tolerance in rats. A 150 MeV proton beam was used to irradiate 50% of the lateral cross-section of the cervical spinal cord resulting in an extremely steep dose gradient (100% to <10% isodose) across the spinal cord. An ED₅₀ (95% CI) of 30 Gy (26.3–31.3) was observed for paralysis compared to an ED_{50} of 20.4 Gy (19.6–21.1) for uniform crosssection irradiation. A more extensive followup study affirmed the lateral volume effect and demonstrated that the lateral white matter is much more radiosensitive than the central part of the white matter[9]. The laterally non-uniform dose distribution used in our companion pig study included a lateral dose gradient of 95% to 10% isodose across the spinal cord with the 50% isodose line bisecting the cord (similar to van Luijk, et. al.[8]); however, the diameter of the pig cervical spinal cord is approximately equivalent to humans (three times greater than for a rat). The reason for the lack of a demonstrable spinal cord sparing effect in pigs irradiated with steep lateral dose gradients is not clear, but the physical size of the spinal cord and/or the steepness of the dose gradient appear to be factors in the repair mechanism. Data from "length effect" studies suggests that migration of remyelinating cells from the unirradiated field edges is at least partially responsible for restoring the damaged glial cell population following irradiation[3, 11, 20]; however, the role of these cells in the development of white matter necrosis in unclear. If viable oligodendrocytes and their precursor cells can only migrate 2-7 mm from unirradiated tissue, as studies suggest[11, 20,

21], the diameter of the human spinal cord could be detrimental to remyelination and the encouraging results from rats may be muted.

Dose-volume effects in the spinal cord have been suggested in the human literature but not studied prospectively[12, 13, 15]. The group at Stanford University[15] provide the strongest evidence for clinical dose-volume effects in the spinal cord in a review of their experience treating hemangioblastomas with radiosurgery. Seventeen tumors were treated with single-session radiosurgery resulting in maximum spinal cord doses ranging from 17.8 to 30.9 Gy (median 22.7 Gy) yet neurologic toxicity was limited to two patients. In contrast, we observed 100% occurrence of motor neurologic deficits after partial irradiation of pig spinal cords with maximum point doses 21.1 Gy[1]. Multiple factors including irradiated length and axial dose gradient may have contributed to the contrasting rates of toxicity observed between the Stanford study and this pig study. A 5 cm length was treated in pigs while lengths are described as "short" in the Stanford series[15]. Many preclinical studies support the hypothesis that dose distribution is a better predictor of neurologic toxicity than dose-volume characteristics [3, 8, 9, 11]. For example, Bijl et. al. [11], observed no response when a dose 36 Gy was delivered to the full cross section of a 4 mm long spinal cord segment while a dose of 35 Gy to the lateral edge of the spinal cord resulted in 100% response when a 20 mm segment was irradiated[8].

Ideally, irradiation conditions would have been consistent for all pigs in this study but dose rate, radiation energy, and irradiated length varied slightly for spinal cords that were uniformly irradiated. None of these differences are believed to have influenced the study results. Dose rate has been demonstrated to modify spinal cord tolerance but only at much lower dose rates than used in this study. The ED₅₀ for paralysis in the rat and mouse spinal cord appears to plateau between dose rates of 0.25 and 1.8 Gy/min and becomes insensitive to further increases [7]. The ED_{50} for paralysis in the pig is consistent with rats at dose rates between 0.2–0.3 Gy/min[4, 22] and the present study indicates that the ED₅₀ for pigs is also consistent with rodents at higher dose rates of 5.0-6.3 Gy/min. Radiation dose response is not known to change between photon energy spectra from 6MV and 10MV linear accelerators. The irradiated length of spinal cord varied between 4.5 and 7.0 cm in the present study. As shown in Table 1, a length of 5.1 cm was irradiated in the majority of pigs in this study but 5 pigs in the 16 Gy prescription group were irradiated to a length of 6.8–7.0 cm to accommodate another companion study. Irradiated length has been shown to affect spinal cord tolerance but only for lengths < 1.6 cm[7]. There is no data to suggest that spinal cord radiosensitivity varies between irradiated lengths from 4.5-7.0 cm.

The follow-up period of the current study (51-56 weeks) is considered sufficient to observe the initial phase of radiation myelopathy as commonly reported for guinea pigs[19], rats[3, 11, 22], mice[18] and pigs[1, 4] (2–6 months). The latent period observed for the onset of motor deficits in the present study (8–13 weeks) is in good agreement with the companion study[1] Potentially more cases of radiation myelopathy would have been observed in a longer follow-up period; a previous spinal cord tolerance study[4] reported two late responding pigs at 65 and 75 weeks following irradiation to doses very close to ED₅₀ but the maximum followup period was 56 weeks in the present study.

In conclusion, no dose-volume effect is observed in pigs for the dose distributions studied and the endpoint of motor neurologic deficit; however, partial spinal cord irradiation results in less debilitating neurologic and histopathologic morbidity. The maximum point dose to the spinal cord was the parameter that best predicted the risk of motor neurologic deficit in this study.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This project was funded entirely by the US National Institute of Neurological Disorders and Stroke, R01 NS049517.

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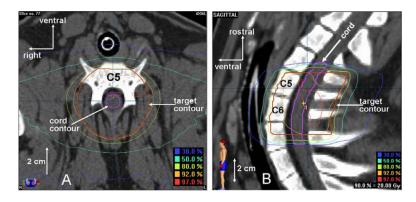


Figure 1. Dose distributions in the axial(A) and sagittal(B) planes.

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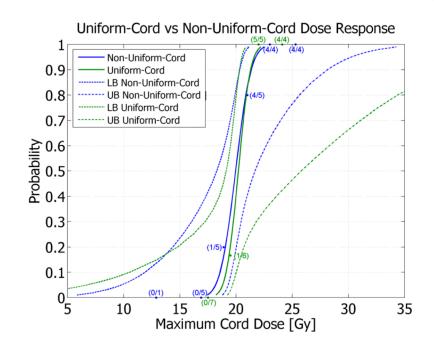


Figure 2.

Dose-response curves with 95% confidence bounds for uniform versus non-uniform dose distributions. *"Lower Bound" and "Upper Bound" have been abbreviated as LB and UB, respectively.

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Table 1

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iii <th< th=""><th>ID #</th><th>Rx Dose (Gy)</th><th>Irradiated Spinal Cord Length (cm)</th><th>Irradiated Spinal Cord Level</th><th>Image- Guidance / Irradiation Platform</th><th>Maximum Spinal Cord Dose (Gy)</th><th>Age at SRS (weeks)</th><th>Follow-up Period (weeks)</th><th>Latency (weeks)</th><th>Overall Tx Time (min)</th></th<>	ID #	Rx Dose (Gy)	Irradiated Spinal Cord Length (cm)	Irradiated Spinal Cord Level	Image- Guidance / Irradiation Platform	Maximum Spinal Cord Dose (Gy)	Age at SRS (weeks)	Follow-up Period (weeks)	Latency (weeks)	Overall Tx Time (min)
iii <th< td=""><td>-</td><td>16</td><td>4.5</td><td>midC4-midC7</td><td>N/X</td><td>17.6</td><td>45</td><td>56</td><td>${ m NA}^{*}$</td><td>14</td></th<>	-	16	4.5	midC4-midC7	N/X	17.6	45	56	${ m NA}^{*}$	14
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iii <th< td=""><td>3</td><td>16</td><td>6.8</td><td>CS-C7</td><td>N/X</td><td>17.4</td><td>45</td><td>52</td><td>NA^{*}</td><td>11</td></th<>	3	16	6.8	CS-C7	N/X	17.4	45	52	NA^{*}	11
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16 7,0 C5-C7 X/N 17,6 63 NA* 17 7.3 C5-C7 X/N 17.8 64 53 NA* 18 5.1 C5-C7 X/N 17.8 64 53 NA* 18 5.1 C5-mid7 X/N 192 102 25 NA* 18 5.1 C5-mid7 X/N 192 102 25 NA* 18 5.1 C5-mid7 X/N 192 102 26 NA* 18 5.1 Nict-mid7 X/N 192 102 NA* 104* 18 5.1 Nict-mid7 V/N 192 140 104* 104* 18 5.1 Nict-mid7 V/N 197 44 104* 104* 104* 19 5.1 Nict-mid7 V/N 107 104 104* 104* 104* 10 5.1 Nict 105 105	5	16	7.0	C5–C7	X/N	17.4	45	53	NA^{*}	8
167.0 $C3-C7$ X/N 17.8 46 53 N^{4} N^{4} 185.1 $C5-midC7$ X/N 19.2 102 53 8^{4} N^{4} 185.1 $C5-midC7$ X/N 19.2 102 54 N^{4} N^{4} 185.1 $C5-midC7$ X/N 19.2 19.2 99 54 N^{4} N^{4} 185.1 $mid-midC7$ V/S 19.7 46 54 N^{4} N^{4} 185.1 $mid-midC7$ V/S 19.7 46 54 N^{4} N^{4} 18 51 $mid-midC7$ V/S 19.7 46 54 N^{4} N^{4} 18 51 $mid-midC7$ V/S 19.7 46 54 N^{4} N^{4} 19 51 $mid-midC7$ V/S 19.7 46 54 N^{4} N^{4} 19 51 $mid-midC7$ V/S 19.7 47 54 N^{4} N^{4} 10 100 100 100 100 100 100 100 100 10 100 100 100 100 100 100 100 100 10 100 100 100 100 100 100 100 100 10 100 100 100 100 100 100 100 100 10 100 100 100 100 100 100 1	9	16	7.0	C5–C7	X/N	17.6	45	53	NA^*	10
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18 5.1 $C5-midC7$ XN 192 99 54 NA^* NA^* 18 5.1 $midC4-midC7$ V/S 19.7 46 54 NA^* NA^* 18 5.1 $midC4-midC7$ V/S 19.7 46 51 NA^* NA^* 18 5.1 $midC4-midC7$ V/S 19.7 46 51 NA^* NA^* 20 5.1 $midC4-midC7$ V/S 21.8 47 52 NA^* NA^* 20 5.1 $midC4-midC7$ XN 22.0 42 14.7 9.0 NA^* 20 5.1 $midC4-midC7$ V/S 22.1 45 11.4 9.0 NA^* 20 5.1 $midC4-midC7$ V/S 22.1 45 11.4 9.0 10.7 20 5.1 $midC4-midC7$ V/S 21.9 47 14.7 9.0 10.7 20 5.1 $midC4-midC7$ V/S 21.9 47 14.7 9.0 10.7 20 5.1 $midC4-midC7$ V/S 21.9 47 14.7 9.9 10.7 21 5.1 $midC4-midC7$ V/S 21.9 47 14.7 90 10.7 22 5.1 $midC4-midC7$ V/S 21.9 47 14.7 90 10.7 22 5.1 $midC4-midC7$ V/S 21.9 47 14.7 90 10.7 22 5.1 $midC4-midC7$ V/S <td>6</td> <td>18</td> <td>5.1</td> <td>C5-midC7</td> <td>N/X</td> <td>19.4</td> <td>66</td> <td>54</td> <td>${ m NA}^{*}$</td> <td>12</td>	6	18	5.1	C5-midC7	N/X	19.4	66	54	${ m NA}^{*}$	12
18 5.1 midc4-midc7 V/S 19.7 46 54 NA* 18 5.1 midc4-midc7 V/S 19.7 46 51 NA* 18 5.1 midc4-midc7 V/S 19.7 47 52 NA* 20 5.1 midc4-midc7 X/N 218 43 55 107 20 5.1 midc4-midc7 X/N 218 43 55 100 20 5.1 midc4-midc7 X/N 22.0 42 14 10 10 20 5.1 midc4-midc7 X/N 22.0 42 14 10 10 20 5.1 midc4-midc7 V/S 22.0 47 14 10<	10	18	5.1	C5-midC7	N/X	19.2	66	54	NA^{*}	10
18 5.1 midc4-midc7 V/S 19.7 46 51 N/A* 18 5.1 midc4-midc7 V/S 19.7 46 51 N/A* 20 5.1 midc4-midc7 X/N 21.8 43 55 10 20 5.1 midc4-midc7 X/N 22.0 42 14 9 16 20 5.1 midc4-midc7 V/S 22.1 45 14 10 10 10 20 5.1 midc4-midc7 V/S 22.1 47 14 13 10 210 5.1 midc4-midc7 V/S 21.9 47 14 13 10	11	18	5.1	midC4-midC7	V/S	19.7	46	54	${ m NA}^{*}$	22
18 5.1 midc4-midc7 V/S 19.7 47 52 NA* 20 5.1 midc4-midc7 X/N 21.8 43 55 10 20 5.1 midc4-midc7 X/N 22.0 43 55 10 20 5.1 midc4-midc7 X/N 22.0 45 14 9 9 20 5.1 midc4-midc7 V/S 22.1 45 14 9 10 20 5.1 midc4-midc7 V/S 21.9 45 14 10 10 210 5.1 midc4-midc7 V/S 21.9 45 14 10 <td< td=""><td>12</td><td>18</td><td>5.1</td><td>midC4-midC7</td><td>S/A</td><td>19.7</td><td>46</td><td>51</td><td>NA^{*}</td><td>25</td></td<>	12	18	5.1	midC4-midC7	S/A	19.7	46	51	NA^{*}	25
20 5.1 $midc4-midc7$ X/N 21.8 43 55 10 10 20 5.1 $midc4-midc7$ X/N 22.0 42 14 9 9 20 5.1 $midc4-midc7$ V/S 22.1 45 11 10 10 20 5.1 $midc4-midc7$ V/S 21.9 47 14 10 10 20 5.1 $midc4-midc7$ V/S 21.9 47 14 10 10 20 5.1 $midc4-midc7$ V/S 21.9 45 11 10 10 22 5.1 $midc4-midc7$ V/S 24.1 46 11 10 10 22 5.1 $midc4-midc7$ V/S 24.1 46 12 12 12 22 5.1 $midc4-midc7$ V/S 24.1 46 12 12 12 22 5.1 $midc4-midc7$ V/S 24.1 46 12 12 12 22 5.1 $midc4-midc7$ V/S 24.1 46 11 11 12 22 5.1 $midc4-midc7$ V/S 24.1 47 12 11 11 22 5.1 $midc4-midc7$ V/S 24.1 47 11 11 11 22 5.1 $midc4-midc7$ V/S 24.1 47 11 11 11	13	18	5.1	midC4-midC7	S/A	19.7	47	52	* NA	25
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20 5.1 midc4-midc7 V/S 22.1 45 11 10 10 20 5.1 midc4-midc7 V/S 21.9 47 14 13 13 20 5.1 midc4-midc7 V/S 21.9 47 14 13 15 20 5.1 midc4-midc7 V/S 21.9 45 11 10	15	20	5.1	midC4-midC7	N/X	22.0	42	14	6	10
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22 5.1 midc4-midc7 V/S 24.0 46 9 9 9 22 5.1 midc4-midc7 V/S 24.1 46 12 12 12 22 5.1 midc4-midc7 V/S 24.1 46 12 12 12 22 5.1 midc4-midc7 V/S 24.1 45 11 11 11 22 5.1 midc4-midc7 V/S 24.4 47 10 10 10	18	20	5.1	midC4-midC7	V/S	21.9	45	11	10	20
22 5.1 midc4-midC7 V/S 24.1 46 12 12 22 5.1 midc4-midC7 V/S 24.1 45 11 11 22 5.1 midc4-midC7 V/S 24.4 45 11 11	19	22	5.1	midC4-midC7	V/S	24.0	46	9	6	26
22 5.1 midc4-midc7 V/S 24.1 45 11	20	22	5.1	midC4-midC7	V/S	24.1	46	12	12	24
22 5.1 midC4-midC7 V/S 24.4 47 10 10	21	22	5.1	midC4-midC7	V/S	24.1	45	11	11	23
	22	22	5.1	midC4-midC7	V/S	24.4	47	10	10	20

X = "Stereoscopic X-ray"

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V = "Volumetric Cone Beam Computed Tomography"N = "Novalis"

S = "Synergy S"

* No motor neurologic deficits were observed in stated follow-up period.

Table 2

Spinal cord dose-volume histogram statistics for animals receiving uniform irradiation.

Rx Dose Group (Gy)	Mean Maximum Point Dose (Gy)	Mean Percent Volume >= 10 Gy	Mean Volume >=14 Gy (cc)	Mean Maximum Dose to 1 cc Volume (Gy)
16 (<i>n</i> =7)	17.5 ± 0.1	91 ± 3	4.12 ± 0.67	17.1 ± 0.0
18 (<i>n</i> =6)	19.5 ± 0.2	94 ± 3	3.74 ± 0.13	19.0 ± 0.4
20 (<i>n</i> =5)	22.0 ± 0.1	93 ± 4	3.87 ± 0.16	21.5 ± 0.2
22 (<i>n</i> =4)	24.1 ± 0.2	92 ± 0	4.00 ± 0.21	23.7 ± 0.1