



# Calcium cycling proteins and heart failure: mechanisms and therapeutics

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**Ca<sup>2+</sup>-dependent signaling is highly regulated in cardiomyocytes and determines the force of cardiac muscle contraction. Ca<sup>2+</sup> cycling refers to the release and reuptake of intracellular Ca<sup>2+</sup> that drives muscle contraction and relaxation. In failing hearts, Ca<sup>2+</sup> cycling is profoundly altered, resulting in impaired contractility and fatal cardiac arrhythmias. The key defects in Ca<sup>2+</sup> cycling occur at the level of the sarcoplasmic reticulum (SR), a Ca<sup>2+</sup> storage organelle in muscle. Defects in the regulation of Ca<sup>2+</sup> cycling proteins including the ryanodine receptor 2, cardiac (RyR2)/Ca<sup>2+</sup> release channel macromolecular complexes and the sarcoplasmic/endoplasmic reticulum Ca<sup>2+</sup> ATPase 2a (SERCA2a)/phospholamban complex contribute to heart failure. RyR2s are oxidized, nitrosylated, and PKA hyperphosphorylated, resulting in “leaky” channels in failing hearts. These leaky RyR2s contribute to depletion of Ca<sup>2+</sup> from the SR, and the leaking Ca<sup>2+</sup> depolarizes cardiomyocytes and triggers fatal arrhythmias. SERCA2a is downregulated and phospholamban is hypophosphorylated in failing hearts, resulting in impaired SR Ca<sup>2+</sup> reuptake that conspires with leaky RyR2 to deplete SR Ca<sup>2+</sup>. Two new therapeutic strategies for heart failure (HF) are now being tested in clinical trials: (a) fixing the leak in RyR2 channels with a novel class of Ca<sup>2+</sup>-release channel stabilizers called Rycals and (b) increasing expression of SERCA2a to improve SR Ca<sup>2+</sup> reuptake with viral-mediated gene therapy. There are many potential opportunities for additional mechanism-based therapeutics involving the machinery that regulates Ca<sup>2+</sup> cycling in the heart.**

## Excitation-contraction coupling

With each beat of the heart, Ca<sup>2+</sup> is released from the sarcoplasmic reticulum (SR) via the ryanodine receptor 2, cardiac (RyR2), raising the cytosolic Ca<sup>2+</sup> concentration about ten-fold (~1 μM) and activating cardiac muscle contraction (Figure 1). The Ca<sup>2+</sup> is then pumped back into the SR by the sarcoplasmic/endoplasmic reticulum Ca<sup>2+</sup> ATPase 2a (SERCA2a), lowering the cytosolic Ca<sup>2+</sup> concentration to baseline levels (~100 nM) and causing relaxation. The Ca<sup>2+</sup> release and reuptake cycle is initiated by the action potential, an electrical signal that depolarizes the plasma membrane and the specialized invagination called the transverse tubule (T tubule). Voltage-gated Ca<sup>2+</sup> channels on the T tubule are activated by depolarization and allow a small amount of Ca<sup>2+</sup> to run down its concentration gradient from mM external Ca<sup>2+</sup> concentration to nM internal Ca<sup>2+</sup> concentration. This entering Ca<sup>2+</sup> binds to and activates RyR2 channels, which release Ca<sup>2+</sup> stored at high concentration (in the millimolar range) in the SR. The Ca<sup>2+</sup> binds to troponin C, allowing actin-myosin cross-bridging and the thick and thin filaments of the sarcomere to slide past each other, shortening the sarcomere and causing cardiac muscle contraction.

## Heart failure

Heart failure (HF) is the leading cause of mortality and morbidity in developed countries. The incidence of HF continues to increase after age 65, affecting nearly 1 in 100 individuals (1). This is despite substantial advances in the care of patients, brought about by coronary care units and the development of devices for the treatment of HF including biventricular pacing and left ventricular assist devices (LVADs) (2). The most common cause of HF in developed

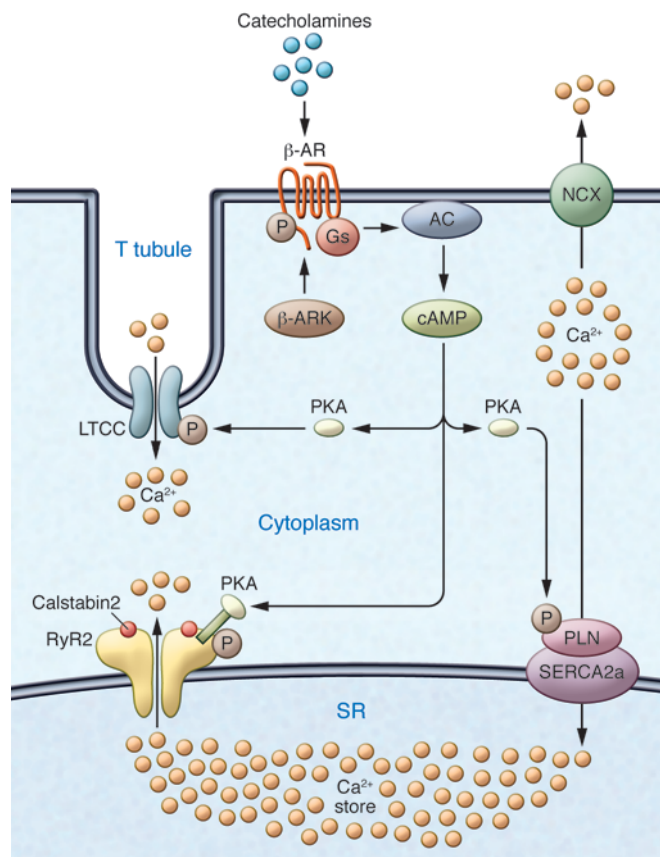
countries is atherosclerosis and concomitant ischemic heart disease. Other causes include hypertension (which leads to hypertrophy that can degenerate to dilated cardiomyopathy and HF), dilated non-ischemic cardiomyopathies, and much rarer genetic causes. While HF initially involves the myocardium, resulting in decreased cardiac performance, it rapidly affects multiple organs including, most prominently, the neurohormonal, circulatory, and renal systems. Indeed, patients with HF have chronic activation of the sympathetic nervous system, which results in a maladaptive attempt to improve cardiac function. Moreover, β-adrenergic agonists or phosphodiesterase (PDE) inhibitors do increase contractility by increasing cAMP and increasing Ca<sup>2+</sup> release, but they also increase mortality. In fact, blocking neurohormonal pathways is the focus of current HF therapy, and while this improves survival, it is limited by side effects and the requirement to titrate drugs to physiological parameters such as heart rate (HR) and blood pressure (3). Additional current therapies are aimed at reducing the symptoms of HF (e.g., diuretics for pulmonary and peripheral congestion), but they do not inhibit HF progression.

The search for novel therapeutics for HF has led investigators to examine the mechanisms underlying HF with the hope that this approach will uncover potential therapeutic targets to slow HF progression, improve quality of life, and reduce mortality.

Much attention has been paid to understanding the role of defects in Ca<sup>2+</sup> regulation in HF (4). This is because, as noted above, Ca<sup>2+</sup> is the signal that regulates cardiac muscle contraction. Cardiac contractility is determined by the amplitude and kinetics of Ca<sup>2+</sup> cycling, which in turn are regulated by phosphorylation and dephosphorylation of key proteins involved in excitation-contraction (EC) coupling by kinases and phosphatases. Stress-induced activation of G protein-coupled β-adrenergic receptors (β-ARs) activates adenylyl cyclase, which in turn leads to cAMP production and PKA activation. PKA phosphorylates many proteins in cardiac muscle, including those

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**Figure 1**

Ca<sup>2+</sup> cycling in cardiomyocytes and regulation by PKA. EC coupling in the heart starts with depolarization of the T tubule, which activates voltage-gated L-type Ca<sup>2+</sup> channels (LTCCs) in the plasma membrane. Ca<sup>2+</sup> influx via LTCCs triggers Ca<sup>2+</sup> release from the SR via RyR2 (SR Ca<sup>2+</sup> release channel). During systole, the free intracellular Ca<sup>2+</sup> concentration increases ten-fold from ~100 nM to ~1 μM, which enables muscle contraction. The β-AR signaling pathway can increase the Ca<sup>2+</sup> transient by activating the trigger (LTCC), release (RyR2), and uptake (SERCA/phospholamban [SERCA/PLN]) pathways. Catecholamine activation of β-ARs allows for the activation of adenylate cyclase (AC), mediated by specific G proteins (Gs), and the generation of cAMP, which in turn activates PKA. Relaxation occurs after intracellular Ca<sup>2+</sup> is pumped out of the cytoplasm by SERCA2a, which is regulated by phospholamban. In addition, Ca<sup>2+</sup> is extruded from the cell by the sarcolemmal NCX. RyR2 is a macromolecular complex comprised of four RyR2 monomers, PP1-spinophilin, PP2A-PR130, PKA-PDE4D3-mAKAP, calstabin2, CaMKII, and calmodulin. Calsequestrin regulates luminal SR free Ca<sup>2+</sup>, and junctin and triadin help maintain the integrity of the T tubule–SR junction. β-ARK, β-AR kinase.

decrease in systolic Ca<sup>2+</sup> transient amplitude secondary to reduced SR Ca<sup>2+</sup> stores was responsible for the decreased contractility and reduced cardiac output in HF (12). Moreover, aberrant diastolic release of SR Ca<sup>2+</sup> can generate a transient inward current that causes delayed afterdepolarizations (DADs) (10, 13).

Impaired SERCA2a function and enhanced Na<sup>+</sup>/Ca<sup>2+</sup> exchanger (NCX) activity have been proposed as causes of reduced SR Ca<sup>2+</sup> load in HF (14). However, over the past 12 years the role of a pathological diastolic SR Ca<sup>2+</sup> leak via calstabin2-depleted leaky RyR2 has been recognized as an important contributor to altered Ca<sup>2+</sup> handling in HF (5). In failing hearts, the RyR2-mediated Ca<sup>2+</sup> leak is caused by the chronic hyperadrenergic state observed in patients with HF, which in turn induces chronic PKA hyperphosphorylation at Ser2808 and oxidation of RyR2, causing depletion of calstabin2 (5) from the channel complex that destabilizes the closed state of the channel (Figure 3). The term hyperphosphorylation describes RyR2 in which 3–4 of the four RyR2 monomers are chronically PKA phosphorylated. PKA-hyperphosphorylated/calstabin2-depleted channels are sensitized to cytosolic Ca<sup>2+</sup>, leading to inappropriate Ca<sup>2+</sup> release during diastole, referred to as a diastolic SR Ca<sup>2+</sup> leak. Ca<sup>2+</sup> leak reduces SR Ca<sup>2+</sup> stores and activates the transient inward current (15).

The RyR2 Ca<sup>2+</sup> leak model of HF is supported by studies demonstrating that: (a) β-adrenergic stimulation causes depletion of calstabin2 from the RyR2 complex; (b) HF patients have PKA-hyperphosphorylated and calstabin2-depleted RyR2 (16–23); and (c) patients whose cardiac function has been normalized by treatment with LVADs have reduced levels of circulating catecholamines (24) and reduced phosphorylation of RyR2 at Ser2809 (5). Indeed, improved cardiac function in patients treated with LVAD is associated with restoration of calstabin2 binding to RyR2 (5).

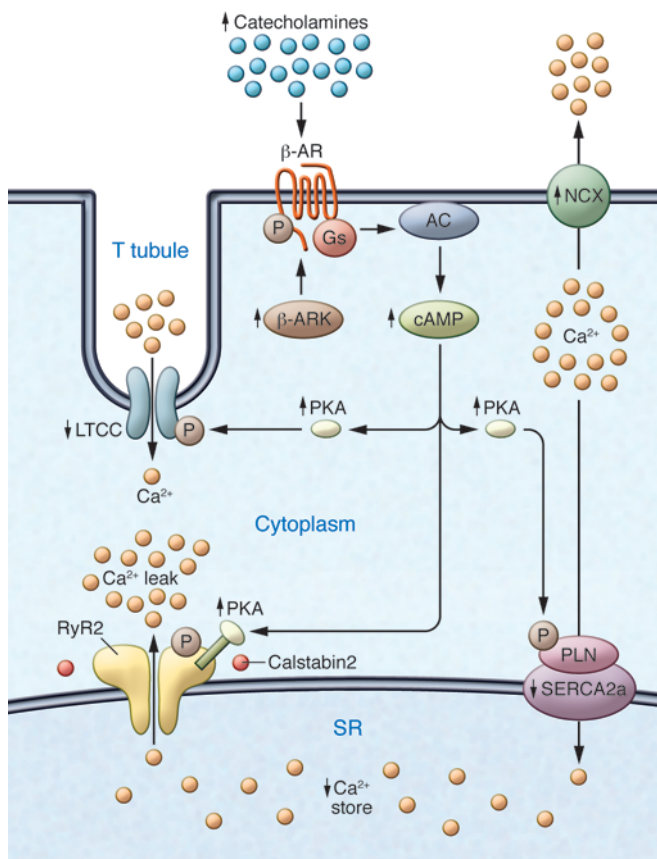
The RyR2 leak is caused by stress-induced remodeling of the RyR2 macromolecular complex due to PKA hyperphosphorylation, nitrosylation, and oxidation of the channel that results in depletion of calstabin2, phosphatases (25), and PDE4D3 (6) from the RyR2 channel in HF (7). Depletion of PDE4D3 and phosphatases results in elevated levels of cAMP at RyR2 (6) and a decreased rate of dephosphorylation of a hyperphosphorylated channel, promoting further PKA hyperphosphorylation (7, 25). The depletion of the channel subunit calstabin2 from the channel results

involved in EC coupling: the L-type Ca<sup>2+</sup> channel, RyR2, and phospholamban (Figure 2), as well as troponin I, myosin binding protein C, and protein phosphatase inhibitor-1. PKA phosphorylation of these proteins (e.g., RyR2) results in increased HR as well as increased amplitude and velocity of Ca<sup>2+</sup> release and reuptake and enhanced contractility (5). Whereas PKA is the “on switch” for cardiac HR and contractility, the off switches are the protein phosphatases, including PP1 and PP2A, as well as PDE4D3, which hydrolyzes cAMP (6).

It is now generally accepted that defective SR Ca<sup>2+</sup> handling plays an important role in HF pathophysiology (7). This defective SR Ca<sup>2+</sup> handling is characterized chiefly by leaky RyR2 channels, due to stress-induced dissociation of the stabilizing RyR2 subunit calstabin2 (also known as FKBP12.6) resulting in a diastolic SR Ca<sup>2+</sup> leak, reduced SR Ca<sup>2+</sup> content, and decreased Ca<sup>2+</sup> transient (6–10). Compounding this problem is impaired SR Ca<sup>2+</sup> uptake due to reduced activity of SERCA2a, as a consequence of both reduced SERCA2a expression and increased inhibition of the pump by phospholamban (11). Thus, these two major players in cardiac EC coupling, the SR Ca<sup>2+</sup> release channel (RyR2) and the SR Ca<sup>2+</sup> uptake pump (SERCA2a), conspire to deplete the SR of Ca<sup>2+</sup> and lead to impaired cardiac contractility (4). Not surprisingly, therefore, both the RyR2 leak and the impaired uptake have been targeted with novel therapeutics, both of which are now undergoing clinical testing in HF patients.

### Ca<sup>2+</sup> signaling defects in HF: leaky RyR2 channels, decreased SR Ca<sup>2+</sup> content, and reduced transients

Over 20 years ago, reports of reduced Ca<sup>2+</sup> transient amplitude, increased Ca<sup>2+</sup> transient duration, prolonged Ca<sup>2+</sup> transient decay time, and more recently reduced SR Ca<sup>2+</sup> load suggested that a



**Figure 2**

Defective  $\text{Ca}^{2+}$  handling in failing hearts due to sympathetic overactivity. Chronic activity of the sympathetic nervous system leads to phosphorylation of the  $\beta$ -AR, activation of  $\beta$ -AR kinase, and desensitization of  $\beta$ -ARs. The LTCC is also phosphorylated, and NCX expression is upregulated. An important contributor to impaired  $\text{Ca}^{2+}$  handling in HF is PKA hyperphosphorylation of RyR2. This leads to a higher sensitivity to  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release at low cytoplasmic  $\text{Ca}^{2+}$  concentrations, resulting in increased RyR2 open probability at low  $\text{Ca}^{2+}$  concentrations and a diastolic SR  $\text{Ca}^{2+}$  leak. The long-term effect of the diastolic  $\text{Ca}^{2+}$  leak is depletion of SR  $\text{Ca}^{2+}$  stores. SERCA2a expression and activity are decreased in HF, which is linked to phospholamban hypophosphorylation. In contrast, NCX expression and activity are upregulated in HF. Arrows indicate increased or decreased expression or activity in HF.

**Arrhythmias: lessons from catecholaminergic polymorphic VT**

Catecholaminergic polymorphic VT (CPVT) is a rare inherited form of exercise-induced sudden cardiac death (SCD) that occurs in individuals with structural normal hearts and normal ECGs. Mutations in RyR2 have been linked to CPVT (13, 34).

We originally reported that RyR2 CPVT mutations reduced the affinity for calstabin2, resulting in leaky channels during exercise (13). Treating mice with a Rycal (a novel class of drugs described below that prevent diastolic SR  $\text{Ca}^{2+}$  leak via RyR2 channels) has been shown to prevent exercise-induced depletion of calstabin2 from the RyR2 complex and reduced VT and SCD (9, 13). Calstabin2-deficient and haploinsufficient mice exhibit CPVT (13) but do not develop HF. Presumably the reason for the lack of cardiac dysfunction is that, in contrast to the post-MI model, in which calstabin2 depletion from the RyR2 complex has been shown to promote HF progression, the calstabin2-deficient mice have otherwise normal cardiac function and are able to compensate for the chronic diastolic SR  $\text{Ca}^{2+}$  leak in the absence of a compromised ventricle (e.g., no MI). However, PDE4D3-deficient mice that have chronic PKA hyperphosphorylation of RyR2 and RyR2-S2808D mice that mimic chronically PKA-hyperphosphorylated RyR2 both have calstabin2 depletion from the RyR2 complex, and both exhibit progressive cardiac dysfunction leading to a dilated cardiomyopathy in the absence of MI (6). This indicates that the combination of calstabin2 deficiency plus another insult (e.g., PKA hyperphosphorylation of RyR2) is sufficient to cause cardiac dysfunction in the absence of MI (7). However, the loss of calstabin2 alone, without another insult to the myocardium, can be compensated for at baseline, although this causes SCD with exercise due to leaky RyR2.

**Fixing leaky RyR2 channels**

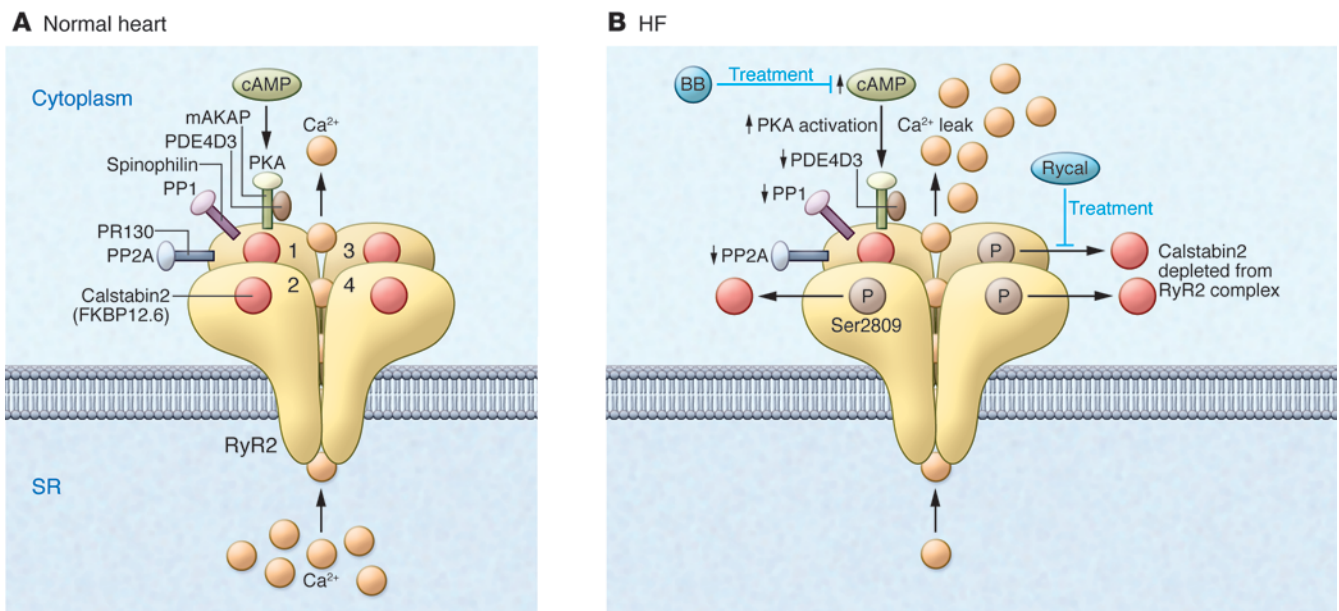
Identification of the diastolic SR  $\text{Ca}^{2+}$  leak via RyR2 as a mechanism underlying HF progression and cardiac arrhythmias has led to novel therapeutic approaches. JTV-519 (K201), a 1,4-benzothiazepine, was noted to have effects on intracellular  $\text{Ca}^{2+}$  (35) and cardioprotective effects (36). However, the target was not known and there was no mechanism of action for JTV-519. Moreover, JTV-519 was found to inhibit  $\text{Na}^+$ ,  $\text{Ca}^{2+}$ , and  $\text{K}^+$  currents (37, 38). Using a canine model of pacing-induced HF, Matsuzaki and colleagues reported that JTV-519 improved cardiac function (19).

Testing the drug in calstabin2-deficient mice showed that the ability of JTV-519 to prevent HF progression and fatal cardiac arrhythmias and improve skeletal muscle function requires stabi-

in destabilization of the channel closed state and the diastolic SR  $\text{Ca}^{2+}$  leak that reduce the SR  $\text{Ca}$  transient, resulting in impaired contractility and trigger fatal cardiac arrhythmias (5, 26). Importantly, the same stress-induced remodeling affects the skeletal muscle RyR1 complex and contributes to weakened skeletal muscle function and impaired exercise capacity in HF (27, 28).

The discovery of leaky RyR2 in failing hearts also provides a mechanism to explain the therapeutic efficacy of  $\beta$ -AR blockers in HF. Indeed,  $\beta$ -blockers inhibit PKA phosphorylation of Ser2808 in murine RyR2 (Ser2809 in human RyR2) by blocking  $\beta$ -ARs, indirectly prevent calstabin2 depletion from the RyR2 complex, and thereby reduce SR  $\text{Ca}^{2+}$  leak in HF patients, resulting in improved contractility and decreased arrhythmogenesis (7, 29, 30).

Multiple animal models have been used to support a role for PKA hyperphosphorylation of RyR2 in HF progression (31). Genetically altered mice harboring RyR2 that cannot be PKA phosphorylated (RyR2-S2808A) were protected against calstabin2 depletion from the RyR2 complex and HF progression 4 weeks after myocardial infarction (MI) (32). PDE4D3-deficient mice develop age-dependent cardiomyopathy and arrhythmias, RyR2 PKA hyperphosphorylation, and calstabin2 depletion. The crossing of PDE4D3-deficient mice and RyR2-S2808A mice has been shown to be protective against HF progression and arrhythmias (6). Transgenic mice expressing a mutant calstabin2-D37V, which remains bound to PKA-phosphorylated RyR2 channels, are also protected against post-MI HF (33). Calstabin2-deficient mice exhibit DADs and exercise-induced ventricular tachycardia (VT) (13), and RyR2s from calstabin2-deficient mice exhibit slightly increased open probability at baseline that increases substantially when the mice are exercised.

**Figure 3**

Dysfunctional RyR2 in failing hearts. **(A)** The RyR2 macromolecular complex includes four identical RyR2 subunits (numerals 1–4 indicate the four monomers). Each RyR2 subunit binds one calstabin2 (also known as FKBP12.6) as well as mAKAP, to which PKA catalytic and regulatory subunits and PDE4D3 are bound; PP2A and its targeting protein PR130; and PP1 and its targeting protein spinophilin (accessory molecules are only shown for one of the four RyR2 subunits, except calstabin2, which is shown for all four RyR2 subunits). The  $\beta$ -adrenergic signaling pathway can activate PKA through the second messenger cAMP. **(B)** In HF, PKA hyperphosphorylation of Ser2809, due to reduced PDE4D3, PP1, and PP2A levels in the RyR2 macromolecular complex, depletes calstabin2 from the RyR2 channel complex. The functional effect of these changes in the macromolecular composition of RyR2 is a pathological increase in Ca<sup>2+</sup>-dependent activation of RyR2 and depletion of SR Ca<sup>2+</sup> stores, as well as functional uncoupling of RyR2 from their neighboring channels. Ca<sup>2+</sup> leak due to abnormal RyR2 channel openings may be prevented by treatment with  $\beta$ -blockers (BBs), which interfere with the upstream  $\beta$ -AR signaling pathway, or with Rycals, which selectively increase the binding affinity of calstabin2 to PKA-phosphorylated and oxidized RyR2.

lization of the closed state of RyR2 by calstabin2 (28, 39). Moreover, JTV-519 had no effect on the gating properties of normal RyR channels and no effects in healthy dogs and mice (9).

My laboratory generated many derivatives of the 1,4-benzothiazepine JTV-519 and have developed a novel class of Ca<sup>2+</sup> release channel stabilizers known as Rycals. An orally available Rycal, S107, improves skeletal muscle force generation and exercise capacity, reduces arrhythmias, and improves muscle function in mice with Duchenne muscular dystrophy by reducing pathologic SR Ca<sup>2+</sup> leak in cardiac and skeletal muscle (7, 9, 40–43). Rycals are protective against post-MI HF progression (7, 44) and have been shown to suppress VT/ventricular fibrillation (VT/VF) and SCD in murine models of human CPVT. S107 also raises the seizure threshold in mice with leaky neuronal RyR2 channels and improves exercise capacity in mouse models of sarcopenia (age-related loss of muscle function) (6, 9, 10, 34, 41).

Dantrolene, a drug used to prevent malignant hyperthermia in patients with mutations in RyR1 who have been exposed to volatile anesthetics, has been proposed to have therapeutic potential in heart disease (45). The Na<sup>+</sup> channel antagonist flecainide prevents ventricular arrhythmias in patients with CPVT mutations (46). Based on single-channel data, flecainide has been proposed as an open channel blocker of RyR2. However, it is unlikely that this could be the mechanism of its anti-arrhythmic activity in CPVT, since blocking the open state of RyR2 would seriously impair cardiac contractility. Moreover, the leak that triggers the fatal ventricular arrhythmias in CPVT is a diastolic SR Ca<sup>2+</sup> leak

that occurs when the channel is supposed to be tightly shut, not open (8, 13, 39). Finally, it is more likely that some other activity of flecainide, such as its well-documented sodium channel blockade (47), is the mechanism underlying its anti-arrhythmic actions in CPVT.

### Focus on the pump

SERCA activity and protein levels are decreased in failing hearts (11). Mice that are haploinsufficient for the SERCA2a gene (*Atp2a2*) develop accelerated HF progression compared with wild-type controls (48). However, patients with an *ATP2A2* mutation and haploinsufficiency (Darier disease) do not have cardiac dysfunction (49). Nevertheless, restoration of normal levels of SERCA2a has been targeted as a novel therapeutic for HF (4). Phospholamban controls the affinity of SERCA2 for Ca<sup>2+</sup>. Unphosphorylated phospholamban inhibits SERCA2 Ca<sup>2+</sup> affinity, and phosphorylation by PKA and Ca<sup>2+</sup>/calmodulin-dependent protein kinase II (CaMKII) relieves this inhibition and results in enhanced SR Ca<sup>2+</sup> uptake. However, a human mutation in phospholamban results in severe dilated cardiomyopathy (50).

### Increasing SR Ca<sup>2+</sup> uptake

Both SERCA overexpression and phospholamban inhibition have been targeted as therapeutic strategies for HF. Cardiac-specific expression of SERCA2a was shown to improve contractility (51). However, results regarding arrhythmias are controversial. Chen et al. reported increased arrhythmias leading to mortality



after MI in transgenic rats (52), whereas decreases in ventricular arrhythmias after postischemic injury were reported (53). Beneficial results observed in preclinical testing have led to clinical trials in patients with HF to enhance SR  $\text{Ca}^{2+}$  uptake, including a first-in-human gene therapy trial, Calcium Upregulation by Percutaneous administration of gene therapy In cardiac Disease (CUPID), using an adeno-associated type 1 vector carrying SERCA2a. In 39 patients with New York Heart Association class III/IV HF, treatment with the SERCA2a adenovirus resulted in improvement or stabilization in the New York Heart Association class, Minnesota Living With Heart Failure Questionnaire, 6-minute walk test, peak maximum oxygen consumption, N-terminal pro-hormone brain natriuretic peptide levels, and left ventricular end-systolic volume, as well as decreased frequency of cardiovascular events and duration of hospitalizations (54). Further trials are planned.

### Controversies in the field

The diastolic SR  $\text{Ca}^{2+}$  leak model for HF and arrhythmias first proposed in 2000 (5) has generated great interest and has led to clinical trials for HF and arrhythmias as noted above. Indeed, many laboratories have examined aspects of our findings. While essentially all of our findings have been reproduced by others (16, 17, 19–23, 55–57), controversy surrounding our work has emerged over the years, with reports challenging our studies as summarized recently (58). I have previously addressed this controversy and pointed out that most of the differences are based either on opposite interpretations of the same findings or on profound differences in experimental approach (59).

We originally reported that RyR2s are PKA hyperphosphorylated in human and canine failing hearts (5) and defined PKA hyperphosphorylation as phosphorylation of 3–4 of the four PKA sites (Ser2808) in the RyR2 homotetramer (60, 61). We also reported that PKA hyperphosphorylation of RyR2 was associated with depletion of the stabilizing subunit calstabin2 from the channel (there are four calstabin2 bound to each RyR2 channel, again one per monomer; ref. 5) and that this renders the channel “leaky,” meaning that at diastolic cytosolic  $\text{Ca}^{2+}$  concentration (~100 nM) when the RyR2 channel should be tightly closed, the PKA-hyperphosphorylated, calstabin2-depleted channels are not tightly closed; that is, HF RyR2 channels have increased single-channel open probability when recorded in planar lipid bilayers (5). This results in a diastolic SR  $\text{Ca}^{2+}$  leak that depletes SR  $\text{Ca}^{2+}$ , contributing to impaired contractility and HF progression, and provides signals that lead to membrane depolarization and triggering of fatal ventricular arrhythmias in failing hearts (13, 34).

Alternative mechanisms have been proposed to explain SR  $\text{Ca}^{2+}$  leak in HF. CaMKII $\delta$  levels are elevated in human HF (62), and there is an increase in CaMKII-dependent phosphorylation of RyR2 in HF. The observation that mice expressing the CaMKII inhibitory peptide AC3-I are protected against HF led to the proposal that CaMKII inhibitors may prevent HF progression (63). It has been reported that PKA hyperphosphorylation of RyR2 is involved in ischemic (post-MI) but not in non-ischemic (aortic banding) cardiomyopathy, and that CaMKII phosphorylation of RyR2 is the mechanism for the leak (64). The aortic banding model is a hypertrophy model in which cardiac function is initially increased and later decreases. It is possible that elevated CaMKII phosphorylation of RyR2 is associated with the hypertrophy.

Moreover, we developed genetically altered mice that harbor RyR2 that cannot be phosphorylated by CaMKII (RyR2-S2814A) and showed that they were not protected against post-MI HF progression (65). Instead, RyR2 CaMKII phosphorylation is required for the rate-related increase in contractility (65). CaMKII is  $\text{Ca}^{2+}$  activated and is exposed to more  $\text{Ca}^{2+}$  at higher HRs, resulting in increased CaMKII phosphorylation of RyR2, increased SR  $\text{Ca}^{2+}$  release, and increased contractility (known as the Bowditch phenomenon) (65). HR is typically elevated in HF due to increased levels of catecholamines, and thus CaMKII activation during HF is likely a result of increased HR.

We originally reported that PKA hyperphosphorylation of RyR2 causes depletion of calstabin2 from the RyR2 complex (5). We subsequently observed that oxidation and nitrosylation of RyR2 also cause depletion of calstabin2 from the RyR2 complex, and that the combination of all three deplete nearly all of the calstabin2 from the channel complex (7, 44). Some of the divergent results reported in the literature concerning the effects of PKA phosphorylation on calstabin binding to the channels could be due to variations in the state of oxidation of the channels. For example, we found that when RyR2-S2808D channels were expressed in HEK cells, there was progressive oxidation of the channel, such that the binding of calstabin2 to the channel decreased each day the transfected cells were in culture (7). This progressive oxidation is likely due to mitochondrial  $\text{Ca}^{2+}$  overload resulting from leaky RyR2 channels and ROS production, which oxidizes the channel (41).

### Future directions: new and old pathways, additional therapeutic targets

It should be evident from Figure 1 that there are many potential therapeutic targets in the key pathways that regulate  $\text{Ca}^{2+}$  handling in cardiac muscle. For example, the cardiac RyR2 is a macromolecular signaling complex with multiple enzymes and targeting proteins (15), each of which should be explored as a potential cause of, and therapeutic target for, HF at the cellular, animal, and human genetic levels (8). Moreover, recent work on mechanisms of SR  $\text{Ca}^{2+}$  leak in different disease models has shown the critical role of nitrosylation of RyR1 in muscular dystrophy (43) and RyR2s (42) and of interactions between RyR1 and mitochondria in skeletal muscle, whereby oxidation of RyR1 causes  $\text{Ca}^{2+}$  overload of mitochondria and further oxidation of the channel, worsening the SR  $\text{Ca}^{2+}$  leak (41). These studies suggest that targeting molecules involved in oxidation/nitrosylation and mitochondria should be explored for future therapeutics.

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