



# Role of dynamin, synaptojanin, and endophilin in podocyte foot processes

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**Podocytes are specialized cells that play an integral role in the renal glomerular filtration barrier via their foot processes. The foot processes form a highly organized structure, the disruption of which causes nephrotic syndrome. Interestingly, several similarities have been observed between mechanisms that govern podocyte organization and mechanisms that mediate neuronal synapse development. Dynamin, synaptojanin, and endophilin are functional partners in synaptic vesicle recycling via interconnected actions in clathrin-mediated endocytosis and actin dynamics in neurons. A role of dynamin in the maintenance of the kidney filtration barrier via an action on the actin cytoskeleton of podocytes was suggested. Here we used a conditional double-KO of dynamin 1 (*Dnm1*) and *Dnm2* in mouse podocytes to confirm dynamin's role in podocyte foot process maintenance. In addition, we demonstrated that while synaptojanin 1 (*Synj1*) KO mice and endophilin 1 (*Sh3gl2*), endophilin 2 (*Sh3gl1*), and endophilin 3 (*Sh3gl3*) triple-KO mice had grossly normal embryonic development, these mutants failed to establish a normal filtration barrier and exhibited severe proteinuria due to abnormal podocyte foot process formation. These results strongly implicate a protein network that functions at the interface between endocytosis and actin at neuronal synapses in the formation and maintenance of the kidney glomerular filtration barrier.**

## Introduction

Nephrotic syndrome is a severe kidney disease involving massive protein loss in the urine due to an impairment of the glomerular filtration barrier between the blood and the urinary space. Podocytes, highly specialized cells that line the capillary loops, play a key role in this barrier via their interdigitated foot processes connected by a thin diaphragm, the slit diaphragm (1). Accordingly, disruption of foot process organization (a process called effacement) results in nephrotic syndrome (2). Several similarities have emerged between protein networks underlying the development and maintenance of podocyte foot processes and those governing the development and function of neuronal synapses (3, 4).

A study based on cathepsin-dependent proteolysis or expression of a dominant-negative dynamin mutant in podocytes provided evidence supporting a role of dynamin in foot process maintenance (5). Dynamin, which is encoded by 3 genes in mammals (dynamin 1 [*Dnm1*] and *Dnm3*, which are expressed primarily in brain, and *Dnm2*, which is ubiquitously expressed), is a GTPase that mediates the fission reaction of clathrin-mediated endocytosis (6, 7). At neuronal synapses, where dynamin is represented primarily by *Dnm1*, this protein plays a critical role in synaptic vesicle endocytosis and recycling (8–10). Dynamin has also been implicated in actin dynamics because of its colocalization with Arp2/3-dependent actin structures and its direct interaction with actin and with actin-regulatory proteins (11–13). Genetic studies in mice and humans have demonstrated the critical dependence of podocyte

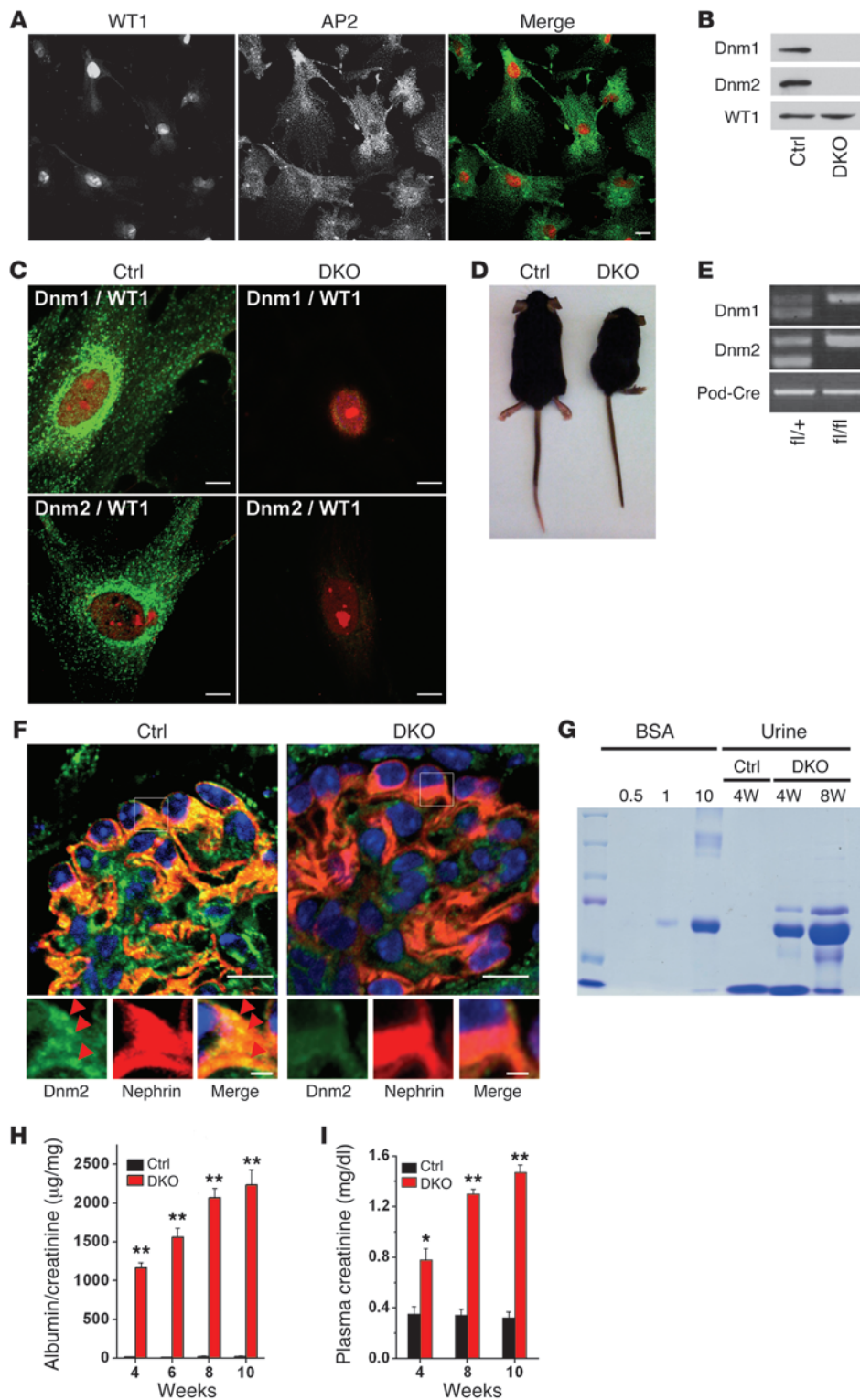
foot process integrity, and thus of kidney filtration barrier function, on actin-regulatory proteins and upstream signaling factors, consistent with actin being a core component for normal podocyte architecture (14). Thus, it has been proposed that the function of dynamin in podocytes may be mediated by its role in the actin cytoskeleton (5, 11). However, clathrin-coated pits and vesicles are frequently observed in podocyte foot processes (15), raising the possibility that endocytic defects may contribute to foot process abnormalities upon disruption of dynamin's function.

In this study, we first addressed the role of dynamin in the stability of podocyte foot processes using a conditional gene KO strategy, and demonstrated that podocyte-selective KO of *Dnm1* and *Dnm2* resulted in severe proteinuria, renal failure, and foot process effacement. Next, we investigated whether 2 major functional partners of dynamin at neuronal synapses, synaptojanin (16–18) and endophilin (19, 20), are also implicated in the formation and maintenance of the kidney glomerular filtration barrier. Synaptojanin, an inositol 5-phosphatase whose preferred substrates are phosphatidylinositol 4,5-bisphosphate (PIP2) and phosphatidylinositol 3,4,5-trisphosphate (PIP3), plays a critical role in clathrin-coated dynamics and in the uncoating of newly formed clathrin-coated vesicles (17). It is also a negative regulator of actin nucleation and signaling (21, 22). Endophilin is an adaptor that binds the membrane bilayer via its N-terminal region (which contains a BAR domain and amphipathic helices) and both dynamin and synaptojanin via its C-terminal SH3 domain, thus helping coordinate the function of these proteins in endocytosis (19, 20, 23). Analysis of mice harboring germline KO mutations of these proteins showed that while the lack of synaptojanin 1 (*Synj1*) or of all 3 endophilins

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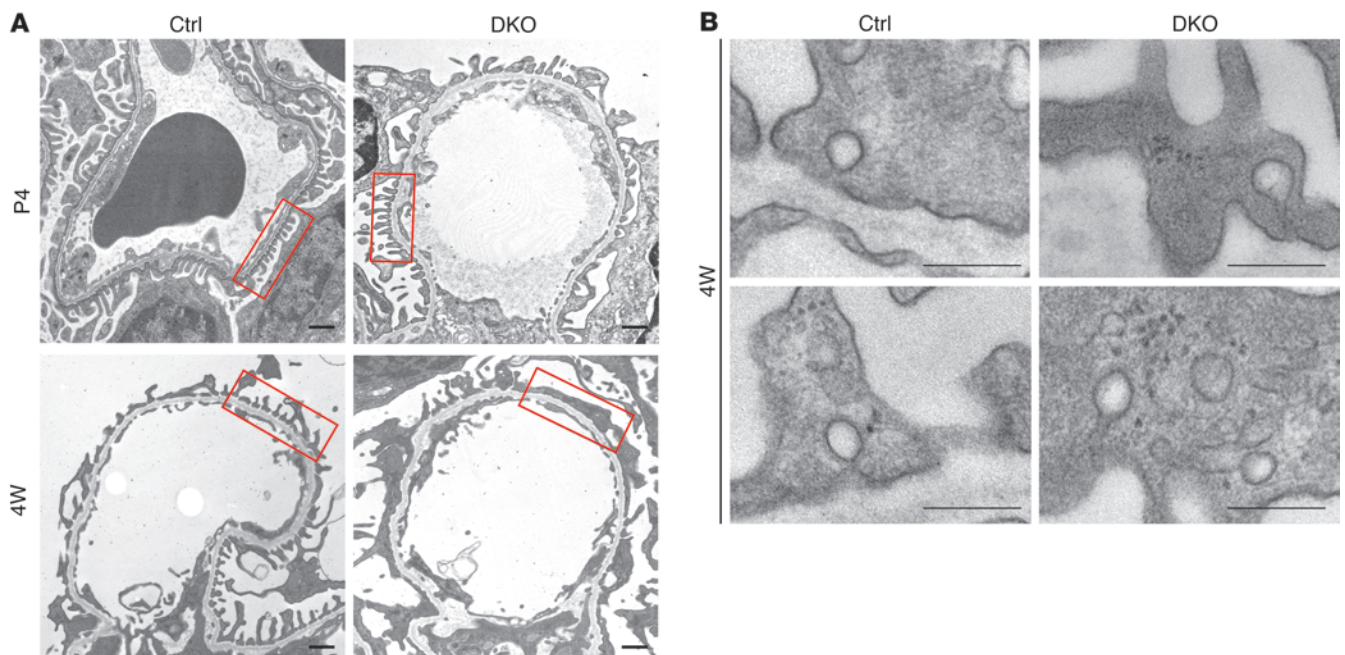
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**Figure 1** Loss of Dnm1 and Dnm2 in podocytes results in proteinuria and renal failure. **(A)** Cultured purified podocytes immunostained for the podocyte-specific nuclear marker WT1 and the endocytic clathrin adaptor AP2. Scale bar: 30 µm. **(B and C)** Expression of both Dnm1 and Dnm2 in purified control podocytes, identified by WT1 staining, and loss of both proteins in pod-*Dnm*-DKO podocytes (age P21), as detected by Western blotting and immunofluorescence. Scale bars: 10 µm. **(D)** Control and pod-*Dnm*-DKO mice at 10 weeks of age. **(E)** *Dnm1*, *Dnm2*, and *Podocin* genotypes, as confirmed by tail genotyping in pod-*Dnm*-DKO mice (age P10). **(F)** Double immunofluorescence detection of nephrin (red) and Dnm2 (green) on kidney sections of the indicated genotypes (age P21). Scale bars: 10 µm (top); 2 µm (bottom). **(G)** SDS-PAGE (Coomassie Blue staining) of 0.5, 1, or 10 µg BSA standard and of urine from pod-*Dnm*-DKO mice, which demonstrated albuminuria at 4 and 8 weeks. 2 µl urine was loaded per lane. **(H)** Quantification of urinary albumin normalized to creatinine at 4, 6, 8, and 10 weeks (*n* = 6). **(I)** Elevated plasma creatinine in pod-*Dnm*-DKO mice at 4, 8, and 10 weeks of age (*n* = 6). \**P* < 0.005, \*\**P* < 0.001 vs. control.

did not produce macroscopic defects in embryonic development, the glomerular filtration barrier was not properly developed at birth in either genotype, resulting in massive proteinuria in newborn mice. We further determined that, as at neuronal synapses, all these proteins functioned at sites of clathrin-mediated endocytosis in podocytes. Interestingly, dynamin, synaptojanin, and

endophilin bound either directly or indirectly to proteins that genetic studies have implicated in human nephrotic syndrome. Our results indicate that a protein network that operates at the interface between endocytosis and actin at neuronal synapses is additionally critical in the formation and maintenance of the kidney glomerular filtration barrier.

**Figure 2**

Podocyte foot process effacement in pod-*Dnm*-DKO mice. **(A)** Electron micrographs illustrating the ultrastructure of glomerular capillaries of control and pod-*Dnm*-DKO mice. In pod-*Dnm*-DKO capillaries, podocyte foot processes had a nearly normal morphology at P4, but were effaced at 4 weeks. Scale bars: 500 nm. Boxed regions illustrate effaced pod-*Dnm*-DKO podocyte foot processes at 4 weeks compared with control. **(B)** High-magnification micrographs from control and mutant glomeruli showing endocytic clathrin-coated pits at 4 weeks of age. Scale bars: 500 nm.

## Results

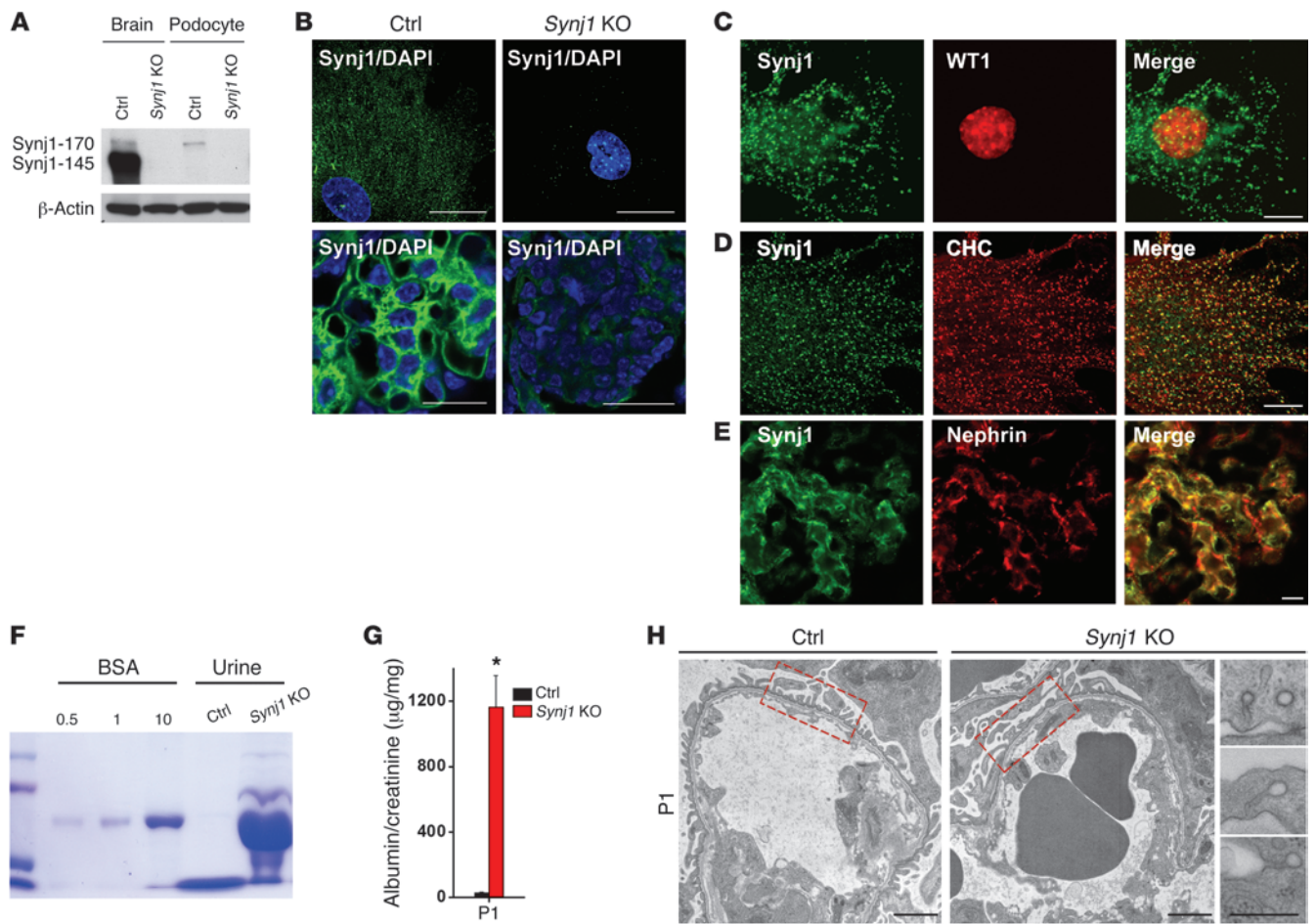
*Loss of Dnm1 and Dnm2 in podocytes results in proteinuria and kidney failure.* We first revisited the role of dynamin in the maintenance of the kidney glomerular filtration barrier using a KO approach. Podocytes were expected to selectively express Dnm2, the house-keeping and ubiquitous dynamin isoform, whose constitutive KO results in embryonic lethality (6). Surprisingly, however, immunoprecipitated podocytes (24), as validated by their reactivity for the podocyte-specific marker WT1 (Figure 1A), were found to express not only Dnm2, but also Dnm1 (Figure 1, B and C), the predominantly neuronal dynamin (9). Thus, we used previously described *Dnm1*<sup>f/f</sup>; *Dnm2*<sup>f/f</sup> mice (6) and mice expressing Cre recombinase under the control of the podocin promoter (Cre expression occurs at E14; ref. 25) to generate podocyte-specific *Dnm1*; *Dnm2* double-KO (referred to herein as pod-*Dnm*-DKO) mice. Such mice appeared normal at birth, but their weight gain fell below that of *Dnm1*<sup>f/f</sup>; *Dnm2*<sup>f/f</sup> control mice in the following days (Supplemental Figure 1B; supplemental material available online with this article; doi:10.1172/JCI65289DS1). By 10 weeks of age, they were severely lethargic and much smaller than controls (Figure 1D). More than 75% of pod-*Dnm*-DKO mice died between 10 and 12 weeks of age.

PCR-based tail genotyping confirmed the presence of the conditional dynamin alleles as well as the podocin Cre transgene (Figure 1E), and Western blot analysis of podocytes purified from pod-*Dnm*-DKO mice revealed that Dnm1 and Dnm2 levels were already somewhat reduced at E16.5 relative to control littermates (Figure 1, B and C, and Supplemental Figure 1A). Gene ablation of Dnm1 and Dnm2 expression was further confirmed by immunofluorescence of purified pod-*Dnm*-DKO podocytes extracted from P21 mice (Figure 1C) and of P21 pod-*Dnm*-DKO

kidney sections counterstained with the podocyte-specific protein nephrin (Figure 1F). Importantly, robust albuminuria was detected in the urine of pod-*Dnm*-DKO mice, as assessed by SDS-PAGE at 4 and 8 weeks (Figure 1G) and quantified by ELISA with normalization to urine creatinine at several ages (Figure 1H and Supplemental Figure 2A). Moreover, measurement of plasma creatinine levels in pod-*Dnm*-DKO mice demonstrated severe kidney failure (Figure 1I and Supplemental Figure 2B).

The kidney/body weight ratio in pod-*Dnm*-DKO mice was significantly reduced at 8 weeks of age (50% reduction; Supplemental Figure 1C), and the kidneys displayed a pale, smaller, and corrugated appearance (Supplemental Figure 1D). While the kidneys were normal at birth, mesangial expansion and evidence of focal segmental glomerulosclerosis was observed by 4 weeks of age (Supplemental Figure 1E). By 10 weeks of age, the glomeruli had undergone a process of global sclerosis, and the kidney further demonstrated severe interstitial fibrosis and tubular dilatation with proteinaceous casts (Supplemental Figure 1F). This time course, with normal development but then loss of foot processes, is consistent with induced rather than constitutive loss of dynamin and with the use of a transgenic mouse in which Cre expression requires podocyte differentiation.

Correspondingly, the glomeruli of kidneys from neonatal pod-*Dnm*-DKO mice appeared relatively normal by electron microscopy, although heterogeneity was observed, with the occurrence of some effaced capillary loops (Figure 2A and Supplemental Figure 3A). However, at 4 weeks of age, kidneys from pod-*Dnm*-DKO mice revealed generalized podocyte foot process effacement (Figure 2, A and B), which became even more severe at 8 weeks (data not shown). Additionally, clathrin-coated pit intermediates were fre-

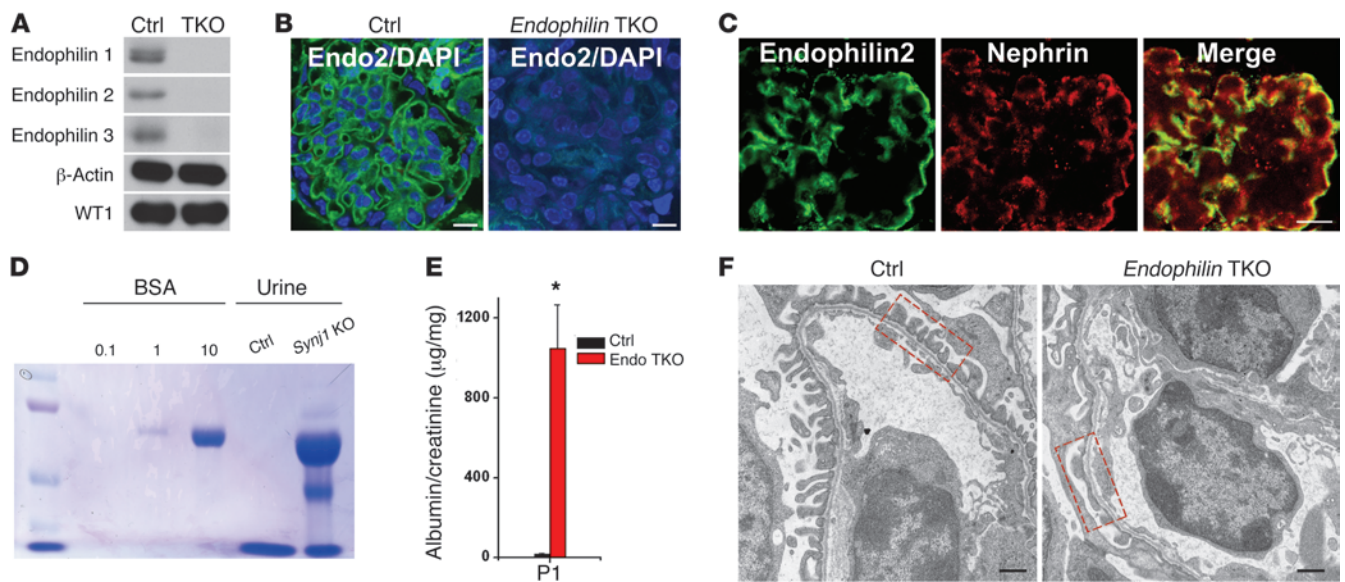


**Figure 3** Lack of Synj1 results in severe proteinuria and foot process effacement. (A) Western blots of total homogenates of brain and purified podocytes of neonatal control and *Synj1* KO mice, demonstrating that podocytes selectively expressed Synj1-170, not Synj1-145.  $\beta$ -Actin served as a loading control. (B) Synj1 immunofluorescence staining of isolated primary podocytes and kidney sections of control and *Synj1* KO mice. Scale bars: 10  $\mu$ m (top); 30  $\mu$ m (bottom). (C) Immunofluorescence for Synj1 and for the podocyte nuclear marker WT1 on an isolated podocyte. Scale bar: 20  $\mu$ m. (D) Colocalization of Synj1 and clathrin heavy chain (CHC) in cultured podocytes. Scale bar: 20  $\mu$ m. (E) Colocalization of Synj1 and nephrin in the glomerulus. Scale bar: 20  $\mu$ m. (F) SDS-PAGE (Coomassie Blue stain) of 0.5, 1, or 10  $\mu$ g BSA standard and of urine (2  $\mu$ l) from control and *Synj1* KO neonatal mice. (G) Quantification of urinary albumin normalized to creatinine ( $n = 12$ ). \* $P < 0.001$  vs. control. (H) Electron micrographs from glomeruli of control and *Synj1* KO neonatal mice revealing foot process effacement and clathrin-coated intermediates in *Synj1* KO mice. Boxed regions illustrate effaced podocytes in *Synj1* KO mice compared with control. Scale bars: 500 nm.

quently observed in such processes (Figure 2B), consistent with the occurrence of clathrin-mediated endocytosis in wild-type podocyte foot processes (Figure 2B, Supplemental Figure 3B, and ref. 15).

*Proteinuria and podocyte foot process effacement in Synj1 KO mice.* The severe effect on glomerular filtration produced by the lack of dynamin selectively in podocytes could in principle be attributed to a critical role of dynamin for the function of any cell in which it is expressed, although cells remain viable in the absence of dynamin (6). Thus, we examined the effect on kidney physiology of Synj1, which functions in close partnership with dynamin at synapses (17, 18, 26), but does not have critical housekeeping functions. *Synj1* KO mice appear grossly normal at birth, although they are neurologically impaired and die within the first few postnatal days (17). Synj1 is a phosphoinositide phosphatase that dephosphorylates PIP2 and PIP3, 2 phosphoinositides that help recruit endocytic clathrin coats to the plasma membrane and whose dephosphorylation is needed

for clathrin uncoating after fission (18, 22). The *Synj1* gene encodes 2 major splice variants. The 170-kDa splice variant Synj1-170, which has a broad tissue distribution (27), contains C-terminal binding sites for clathrin as well as other endocytic adaptors, is recruited early to endocytic-coated pits, and remains associated with them throughout their growth (28). The 145-kDa synaptic variant Synj1-145, which is selectively expressed in the developed nervous tissue and is present at very high concentrations at neuronal synapses, is recruited only to late-stage endocytic clathrin-coated pits (28). Western blotting of isolated podocytes revealed the presence of Synj1, specifically the Synj1-170 isoform (Figure 3A). Correspondingly, Synj1 immunofluorescence demonstrated the punctate pattern of immunoreactivity expected for the clathrin-coated pit localization of this isoform, whereas such reactivity was lost in the podocytes and kidneys of *Synj1* KO mice (Figure 3, B–D). Moreover, Synj1 colocalized with nephrin in podocytes within the glomerulus (Figure 3E).

**Figure 4**

Lack of endophilin results in severe proteinuria and foot process effacement. (A) Western blots of purified podocytes of neonatal control and *endophilin* TKO mice (P0) demonstrating loss of endophilin 1, 2, and 3 in *endophilin* TKO mice. WT1 staining was used as a loading control. (B) Endophilin 2 immunofluorescence staining of glomeruli within kidney sections of control and *endophilin* TKO mice. Scale bar: 10  $\mu\text{m}$ . (C) Colocalization of endophilin 2 with nephrin in the glomerulus. Scale bar: 10  $\mu\text{m}$ . (D) SDS-PAGE (Coomassie Blue staining) of 0.5, 1, or 10  $\mu\text{g}$  BSA standard and of urine (2  $\mu\text{l}$ ) from control and *endophilin* TKO neonatal mice. (E) Quantification of urinary albumin normalized to creatinine ( $n = 7$ ).  $*P < 0.001$  vs. control. (F) Electron micrographs from glomeruli of control and *endophilin* TKO neonatal mice revealing foot process effacement. Boxed regions illustrate effaced podocytes in *endophilin* TKO mice compared with control. Scale bars: 500 nm.

We next analyzed the effect of the lack of *Synj1* on kidney function in *Synj1* KO mice. Histological examination of neonatal kidneys revealed that some glomeruli appeared to have mesangial matrix accumulation (Supplemental Figure 4A). Dilated tubules with proteinaceous casts were also present (Supplemental Figure 4B), suggestive of severe proteinuria. Moreover, PIP2 levels, as detected by an HPLC-based method, were 25% higher in kidneys of newborn KO mice relative to kidneys of control littermates (Supplemental Figure 4C), demonstrating an important effect of *Synj1* on PIP2 metabolism in this tissue.

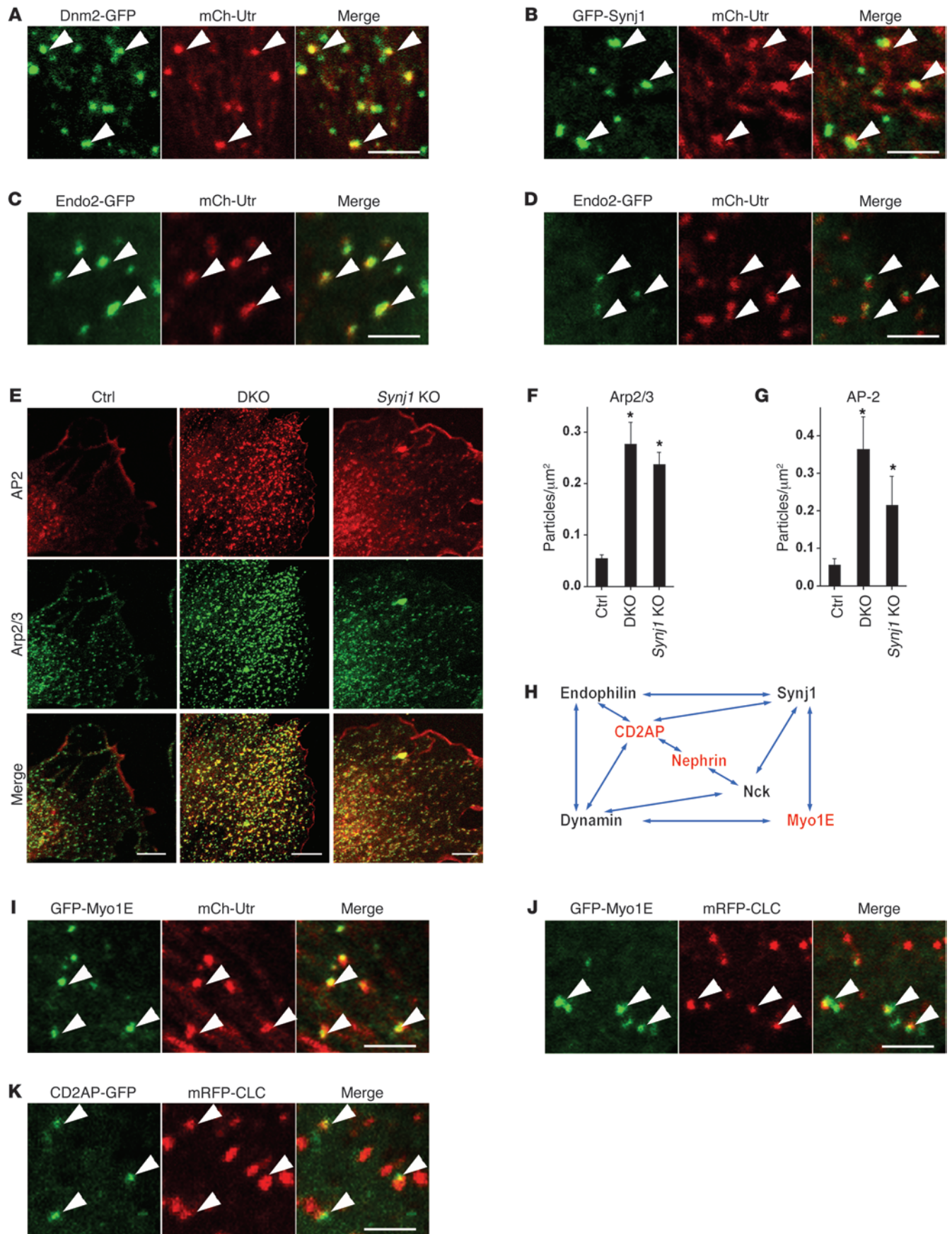
Importantly, urine analysis of P1–P4 *Synj1* KO mice revealed prominent albuminuria (Figure 3F) that was confirmed by normalization to urine creatinine (Figure 3G and Supplemental Figure 2C), consistent with disruption of the glomerular filtration barrier. Accordingly, electron microscopy of kidneys of P1–P4 *Synj1* KO mice revealed massive foot process effacement (Figure 3H and Supplemental Figure 3A). As in pod-*Dnm*-DKO mice, the podocytes of *Synj1* KO mice also exhibited numerous clathrin-coated endocytic intermediates. Significant foot process effacement was already observed in E16.5 embryos (Supplemental Figure 4D), which indicates that *Synj1* is involved not only in the maintenance, but also in the development, of the filtration barrier.

*Proteinuria and podocyte foot process effacement in endophilin triple-KO (TKO) mice.* In the nervous system, the effect of the lack of *Synj1* (primarily clathrin uncoating) is strikingly phenocopied by the lack of endophilin, which indicates that these 2 proteins work closely together (20, 29–31). Endophilin is the collective name for 3 very similar proteins, endophilin 1, 2, and 3 (encoded by *Sh3gl2*, *Sh3gl1*, and *Sh3gl3*, respectively). Endophilin is recruited to the necks of endocytic clathrin-coated pits by its curvature sensitive

BAR domain (32, 33) and binds both dynamin and synaptojanin (with 10-fold higher affinity for synaptojanin than for dynamin; refs. 19, 23) via its C-terminal SH3 domain, thus helping recruit these proteins to their sites of action. Thus, it was of interest to determine whether the lack of endophilin phenocopies the lack of *Synj1* in the organization of the glomerular filtration barrier.

Like *Synj1* KO mice, *endophilin* TKO mice (with triple-KO of *Sh3gl2*, *Sh3gl1*, and *Sh3gl3*) have grossly normal embryonic development, which argues against essential housekeeping functions of endophilin. Like *Synj1* KO mice, they die perinatally, at least in part because of impaired neuronal function. Western blot analysis of isolated podocytes demonstrated expression of all 3 endophilins that was lost in the *endophilin* TKO, as expected (Figure 4A). Immunofluorescence staining of kidney sections from wild-type newborn mice revealed endophilin 2 expression in control, but not *endophilin* TKO, podocytes, which were identified by the presence of immunoreactivity for nephrin (Figure 4, B and C). As observed in *Synj1* KO mice, histological examination of the *endophilin* TKO kidney cortex showed abnormalities in the glomeruli and dilated tubules with proteinaceous casts (Supplemental Figure 5, A and B). Urine obtained from these mice revealed robust proteinuria, as assessed by Coomassie Blue staining and further validated by normalization to urine creatinine (Figure 4, D and E, and Supplemental Figure 2D). Importantly, severe foot process effacement with a thickened glomerular basement membrane was observed by electron microscopy at birth (Figure 4F and Supplemental Figure 3A), as in *Synj1* KO mice.

*Clathrin-mediated endocytosis and actin in podocytes.* Collectively, the findings described here suggest that the functional partnership among dynamin, synaptojanin, and endophilin at endocytic sites,





### Figure 5

Defects in actin regulation and endocytosis in pod-*Dnm*-DKO, *Synj1* KO, and *endophilin* TKO podocytes. (A–D and I–K) High-magnification images acquired by spinning disk confocal microscopy of podocytes transfected with the indicated pairs of fluorescent fusion proteins. Arrowheads highlight examples of colocalization. Scale bars: 2.5  $\mu\text{m}$ . (E) Double immunofluorescence labeling demonstrating the increased number of endocytic clathrin-coated intermediates AP2 (alpha-adaptin subunit), and the accumulation of Arp2/3 (p34 subunit) at such intermediates, in pod-*Dnm*-DKO and *Synj1* KO podocytes. Scale bars: 10  $\mu\text{m}$ . (F and G) Quantification of the abundance of Arp2/3 and AP2 puncta ( $n = 5$  experiments). \* $P < 0.05$  vs. control. (H) Network of proteins found at endocytic clathrin-coated pits whose loss or functional impairment results in nephrotic syndrome in mice and/or humans. Proteins indicated in red were linked to genetic forms of nephrotic syndrome and focal segmental glomerulosclerosis in humans. Direct interactions are indicated by arrows.

which in neurons supports neurotransmission, is used in podocytes to support formation and maintenance of the glomerular filtration barrier. Akin to what has been proposed for dynamin (5, 11), the action of synaptojanin at podocyte foot processes may be due primarily to an effect on actin, because the substrates of the phosphatase activity of *Synj1* (namely, PIP2 and PIP3) are critical and potent regulators of actin nucleation (22, 34). Likewise, endophilin could have a role in actin function via its property to recruit dynamin, synaptojanin (19, 23), and actin-regulatory proteins (6, 35). However, it is quite possible that the effects produced by perturbing dynamin, synaptojanin, and endophilin function in podocytes may be mediated primarily by the endocytic roles of these proteins. An even more attractive possibility is that the 2 actions may be tightly interrelated, because actin has been strongly implicated in dynamics of late-stage clathrin-coated pits in a variety of model cell types (6, 36, 37).

The localization and dynamics of fluorescently tagged *Dnm2* (*Dnm2*-GFP), *Synj1*-170 (GFP-*Synj1*), endophilin 2 (*Endo2*-GFP), and actin in cultured podocytes were examined by spinning disk confocal microscopy. As previously shown in fibroblasts (28, 37, 38), dynamin selectively colocalized with clathrin at late-stage clathrin-coated pits, just prior to the disappearance of the clathrin signal due to membrane fission and movement of the vesicle away from the plane of imaging concomitant with uncoating (Supplemental Figure 6, A and B, Figure 5C, and Supplemental Video 1). Likewise, as in fibroblasts (28), *Synj1*-170 was observed at all clathrin-coated pits regardless of maturation state (Supplemental Figure 6, D–F, and Supplemental Video 2), whereas endophilin appeared along with dynamin just before fission (*Synj1*-170 is not dependent on endophilin for its recruitment, but contains the endophilin binding sites, consistent with a functional interaction of the 2 proteins at late-stage pits; refs. 28, 39).

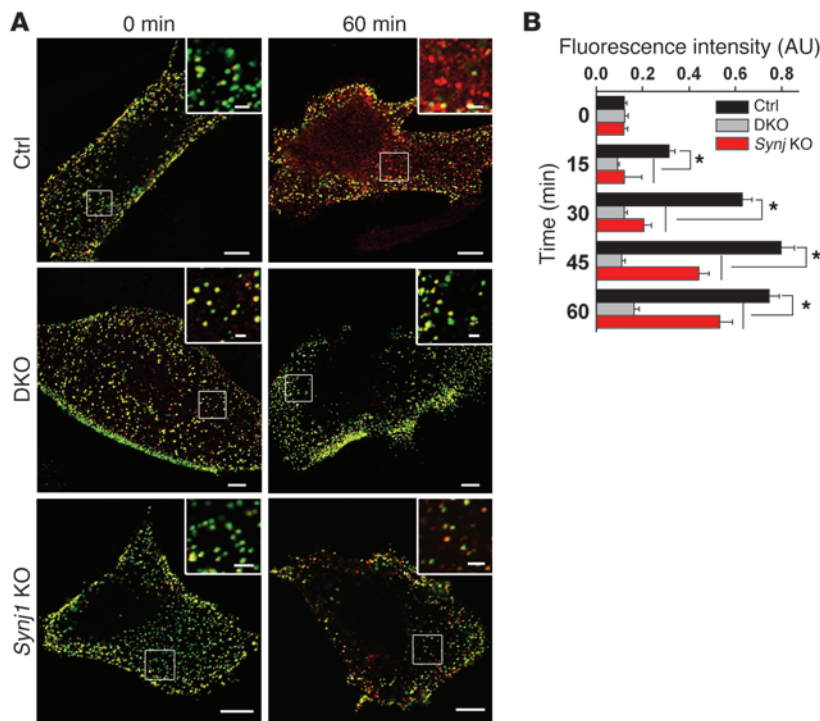
As shown in Figure 5A and Supplemental Figure 7A, robust colocalization was observed between *Dnm2*-GFP-positive spots and spots positive for the mCherry-tagged calponin homology domain of utrophin (mCh-Utr; a probe for F-actin in living cells; ref. 40), demonstrating a transient accumulation of actin, in parallel with dynamin, at the neck of endocytic clathrin pits prior to their fission, as described previously (37). mCh-Utr spots also overlapped with a subset of GFP-*Synj1* spots (Figure 5B), representative of late-stage pits (Supplemental Figure 7B), and completely overlapped with *Endo2*-GFP spots, which also colocalized with mRFP-tagged clathrin (mRFP-CLC; Figure 5, C and D, and Supplemental

Figure 7, C and D). Thus, while the obligatory recruitment of actin at late-stage endocytic clathrin-coated pits and a cooperative role of dynamin and actin in fission is a matter of debate (38, 41, 42), a link between actin and clathrin-mediated endocytosis clearly existed in podocytes (Supplemental Video 3). Importantly, *Dnm2*-GFP, GFP-*Synj1*, and *Endo2*-GFP colocalized with F-actin at clathrin-coated pits, not on actin stress fibers (Figure 5, A–C), emphasizing the link between these 3 endocytic proteins and the endocytic function of actin.

The relationship between endocytic clathrin-coated pits and actin in podocytes was further demonstrated by the massive accumulation of Arp2/3 at arrested clathrin-coated pits of cultured podocytes from pod-*Dnm*-DKO mice (Figure 5, E–G) that were also positive for endophilin (Supplemental Figure 6G), as shown previously in fibroblasts (6). A less pronounced but significant accumulation of Arp2/3 was observed at endocytic clathrin spots of *Synj1* KO podocytes (Figure 5, E–G), possibly reflecting a consequence of elevated PIP2 levels at these sites, as previously shown after perturbation of *Synj1* function at synapses (21).

*Dynamin, synaptojanin, and endophilin interactors play critical functions in podocyte foot processes.* In light of the effect that the lack of dynamin, synaptojanin, and/or endophilin has on the glomerular filtration barrier, it is quite striking that other proteins that interacted with dynamin, synaptojanin, and endophilin (Figure 5H), when lost, also result in proteinuria (43–45), although their function at foot processes was not attributed to an endocytic function. One such protein is myosin 1E (*Myo1E*), a monomeric myosin that binds both dynamin and synaptojanin via a COOH-terminal SH3 domain (45), and whose absence in mice and mutations in humans result in nephrotic syndrome (44–46). When *Myo1E* was expressed along with either mRFP-CLC or mCh-Utr in podocytes, it accumulated at late-stage clathrin-coated pits, where it colocalized with the pool of F-actin present at these sites (Figure 5, I and J, Supplemental Figure 7, E and F, and Supplemental Video 4). CD2AP, whose functional impairment also results in nephrotic syndrome in both mice and humans (43, 47), is a major binding partner of endophilin (48, 49) and interacts (like its homolog CIN85) with dynamin and synaptojanin (50, 51) both directly and indirectly through endophilin. Accordingly, GFP-tagged CD2AP (CD2AP-GFP), when expressed in podocytes, was localized at late-stage clathrin-coated pits along with endophilin (Figure 5K and Supplemental Figure 7G). Nephrotic syndrome is also produced by loss of *Nck*, an endocytic adaptor that binds to both dynamin and synaptojanin (52, 53). Finally, it has been recently shown that the mouse KO of *PI3KC2 $\alpha$* , the gene encoding a class II PI3K that binds clathrin and participates in clathrin-mediated endocytosis (28), develops glomerulonephropathy with proteinuria and renal failure (54).

*Endocytic defect in pod-*Dnm*-DKO and *Synj1* KO podocytes.* To determine whether the protein network discussed above plays a role in mediating the internalization of key podocyte cargo proteins, we performed internalization assays in isolated podocytes of 2 mutant genotypes, *Dnm1*;*Dnm2* DKO and *Synj1* KO. We focused on nephrin as the cargo protein because endocytosis was previously shown to play a role in nephrin regulation (55). Nephrin is an integral membrane protein that participates in the formation of the slit diaphragm that connects adjacent foot processes (56). Podocytes were transfected with a protein chimera comprising the transmembrane and cytoplasmic domain of nephrin and the



**Figure 6** Nephrin endocytosis is reduced in pod-*Dnm*-DKO and *Synj1* KO podocytes. **(A)** Internalized (red) versus cell surface (green) CD8-nephrin chimera in control, pod-*Dnm*-DKO, and *Synj1* KO podocytes. Scale bars: 10  $\mu$ m; 2  $\mu$ m (insets). **(B)** Quantification of data in **A** ( $n = 3$ ; 20 cells evaluated per experiment). \* $P < 0.01$ , pod-*Dnm*-DKO and *Synj1* KO vs. control.

extracellular domain of CD8. Using an anti-CD8 antibody-based internalization assay, we found that the internalization of this probe was significantly delayed in pod-*Dnm*-DKO and *Synj1* KO podocytes (Figure 6, A and B).

**Discussion**

Our present findings support an important role of dynamin in the maintenance of the permeability barrier of the kidney (5). We further demonstrated that synaptojanin and endophilin, which are functional partners of dynamin in synaptic vesicle endocytosis at neuronal synapses, were critically implicated in the development of the permeability barrier of the kidney. Most likely, dynamin is also essential for the development of this barrier, but the early embryonic lethality of *Dnm2* KO mice did not allow us to explore this possibility. In the genetic model used by us, the expression of *Dnm1* and *Dnm2* was suppressed after podocyte foot processes have already developed. In contrast, the *Synj1* KO and *endophilin* TKO mice used for our investigations were constitutive KOs. Interestingly, these mice, although greatly neurologically impaired, were macroscopically normal at birth, which indicates that these 2 proteins are critically required only for the development of selected cell types.

Dynamin, synaptojanin, and endophilin have well-established endocytic functions. Endocytosis may help control the steady-state level and turnover of cell surface molecules, which, via actions on signaling pathways, may affect foot process structure. An effect mediated by defective endocytosis, however, does

not rule out additional direct effects on actin. All these proteins have been linked to actin function and endocytosis, which are tightly interconnected with actin (Figure 7 and refs. 6, 13, 37, 38, 42).

Interestingly, even at synapses, disruption of the function of dynamin, synaptojanin, and endophilin, as well as that of other endocytic proteins, was shown to not only impair endocytosis, but also affect the architecture of nerve terminals, which prompted Dickman et al. to suggest that “endocytosis plays a critical role in sculpting the structure of synapses, perhaps through the endocytosis of unknown regulatory signals that organize morphogenesis at synaptic terminals” (57). In conclusion, our study provides a striking demonstration of how a similar and evolutionary conserved endocytosis machinery, functionally linked to the actin cytoskeleton, has been used in different cellular contexts to control highly specialized physiological process, such as synaptic vesicle recycling and the formation of the glomerular filtration barrier.

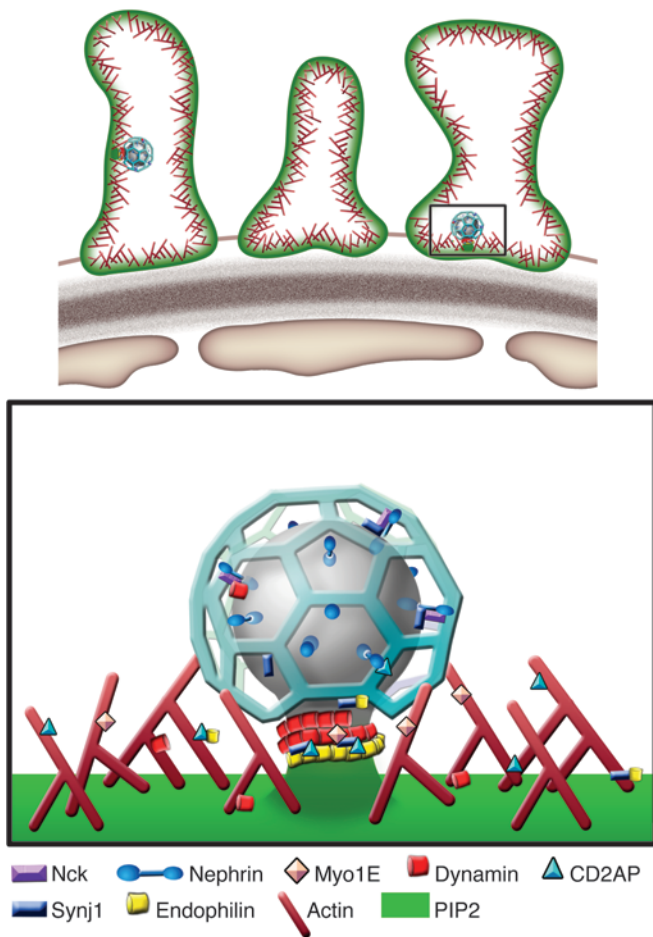
**Methods**

**Antibodies and plasmids.** Mouse anti-human CD8 (BD Biosciences — Pharmingen); rabbit anti-WT1, mouse anti-clathrin heavy chain, and rabbit anti-endophilin 3 (Santa Cruz); rabbit anti-p34 Arc/ARPC (Millipore); mouse anti-actin (Sungene Biotech); guinea pig anti-nephtrin (Fitzgerald); rabbit anti-Dnm1 (Epitomics); mouse anti- $\alpha$ -adaptin (AP6) (Abcam); Alexa Fluor 488 goat anti-rabbit IgG (Molecular Probes); and Alexa Fluor 594 goat anti-mouse IgG (Molecular Probes) antibodies were obtained commercially. Rabbit anti-Dnm2, rabbit anti-Synj1, rabbit anti-endophilin 1, and rabbit anti-endophilin 2 antibodies were previously described (6, 17). Rabbit polyclonal anti-nephtrin antibody was provided by Y. Harita (University of Tokyo, Tokyo, Japan; ref. 58). Mammalian expression vectors encoding GFP-Flag-human Synj1-170, mRFP-CLC, and Endo2-GFP were previously reported (28). Dnm2-GFP was provided by M. McNiven (Mayo Clinic, Rochester, Minnesota, USA; ref. 59). mCh-Utr was provided by W. Bement (University of Wisconsin, Madison, Wisconsin, USA; ref. 40). CD8-rat nephtrin-Flag construct was provided by Y. Harita (University of Tokyo, Tokyo, Japan; ref. 58). GFP-Myo1E was a gift from M. Mooseker (Yale University, New Haven, Connecticut, USA; ref. 60). CD2AP-GFP was a gift from A. Shaw (Washington University, St. Louis, Missouri, USA; ref. 61).

**Generation of mice.** *Dnm1* <sup>$\beta/\beta$</sup>  and *Dnm2* <sup>$\beta/\beta$</sup>  mice (6) were mated with pod-Cre mice, obtained as previously described (25), to generate the pod-*Dnm*-DKO mice with selective glomerular podocyte-specific KO of *Dnm1* and *Dnm2*. Constitutive *Synj1* KO (17) and *endophilin* TKO mice (20) were previously described. Genotyping was performed by PCR using previously described protocols (6, 17, 25).

**Biochemical measurements: plasma creatinine, urine albumin, and urine creatinine.** Urine samples were collected from P1–P4 *Synj1* KO, P0 *endophilin* TKO, and littermate control mice and from pod-*Dnm*-DKO and littermate control mice at various ages. Albuminuria was qualitatively assessed by SDS-PAGE followed by Coomassie Blue staining. Quantitatively, urine albumin was measured using Albumin ELISA Quantitation kit, according to the manufacturer’s protocol, in duplicates for





**Figure 7**

Synaptojanin, dynamin, and endophilin within podocyte foot processes. All the actin-associated proteins shown have interconnected roles in endocytosis, actin regulation, and signaling, and their genetic disruption results in nephrotic syndrome.

each sample (Bethyl Laboratories Inc.). Urine and plasma creatinine were measured in duplicate for each sample by the core facility of the George O'Brien Kidney Center at Yale University.

**Cell culture.** For *Synj1* KO, *endophilin* TKO, and littermate control mice, whole kidneys were isolated, minced with a razor, and digested with collagenase (5 mg/ml) containing DNase (0.2 mg/ml) for 1 hour at 37°C. The digested kidneys were filtered through a 100- $\mu$ m (Falcon) cell strainer, plated on type 1 collagen-coated dishes, and cultured at 37°C in RPMI 1640 medium supplemented with 10% FBS, 100 U/ml penicillin/streptomycin, 10 mM HEPES, 1 mM sodium bicarbonate, and 1 mM sodium pyruvate. Subculture of primary podocytes was performed from the isolated kidney cells described above by trypsinization with 0.25% trypsin/EDTA (Invitrogen) at culture day 3, followed by sieving through a 40- $\mu$ m cell strainer (Falcon) to isolate an enriched podocyte population as previously described (62). For pod-*Dnm*-DKO mice, glomeruli were isolated by Dynabeads (Invitrogen) perfusion followed by podocyte isolation as previously described (24). Transfections were performed by electroporation using the Amaxa nucleofactor protocol for podocytes (Lonza).

**Immunoblotting.** Cells or kidney homogenates were lysed with ice-cold buffer (for Dnm1, Dnm2, or *Synj1*, 20 mM Tris [pH 7.5], 150 mM NaCl, 1% Nonidet P-40, 5 mM EDTA, 1 mM  $\text{Na}_3\text{VO}_4$ , and 50 mM NaF; or 10 mM

HEPES-OH [pH 7.4], 1 mM EDTA, 0.32 M sucrose, and 1% TritonX-100; for endophilin, 10 mM HEPES, 320 mM sucrose, 1 mM EDTA, and 150 mM NaCl with protease inhibitor cocktail [Roche Diagnostics]), and protein concentrations were quantified with the Bio-Rad Protein Assay (Bio-Rad). Proteins were separated by SDS-PAGE under reducing conditions, transferred to PVDF membrane (Millipore), blocked with 5% milk in TBST, and incubated with appropriate primary antibodies in this buffer. After 3 washes with PBS, membranes were incubated with horseradish peroxidase-conjugated secondary antibodies, and signals were detected by enhanced chemiluminescence reagents (GE).

**Histology, immunohistochemistry, and immunofluorescence staining.** Mice were anesthetized by intraperitoneal injection of ketamine and xylazine followed by perfusion fixation with 4% paraformaldehyde with or without glutaraldehyde through the left ventricle. For histology, sections were sent to Yale Pathology Core Tissue Services for H&E, periodic acid-Schiff, and Masson's trichrome staining as well as electron microscopy.

For immunohistochemistry, kidney cryosections (4  $\mu$ m) were subjected to antigen retrieval (Retrievagen; BD Biosciences), followed by blocking with 3% BSA in PBS for 1 hour. Immunostaining was performed with primary antibodies overnight at 4°C, followed by Alexa Fluor 488- and/or Alexa Fluor 594-conjugated secondary antibodies, washing, and mounting with Vectashield with DAPI mounting medium (Vector Labs). Cultured cells were fixed with 4% paraformaldehyde for 10 minutes at room temperature, washed with PBS, then incubated for 5 minutes in 0.1% Triton X-100 in PBS, followed by blocking with 5% BSA for 30 minutes. Cells were then incubated with primary antibodies for 1 hour at room temperature, followed by the appropriate secondary antibody, and imaged using a Perkin Elmer Ultraview Vox spinning disc confocal microscope with either  $\times 60$  CFI PlanApo VC (NA 1.4) or  $\times 100$  CFI PlanApo VC (NA 1.4) objectives or a Zeiss LSM 710 laser scanning confocal microscope using a  $\times 63$  Plan Apo (NA 1.4) oil immersion objective. We used NIH Image J software to analyze raw images and quantitate particles as previously described (28). For measurement of foot processes/glomerular basement membrane, 4 random glomeruli capillary loops in 3 different animals for each condition were chosen, and glomerular basement membrane length was analyzed by Image J software followed by manually counting the number of foot processes for each loop.

**Nonradioactive phosphatidylinositol assay.** To evaluate PIP2 levels, a nonradioactive phosphatidylinositol assay was used (63). Briefly, 9 pairs of *Synj1* KO and wild-type kidneys were snap-frozen in liquid  $\text{N}_2$ , pulverized, and transferred to a glass tube with 1 ml cold chloroform/MeOH/10 N HCl (20:40:1) containing 2 mM  $\text{AlCl}_3$ . Following homogenization, chloroform and water was added at a 1:2 ratio, the samples were centrifuged, and the lower phase was collected and dried under nitrogen. Phospholipids were deacylated with monomethylamine, resuspended in 0.5 ml water, and extracted twice with an equal volume *n*-butanol/petroleum ether/ethyl formate (20:4:1) to remove fatty acids. The deacylated lipids were recovered by addition of 0.5 ml water, vortexing, centrifugation, and collection of the aqueous phase. The butanol phase was re-extracted with 0.5 ml water. The second aqueous phase was combined with the first, and the pool was dried under vacuum and resuspended in water. The anionic glycerol head groups were then separated via anion exchange using a Dionex ICS-3000 HPLC system.

**Live cell imaging and analysis.** Live cell imaging were performed as described previously (28). Briefly, GFP-, mRFP-, and/or mCherry- tagged proteins were coexpressed in primary podocytes by electroporation (Amaxa nucleofactor kit R). Transfected cells were seeded in glass-bottomed 35-mm dishes (no. 1.5 thickness; MatTek) and imaged 24 hours later. Before imaging, medium was replaced with an imaging buffer (containing 136 mM NaCl, 2.5 mM KCl, 2 mM  $\text{CaCl}_2$ , 1.3 mM  $\text{MgCl}_2$ , and 10 mM HEPES [pH 7.4]). Cells were imaged using a Perkin Elmer spinning confocal microscope



(Perkin Elmer) as previously described (6). NIH Image J software was used to analyze images, to generate intensity plots of areas of interest, and to quantitate particles. We calculated the lifetimes of fluorescently tagged proteins by manual assessment of fluorescent spots from images acquired during 3 separate experiments (29). Colocalization between 2 proteins was determined by randomly selecting 20 spots in the green channel, followed by manually scoring for colocalization in the red channel. All data were analyzed for statistical significance using Student's *t* test.

**CD8-nephrin chimera endocytosis assay.** For internalization studies, the cells expressing CD8-rat nephrin-Flag were washed with cold PBS, and labeled for 1 hour at 4°C with mouse anti-CD8 antibody in serum free RPMI. Unbound antibody was removed by 3 washes with cold PBS prior to incubating the cells at 37°C in prewarmed complete RPMI. After internalization of bound label for varying time intervals, the cells were washed with PBS, and surface-retained mouse anti-CD8 antibody was labeled at 4°C for 40 minutes with Alexa Fluor 488-conjugated anti-mouse antibody in serum-free RPMI medium, followed by extensive washing with cold PBS and fixation with 4% paraformaldehyde. The cells were then permeabilized with 0.1% Triton-X 100 and subsequently blocked with 5% BSA in PBS, and mouse anti-CD8 antibody was detected with Alexa Fluor 594 anti-mouse antibody. The cells were washed 3 times with PBS, followed by mounting with Vectashield (Vector Laboratories). Images were acquired with a Zeiss LSM 710 laser scanning confocal microscope using a x63 Plan Ao (NA 1.4) oil immersion objective. To quantify CD8-nephrin endocytosis, fluorescence intensities of cell surface-retained nephrin (Alexa Fluor 488 signal originating from the cell surface) and internalized nephrin (Alexa Fluor 594 signal from the cell interior) were measured with NIH Image J software by examining 20 cells per experiment (*n* = 3) from each indicated time point. We defined the endocytosed nephrin by subtracting the Alexa Fluor 488 image from the Alexa Fluor 594 image and normalizing to surface-derived fluorescence intensity ( $[Alexa\ Fluor\ 594 - Alexa\ Fluor\ 488]/Alexa\ Fluor\ 488$ ). Anti-CD8 antibody staining on untransfected cells was evaluated as a negative control and found to be minimal.

**Statistics.** All data are represented as mean ± SEM. In all cases, group differences were assessed by 2-tailed Student's *t* test. A *P* value less than 0.05 was considered significant.

**Study approval.** All procedures were approved by the Yale University IACUC and followed the guidelines of the NIH Guide for the Care and Use of Laboratory Animals.

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