

RESEARCH ARTICLE

Open Access

# In vitro antioxidant and antiproliferative activities of plants of the ethnopharmacopeia from northwest of Mexico

Manuel Jiménez-Estrada<sup>1</sup>, Carlos Velázquez-Contreras<sup>2</sup>, Adriana Garibay-Escobar<sup>2</sup>, Davisela Sierras-Canchola<sup>2</sup>, Ricardo Lapizco-Vázquez<sup>2</sup>, Carolina Ortiz-Sandoval<sup>2</sup>, Armando Burgos-Hernández<sup>3</sup> and Ramón Enrique Robles-Zepeda<sup>2\*</sup>

## Abstract

**Background:** The aim of this study, is to investigate the in vitro antioxidant activity, the total phenols content, the flavonoids content and the antiproliferative activity of methanolic extracts of the plants: *Krameria erecta*, *Struthanthus palmeri*, *Phoradendron californicum*, *Senna covesii* and *Stegnosperma halimifolium*, used by different ethnic groups from northwestern Mexico in the treatment and cure of various diseases.

**Methods:** The in vitro antioxidant activity was measured by 2,2-diphenyl-1-picrylhydrazyl (DPPH) and Ferric Reducing/Antioxidant Power assay (FRAP), the total phenols content was measured by Folin-Ciocalteu assay, the flavonoids content by the  $AlCl_3$  colorimetric method and the antiproliferative activity (line cells HeLa, RAW 264.7, M12A<sup>k</sup>.C3.F6 and L929) using MTT method.

**Results:** The *K. erecta* extract showed the higher radical scavenging activity (67.88%), antioxidant activity by FRAP (1.41 mg Trolox Eq), the highest total phenols content (598.51 mg Galic Acid Eq/g extract), the highest flavonoids content (3.80 mg Quercetin Eq/g extract) and the greatest antiproliferative activity in a dose dependent manner against most Cell line evaluated. A positive correlation was found between the antioxidant activity and the flavonoids content.

**Conclusions:** This study is the first report on the antioxidant and antiproliferative activities of the five species evaluated. The results demonstrate that there is a positive correlation between antioxidant activity and the flavonoids content, indicating that these type of polyphenols could be the major contributors to the observed antioxidant activity in the evaluated plant extracts. Of the extracts evaluated, that of *Krameria erecta* showed the greatest antioxidant and antiproliferative activities, a discovery that makes this species a promising candidate for future research.

**Keywords:** DPPH, Total phenol content, Flavonoids, Methanolic plants extracts, Medicinal plants Mexican

## Background

The knowledge of traditional medicine has always guided the search for new cures. In spite of the advent of modern high throughput drug discovery and screening techniques, traditional knowledge systems have given clues for the discovery of valuable drugs [1,2]. The medicinal use of

natural products derived from plants, animals or microorganisms, has a long history.

Free radicals, produced as a result of normal biochemical reactions in the body, play important role in the human body and become harmful only when they are produced in high amounts. Free radicals are implicated in cancer, atherosclerosis, aging, immunosuppression, inflammation, ischemic heart disease, diabetes, hair loss, and neurodegenerative disorders such as Alzheimer's disease and Parkinson's disease [3-5]. The human body possesses innate defense mechanisms to counter free

\* Correspondence: rrobles@guayacan.uson.mx

<sup>2</sup>Departamento de Ciencias Químico Biológicas, División de Ciencias Biológicas y de la Salud, Universidad de Sonora, Blvd. Luis Encinas y Rosales Hermosillo, Sonora 83000, México

Full list of author information is available at the end of the article

radicals in the form of enzymes such as superoxide dismutase, catalase, and glutathione peroxidase. Vitamin C, vitamin E, selenium,  $\beta$ -carotene, lycopene, lutein and other carotenoids have been used as supplementary antioxidants. Plant secondary metabolites such as flavonoids and terpenoids also play an important role in the defense against free radicals [6-8].

Free radicals, produced in oxidation processes, are essential for the production of energy to fuel biological processes in most living organisms. However, the excessive production of free radicals, such as superoxide radicals, hydroxyl radicals, hydrogen peroxide and nitric oxide has been associated with carcinogenesis, coronary heart disease, and many other health issues [9]. Cancer is a leading cause of death and may result from chronic injury to the epithelium by oxidants and other carcinogens.

From about 250,000 species of plants worldwide, all with potential to obtain natural products for the development of new drugs, only 1% of tropical species have been studied for their pharmaceutical potential [10]. About 4,000 species of Mexican plants have medicinal attributes, meaning roughly that one in seven species has a healing property. However it is estimated that the chemical, pharmacological and biomedical validations have been carried out in only 5% of these species [11].

In the state of Sonora in northwestern Mexico, it is estimated that the various indigenous groups used for medicinal purposes, about 450 different plants, including native and introduced species [12]. The aim of this work was to investigate the antiproliferative and antioxidant properties of five wild plants from northwest Mexico and used in the ethnopharmacopeia from state of Sonora, Mexico.

## Methods

### Plant samples

Wild plants were selected based upon their reported traditional use for the treatment of several diseases. General characteristics of the plant species used in this study are described in Table 1. The plants were collected in a region of the state of Sonora, northwest of México (Ranch "El Chaparral", 111° 09' 09" W; 28° 18' 28" N), during February 2008. The plants were taxonomically identified by Professor Jesús Sánchez Escalante at the

Universidad de Sonora Herbarium and a voucher of classification was assigned (Table 1).

### Preparation of plant extracts

The plant extracts were obtained based on the procedures described by Ruiz-Bustos et al. [13]. Plant extracts of *Krameria erecta*, *Struthanthus palmeri*, *Phoradendron californicum*, *Senna covesii* and *Stegnosperma halimifolium* were prepared from air-dried samples (100 g) finely ground with a Wiley mill (200 mesh) and extracted with methanol (1:10 wt/vol) at room temperature. Extractions were carried out over a 10-day period with brief manual shaking twice daily. The extracts were passed through filter paper, dried under reduced pressure at 45°C, and weighed. These "crude" methanolic extracts were dissolved in dimethyl sulfoxide to a final concentration of 10 mg of dry matter/mL. All extracts were stored to -4°C in amber glass vials until use.

### Free-radical scavenging activity (DPPH assay)

Free-radical scavenging activity was measured as described previously with minor modifications [14]. Briefly, plant extract samples dissolved in absolute ethanol (0.6 mL) were mixed with an equal volume of a DPPH solution (300  $\mu$ M) followed by vigorously vortexing for 10 s. After 30 min of incubation at room temperature in the dark, absorbance of the plant extract samples /DPPH solution was read at 517 nm in a spectrophotometer. The scavenging activity was determined by comparing the absorbance with that of the blank containing only DPPH and ethanol. Results were expressed as a percentage decrease with respect to control values. Plant extract samples and Trolox (used as antioxidant standard) were evaluated at different concentrations (12.5–100  $\mu$ g/mL).

### Ferric reducing antioxidant power assay (FRAP assay)

The FRAP assay was carried out according to Benzie and Strain with some modifications [15]. The stock solutions included 300 mM acetate buffer (3.1 g  $C_2H_3NaO_2 \cdot 3H_2O$ ) and 16 mL acetic acid), pH 3.6, 10 mM TPTZ (2, 4, 6-tripyridyl-s-triazine) solution in 40 mM HCl, and 20 mM  $FeCl_3 \cdot 6H_2O$  solution. The fresh working solution was prepared by mixing 25 mL acetate buffer, 2.5 mL TPTZ solution, and 2.5 mL  $FeCl_3 \cdot 6H_2O$  solution. Plant

**Table 1 List of plants of the ethnopharmacopeia from Northwestern Mexico screened for the in vitro antiproliferative and antioxidant activities**

Plant	Common Name	Voucher	Family	Medicinal Use
<i>Krameria erecta</i> Willd. ex Schult	Cosahui	USON2008-9	Krameriaceae	Diarrhea, colitis, enteritis, stomach and intestinal cancer
<i>Phoradendron californicum</i> Nutt	Toji de mesquite	USON2008-1	Viscaceae	Diarrhea
<i>Struthanthus palmeri</i> Kuijt	Toji	USON2008-7	Loranthaceae	Diarrhea
<i>Stegnosperma halimifolium</i> Benth	Chapacolor	USON2008-3	Phytolaccaceae	Headache, bite snake, rabies
<i>Senna covesii</i> (A. Gray) Irwin & Barneby	Hoja Sen	USON2008-5	Fabaceae	Chickenpox, diseases of kidney and liver, measles

extract samples (50  $\mu$ L, 1 mg/mL) were mixed with 1.5 mL of the FRAP solution and allow to react for 5 min in the dark. Readings of the colored product [ferrous tripyridyltriazine complex] were then taken at 593 nm. The results were expressed as milligrams of trolox equivalents (mg TE/g).

#### Determination of total phenols content

The total phenolic content of plant extracts was determined using the Folin-Ciocalteu reagent as described previously [16]. Briefly, 40  $\mu$ L of plant extract samples (1 mg/mL) were diluted with distilled water (300  $\mu$ L). Eighty microlitres of Folin-Ciocalteu reagent and 120  $\mu$ L of a 20% sodium carbonate solution (w / v) were added. The volume was brought up to 1 mL with distilled water. The sample was left for 2 h and the absorbance was measured at 760 nm in a spectrophotometer. Based in a curve type realized previously, the results were expressed as milligrams of galic acid equivalents (mg GAE/g).

#### Determination of total flavonoid content

The total flavonoids were determined using a colorimetric method as described previously [17]. Briefly, 0.5 mL of the plant extract (1 mg/mL) was diluted with 2 mL of distilled water. Then, 0.15 mL of a 5% NaNO<sub>2</sub> solution was added to the mixture. After 6 min, 0.15 mL of a 10% AlCl<sub>3</sub> solution was added and the mixture was allowed to stand for 6 min; 2 mL of 1M NaOH were added and the total mixture was made up to 5 mL using distilled water. The solution was mixed well and was allowed to stand for 15 min. Then, the absorbance was measured at 510 nm in a spectrophotometer. The results were expressed as milligrams of quercetin equivalents (mg QE/g).

#### Cell lines and cell culture

Cell lines NCTC clone L929 (normal subcutaneous connective tissue) and HeLa (human cervix carcinoma) were purchased from the American Type Culture Collection (ATCC, Rockville, MD). The M12A<sup>k</sup>.C3.F6 (murine B-cell lymphoma) and RAW (macrophage, Abelson murine leukemia virus transformed) cells lines were kindly provided by Dr. Emil R. Unanue (Department of Pathology and Immunology, Washington University in St. Louis, MO). All cell cultures were carried out in Dulbecco's modified Eagle's medium (DMEM) supplemented with 5% heat inactivated fetal calf serum and grown at 37°C in an atmosphere of 5% CO<sub>2</sub>.

#### Cell proliferation assay

To evaluate the effect of plant extracts on the proliferation of four cell lines, cell proliferation was determined using the standard MTT assay (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) [18]. Briefly, 10,000

cells (50  $\mu$ L) were plated into each well of a flat 96 well plate. After 12 h incubation at 37°C in an atmosphere of 5% CO<sub>2</sub> to allow cell attachment, the cell cultures were incubated with 50  $\mu$ L of medium containing various concentrations of either crude extract or fraction, and the cell cultures were incubated for 48 h. The crude extract or fraction was first pre-suspended in DMSO and then diluted in DMEM media. Control cell cultures were incubated with DMSO (final concentrations of DMSO 0.06%-0.5%). Control cell cultures did not show any evidence of cell damage. In the last 4 h of the cell culture, 10  $\mu$ L of MTT stock solution (5 mg/mL) were added to each well. Formazan crystals were dissolved with acidic isopropanol, and the plates were read in an ELISA plate reader, using a test wavelength of 570 nm and a reference wavelength of 630 nm. Plates were normally read within 15 min of adding isopropanol. The antiproliferative activity of extracts was reported as IC<sub>50</sub> values (IC<sub>50</sub> was defined as the concentration of extract evaluated and that to inhibit cell proliferation by 50%).

#### Statistical analysis

All data were expressed as mean  $\pm$  SD. Statistical analysis was performed by ANOVA procedures. A post-hoc test (Tukey) was carried out when the differences shown by data were significant ( $p < 0.05$ ). SPSS (version 16.0) statistical program was used for all analysis included the Pearson's correlation coefficient.

#### Results and discussion

There are no previous reports on the antioxidant activity, total phenols content, flavonoid content and antiproliferative activity of the five species reported here.

#### Antioxidant activity

The DPPH method use organic radicals [19] and the FRAP assay can react with iron (II) and SH-group containing antioxidants [15], so it is expected that using these two methods accurately reflects all of the antioxidants in a sample.

#### DPPH radical scavenging activity

Antioxidants are compounds that can delay or inhibit the oxidation of lipids and other molecules by inhibiting the initiation or propagation of oxidizing chain reactions [20]. The antioxidant activity is believed to be mainly due to their redox properties, which play an important role in adsorbing and neutralizing free radicals, quenching singlet and triplet oxygen, or decomposing peroxides [21]. The DPPH radical scavenging assay, based in the use DPPH, a stable free radical that decreases significantly on exposure to proton radical scavengers has been used to evaluate the free radical scavenging activity in foods, plant medicinal extracts and natural antioxidants [22-24]. The free radical scavenging activity of

*Krameria erecta*, *Struthanthus palmeri*, *Phoradendron californicum*, *Senna covesii* and *Stegnosperma halimifolium* methanolic extracts was assessed by DPPH assay. Trolox was used as control.

The results show that there is a dose-dependent radical scavenging activity (Figure 1). At all concentrations tested, the highest DPPH radical scavenging activity (%) was shown by *Krameria erecta* ( $92.74 \pm 0.31$ ) and *Struthanthus palmeri* ( $85.13 \pm 3.50$ ) at 100  $\mu\text{g/mL}$ , which were at the same level with the standard trolox. By contrast *Senna covesii* and *Stegnosperma halimifolium* showed no activity at any of the concentrations evaluated. At the lowest concentration tested (12.5  $\mu\text{g/mL}$ ), *K. erecta* was the one that showed higher radical scavenging activity (67.88%), with a significant difference (Tukey,  $p < 0.05$ ) with the rest of the extracts, including the control Trolox. The DPPH scavenging activity is arranged in the following descending order: *K. erecta* > *S. palmeri* > *P. californicum* > *S. covesii* > *S. halimifolium*.

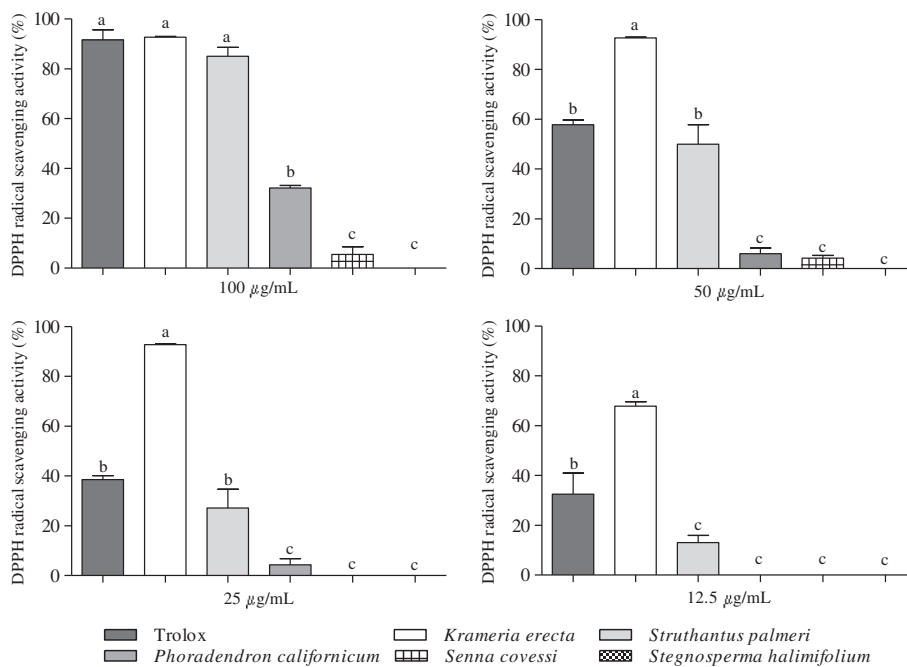
The *Krameria erecta* specie is one of 17 belonging to the only genus of the Family Krameriaceae. It has been reported for this genus the presence of phenolic compounds like tannins and lignans and various biological activities such as hepatoprotective effect, antioxidant and anti-inflammatory [25-29]. The prevention of the chain initiation step by scavenging various reactive species such as free radicals is considered to be an

important antioxidant mode of action [30]; in this context, *Krameria erecta* is likely to be acting through this mechanism.

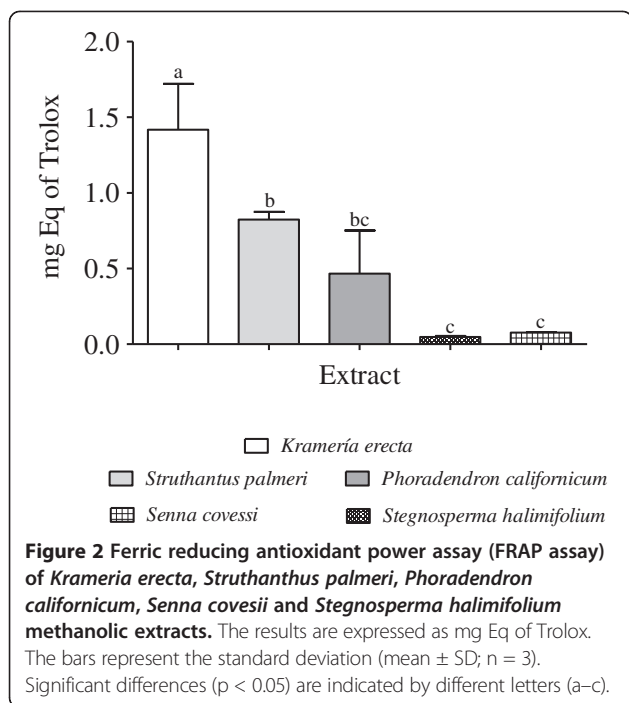
#### Ferric reducing antioxidant power assay (FRAP assay)

A FRAP method is based on the ability of an antioxidant to reduce (electron transfer)  $\text{Fe}^{3+}$  to  $\text{Fe}^{2+}$  ions in the presence of TPTZ (2,4,6-tris(2-pyridyl)-s-triazine), forming an intense blue  $\text{Fe}^{2+}$ -TPTZ complex with an absorption maximum at 593 nm. The reaction is pH-dependent (optimum pH 3.6). The absorbance decrease is proportional to the antioxidant (reductant) content. Compounds that are active in  $\text{Fe}^{3+}$  reduction also stimulate the formation of  $\text{OH}^{\cdot}$  (prooxidation activity) [31].

The FRAP assay results of the five methanolic extracts expressed as mg equivalent of trolox are shown in the Figure 2. In accordance with findings from the DPPH assay, all extracts showed a similar antioxidant activity by FRAP assay, the *Krameria erecta* extract showed the highest reducing power among the extracts with 1.41 mg Trolox Eq (mg TE), followed by *Struthanthus palmeri* (0.82 mg TE). The FRAP values varied from 1.41 to 0.04 (mean 0.56) mg TE, with a significant difference (Tukey,  $p < 0.05$ ) with the rest of the extracts. The methanol extracts with lower antioxidant activity by FRAP, were from *Stegnosperma halimifolium* and *Senna covesii* with 0.047 and 0.075 mg TE, respectively. The extracts with



**Figure 1** Radical scavenging activity in the DPPH assay of *Krameria erecta*, *Struthanthus palmeri*, *Phoradendron californicum*, *Senna covesii* and *Stegnosperma halimifolium* methanolic extracts. Trolox was used as control. The bars represent the standard deviation (mean  $\pm$  SD;  $n = 3$ ). Significant differences ( $p < 0.05$ ) are indicated by different letters (a–c).

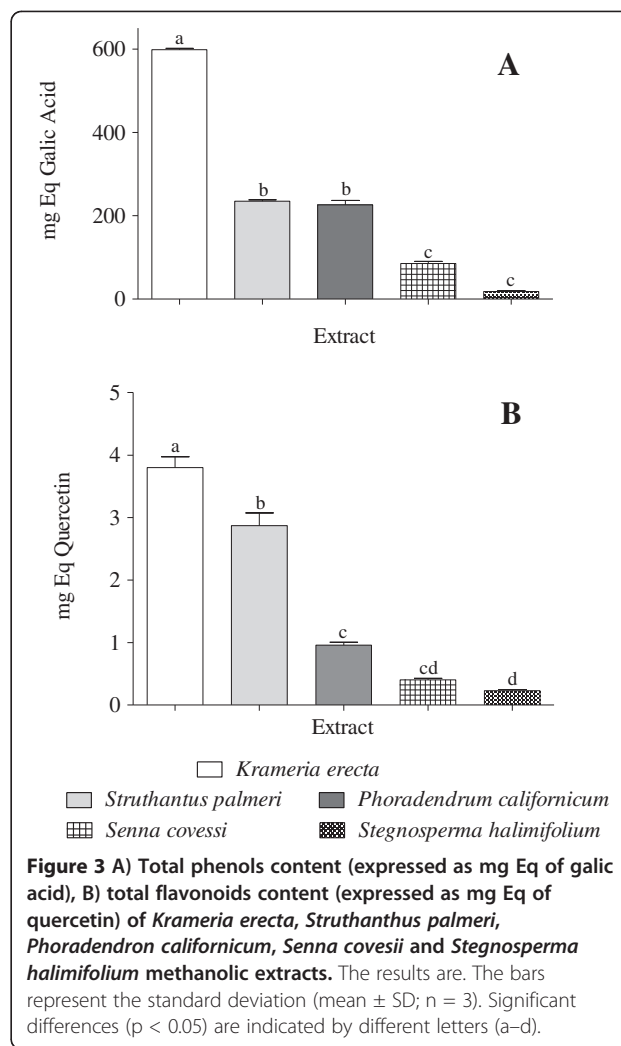


lower antioxidant activity by both methods (DPPH and FRAP) were those with a lower totals phenols content and flavonoids (*S. halimifolium* and *S. covesii*, Figures 3A and B), which indicates that there is a relationship between the antioxidant activity and content of phenolic type compounds in both plant extracts.

Although the DPPH and FRAP methods have different molecular targets, we observe a high correlation between the results of both methods, with a correlation coefficient  $r = 0.9501$  (Table 2).

### Total phenols content

Phenolic compounds are highly effective free radical scavengers and exhibit strong antioxidant activity. The contents of phenolic compounds determined spectrometrically by the Folin-Ciocalteu method of the different analyzed methanolic extracts, are presented in Figure 3A and values are expressed as mg Gallic Acid Eq/g extract (mg GAE/g extract). The total phenols content varied from 598.51 to 18.06 mg GAE/g extract. *Krameria erecta* extract had the highest total phenols content (598.51 mg GAE/g extract) with a significant difference (Tukey,  $p < 0.05$ ) with the rest of the extracts, followed by *S. palmeri* extract (234.69 mg GAE/g extract). The extract of *S. covesii* and *S. halimifolium* showed the lowest content of phenols with 85.11 mg GAE/g extract and 18.06 mg GAE/g extract, respectively. Similar results to those of *K. erecta* have been reported for other species such as *Polygonum aviculare* (677.4 GAE/g extract) [32], but it has a higher content than other species reported [33,34].



**Figure 3** A) Total phenols content (expressed as mg Eq of gallic acid), B) total flavonoids content (expressed as mg Eq of quercetin) of *Krameria erecta*, *Struthanthus palmeri*, *Phoradendron californicum*, *Senna covesii* and *Stegnosperra halimifolium* methanolic extracts. The results are. The bars represent the standard deviation (mean  $\pm$  SD; n = 3). Significant differences ( $p < 0.05$ ) are indicated by different letters (a–d).

It is well known that plant polyphenols are widely distributed in the vegetal kingdom and that they are sometimes present in surprisingly high concentrations; such is the case of *K. erecta*. This plant, commonly known by the name of cosahui, is widely used by ethnic groups in northwestern Mexico, for the treatment of various gastrointestinal diseases such as diarrhea and stomach and intestinal cancer [35]. Antioxidants derived from plant extracts have been shown to have multiple

**Table 2** Pearson's correlation coefficients (r) between the analysis parameters

	Flavonoides	Total phenols	DPPH	FRAP
Flavonoids	1			
Total phenols	0.895862508	1		
DPPH	0.983311003	0.844512931	1	
FRAP	0.976039557	0.965356286	0.950100916	1

biological activities including vasodilatory, anti-inflammatory, antiviral, antiproliferative, and antimicrobial effects [16,36,37].

Since the structural features of phenolic compounds are commonly reported as responsible of the antioxidant activity, the measurements of phenols in extracts may be related to their antioxidant properties [38]. In this study, DPPH scavenging activity and FRAP activity of extracts showed similar trend with the result of total phenolic content,  $r=0.8445$  and  $r=0.9653$ , respectively, indicating that these activities of extracts are highly related to the amount of phenolic compounds, especially flavonoids that present in the extracts (Table 2).

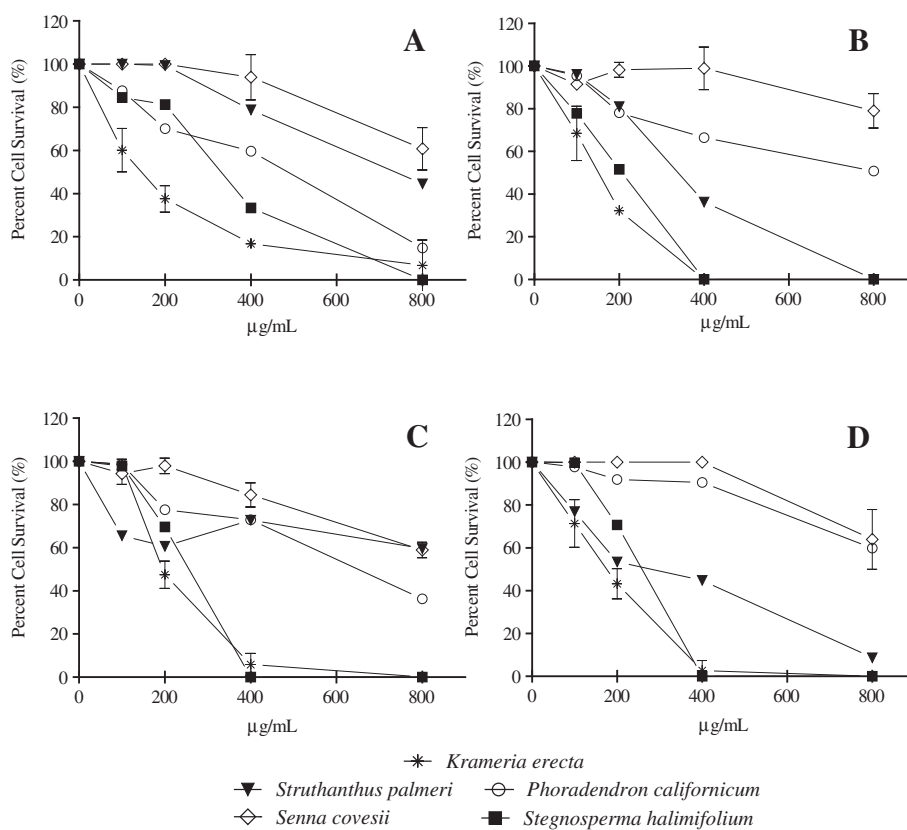
### Total flavonoid content

Flavonoids are the most common and widely distributed group of plant phenolic compounds, which are usually attributed to their antioxidant properties [34]. In this study, the total flavonoid content from different methanolic extracts was evaluated by  $AlCl_3$  colorimetric assay. Quercetin was used as a standard and the total flavonoid content was expressed in mg Quercetin Eq per gram of

extract (mg QE/g extract). The results are showed in the Figure 3B.

Similar to that observed in total phenols content, *Krameria erecta* showed the highest content of flavonoids (3.80 mg QE/g extract), followed by *S. palmeri* (2.87 mg QE/g extract), with a significant difference (Tukey,  $p < 0.05$ ). The lowest total flavonoid content was observed in *S. halimifolium* (0.23 mg QE/g extract). The total flavonoid content is arranged in the following descending order: *K. erecta* > *S. palmeri* > *P. californicum* > *S. covesii* > *S. halimifolium*.

Flavonoids are described as hydrogen donating antioxidants by virtue of the reducing properties of the multiple hydroxyl groups attached to their aromatic ring system, along with their ability to delocalize the resulting phenoxyl radical within the structure. It is recognized that polyphenolic flavonoids are able to scavenge different reactive oxygen radicals such as the hydroxyl and superoxide radicals [39]. In this study, it is likely that the antioxidant activity observed is related to the flavonoid content, since the results from both methods used to determine the antioxidant activity, DPPH and FRAP, the total flavonoid content showed a better correlation with  $r$  values of 0.9833 and 0.9760, respectively (Table 2). The



**Figure 4** Antiproliferative activity of the extracts on A) HeLa, B) RAW 264.7, C) M12A<sup>k</sup>.C3.F6 and D) L929 cells of *Krameria erecta*, *Struthanthus palmeri*, *Phoradendron californicum*, *Senna covesii* and *Stegnosperma halimifolium* methanolic extracts. The bars represent the standard deviation (mean  $\pm$  SD;  $n = 3$ ).

correlation between flavonoids and total phenols was  $r=0.8958$  (Pearson's correlation coefficients, Table 2), due to this, flavonoids probably not the main component in total content of phenols.

#### Cell proliferation assay

The antiproliferative activity of the extracts on HeLa, RAW 264.7, M12A<sup>k</sup>.C3.F6 and L929 cells was evaluated using the MTT assay, which demonstrates mitochondrial activity of cells and is conventionally used as a measure of cell viability. The results are shown in Figure 4.

The extract of *K. erecta* showed the greatest antiproliferative activity in a dose dependent manner, being more active against HeLa and RAW 264.7 cells ( $IC_{50} < 200 \mu\text{g/mL}$ ), while at  $400 \mu\text{g/mL}$  showed a 100% inhibition of cell growth. Additionally, to determine whether the extracts had unspecific cytotoxicity on any cell type, we also tested the effect on the growth of L929, a normal line of subcutaneous connective tissue. On this cell line, *K. erecta* extract showed an  $IC_{50}$  of  $< 200 \mu\text{g/mL}$ , and at  $400 \mu\text{g/mL}$  completely inhibited growth (Figure 4).

The extract of *S. halimifolium* showed an  $IC_{50}$  for antiproliferative activity of between  $200\text{--}400 \mu\text{g/mL}$  with a 100% inhibition of cell growth at  $400 \mu\text{g/mL}$  for RAW 264.7, M12Ak.C3.F6 and L929 cell lines. The extract of *S. palmeri*, presented an  $IC_{50}$  of between  $200\text{--}400 \mu\text{g/mL}$  only for cell lines RAW 264.7 and L929. The remaining two extracts, *P. californicum* and *S. covesii* showed no significant antiproliferative activity at concentrations evaluated in this study.

Lignans are involved in the inhibition of phospholipase C $\gamma$ 1, a key enzyme involved in signaling pathways and essential for proliferation by mitogens [40]. Lignan-type compounds have been isolated from different species of *Krameria* [27,28,41,42]. Considering the antioxidant activity, the content of total phenols, flavonoids and antiproliferative activity observed in the methanolic extract of *K. erecta*, it is likely that these biological activities are related to the presence of lignan-type compounds already observed in other species of the genus.

In addition to the significant antiproliferative activity, as mentioned above, the methanolic extract of *K. erecta* presented a great antioxidant activity. The events of oxidative cell damage are often correlated with the oxidative stress, so that the presence of both properties in this extract could be beneficial for preventive or therapeutic purposes. Since the preliminary results reported here are very promising, it is very important to explore the isolation of pure compounds responsible for these activities.

#### Conclusions

This study is the first report on the antioxidant and antiproliferative activities of the five species evaluated. The results demonstrates that there is a positive correlation

between antioxidant activity and the flavonoids content, indicating that these type of polyphenols could be the major contributors to the observed antioxidative activity in the evaluated plant extracts. Of the extracts evaluated, that of *Krameria erecta* showed the greatest antioxidant and antiproliferative activities, a discovery that makes this species a promising candidate for future research.

#### Competing interests

The authors declare that they have no competing interests.

#### Authors' contributions

MJE, CVC conceived the study, analysis of data and drafting of the manuscript. AGE, DSC, RLV, COS, were involved in cell culturing, MTT, DPPH and total phenolic content assays. ABH was involved in the flavonoids assay and analysis of data. RERZ, was responsible for conception and design, acquisition of data, analysis and interpretation of data and drafted the manuscript. All authors have read and approved the final manuscript.

#### Acknowledgement

This work was supported by a grant from National Council for Science and Technology of Mexico (CONACYT, 83462).

#### Author details

<sup>1</sup>Departamento de Productos Naturales, Instituto de Química, Universidad Nacional Autónoma de México, Cd. Universitaria, México, Distrito Federal 04510, México. <sup>2</sup>Departamento de Ciencias Químico Biológicas, División de Ciencias Biológicas y de la Salud, Universidad de Sonora, Blvd. Luis Encinas y Rosales Hermosillo, Sonora 83000, México. <sup>3</sup>Departamento de Investigación y Posgrado en Alimentos, División de Ciencias Biológicas y de la Salud, Universidad de Sonora, Encinas y Rosales Hermosillo, Sonora 83000, México.

Received: 20 September 2012 Accepted: 4 January 2013

Published: 10 January 2013

#### References

1. Walaszek Z: Metabolism, uptake and excretion of a D-glucaric acid salt and its potential use in cancer prevention. *Cancer Detect Prev* 1997, **21**:178–90.
2. Lai LS, Chou ST, Chao WW: Studies on the antioxidative activities of Hsian-tiao (*Mesona procumbens* Hems) leaf gum. *J Agr Food Chem* 2001, **49**:963–968.
3. Beal MF: Aging, energy, and oxidative stress in neurodegenerative diseases. *Ann Neurol* 1995, **38**:357–366.
4. Maxwell SR: Prospects for the use of antioxidant therapies. *Drugs* 1995, **49**:345–361.
5. Poulsen HE, Prieme H, Loft S: Role of oxidative DNA damage in cancer initiation and promotion. *Eur J Cancer Prev* 1998, **7**:9–16.
6. Devasagayam TP, Sainis KB: Immune system and antioxidants, especially those derived from Indian medicinal plants. *Indian J Exp Biol* 2002, **40**:639–655.
7. Govindarajan R, Vijayakumar M, Pushpangadan P: Antioxidant approach to disease management and the role of 'Rasayana' herbs of Ayurveda. *J Ethnopharmacol* 2005, **99**:165–78.
8. Park EJ, Pezzutto JM: Botanicals in cancer chemoprotection. *Canc Metast Rev* 2002, **21**:231–255.
9. Kim M, Park J, Lim S: Antioxidant activity and cell toxicity of pressurised liquid extracts from 20 selected plant species in Jeju, Korea. *Food Chem* 2010, **122**:546–552.
10. Sanjay J, Saklani A: Challenges and opportunities in drug discovery from plants. *Curr Sci India* 2007, **92**:1251–1257.
11. Ocegueda S, Moreno E, Koleff P: Plantas Utilizadas en la Medicina Tradicional y su Identificación Científica. CONABIO. *Biodiversitas* 2005, **62**:12–15.
12. López-Estudillo R, Hinojosa-García A: *Catálogo de plantas medicinales sonorenses*. Hermosillo, México: Ed. Universidad de Sonora; 1988:130.
13. Ruiz-Bustos E, Velázquez C, Garibay-Escobar A, García Z, Plascencia-Jatomea M, Cortez-Rocha MO, Robles-Zepeda RE: Antibacterial and antifungal

- activities of some Mexican medicinal plants. *J Med Food* 2009, **12**:1398–1402.
14. Usia T, Banskota AH, Tezuka Y, Midorikawa K, Matsushige K, Kadota S: **Constituents of Chinese propolis and their antiproliferative activities.** *J Nat Prod* 2002, **65**:673–676.
  15. Benzie IFF, Strain JJ: **The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay.** *Anal Biochem* 1996, **239**:70–76.
  16. Velazquez C, Navarro M, Acosta A, Angulo A, Domínguez Z, Robles R, Robles-Zepeda R, Lugo E, Goycoolea FM, Velazquez EF, Astiazaran H, Hernandez J: **Antibacterial and free-radical scavenging activities of Sonoran propolis.** *J Appl Microbiol* 2007, **103**:1747–1756.
  17. Yanping Z, Yanhua L, Dongzhi W: **Antioxidant activity of a Flavonoid-Rich extract of *Hypericum perforatum* L. in vitro.** *J Agr Food Chem* 2004, **52**:5032–5039.
  18. Mosmann T: **Rapid colorimetric assay for cellular growth and survival-application to proliferation and cyto-toxicity assays.** *J Immunol Methods* 1983, **65**:55–63.
  19. Chandrasekar D, Madhusudhana K, Ramakrishna S, Diwan PV: **Determination of DPPH free radical scavenging activity by reversed-phase HPLC: a sensitive screening method for polyherbal formulations.** *J Pharmaceut Biomed* 2006, **40**:460–464.
  20. Velioglu YS, Mazza G, Gao L, Oomah BD: **Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products.** *J Agr Food Chem* 1998, **46**:4113–4117.
  21. Zheng W, Wang SY: **Antioxidant activity and phenolic compounds in selected herbs.** *J Agr Food Chem* 2001, **49**:5165–5170.
  22. Sánchez-Moreno C: **Review: Methods used to evaluate the free radical scavenging activity in food and biological systems.** *Food Sci Technol Int* 2002, **8**:121–137.
  23. Singh N, Rajini PS: **Free radical scavenging activity of an aqueous extract of potato peel.** *Food Chem* 2004, **85**:611–616.
  24. Maisuthisakul P, Suttajit M, Pongsawatmanit R: **Assessment of phenolic content and free radical-scavenging capacity of some Thai indigenous plants.** *Food Chem* 2007, **100**:1409–1418.
  25. Achenbach H, Utz W, Lozano B, Touche EMG, Moreno S: **Studies on Krameriaceae .11. Lignans and neolignans from *Krameria parvifolia*.** *Phytochemistry* 1996, **43**:1093–1095.
  26. Facino RM, Carini M, Aldini G, De Angelis L: **A rapid screening by liquid chromatography mass spectrometry and fast-atom bombardment tandem mass spectrometry of phenolic constituents with radical scavenging activity, from *Krameria triandra* roots.** *Rapid Commun Mass Sp* 1997, **11**:1303–1308.
  27. Carini M, Aldini G, Orioli M, Facino RM: **Antioxidant and photoprotective activity of a lipophilic extract containing neolignans from *Krameria triandra* roots.** *Planta Med* 2002, **68**:193–197.
  28. Castro JC, Silva MS, Cortes SF, Lemos VS: **Inhibitory effect of the norlignan 2-(2'-hydroxy-4',6'-dimethoxyphenyl)-5-[(E)-propenyl]benzofuran from *Krameria tomentosa* on acetylcholine-induced relaxation of the rat aorta.** *Planta Med* 2006, **72**:78–81.
  29. Torres-González L, Muñoz-Espinosa LE, Rivas-Estilla AM, Trujillo-Murillo K, Salazar-Aranda R, Waksman TN, Cordero-Pérez P: **Protective effect of four Mexican plants against CCl4-induced damage on the Huh7 human hepatoma cell line.** *Ann Hepatol* 2011, **10**:73–79.
  30. Oboh G, Ademosun AO: **Characterization of the antioxidant properties of phenolic extracts from some citrus peels.** *J Food Sci Tech* 2011, **10**:1007/ s13197-010-0222-y.
  31. Stratil P, Klejduš B, Kubáň V: **Determination of total content of phenolic compounds and their antioxidant activity in vegetables—evaluation of spectrophotometric methods.** *J Agr Food Chem* 2006, **54**:607–616.
  32. Hsu CY: **Antioxidant activity of extract from *Polygonum aviculare* L.** *Biol Res* 2006, **39**:281–288.
  33. Ślusarczyk S, Hajnos M, Skalicka-Woźniak K, Matkowski A: **Antioxidant activity of polyphenols from *Lycopus lucidus* Turcz.** *Food Chem* 2009, **113**:134–138.
  34. Ismail HI, Wei CK, Mariod AA, Ismail M: **Phenolic content and antioxidant activity of cantaloupe (*Cucumis melo*) methanolic extracts.** *Food Chem* 2010, **119**:643–647.
  35. Moreno-Salazar SF, Enríquez-Verdugo A, Cuamea-López C, Bolado-Martínez E, Medrano-Candelas T, Robles-Zepeda RE: **Activity of Medicinal Plants, Used by Native Populations from Sonora, Mexico, Against Enteropathogenic Bacteria.** *Pharm Biol* 2008, **46**:732–737.
  36. Bendini A, Cerretani L, Pizzolante L, Gallina TT, Guzzo F, Ceoldo S, Marconi AM, Andreetta F, Levi M: **Phenol content related to antioxidant and antimicrobial activities of *Passiflora* spp. Extracts.** *Eur Food Res Technol* 2006, **223**:102–109.
  37. Hernández J, Goycoolea FM, Quintero J, Acosta A, Castañeda M, Domínguez Z, Robles R, Vazquez-Moreno L, Velazquez EF, Astiazaran H, Lugo E, Velazquez C: **Sonoran Propolis: chemical composition and antiproliferative activity on cancer cell lines.** *Planta Med* 2007, **73**:1469–1474.
  38. Katalinic V, Milos M, Kulisic T, Jukic M: **Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols.** *Food Chem* 2006, **94**:550–557.
  39. Teixeira S, Siquet C, Alves C, Boal I, Marques M, Borges F, Lima JLF, Reis S: **Structure–property studies on the antioxidant activity of flavonoids present in diet.** *Free Radical Bio Med* 2005, **39**:1099–1108.
  40. Lee JS, Kim J, Uck YY, Choong KY: **Inhibition of Phospholipase Cy1 and Cancer Cell Proliferation by Lignans and Flavans from *Machilus thunbergii*.** *Arch Pharm Res* 2004, **27**:1043–1047.
  41. Domínguez XA, Rombold C, Utz W, Achenbach H: **Neolignans, Norneolignans, and Other Components from *Krameria sonorae*.** *Planta Med* 1992, **58**:382–383.
  42. Baumgartner L, Sosa S, Atanasov AG, Bodensieck A, Fakhrudin N, Bauer J, Del Favero G, Ponti C, Heiss EH, Schwaiger S, Ladurner A, Widowitz U, Loggia RD, Rollingner JM, Werz O, Bauer R, Dirsch VM, Tubaro A, Stuppner H: **Lignan Derivatives from *Krameria lappacea* Roots Inhibit Acute Inflammation in Vivo and Pro-inflammatory Mediators in Vitro.** *J Nat Prod* 2011, **74**:1779–1786.

doi:10.1186/1472-6882-13-12

**Cite this article as:** Jiménez-Estrada et al.: In vitro antioxidant and antiproliferative activities of plants of the ethnopharmacopeia from northwest of Mexico. *BMC Complementary and Alternative Medicine* 2013 **13**:12.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

