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Application of ^1H DOSY for Facile Measurement of Polymer Molecular Weights

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Abstract

To address the practical issues of polymer molecular weight determination, the first accurate polymer weight-average molecular weight determination method in diverse living/controlled polymerization via DOSY (diffusion-ordered NMR spectroscopy) is reported. Based on the linear correlation between the logarithm of diffusion coefficient (log D) and the molecular weights (log Mw), external calibration curves were created to give predictions of molecular weights of narrowly-dispersed polymers. This method was successfully applied to atom transfer radical polymerization (ATRP), reversible addition–fragmentation chain transfer (RAFT), and ring-opening metathesis polymerization (ROMP), with weight-average molecular weights given by this method closely correlated to those obtained from GPC measurement.

Keywords

DOSY; diffusion NMR; living polymerization; ATRP; ROMP; RAFT; weight-average molecular weight

The physical properties of polymers are intimately linked to molecular weight (MW).^{1,2} Methods for polymer synthesis are considered “controlled” when they provide polymers with defined MWs and narrow molecular weight distributions (i.e., low polydispersities, PDIs). These key parameters drastically affect the utility of a polymer for a given application. Because synthetic polymers have polydispersity, we speak of their MWs in terms of averages; choice of a relevant average MW is defined by the physical property under consideration. The most common average MWs can be measured via a variety of different techniques such as gel permeation chromatography (GPC, for number-average molecular weight, Mn), osmometry (Mn), static light scattering (weight-average molecular weight, Mw), matrix-assisted laser desorption-ionization mass spectrometry (Mn and Mw), viscometry (Mv), field-flow fractionation, small-angle X-ray scattering, small-angle neutron

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ASSOCIATED CONTENT

Supporting Information

NMR spectra and experimental procedures. This information is available free of charge via the Internet at <http://pubs.acs.org/>.

scattering, ultracentrifugation, cryoscopy, ebulliometry and end-group analysis.³⁻⁸ The weight-average molecular weight is particularly useful to determine certain bulk properties of polymer materials including mechanical properties, thermal properties, and viscoelastic properties. The most common way to determine Mw is via static light scattering, which relates the intensity of scattered light to molar mass. LS systems are not routinely available in most synthetic labs; analysis of Mw is typically achieved via comparison of GPC retention times to polymer standards.

Although it is easy to obtain molecular weights and molecular weight distributions by GPC analysis, this characterization has many practical drawbacks. For instance, GPC experiments are often time-consuming (typically ~15 min – 60 min per sample), furthermore, GPC instruments often consume large amounts of organic solvent for a single measurement. Also, GPC requires regular calibration to get reliable information without a LS detector. The expensive columns and detectors are additional burden for GPC maintenance. Additionally, potential solubility problems could rise when changing solvents. Lastly, GPC is incapable of yielding accurate Mw results at low Mw range (<2000 g/mol).

In polymer science, 1-D and 2-D NMR techniques have been widely utilized to characterize number-average molecular weight (Mn), chemical composition, and microstructures of polymers.^{9,10} Although NMR is very powerful in measuring Mn via end-group analysis, this method is limited to low molecular weights and situations where polymer backbone resonances do not overlap with end groups. We reasoned that diffusion-ordered NMR spectroscopy (DOSY), which relates the chemical shifts of NMR resonances from a given molecular species to the translational diffusion coefficient of that species, could offer a facile method for accurate determination of molecular weights.

DOSY has been utilized as a powerful tool in recent years to investigate solution state structures and interactions of small organic molecules, organometallic aggregates, and macromolecules.¹¹⁻¹⁸ It provides diffusion coefficients of molecules, which are related to hydrodynamic radius and molecular weights.¹⁹⁻²² Johnson invented and applied DOSY to determine polymer weight distribution.¹⁴ Delsuc reported a DOSY-based method to calculate PDIs of linear polymers.²³ DOSY was also utilized by Matyjaszewski to evaluate solvent effects in the atom transfer radical polymerization (ATRP).²⁴ Our lab has recently utilized DOSY to characterize a molecular charm bracelet prepared via ring-closing metathesis and polyrotaxanes prepared via multicomponent ADMET polymerization.^{25,26}

Internal reference systems for ¹H, ³¹P, and ⁶Li DOSY have been successfully applied to determine the molecular weights of a panel of air/moisture sensitive organometallic complexes and self-assembly systems.²⁷⁻³⁵ Given appropriate internal references, formula weights of organic compounds can be estimated within ± 10 % deviation through diffusion coefficient - molecular weight (D-Mw) analysis, which is based on the linear correlation of the logarithm of diffusion coefficients (log D) to the logarithm of the molecular weights of the references (log Mw).^{22,31-35} By evaluating the concentration, viscosity and density effects on DOSY measurements respectively, accurate molecular weight information has been obtained when the internal reference solution was physically separated from the analyte solution by means of a glass capillary as an insert into the NMR tubes.^{32,33} This was most easily accomplished when the reference and analyte solution concentrations were at or below μM level, and therefore the solution properties approached those of the pure solvent.

We envisioned that the principles of the aforementioned reference systems for DOSY could be adopted to create external calibration curves for polymer solutions that would allow one to predict weight-average molecular weights of polymers and monitor the kinetics of living

polymerization. For ^1H DOSY, the acquisition time can be as short as 4 minutes and the concentration can be at μM range, which are ideal for polymer measurements.

This report presents the first use of DOSY technology for the determination of M_w values for kinetic samples from various living/controlled polymerization. To demonstrate the generality of this approach, we synthesized model polymers of varied composition via three different controlled polymerization methods: atom transfer radical polymerization (ATRP),^{36–38} reversible addition-fragmentation chain transfer (RAFT),^{39–42} and ring-opening metathesis polymerization (ROMP).^{43–47} Kinetic studies were performed on model polymerizations using each of these techniques; the resulting data highlighted the strength of DOSY for convenient monitoring of polymer synthesis.

Results and discussions

For a spherical particle of radius R_H , the Stokes-Einstein equation¹¹ describes the diffusion coefficient in eq 1,

$$D = \frac{kT}{6\pi\eta R_H} \quad (1)$$

where k is the Boltzmann constant, T is the absolute temperature, and η is the solvent viscosity. Based on eq 1, Johnson noted that for a monodispersed polymer system, D can be correlated to molar mass M described in eq 2,¹⁴

$$D = AM^\alpha \quad (2)$$

where A and α are constants that correlates to different polymers. Equation 2 can be linearized by taking the logarithm of both sides to yield equation 3,

$$\log D = \alpha \log M + \log A \quad (3)$$

For a class of macromolecules with an approximate constancy in density and similarity in shape, the relation of D and M is shown in eq 4,⁴⁸

$$M \approx (C/D)^{d_f} \quad (4)$$

where d_f is the fractal dimension of molecules and C is a constant. Taking the logarithm of both sides results in eq 5,

$$\log D = d_f \log M + \log C \quad (5)$$

Equation 5 is empirically identical to equation 3 on condition that the shape of a set of analyte compounds remains similar.⁴⁸ Moreover, introducing equation 6: $R_H = (3M/4\pi\rho N_A)^{1/3}$ (ρ is the density of the liquid, N_A is the Avogadro constant) to the Stokes-Einstein equation 1 resulted in equation 7,²²

$$\log D = -\frac{1}{3} \log M + \frac{1}{3} \log \rho - \log \eta - \frac{1}{3} \log \frac{162\pi^2}{k^3 T^3 N_A} \quad (7)$$

To obtain a linear relationship between $\log D$ and $\log M$, convection, density change, and viscosity variation must be well controlled. Since the Stokes-Einstein equation is valid for

particles at least five times larger than the solvent molecules and under infinitely diluted condition, it is highly applicable to diluted polymer solutions. Diluted solvent will assure constant viscosity and density conditions, so that a linear correlation between log D and log M can be achieved.

We therefore postulated that for all our measurements towards external calibration curves and kinetic analysis of polymerization, highly diluted reaction mixture samples can be measured separately to give reliable and comparable diffusion coefficients by DOSY. By creating the linear calibration curve between log D and log Mw of known narrowly dispersed polymers, the molecular weight of unknown polymers can be calculated with the measured diffusion coefficient. In order to prove this hypothesis, calibration curves between log D and log Mw of a set of five selected narrowly dispersed polymers were determined by D-Mw analysis as a first step.

Polystyrene (PS) commercial standards were selected to construct the calibration curve due to its general use as GPC standards. Narrowly dispersed polymers were used to generate the D-Mw plot calibration to reduce the discrepancy of number average molecular weight and weight average molecular weight. Benzene was selected as the solvent due to its relatively high viscosity (0.603 cp at 25 °C)⁴⁹ and its ability to solvate a wide variety of organic polymers.

Considering the large molecular weights of polystyrene standards, full signal attenuation during DOSY experiments were achieved (<5%) by prolonged diffusion time to ensure valid values of diffusion coefficients. Table 1 and Figure 1 showed the detailed results of the PS calibration curve in benzene. The high r^2 value of 0.9991 demonstrated excellent linearity of the log D-log Mw curve. Remarkably, if extrapolating the calibration curve from 9000 g/mol to low molecular range, the Mw of benzene was predicted as 73 g/mol, with only 6.4% deviation of its calculated value. The successful establishment of the polystyrene-based calibration curve in organic system facilitated further kinetic analysis for living polymerization.

We then performed a model ATRP experiment to examine our method. ATRP is the most widely used living radical polymerization because of its high control in polymerization and versatile applications in material science. In ATRP, good control of radical polymerization can be obtained through reversible and repeatable activation/deactivation of dormant species containing propagating radicals in the presence of transition metal complex.³⁶ In our experiment, ATRP of styrene **1** took place with commonly used copper/nitrogen ligand complex, CuBr and N,N,N',N'',N''-pentamethyldiethylenetriamine (PMDETA), and alkyl halide, methyl 2-bromoisobutyrate in anisole. Kinetic aliquots were taken from the polymerization solution at each designated time interval. Mw of each aliquot was characterized by both GPC and DOSY. Diffusion coefficients were obtained after full attenuation of NMR signals. Diffusion coefficients of the samples were fitted to the PS calibration curve (Figure 1) to calculate the Mw. The aliquot was analysed by GPC with multi-angle laser light scattering in THF eluent to determine absolute molecular weights of polymers.

The molecular weights of polystyrene **3** linearly increased with conversions showing a typical ATRP behavior (Table 2, Figure 2). Between the range of 5,500 to 26,900 g/mol (data from GPC), the deviation ranged from 0.4% to 6.4%. Most importantly, the results from both GPC and DOSY were closely correlated. The number average Mn of the final polymer sample at 2700 min was determined to be 265000 g/mol by end-group analysis, based on the purified polystyrene sample redissolved in benzene. Thus, the Mw and Mn given from NMR data can yield a PDI value of 1.02, consistent with GPC measurement.

We stacked six DOSY spectra of ATRP aliquots at different time points ranging from 2 h to the end point of 45 h (Figure 3). The diffusion coefficient of styrene monomer **1** remained unchanged at $17.65 \times 10^{-10} \text{ m}^2/\text{s}$, which demonstrated a good viscosity control by solvent dilution. The polystyrene **3** chain continued elongating with increasing values of hydrodynamic radii and reached a Mw of 26900 g/mol after 45 h. Since the chemical shifts of monomer and polymer overlapped at aromatic regions, only olefinic and alkyl regions were displayed. The stacked DOSY spectrum was a vivid and informative portrait of this living polymerization process.

This model kinetic study of styrene ATRP demonstrated a close correlation between conventional GPC molecular weight determination method and the DOSY molecular weight determination method over a wide range of molecular weights. Compared to conventional GPC molecular weight measurement, the measuring time for DOSY was 4 minutes, about one tenth of the time commonly required for GPC. Moreover, since we used highly diluted samples for ^1H DOSY measurement, the amount of reaction sample required was less than 10% of that for GPC. As such, this DOSY technique is advantageous to perform large quantity measurement for kinetic analysis as well as smaller reaction setup for catalyst evaluation.

Based on the exciting results from ATRP, we continued to assess reversible addition-fragmentation chain-transfer polymerization (RAFT) kinetic aliquot as another example of living polymerization. The PS calibration curve with the region between 5,000 and 30,000 g/mol was enhanced by incorporating four data points obtained from ATRP measurements for more accurate Mw prediction of RAFT polymers. The enhanced calibration curve ($y = -0.537x - 7.695$) showed an increased r^2 value of 0.9997 with improved accuracy in the lower Mw range (Figure S26, Table S4).

We next analyzed a typical RAFT polymerization of styrene **1** in a similar fashion. In our experiment, trithiocarbonate CTA, 2-(dodecylthiocarbonothioylthio)-2-methylpropionic acid **4** were used to perform RAFT which was initiated by thermal initiator azobisisobutyronitrile. We randomly sampled the reaction crude at four time points to verify agreement in molecular weights for both GPC and DOSY experiments (Table 3). The molecular weights obtained from GPC and DOSY were within less than 7% deviation between 5,000 g/mol and 90,000 g/mol. In general, higher molecular weight polymer indicated higher agreement between DOSY and GPC values than the lower molecular weight instance. The values of diffusion coefficient of styrene were all close to $17.65 \times 10^{-10} \text{ m}^2/\text{s}$, suggesting good viscosity control between each sample. The chain growth of polystyrene **5** over time was conveniently monitored by the decreasing values of diffusion coefficient (Figure 4).

Living polymerization ATRP and RAFT of styrene were proved to be highly agreeable in molecular weight determination by both GPC and DOSY measurement over a wide range of molecular weights. Finally, ROMP was examined to broaden the utility of our method for controlled polymerization.

To begin, two representative polycyclooctenes with Mw of 9,800 g/mol and 71,800 g/mol (GPC data) were prepared separately with the 2nd generation catalyst **8** in dichloromethane. The molecular weights of the polycyclooctene samples were predicted to be 9,700 g/mol and 68,900 g/mol by the PS calibration curve (Figure 1), with 1.2 % and 3.9 % deviation from the GPC values, respectively. The PS-based calibration plot was tailored again for ROMP type linear polymers by incorporating the two polycyclooctene samples into the existing curve (Figure S27). The r^2 of overall calibration plot increased from 0.9991 to 0.9998, and

the molecular weight prediction differences of the polycyclooctene samples were reduced to 0.4% and 3.1% (Table S6).

Stacking five DOSY spectra of polystyrene and polycyclooctene (poly(COE)) standards resulted in a spectrum resembling that from a typical gel electrophoresis (Figure 5). The benzene and polystyrene standards resonances at δ 7.15 ppm and 7.00 ppm lined up at various diffusion coefficients according to their molecular weights. The polycyclooctene resonances at δ 5.50 ppm, 2.12 ppm, and 1.35 ppm diffused at the same level, with one set of resonances close to polystyrene with a Mw of 9,000 g/mol, and the other set higher than polystyrene with a Mw of 600,000 g/mol. This intriguing observation implies the possibility of using this method on the determination of molecular weights of biological relevant macromolecules.

The polymerization process of ROMP of 1, 5-cyclooctadiene (COD) **6** was initiated by the 2nd generation catalyst **8** after 5 minutes (Figure 6). The conversion rate was proportional to the increase in molecular weight. Over a period of 45 minutes, the molecular weight of polyCOD **7** reached 29,600 g/mol with 100% conversion (Figure 6 and Figure 7). The Mw – conversion plot yielded a non-linear curve with a slope change after 20% conversion. The diffusion coefficients of the newly generated polyCOD **7** resonance at δ 5.50 ppm decreased from 10 minutes to 45 minutes from the stacked DOSY spectrum (Figure 8).

The DOSY polymer molecular weight determination method was utilized to evaluate polymerizations process of various olefin metathesis catalysts as an additional application. We used the 2nd generation **8** and 3rd generation **9** catalysts to polymerize cyclooctene **10** by ROMP in DCM and THF (Figure 9). The 3rd generation catalyst **9** is known to be a fast initiating catalyst for ROMP. The resulting data indicated that **9** initiated the polymerization of COE **10** immediately upon addition to the reaction mixture and completed the reaction in 20 min, generating **11** with a predicted Mw of 52,700 g/mol. In contrast, the polymerization catalyzed by **8** did not commence until 20 min. However, the final molecular weight of **11** polymerized by **9** was 56,700 g/mol, lower than that catalyzed by **8** (84,900 g/mol). Additionally, ¹H DOSY and ¹H NMR experiments showed that ROMP by **8** in THF was slow with low conversion, due to the coordinating nature of THF towards active catalytic species in solution.⁵⁰ These successful examples of ROMP revealed a facile access to new olefin polymerization catalysts evaluation and reaction condition optimization by this method.

Conclusions

A new diffusion NMR-based method to determine the molecular weight of linear and narrowly dispersed polymers is reported. Narrowly dispersed polystyrene-based calibration curves have been established using linear relation of log D and log Mw to determine weight-average molecular weight of linear flexible polymers. The calibration curve can be utilized to accurately monitor the polymerization proceeding of three commonly used polymerizations: ATRP, RAFT and ROMP. The Mw of polymers determined by DOSY showed excellent agreement with the corresponding GPC data. The calibration curve of PS was tailored twice with ATRP PS and ROMP polyCOE to adapt to various types of linear polymers for more accurate prediction. Moreover, this method was successfully applied to ROMP to monitor kinetics of polymerization catalyst efficiency. It may replace many conventional polymer molecular weight determination methods including GPC, owing to its short measurement time, low sample loading, and high accuracy at low molecular range. One example of measurement of polydispersity index (defined as Mw/Mn) was given in ATRP experiments using DOSY (Mw) and end-group analysis (Mn) data. More accurate

results of PDI can be determined by designing and applying specially modified catalysts or initiators to enhance NMR signals and reduce integration errors.

EXPERIMENTAL SECTION

Materials and General Methods

Benzene- d_6 was kept with 4 Å molecular sieves under argon. 1, 5- cyclooctadiene, cyclooctene, Cu(I)Br (99.999%), N,N,N',N'',N'''-pentamethyldiethylenetriamine (PMDETA), methyl 2-bromoisobutyrate, 2-(Dodecylthiocarbonothioylthio)-2-methylpropionic acid (DDMAT) azobisisobutyronitrile (AIBN) and styrene were obtained from Aldrich Chemical Co., and used as received. Polystyrene commercial standards were purchased from Polysciences Inc. (9000 g/mol, 60000 g/mol, and 80000 g/mol), Pressure Chemical Co. (30000 g/mol and 200000 g/mol), and Shodex Denko America (156000 g/mol).

ATRP experiments

In a Schlenk flask, CuBr (50 mg), PMDETA (60.4 mg) and methyl 2-bromoisobutyrate (63 mg) were placed. The solid reagents containing Schlenk flask was repeatedly vacuumed and backfilled with dry argon for three times. Degassed styrene (7.3 g) and Anisole (8 ml) were added to the solid reagent containing Schlenk flask. The first aliquot was taken prior to placing the reaction mixture in 90 °C oil bath. Kinetic aliquots (0.3 ml) were taken at designated times for DOSY and GPC characterization. For each aliquot, the polymerization was stopped by removing heat followed by exposure the reaction solution in the air. The polymer solution was filtered through neutral alumina packed column to remove copper complex.

RAFT experiments

All solid reagents, DDMAT (100 mg), AIBN (9 mg) were placed in Schlenk flask and the Schlenk flask was repeatedly vacuumed and backfilled with dry argon for three times. Degassed styrene (5.7 g) and anisole (6.3 ml) were added to the Schlenk flask and then the reaction mixture was degassed by argon bubbling. The reaction mixture was heated at 70 °C with stirring. Aliquots (0.3 ml) were taken at designated times for DOSY and GPC characterization.

ROMP experiments

In a flame-dried and argon-charged 50 mL flask was added 1.696 mg (0.002 mmol) The 2nd generation “Sheng Ding” catalyst **8** and 20 mL dry dichloromethane at room temperature. Timing was started as soon as 1.230 mL (10 mmol) 1, 5- cyclooctadiene was injected into the solution. 1 mL aliquots were extracted via syringe from the reaction mixture at designated times into a scintillation vial with two drops of ethyl vinyl ether. 40 µL of the quenched reaction mixture were transferred to an NMR tube and diluted with 1 mL of benzene- d_6 for NMR experiments.

NMR experiments

All NMR tubes were flame dried and evacuated *in vacuo* in advance. NMR experiments were performed at 25 ± 1 °C. All NMR samples were stabilized at 25 °C for 5 min before data collection. For polystyrene standard samples, each NMR tube contained 0.5 mg polystyrene and 1 mL benzene- d_6 . For ATRP, RAFT, and ROMP samples, each NMR tube contained 40 µL reaction mixture and 1 mL benzene- d_6 . DOSY experiments were performed on a Bruker DRX 300 spectrometer equipped with an Accustar z-axis gradient amplifier and an ATMA BBO probe with a z-axis gradient coil. All experiments were run without

spinning to avoid convection. Maximum gradient strength was 0.214 T/m. The standard Bruker pulse program, *step1s*, employing a stimulated echo sequence and 1 spoil gradient was utilized. Bipolar rectangular gradients were used with a total duration of 0.5–10 ms (7 ms for polystyrene measurement). Gradient recovery delays were 0.5–1 μ s. Diffusion times were between 100–2000 ms (200 ms for polystyrene measurement). The number of gradient steps was set to be 32. Individual rows of the quasi-2-D diffusion databases were phased and baseline corrected. DOSY spectra were processed by Topspin 1.3 software. Diffusion dimension was generated using inverse Laplace Transform driven by maximum entropy method.²² Diffusion coefficients of a chosen narrow chemical shift range were extracted from T1/T2 analysis module of Topspin 1.3 calculated by the maximum entropy (GIFA) method developed by Delsuc *et al.*⁵¹ Unless otherwise mentioned, the DOSY spectrum was processed without additional change of processing parameters.

GPC characterization

GPC analysis of polymer was performed by two PLgel 10 μ m mixed-B LS columns (Polymer Laboratories) connected in series with a DAWN EOS multiangle laser light scattering (MALLS) detector (Wyatt Technology), an Optilab DSP differential refractometer (Wyatt Technology), and a ViscoStar viscometer (Wyatt Technology) in THF eluent. The analysis was performed with a flow rate of 1 ml/min at room temperature. Absolute molecular weights and PDI were calculated from dn/dc values that were obtained by assuming 100% mass elution from the GPC columns.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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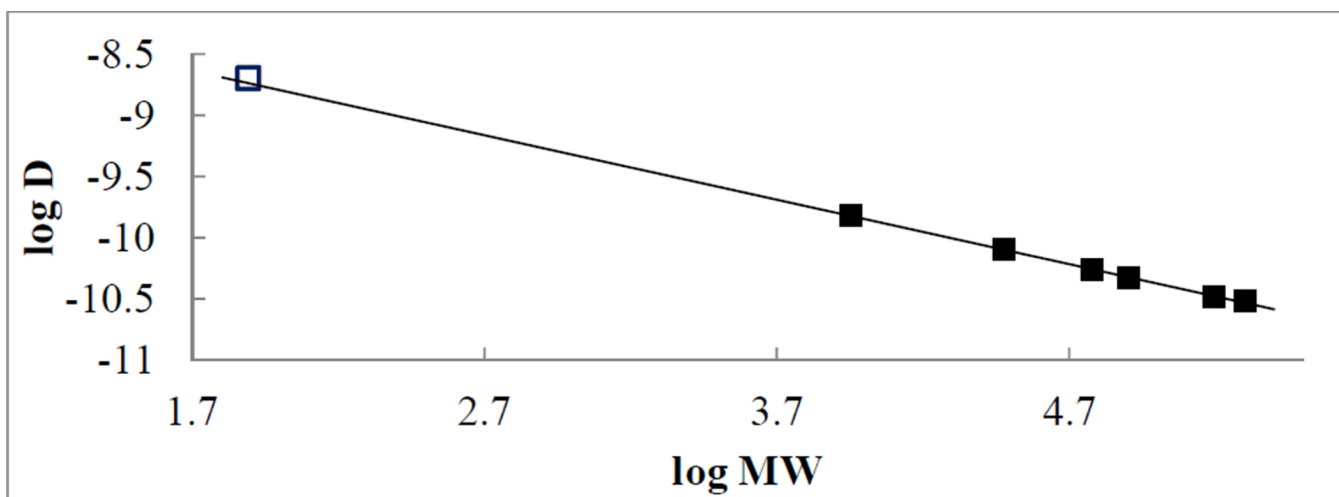


Figure 1. PS calibration curve in benzene- d_6 for Mw prediction. Black solid square: PS standards, blue open square: benzene.

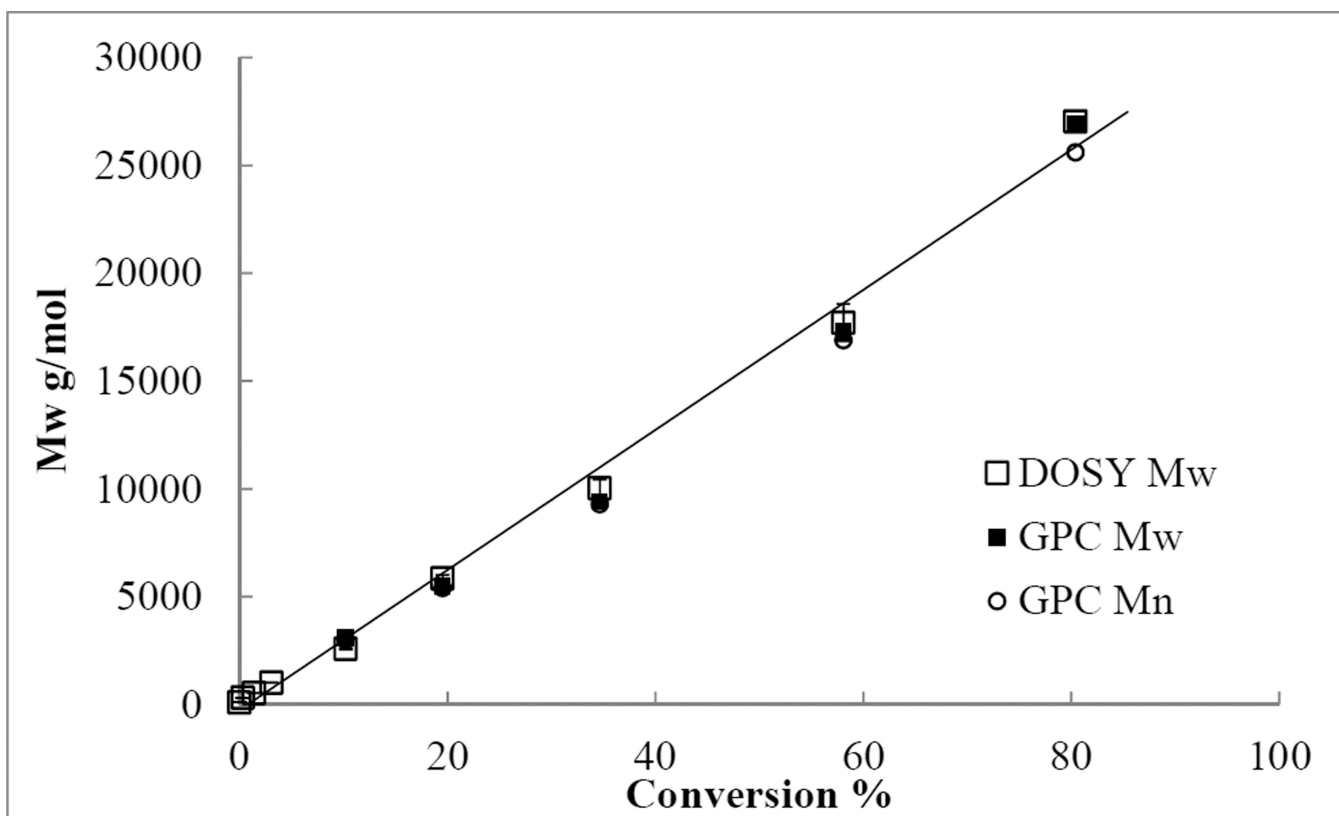


Figure 2. Conversion-Mw plot of polymerization of styrene by ATRP. Black solid square: DOSY Mw, black open square: GPC Mw, black open circle: GPC Mn.

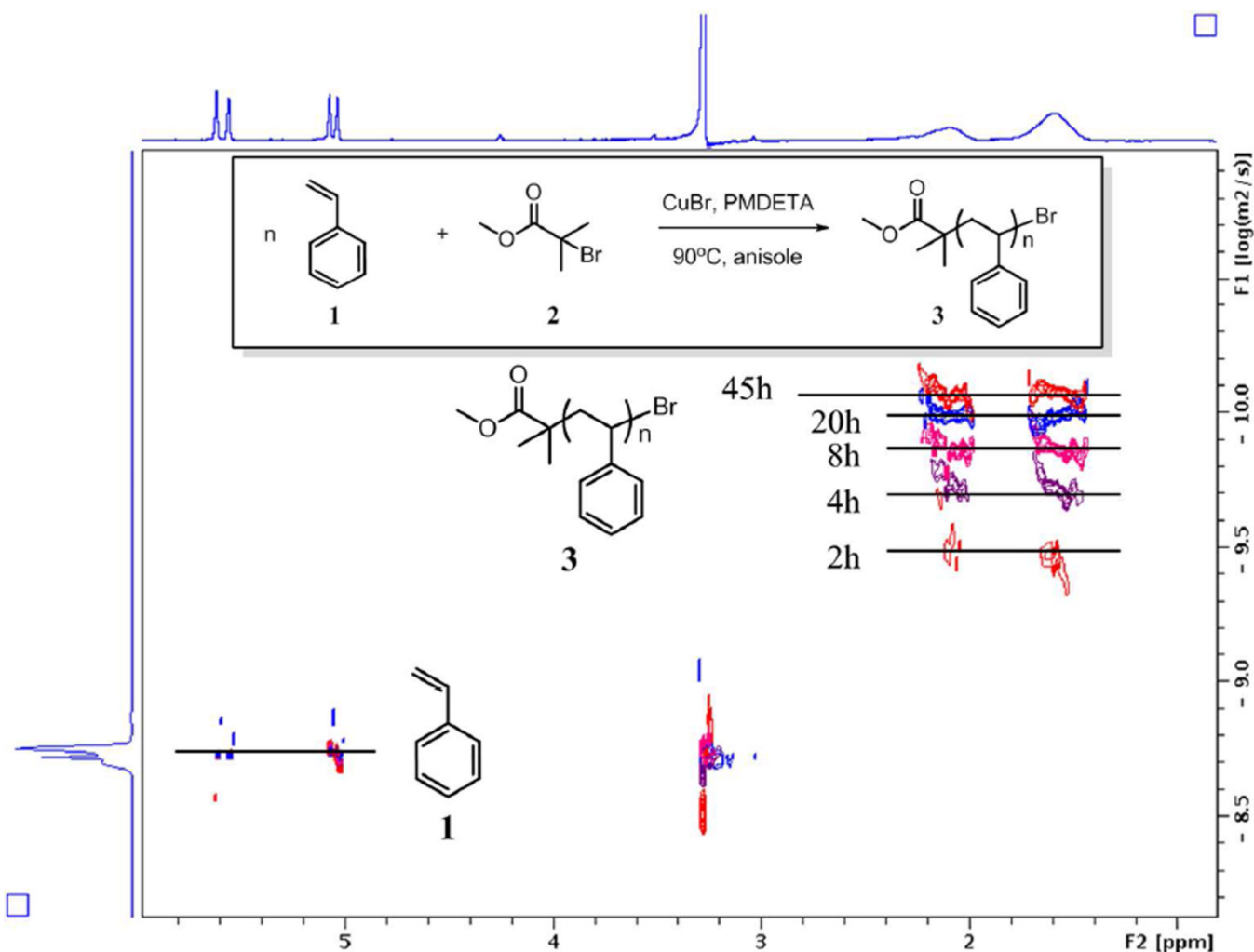


Figure 3. Stacked DOSY spectra of reaction crude of ATRP. DOSY spectra of 0h, 2h, 4h, 8h, 20h, and 45h were stacked. X-axis demonstrates all the ^1H resonances of the components in solution. Y-axis is the diffusion dimension. Olefinic peaks of styrene were used as internal references to monitor the viscosity change of the dilute crude solution. Due to the heavy overlap of the aromatic regions of styrene and polystyrene, they are not shown in the stacked spectra. Representative ATRP ^1H DOSY spectrum can be found in Figure S20.

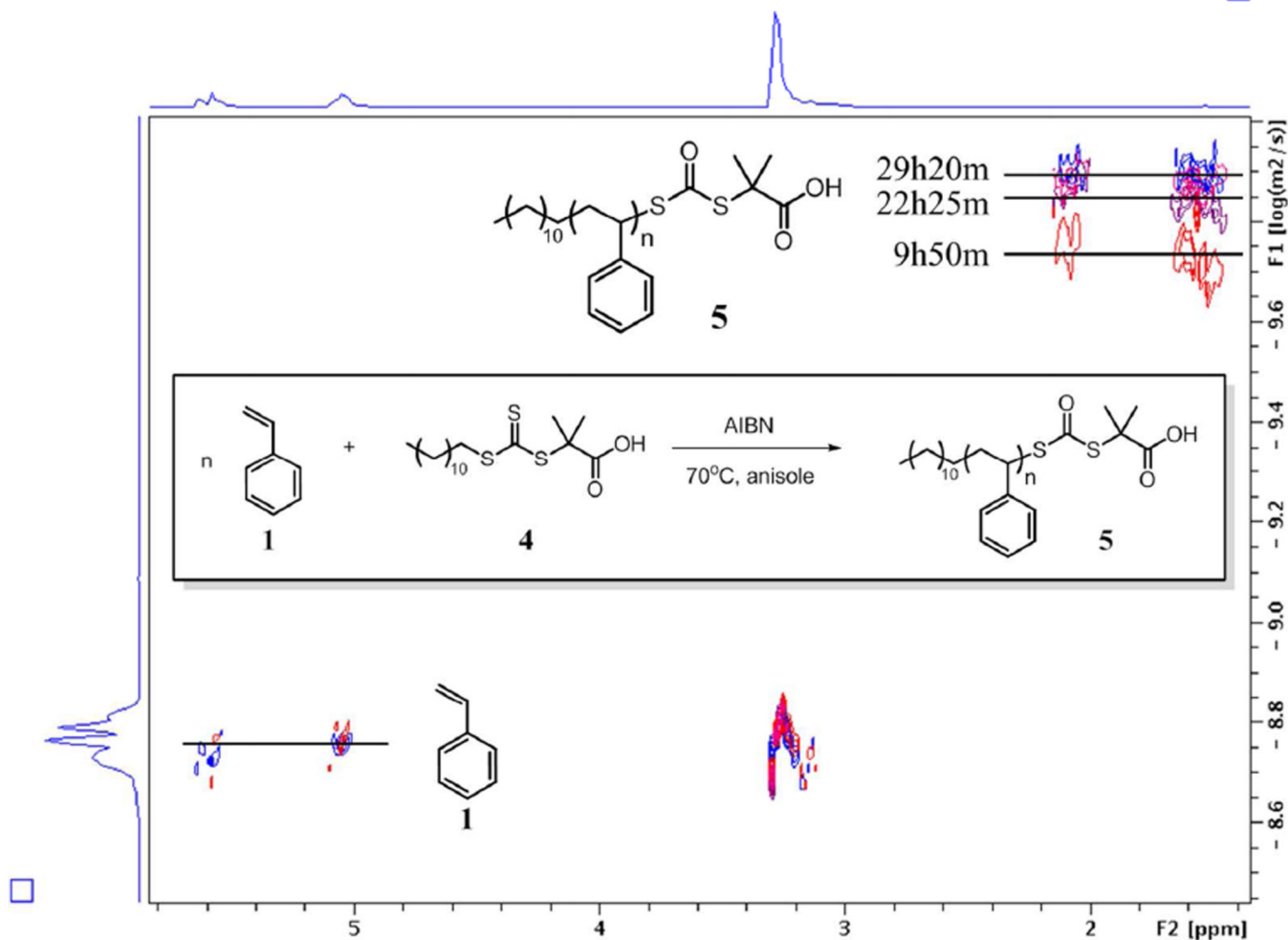


Figure 4. Stacked DOSY spectra of reaction crude of RAFT. DOSY spectra of 0h, 9h50m, 22h25m, and 29h20min were stacked. Olefinic peaks of styrene were used as internal references to monitor the viscosity change of the dilute crude solution. Due to the heavy overlap of the aromatic regions of styrene and polystyrene, they are not shown in the stacked spectra. Representative RAFT ^1H DOSY spectra can be found in supplementary Figure S21.

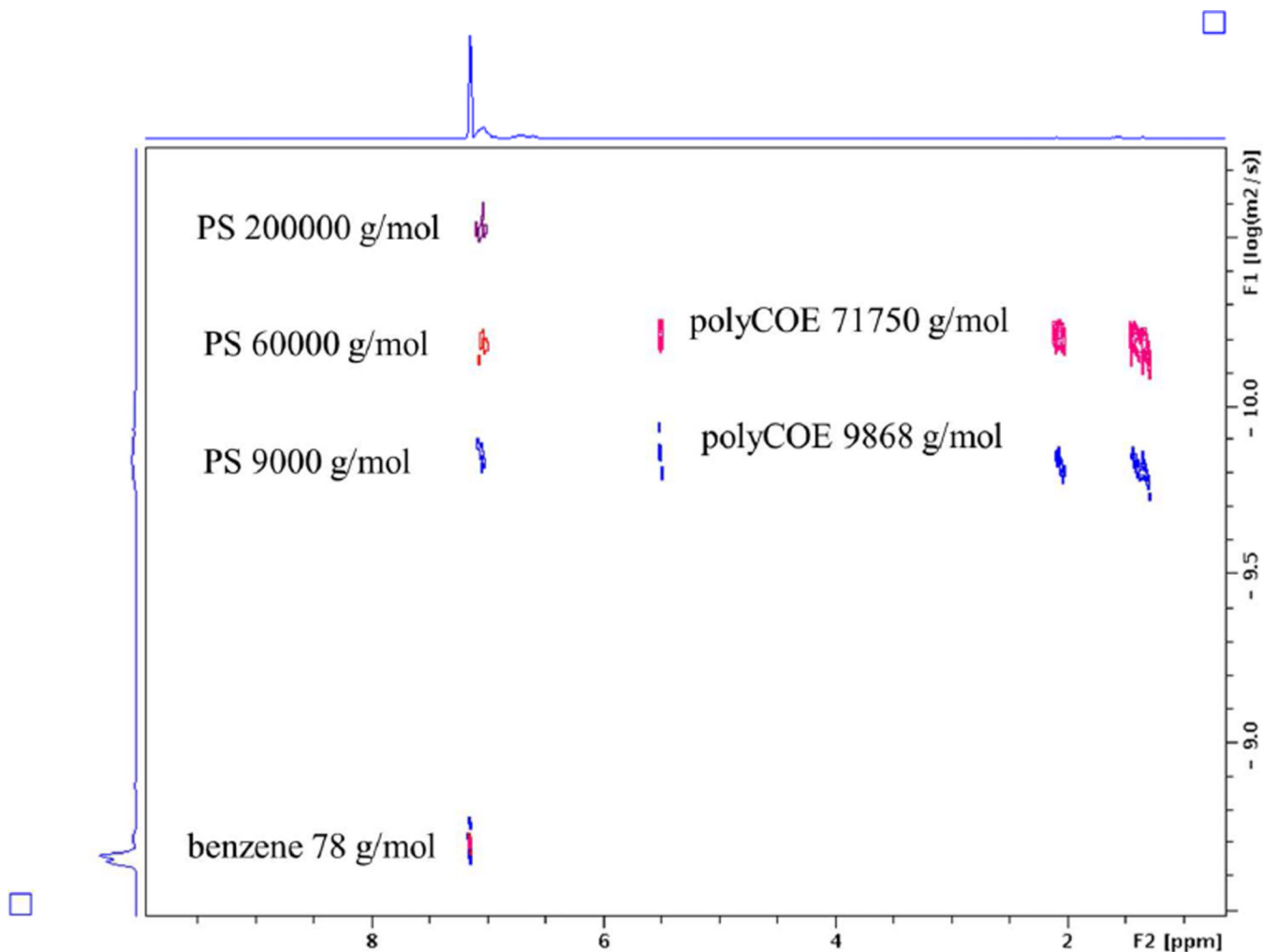


Figure 5. Stacked DOSY spectra of polystyrene standards and polycyclooctene standards. We chose polystyrene with molecular weight of 9000 g/mol, 60000 g/mol, and 200000 g/mol and polycyclooctene with molecular weight of 9800 g/mol and 71800 g/mol as representative examples.

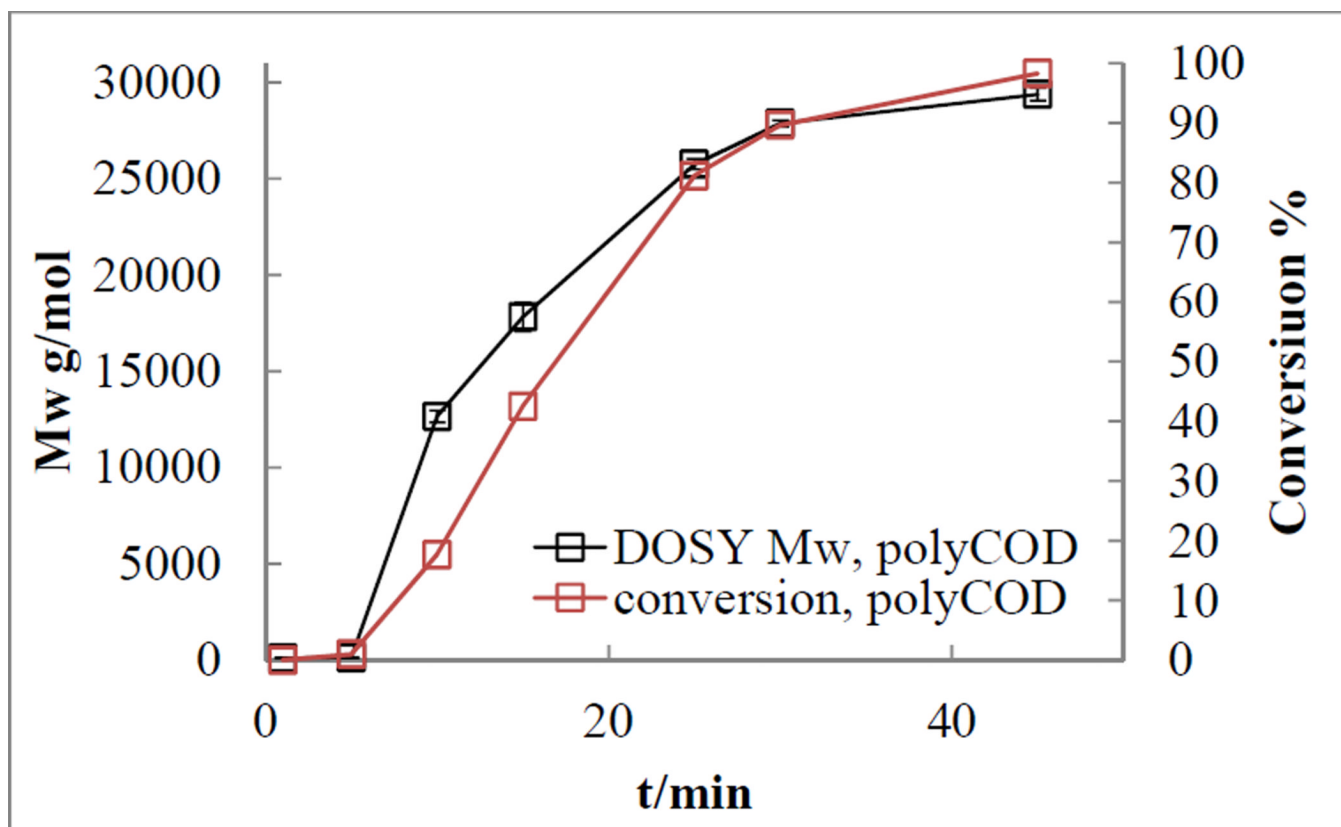


Figure 6. Mw-t plot and conversion-t plot of polymerization of 1,5-cyclooctadiene by ROMP. Black open square: weight-average molecular weight provided by ^1H DOSY. Red open square: conversion calculated from ^1H NMR.

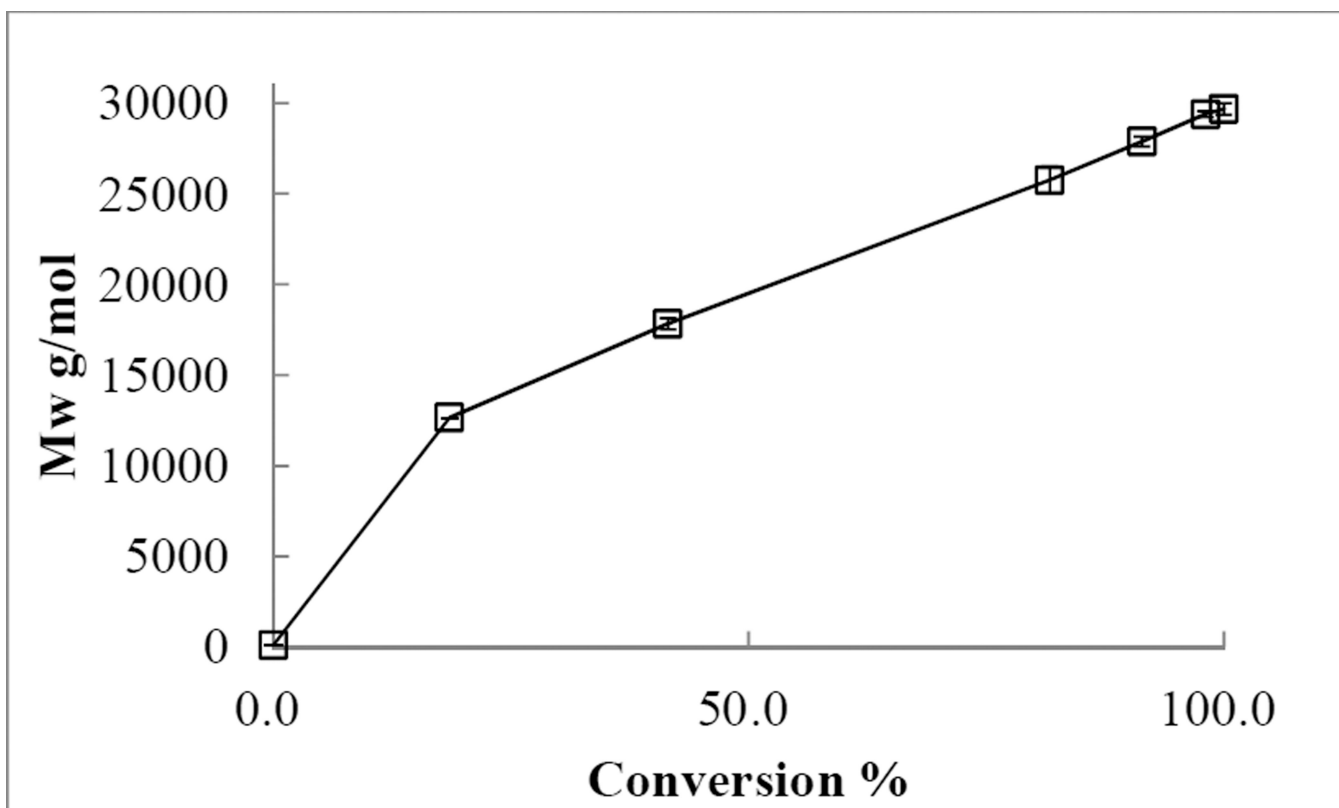


Figure 7. Mw-conversion plot of polymerization of COD by ROMP under the experimental conditions.

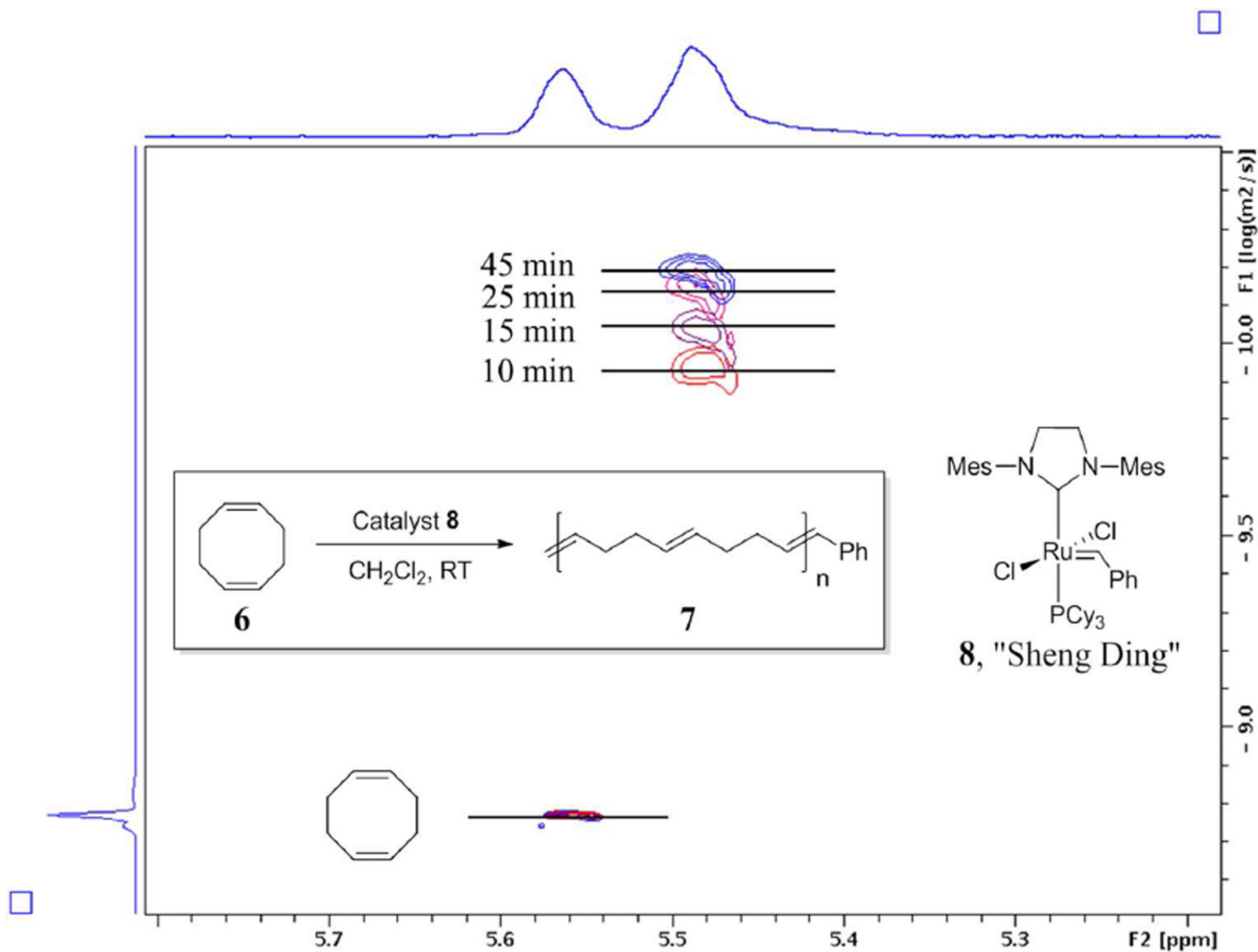


Figure 8. Stacked DOSY spectra of the reaction crude of ROMP. DOSY spectra of 0 min, 10 min, 15 min, and 25 min, and 45 min were stacked. Olefinic peaks of cyclooctadiene were used as internal references to monitor the viscosity change of the dilute crude solution.

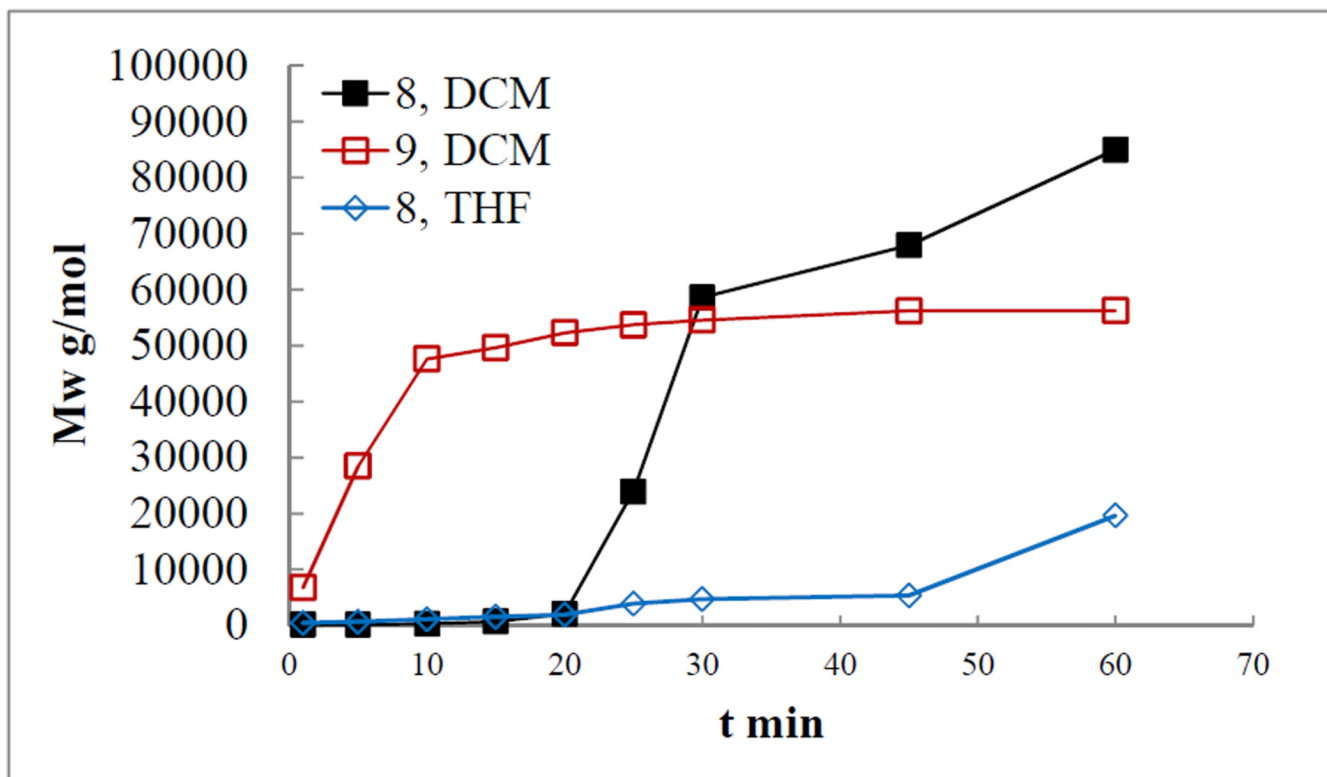
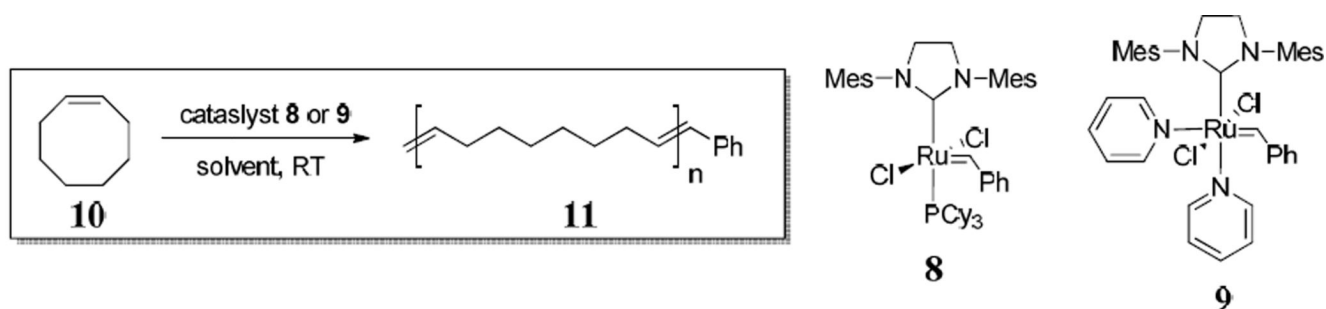


Figure 9. Mw-t plot of polymerization of cyclooctene by ROMP. The monomer to catalyst ratio is 5000/1. All reactions were performed in dry dichloromethane or THF at room temperature in 0.5 M concentration.

Table 1

D-Mw results of PS standard calibration curve and predictions.

Samples	Mw (GPC) g/mol	D ^a (10 ⁻¹¹)	ave. Mw (DOSY) ^b g/mol	Diff % ^c	PD ^d
PS	9000	15.02 ± 0.03	9100	1.1	1.06
PS	30000	8.08 ± 0.01	29000	-3.3	1.06
PS	60000	5.40 ± 0.02	61000	1.7	1.04
PS	80000	4.69 ± 0.02	80000	0.0	1.07
PS	156000	3.31 ± 0.03	153000	-1.9	1.06
PS	200000	3.00 ± 0.03	183000	-8.5	1.06
BEN	78	200.0	73	-6.4	-

^a¹H DOSY measurement was performed at 25 °C in benzene-*d*₆.

^b Equation of the PS standard calibration curve is $y = -0.537x - 7.697$ ($R^2 = 0.9991$). Mw (DOSY) was calculated from the calibration curve using the experimental values of D.

^c Calculated from Mw (GPC) and Mw (DOSY).

^d According to commercial samples. For more details please see Table S1.

Table 2

ATRP results from GPC and externally referenced DOSY.

t min	M _w (GPC) g/mol	D ^a (10 ⁻¹⁰)	ave. M _w (DOSY) ^b g/mol	Diff ^c %	Conv ^d %	PDI
0	-	17.6 ± 0.5	96	-	0.0	-
5	-	9.17 ± 0.02	300	-	0.3	-
20	-	7.00 ± 0.09	500	-	1.4	-
50	-	4.97 ± 0.03	1000	-	3.1	-
120	3100	3.00 ± 0.02	2600	-16.1	10.2	1.03
240	5500	1.93 ± 0.03	5800	5.4	19.5	1.02
480	9400	1.45 ± 0.03	10000	6.4	34.7	1.01
1200	17300	1.07 ± 0.03	17700	2.3	58.1	1.02
2700	26900	0.849 ± 0.002	27000	0.4	80.4	1.03

^a¹H DOSY measurement was performed at 25 °C in benzene-*d*₆.

^b Equation of the PS standard calibration curve is $y = -0.537x - 7.697$ ($R^2 = 0.9991$). M_w (DOSY) was calculated from the calibration curve using the experimental values of D.

^c Calculated from M_w (GPC) and M_w (DOSY).

^d Conversion was calculated by ¹H NMR. GPC molecular weight data below 3000 g/mol is not shown due to possible GPC errors at low molecular weight range. For more details please see Table S2.

Table 3

RAFT results from GPC and externally referenced DOSY.

t/min	Mw (GPC) g/mol	D ^a (10 ⁻¹⁶)	ave. Mw (DOSY) ^b g/mol	Diff % ^c	Conv % ^d	PDI
230	2900	2.60 ± 0.06	3300	13.8	18.7	1.04
590	6200	1.80 ± 0.04	6600	6.4	46.0	1.04
1345	9700	1.41 ± 0.02	10300	6.2	49.4	1.06
1760	10600	1.36 ± 0.03	11000	3.8	53.9	1.06

^a¹H DOSY measurement was performed at 25 °C in benzene-*d*₆.

^bEquation of the PS standard calibration curve is $y = -0.5369x - 7.6947$ ($r^2 = 0.9997$). Mw Pred is based on measured D and the calibration curve.

^cCalculated from Mw (GPC) and Mw (DOSY).

^dConversion was calculated by ¹H NMR. For more details please see Table S3.