

Depletion of the Ubiquitin-binding Adaptor Molecule SQSTM1/p62 from Macrophages Harboring *cftr* Δ F508 Mutation Improves the Delivery of *Burkholderia cenocepacia* to the Autophagic Machinery*

Received for publication, August 21, 2012, and in revised form, October 31, 2012. Published, JBC Papers in Press, November 12, 2012, DOI 10.1074/jbc.M112.411728

Basant A. Abdulrahman^{†§}, Arwa Abu Khweek[†], Anwari Akhter[†], Kyle Caution[†], Mia Tazi[†], Hoda Hassan[†], Yucheng Zhang[†], Patrick D. Rowland[†], Sankalp Malhotra[¶], Famke Aeffner^{||}, Ian C. Davis^{||}, Miguel A. Valvano^{**††1}, and Amal O. Amer^{‡2}

From the [†]Department of Microbial Infection and Immunity, Department of Internal Medicine and Center for Microbial Interface Biology, Ohio State University, Columbus, Ohio, the [§]Department of Biochemistry and Molecular Biology, Faculty of Pharmacy, Helwan University, Cairo, Egypt, the [¶]The Ohio State University Medical Scientist Training Program, Ohio State University, Columbus, Ohio, the ^{||}Department of Veterinary Biosciences, Ohio State University, Columbus, Ohio 43210, the ^{**}Centre for Human Immunology, Department of Microbiology and Immunology, University of Western Ontario, London, Ontario, N6A 5C1, Canada, and the ^{††}Centre for Infection and Immunity, Queen's University, Belfast BT9 7BL, United Kingdom

Background: Cystic fibrosis is characterized by defective autophagy and increased *Burkholderia cenocepacia* infection.

Results: The depletion of SQSTM1/p62 from Δ F508 macrophages improves bacterial clearance via autophagy.

Conclusion: p62 expression level determines the fate of *B. cepacia* infection in Δ F508 macrophages.

Significance: Our study reveals the role of p62 in diseases characterized by protein aggregates that compromise autophagy by consuming essential autophagy molecules.

Cystic fibrosis is the most common inherited lethal disease in Caucasians. It is caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR), of which the *cftr* Δ F508 mutation is the most common. Δ F508 macrophages are intrinsically defective in autophagy because of the sequestration of essential autophagy molecules within unprocessed CFTR aggregates. Defective autophagy allows *Burkholderia cenocepacia* (*B. cepacia*) to survive and replicate in Δ F508 macrophages. Infection by *B. cepacia* poses a great risk to cystic fibrosis patients because it causes accelerated lung inflammation and, in some cases, a lethal necrotizing pneumonia. Autophagy is a cell survival mechanism whereby an autophagosome engulfs non-functional organelles and delivers them to the lysosome for degradation. The ubiquitin binding adaptor protein SQSTM1/p62 is required for the delivery of several ubiquitinated cargos to the autophagosome. In WT macrophages, p62 depletion and overexpression lead to increased and decreased bacterial intracellular survival, respectively. In contrast, depletion of p62 in Δ F508 macrophages results in decreased bacterial survival, whereas overexpression of p62 leads to increased *B. cepacia* intracellular growth. Interestingly, the depletion of p62 from Δ F508 macrophages results in the release of the autophagy molecule beclin1 (BECN1) from the mutant CFTR aggregates and allows its redis-

tribution and recruitment to the *B. cepacia* vacuole, mediating the acquisition of the autophagy marker LC3 and bacterial clearance via autophagy. These data demonstrate that p62 differentially dictates the fate of *B. cepacia* infection in WT and Δ F508 macrophages.

Cystic fibrosis (CF)³ is the most common inherited lethal disease among Caucasians, which is caused by mutations in the *cftr* gene encoding the cystic fibrosis transmembrane conductance regulator (CFTR). The most common CFTR mutation results in a deletion of phenylalanine at position 508 (Δ F508), which affects the processing of the CFTR protein in such way that it cannot reach the epithelial cell surface. This mutation results in an aggresome-prone protein that forms intracellular aggregates (1–4).

Autophagy is a conserved physiological process that eliminates non-functional organelles and recycles cytosolic components to generate nutrients during periods of stress or starvation (5, 6). Autophagy also targets cytosolic long-lived proteins and organelles for lysosomal degradation in eukaryotic cells and plays a role in innate immunity (7). Loss of autophagy in murine tissues is accompanied by accumulation of protein aggregates and disordered organelles, leading to life-threatening diseases (8). Autophagy plays a key role in protecting the cytosol from bacterial infection. The mechanisms of bacterial recognition by this pathway are starting to be elucidated. Some cellular cargos are marked for autophagy by acquiring adaptor

* This work was supported by a doctoral fellowship from the Egyptian Bureau of Education (to B. A. A.). This work was supported, in whole or in part, by the National Institutes of Health (Grant R01HL094586 (to A. O. A.)) and a Cystic Fibrosis Canada grant (to M. A. V.).

¹ Canada Research Chair in Infectious Diseases and Microbial Pathogenesis.

² To whom correspondence should be addressed: Department of Microbial Infection and Immunity, Department of Internal Medicine and Center for Microbial Interface Biology, Ohio State University, Biological Research Tower, 460 W. 12th Ave., Room 1014, Columbus, OH 43210. Tel.: 614-247-1566; Fax: 614-292-9616; E-mail: amal.amer@osumc.edu.

³ The abbreviations used are: CF, cystic fibrosis; CFTR, cystic fibrosis transmembrane regulator; q-PCR, quantitative PCR; CFU, colony-forming unit; RFP, red fluorescent protein.

p62 Dictates the Fate of *B. cepacia* in Murine Macrophages

proteins such as Calcoco2 (also known as NDP52) and neighbor of BRCA1 gene product (NBR1) (9–14). In addition, SQSTM1 (also known as p62) is required for targeting *Salmonella enterica* serovar Typhimurium (*Salmonella typhimurium*), intracytosolic *Shigella*, and *Listeria* to the autophagic pathway (9, 10).

The adaptor molecule p62 is a ubiquitously expressed cellular protein that is conserved in metazoa but not in plants or fungi (15, 16). The quantity of p62 is critical for cell viability and is strictly controlled (17). p62 has multiple protein-protein interaction domains, including the ubiquitin-associated domain for ubiquitinated cargo binding and the LC3 interaction region for binding LC3 (10). Accordingly, impaired autophagy is accompanied by accumulation of p62 followed by formation of aggregates containing p62 and ubiquitinated proteins. This accumulation occurs because of the nature of both self-oligomerization and ubiquitin binding of p62 (18, 19).

Burkholderia cenocepacia (*B. cepacia*) is an opportunistic Gram-negative bacterium that infects CF patients and leads to severe lung inflammation and lung tissue destruction. Occasionally, this infection results in a lethal necrotizing pneumonia (20–22). Unfortunately, *B. cepacia* is resistant to most known antibiotics and, thus, is nearly impossible to treat. *B. cepacia* adopts an extracellular or intracellular lifestyle (23, 24). This bacterium can survive within a variety of eukaryotic cells such as amoebae, epithelial cells, and macrophages (25–28).

We have demonstrated previously that in WT macrophages, the majority of *B. cepacia*-containing vacuoles slowly acquire the specific autophagy marker LC3 within 2 h of infection. Subsequently, these vacuoles fuse with the lysosomes, and the bacterium is degraded. In Δ F508 macrophages, *B. cepacia*-containing vacuoles do not acquire autophagosome markers and do not fuse with the lysosomes.

Here, we demonstrate that in WT macrophages, p62 is required for targeting *B. cepacia* to the autophagosome. Upon p62 down-regulation, bacterial growth increases, whereas the overexpression of p62 results in a significant decrease in *B. cepacia* replication. On the contrary, down-regulation of p62 in Δ F508 macrophages is associated with decreased bacterial growth, and p62 overexpression results in increased *B. cepacia* replication. p62 down-regulation in Δ F508 macrophages releases the trapped BECN1 from CFTR aggregates, allowing its recruitment to the *B. cepacia* vacuole. BECN1 acquired by the *B. cepacia*-containing vacuole subsequently attracts LC3, thereby mediating the fusion of the maturing autophagosome containing *B. cepacia* with the lysosome via the autophagic machinery. These data provide mechanistic insight on how *B. cepacia* persists in Δ F508 macrophages. This report also suggests that p62 may be an attractive drug target to improve *B. cepacia* clearance by autophagic machinery.

EXPERIMENTAL PROCEDURES

Bone Marrow-derived Macrophages—Animal experiments were performed according to protocols approved by the Animal Care and Use Committee of the Ohio State University College of Medicine. WT C57BL/6 were purchased from The Jackson Laboratory. Δ F508 mice on a C57BL/6 background were obtained from Case Western University and housed in the Ohio State University vivarium. Bone marrow-derived macrophages

were isolated from the femurs of 6- to 12-week-old mice and were cultured in Iscove's modified Dulbecco's medium (Invitrogen, catalog no. 12440) containing 10% heat-inactivated FBS (Invitrogen, catalog no. 16000), 20% L cell-conditioned medium, 100 units/ml penicillin, and 100 mg/ml streptomycin (Invitrogen, catalog no. 15140) at 37 °C in a humidified atmosphere containing 5% CO₂. Macrophages were infected with *B. cepacia* K56-expressing m-RFP or the corresponding gentamicin-sensitive strain MHK1 at a multiplicity of infection of 10.

Bacterial Strains and Culture—*B. cenocepacia* strain K56-2 is a clinical isolate from a CF patient. The corresponding gentamicin-sensitive strain MHK1 was described previously (29). All bacterial strains were grown in Luria-Bertani broth at 37 °C overnight with high-amplitude shaking. To kill extracellular bacteria, Iscove's media (Invitrogen, catalog no. 12440) plus FBS (Invitrogen, catalog no. 16000) containing 50 μ g/ml gentamicin (Invitrogen, catalog no. 3564) were added for 0.5 h, as described previously (29). To enumerate intracellular bacteria, infected macrophages were lysed with ice-cold PBS (Invitrogen, catalog no. 14190) at designated times. Recovered bacteria were quantified by plating serial dilutions on Luria-Bertani agar plates and counting colonies using the Acolyte Colony Counter, 5710/SYN.

Immunoblotting—Macrophages were stimulated with *B. cepacia*, and the culture supernatant was removed. Cells were lysed in lysis buffer solution supplemented with a protease inhibitor mixture (Roche Applied Science, catalog no. 10-519-978-001). The protein concentration was adjusted to 30 μ g/ml. Proteins were separated on a sodium dodecyl sulfate 15% polyacrylamide gel and transferred to a PVDF membrane (Bio-Rad, catalog no. 1p62-0117). Membranes were immunoblotted for p62 (Sigma-Aldrich, catalog no. P0067), LC3 (Sigma-Aldrich, catalog no. L8918), Calreticulin (Stressgen, catalog no. SPA600), BECN1 (Abcam, catalog no. ab55878), NDP52 (Millipore, catalog no. MAB4386), NBR1 (Santa Cruz Biotechnology, Inc., SC-130380), and Actin (Abcam, catalog no. ab8299, and Atg7 (Sigma, A2856)). Protein bands were detected with secondary antibodies conjugated to horseradish peroxidase, followed by enhanced chemiluminescence reagents (Amersham Biosciences, ECL Western blotting detection reagents; GE Healthcare, catalog no. RPN2106).

siRNA Treatment and Plasmid Transfection—siRNA treatment was performed using siRNA against p62 (Dharmacon, catalog no. 18412) ACAGAUGCCAGAAUCGGAA, CUGCUCAGGAGGAGACGAU, GAACAGAUGGAGUCGGGAA, and CCAUGGGUUUCUCGGAUGA; siRNA against NDP52 (Dharmacon, catalog no. 76815) CAACACAGGGGUCAAUAA, CAGAAGAGGACAUCGGGAU, CCAAGGAUGGAGCGCUA, and GAGUUGAGGUGUCCGUGUA; siRNA against NBR1 (Dharmacon, catalog no. 17966) GAAAUGGG-AUUCUGCGACA, AGUCCGUGGAAGCGAGUAA, CAAGCAAAGCUGACGAUUU, and ACAGGAGGCAUUCGGGUUA; and siRNA against Atg7 (Dharmacon, catalog no. 49953) CAUCAUCUUUGAAGUGAAA, GCUAGAGACGUGACA-CAUA, AGCGAAAGCUGGUCAUCAA, and GGUCGUGUCUGUCAAGUGC. siRNA was nucleofected into primary macrophages 48 h before infection using a Lonza nucleofection

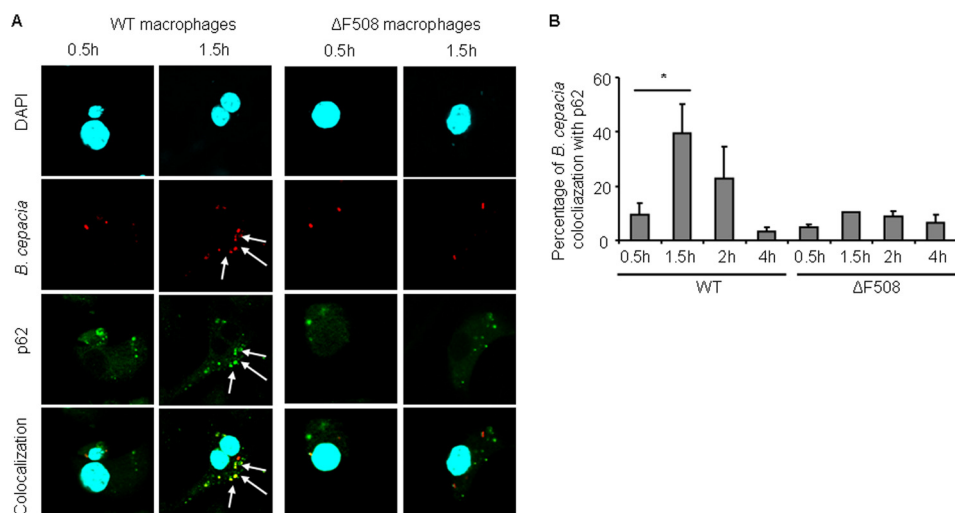


FIGURE 1. **More *B. cepacia* vacuoles colocalize with p62 in WT macrophages than in Δ F508 macrophages.** *A*, confocal microscopy for WT and Δ F508 macrophages infected with *B. cepacia*-expressing m-RFP for 0.5 or 1.5 h. p62 stained green, whereas nuclei were stained with DAPI. *B*, the percentage of colocalization of *B. cepacia* with p62 was quantified at the indicated time points. More than 200 cells were scored. The white arrows indicate the sites of colocalization. Data in *B* are presented as means \pm S.D. *, $p < 0.05$.

kit and Amaxa equipment, as described previously (30, 31). Successful knockdown was confirmed by immunoblot analysis for each experiment. The DsRed-p62 plasmid was obtained from Addgene (32) and was nucleofected into primary macrophages using a Lonza nucleofection kit and Amaxa equipment. The plasmid was nucleofected 24 h before the infection. Successful p62 overexpression was confirmed by immunoblotting.

Real-time PCR—Total RNA was isolated from cells lysed in TRIzol (Invitrogen, catalog no. 15596-026) and then converted to cDNA. Gene expression was calculated as relative copy numbers, as described previously (30, 33). Briefly, C_t values of the p62 gene were subtracted from the average C_t of two housekeeping genes (GADPH and CAP1), and the resulting ΔC_t was used in the following equation: $RCN = (2^{-\Delta C_t}) 100$. The relative copy number of a gene is represented as the number of copies relative to the 100 copies of average housekeeping genes (30, 33).

Confocal Microscopy—Immunofluorescence experiments for colocalization with autophagy markers were performed as described previously (34, 6). Rabbit anti-LC3 (Abgent, catalog no. AP1805a), mouse anti-p62 (BD Biosciences, catalog no. 610832), FK2 mAb (Enzo Bioscience, catalog no. BML-PW8810), and rabbit anti-BECN1 (Abcam, catalog no. ab55878) were used, followed by fluorescent secondary antibodies (Molecular Probes, catalog no. A11008). Nuclei were stained with the nucleic acid dye DAPI (6, 35). Samples were analyzed with an Olympus Fluoview FV10i confocal microscope at the Ohio State University, Department of Microbial Infection and Immunity.

Statistical Analysis—All experiments were performed at least three times independently and yielded similar results. Comparisons of groups for statistical difference were conducted using Student's two-tailed t test. $p \leq 0.05$ was considered significant.

Ethics Statement—This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes

of Health and Ohio State University. The Institutional Animal Care and Use Committee has approved our protocol number 2007A0070. All efforts were made to minimize suffering.

RESULTS

More *B. cepacia* Colocalized with p62 in WT Macrophages Than in Δ F508 Macrophages—We demonstrated previously that *B. cepacia* is cleared by the autophagy machinery in WT macrophages but not in their Δ F508 counterparts. To determine why the *B. cepacia* vacuole is not efficiently recognized by the autophagy machinery in Δ F508 macrophages, we followed the trafficking within WT and Δ F508 macrophages. Recent studies showed that p62 is required for targeting *S. typhimurium*, *Shigella*, and *Listeria* to the autophagic pathway (9, 10). Therefore, we examined the colocalization of *B. cepacia* with p62 in WT and Δ F508 macrophages. The time course for infection was 0.5, 1.5, 2, and 4 h. In WT macrophages, a significant percentage of *B. cepacia* colocalized with p62 at 1.5 h post-infection. Colocalization then declined at later time points (Fig. 1, *A* and *B*). However, *B. cepacia* vacuoles in Δ F508 macrophages did not colocalize with p62 at any time point throughout infection (Fig. 1, *A* and *B*). Together, these data show that p62 labels the *B. cepacia* vacuole in WT but not in Δ F508 macrophages.

The *B. cepacia* Vacuole Efficiently Acquires Ubiquitin in Δ F508 Macrophages—Autophagy recognizes cargo for uptake and degradation when it becomes ubiquitinated and bound to an autophagy adaptor molecule (10). The lack of p62 acquisition by the *B. cepacia* vacuole in Δ F508 macrophages could be due to defective ubiquitination of the *B. cepacia*-containing vacuole or because of lack of p62 expression in Δ F508 macrophages. To differentiate between these possibilities, we first infected WT and Δ F508 macrophages with *B. cepacia*-expressing m-RFP for 0.5 h or 2 h and examined the colocalization of *B. cepacia* with ubiquitin. There was no significant difference in the colocalization of *B. cepacia* with ubiquitin between WT and Δ F508 macrophages (Fig. 2, *A* and *B*). These data demonstrate

p62 Dictates the Fate of *B. cepacia* in Murine Macrophages

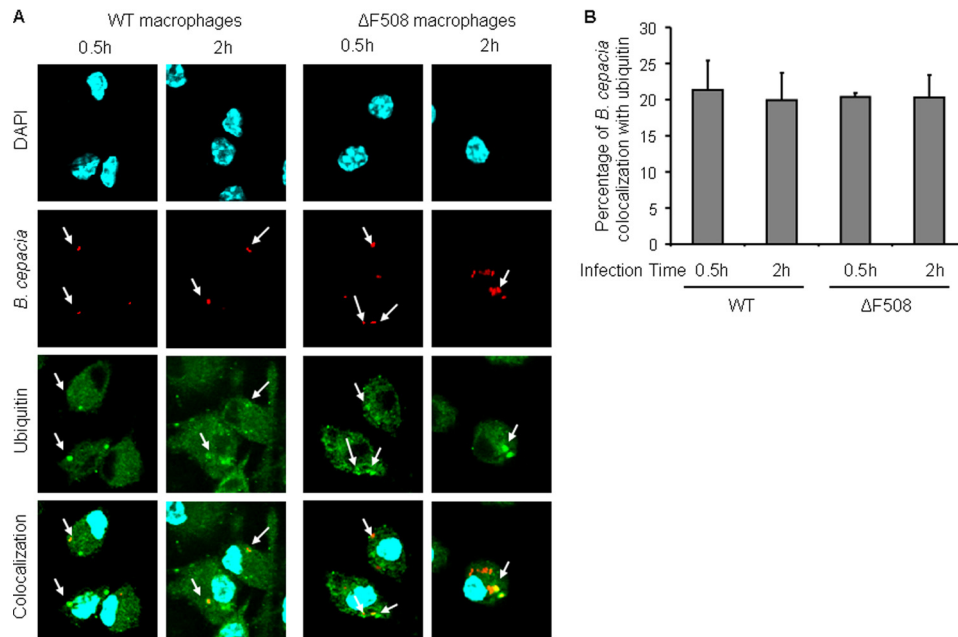


FIGURE 2. The ubiquitination of the *B. cepacia* vacuole is similar in WT and ΔF508 macrophages. *A*, confocal microscopy for WT and ΔF508 macrophages infected with m-RFP-expressing *B. cepacia* for 0.5 or 2 h. Ubiquitin stained green, and nuclei were stained with DAPI (the white arrow indicates the sites of colocalization). *B*, the percentage of colocalization of *B. cepacia* with ubiquitin was quantified. More than 200 cells were scored.

that equivalent numbers of *B. cepacia* vacuoles acquired ubiquitin in WT and ΔF508 macrophages. Therefore, the lack of colocalization of *B. cepacia* with autophagosomes in ΔF508 macrophages is not due to the absence of ubiquitin around the *B. cepacia* vacuole.

Next, to determine whether the failure of the autophagy machinery to target the *B. cepacia* vacuole is due to the lack of p62 expression in ΔF508 macrophages, we examined the level of p62 within WT and ΔF508 macrophages. Immunoblot analysis using an antibody against p62 revealed that murine macrophages harboring the ΔF508 mutation exhibited a higher level of p62 compared with WT macrophages (Fig. 3A). Quantitative PCR (q-PCR) was performed to determine whether the increase in p62 protein level in ΔF508 macrophages, compared with WT macrophages, is due to regulation of gene expression or accumulation of the p62 protein. There was no significant difference in the p62 mRNA level in both types of macrophages (Fig. 3B). Together, these data show that the increase in p62 level in ΔF508 macrophages is due to accumulation of the protein inside the cell, suggesting defective autophagy activity.

B. cepacia Infection Elevates p62 Expression within WT and ΔF508 Macrophages—p62 is well expressed in ΔF508 macrophages. However, *B. cepacia* infection down-regulates autophagy in both WT and ΔF508 macrophages. Thus, it is possible that *B. cepacia* infection is accompanied by depletion of p62 from ΔF508 macrophages upon infection. To examine this possibility, we examined the effect of *B. cepacia* on p62 expression upon infection in WT and ΔF508 macrophages by q-PCR and immunoblot analysis. At 4 h post infection, q-PCR analysis demonstrated increased expression of the p62 gene level in both WT and ΔF508 macrophages compared with non-infected macrophages (Fig. 3C). Similarly, immunoblotting showed a higher p62 level in both types of macrophages (Fig. 3D). Together, these data show that *B. cepacia* infection

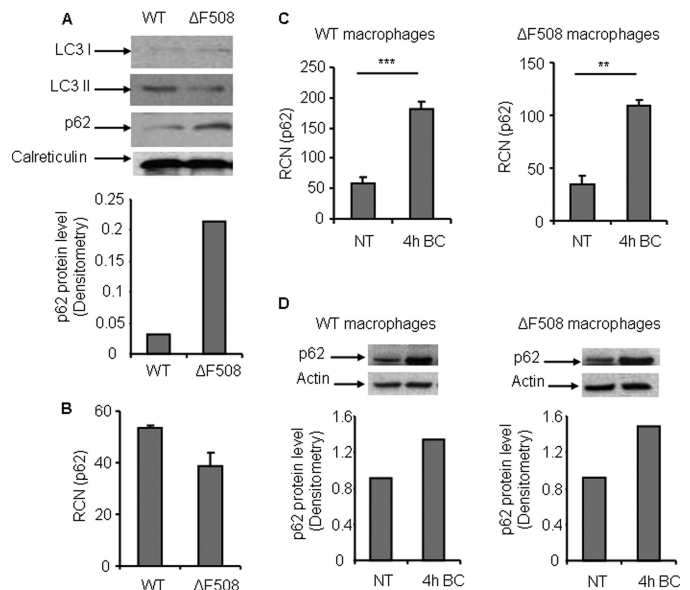


FIGURE 3. Murine bone marrow-derived macrophages harboring the ΔF508 mutation have a higher level of p62 than WT macrophages.

A, upper panel, immunoblot analysis of WT and ΔF508 macrophage lysates showing the expression level of LC3 (I/II) and p62, respectively. Lower panel, densitometry of the p62 protein level. *B*, q-PCR expression profile of p62 in WT and ΔF508 macrophages. *C*, q-PCR expression profile of p62 in WT and ΔF508 macrophages non-infected (NT) or infected with *B. cepacia* for 4 h (4h BC). **, $p < 0.01$; ***, $p < 0.001$. *D*, upper panel, immunoblot analysis for WT and ΔF508 macrophages using p62 antibody prior and at 4 h post-infection with *B. cepacia*. Lower panel, densitometry of the p62 protein level. Data in *B* and *C* are expressed as relative copy numbers (RCN) and shown as means \pm S.D. of three independent experiments.

increases the expression level of p62 in WT and ΔF508 macrophages.

Overexpression of p62 Conversely Affects B. cepacia Replication in WT and ΔF508 Macrophages—To determine the role of p62 in *B. cepacia* replication in WT and ΔF508 macrophages, we examined *B. cepacia* survival in the presence of ectopically

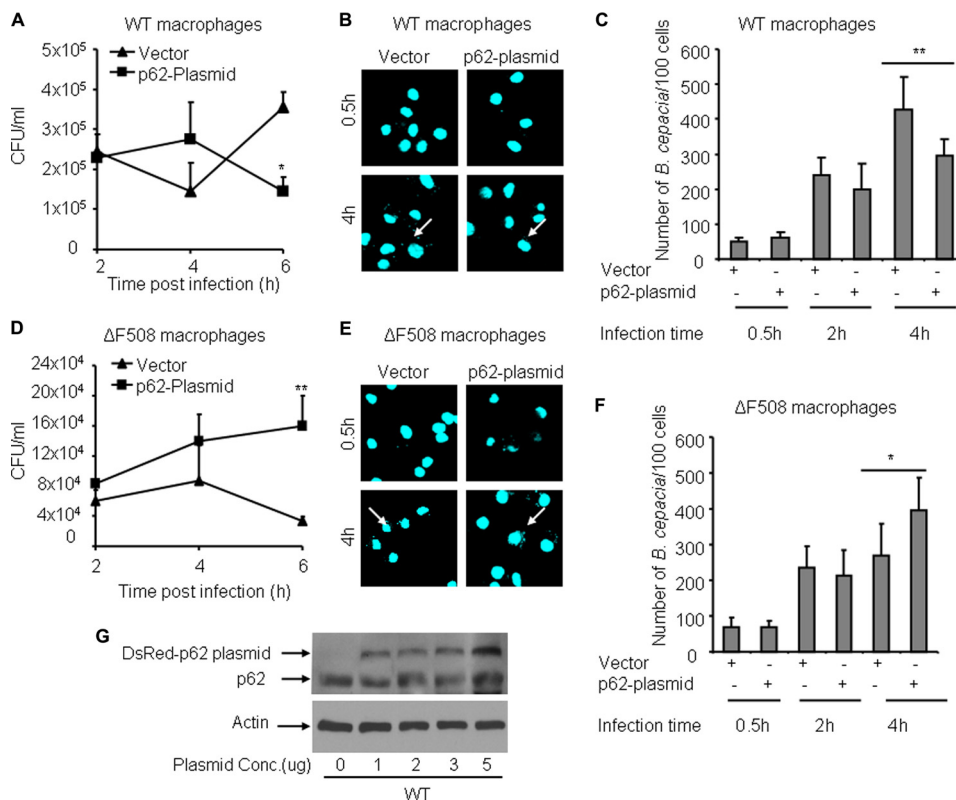


FIGURE 4. p62 overexpression decreases growth of *B. cepacia* in WT macrophages, whereas increases bacterial growth in Δ F508 macrophages. *A* and *D*, WT and Δ F508 macrophages were nucleofected with the p62 plasmid or vector control 24 h prior to infection and then infected with *B. cepacia* for 2, 4, and 6 h. CFUs were enumerated. *B* and *E*, confocal microscopy for WT and Δ F508 macrophages overexpressing p62 at 0.5 or 2 h post-infection. The white arrows indicate *B. cepacia* stained with DAPI. *C* and *F*, both types of macrophages overexpressing p62 were infected for 0.5, 2, and 4 h, and the number of bacteria/100 cells was quantified. More than 200 cells were scored. *G*, immunoblot analysis for WT macrophages after 24-h nucleofection with the DsRed-p62 plasmid showing the overexpression of p62. Data are representative of three different experiments and presented as the means \pm S.D. *A*, *C*, *D*, and *F*, *, $p < 0.05$; **, $p < 0.01$; significant differences from the vector at the respective time points.

expressed p62. WT and Δ F508 macrophages were nucleofected with p62 plasmid or vector control and after 24 h, cells were infected with *B. cepacia* for 2, 4, and 6 h (Fig. 4G). In WT macrophages harboring the p62 plasmid, recovered *B. cepacia* CFUs decreased at 6 h post-infection compared with the cells harboring the vector alone (Fig. 4A). Confocal microscopy revealed significantly less bacterial accumulation upon overexpression of p62 (Fig. 4, B and C). In contrast, Δ F508 macrophages harboring the p62 plasmid allowed significantly increased *B. cepacia* accumulation after 6 h post-infection (Fig. 4D). Confocal microscopy confirmed increased bacterial accumulation (Fig. 4, E and F). Together, these results demonstrate that the availability of p62 differentially determines the fate of *B. cepacia* in WT and Δ F508 macrophages.

Down-regulation of p62 Decreases the Growth of *B. cepacia* in Δ F508 Macrophages—To determine whether p62 targets *B. cepacia* vacuoles to autophagosomes for degradation, we nucleofected WT and Δ F508 macrophages with p62 siRNA or scrambled siRNA (Fig. 5G). After 48 h, cells were infected with *B. cepacia* for 2, 4, and 6 h. In WT macrophages, *B. cepacia* CFUs significantly increased upon down-regulation of p62 (Fig. 5A). In addition, confocal microscopic analysis demonstrated significantly increased bacterial numbers at 2 h post-infection (Fig. 5, B and C). In contrast, Δ F508 macrophages showed decreased *B. cepacia* CFUs upon down-regulation of p62 (Fig. 5D). Furthermore, confocal microscopy revealed significantly

low bacterial accumulation 2 h after *B. cepacia* infection upon down-regulation of p62 (Fig. 5, E and F). Therefore, these data demonstrate that p62 controls *B. cepacia* infection in WT macrophages but not in Δ F508 macrophages. The details of this differential role are not clear.

Decreased p62 Expression Promotes LC3 Acquisition by *B. cepacia* Vacuole in Δ F508 Macrophages—LC3 is the main marker for autophagosomes. The conversion of LC3-I to LC3-II denotes autophagy stimulation and autophagosome formation (7, 36). We have demonstrated previously that *B. cepacia* colocalization with LC3 is markedly decreased in Δ F508 macrophages compared with WT macrophages (37, 38). To determine the underlying mechanism, WT and Δ F508 macrophages were nucleofected with either siRNA against p62 to down-regulate p62 or with scrambled siRNA, and after 48 h, nucleofected macrophages were infected with *B. cepacia*-expressing m-RFP for 0.5 and 2 h. Confocal microscopy showed that in WT macrophages, *B. cepacia* colocalization with LC3 decreased significantly when p62 was down-regulated compared with the siRNA control-treated cells (Fig. 6, A and C). In contrast, Δ F508 macrophages allowed significantly more *B. cepacia* colocalization with LC3 after the down-regulation of p62 compared with the siRNA control-treated cells (Fig. 6, B and D). Together, these data suggest that p62 is required for the delivery of *B. cepacia* to the autophagosomes in WT macro-

p62 Dictates the Fate of *B. cepacia* in Murine Macrophages

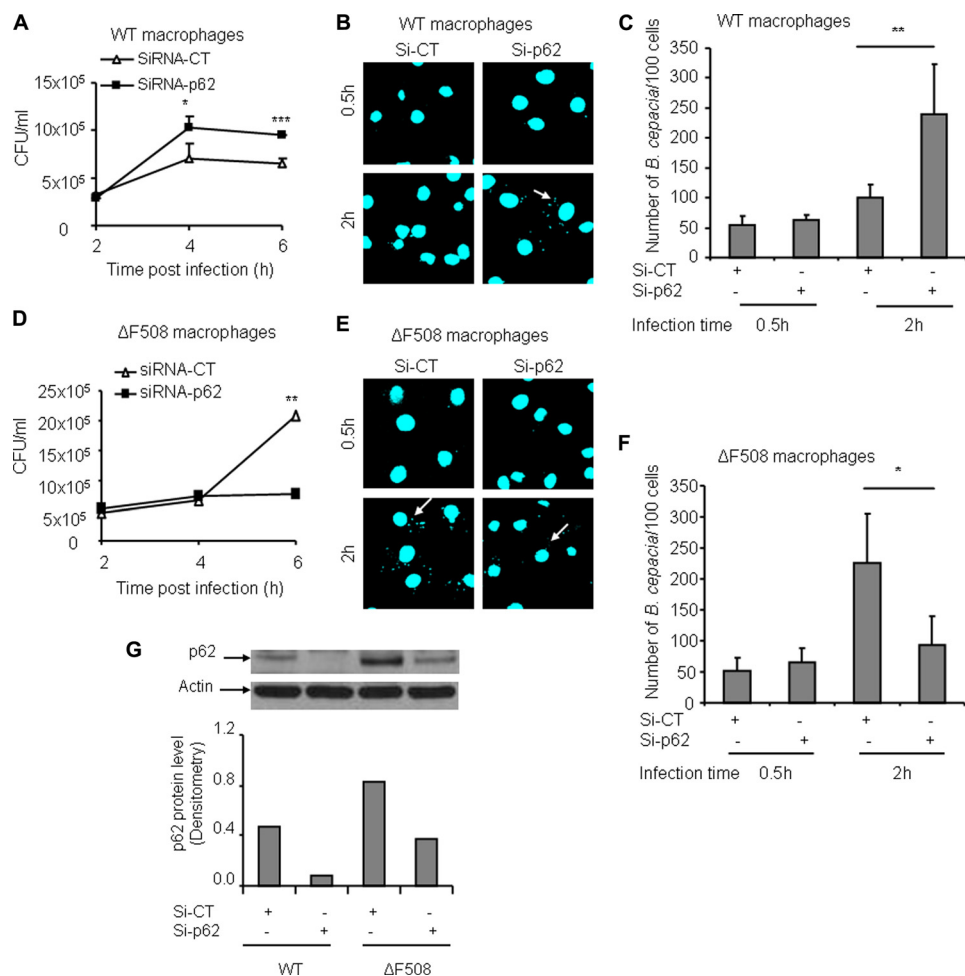


FIGURE 5. Down-regulation of p62 results in increased growth of *B. cepacia* in WT murine macrophages, whereas in Δ F508 macrophages it leads to decreased growth. A and D, WT and Δ F508 macrophages were nucleofected with siRNA against p62 (siRNA-p62) or control siRNA (siRNA-CT) 48 h prior to infection and then infected with *B. cepacia* for 2, 4, and 6 h. CFUs were enumerated. B and E, confocal microscopy of p62-depleted WT and Δ F508 macrophages infected with *B. cepacia* for 0.5 or 2 h. The white arrows show *B. cepacia* stained with DAPI. C and F, both types of macrophages were infected for 0.5 and 2 h after depletion of p62. The number of bacteria/100 cells was quantified. More than 200 cells were scored. G, upper panel, immunoblot analysis for WT and Δ F508 macrophages after 48-h nucleofection with siRNA against p62 (si-p62) or control siRNA (si-CT). Lower panel, densitometry of the p62 protein level. Data are representative of three different experiments and presented as the means \pm S.D. A, C, D, and F, *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$; significant differences from the siRNA-CT at the respective time points.

phages, a role that is compromised for unknown reasons in Δ F508 macrophages.

Depletion of p62 Liberates BECN1, Allowing Its Redistribution and Recruitment by the *B. cepacia* Vacuole in Δ F508 Macrophages—A growing body of evidence indicates that BECN1 is sequestered within the mutant CFTR aggregates (1, 2). BECN1/Atg6 is a member of the class III PI3K complex and is essential for the early stages of autophagosome formation (5, 39). Thus, its unavailability leads to defective autophagic activity (1, 2). Mutant CFTR aggregates sequester autophagy molecules such as BECN1, depleting them from their storage areas, leading to defective autophagy. We examined the colocalization of *B. cepacia* with BECN1 in WT and Δ F508 macrophages. Confocal microscopy showed that in WT macrophages, high numbers of *B. cepacia* colocalized with BECN1 compared with Δ F508 macrophages (Fig. 7, A and B, arrows). In WT macrophages, BECN1 was distributed throughout the cytosol, whereas in Δ F508 macrophages, BECN1 was condensed in patches (Fig. 7 A, arrowheads).

Because the sequestration of BECN1 in CFTR aggregates requires p62 (1, 2), we examined the effect of p62 depletion on BECN1 distribution within the cytosol and around the *B. cepacia* vacuole inside Δ F508 macrophages. Δ F508 macrophages nucleofected with p62 siRNA showed significantly more colocalization of *B. cepacia* with BECN1 compared with the siRNA control (Fig. 7, C, arrows, and D). Additionally, within the Δ F508 macrophages nucleofected with siRNA against p62, BECN1 was redistributed within the cytosol with the disappearance of BECN1-containing patches (Fig. 7C) after down-regulation of p62 (B). Notably, our immunoblot analysis using antibody specific to BECN1 showed equal amounts of the total BECN1 in the Δ F508 macrophages before and after p62 depletion (Fig. 7E). Together, these data show that depletion of p62 from Δ F508 macrophages allows the redistribution of BECN1 throughout the cell and increases its availability for the *B. cepacia*-containing vacuole.

Together, these data suggest that depletion of p62 from Δ F508 macrophages mediates *B. cepacia* clearance via recuper-

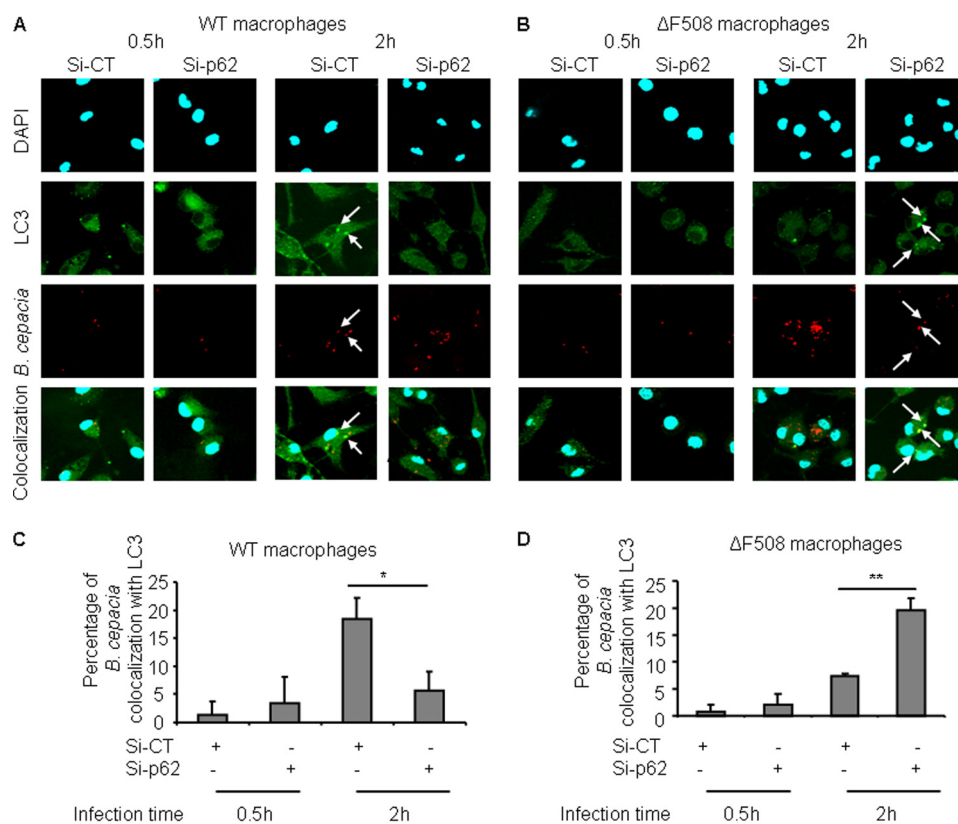


FIGURE 6. Down-regulation of p62 decreases *B. cepacia* colocalization with LC3 in WT macrophages but increases the colocalization in ΔF508 macrophages. *A* and *B*, confocal microscopy for WT macrophages and ΔF508 macrophages infected with *B. cepacia*-expressing m-RFP for 0.5 or 2 h. LC3 stained green, whereas nuclei were stained with DAPI. The percentage of colocalization of *B. cepacia* with LC3 at the indicated time points was scored in both WT and ΔF508 macrophages, respectively. *C* and *D*, more than 200 cells were scored. Data are presented as means ± S.D. *, $p < 0.05$; **, $p < 0.01$.

ated autophagy. To confirm this conclusion, ΔF508 macrophages were depleted of p62 and Atg7 (an essential autophagy molecule) (40) to disrupt the autophagy machinery and then infected with *B. cepacia*. Depletion of p62 alone from ΔF508 macrophages improved *B. cepacia* clearance, yet concomitant depletion of Atg7 hindered bacterial clearance (Fig. 7, *F* and *G*). Thus, improved bacterial clearance upon depletion of p62 from ΔF508 macrophages is mediated by autophagy.

NBR1 and NDP52 Contribute to the Delivery of B. cepacia to Autophagosomes after Down-regulation of p62 in ΔF508 Macrophages—Thus, down-regulation of p62 in ΔF508 macrophages improves *B. cepacia* clearance by restoring autophagy activity. However, what targets the *B. cepacia* vacuole for autophagy in the absence of adequate amounts of p62 is unknown. Recently, it has been shown that p62 and NDP52 act cooperatively to drive efficient antibacterial autophagy of *Salmonella*, *Shigella*, and *Listeria* (9, 41). Furthermore, it was revealed that NBR1 and p62 mark ubiquitinated cargo for autophagy (11, 42). To determine whether NDP52 or NBR1 contribute to the delivery of *B. cepacia* to autophagosomes, we nucleofected WT and ΔF508 macrophages with siRNA against NDP52 or NBR1 prior to infection with *B. cepacia*. Our results showed that in WT macrophages, down-regulation of NDP52 did not affect *B. cepacia* recovered CFUs, whereas that of NBR1 resulted in a significant increase in *B. cepacia* growth. In ΔF508 macrophages, the down-regulation of either NDP52 or NBR1 resulted in an increase in *B. cepacia* growth. This result demonstrates that both NDP52 and NBR1 facilitate the delivery of

B. cepacia to autophagosomes in ΔF508 macrophages. In WT macrophages, however, only NBR1 contributes to the delivery of *B. cepacia* to autophagosomes (Fig. 8, *A*, *B*, and *D*). To determine whether NDP52 and NBR1 mark the *B. cepacia* vacuole for autophagy uptake in p62-depleted ΔF508 macrophages, we nucleofected ΔF508 macrophages with siRNA against p62 alone or in combination with either NDP52 or NBR1. Simultaneous down-regulation of p62 and NDP52 or p62 and NBR1 in ΔF508 macrophages resulted in a significant increase in *B. cepacia* growth (Fig. 8, *C* and *E*). Together, our data suggest that both NDP52 and NBR1 contribute to labeling the *B. cepacia* vacuole for autophagy uptake when p62 is unavailable.

DISCUSSION

Human and mouse CF macrophages and airway epithelial cells exhibit impaired autophagy that is associated with formation of aggregates, including p62 and ubiquitinated mutant CFTR protein. This is due to both the self-oligomerization and ubiquitin-binding nature of p62 (1, 2, 10, 37, 38). Similar aggregates have been identified in various neurodegenerative diseases, such as Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis, and cancer (1, 2, 43, 44). Impaired turnover of p62 is a major cause of the pathogenic changes seen in the autophagy-deficient mice, as the loss of Atg7 in mouse livers results in severe p62 accumulation. Loss of p62 greatly attenuates liver injury resulting from autophagy deficiency (18).

Here, we found that ΔF508 macrophages express more p62 than WT macrophages. This could be due to p62 accumulation

p62 Dictates the Fate of *B. cepacia* in Murine Macrophages

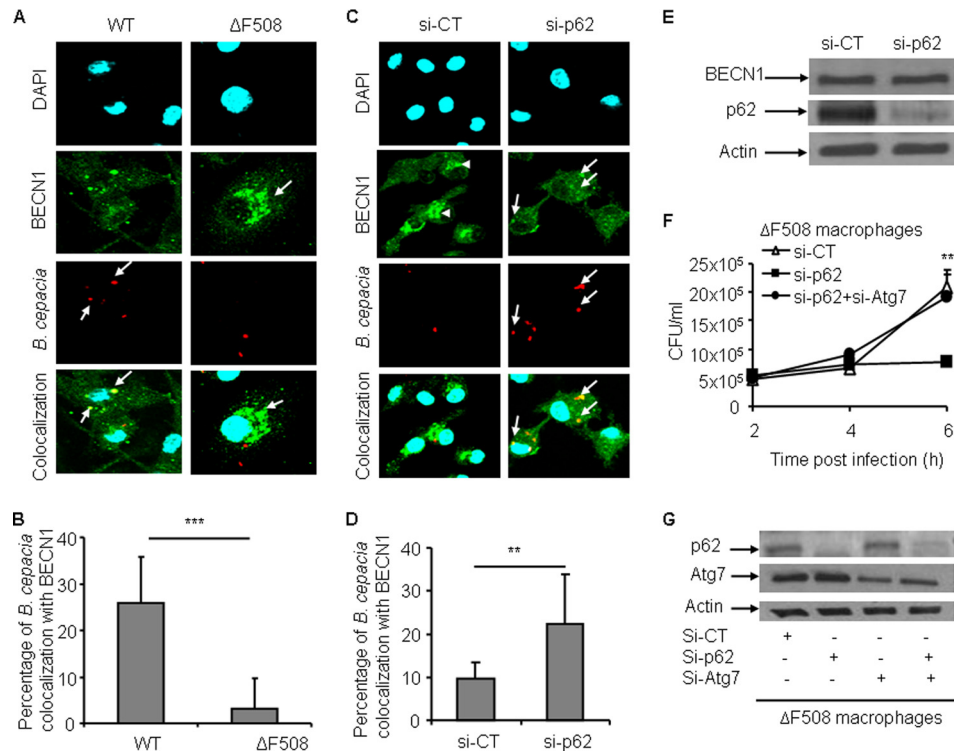


FIGURE 7. Colocalization of *B. cepacia* with BECN1 is increased in Δ F508 macrophages upon depletion of p62. *A*, confocal microscopy for WT and Δ F508 macrophages infected with *B. cepacia*-expressing m-RFP for 2 h. BECN1 stained green, and nuclei were stained with DAPI. The white arrows indicate *B. cepacia*, whereas the arrowheads indicate BECN1 aggregates. *B*, the percentage of colocalization of *B. cepacia* with BECN1 was scored by examining more than 400 cells. *C*, confocal microscopy for Δ F508 macrophages nucleofected with siRNA against p62 (*Si-p62*) or scrambled siRNA control (*si-CT*) 48 h prior to infection. Nucleofected macrophages were infected with *B. cepacia*-expressing m-RFP for 2 h. BECN1 stained green, whereas nuclei were stained with DAPI. The arrows indicate *B. cepacia*, whereas the arrowheads indicate BECN1 aggregates. *D*, the percentage of colocalization of *B. cepacia* with BECN1 at the assigned time point was scored. More than 400 bacteria were scored. *E*, immunoblot for Δ F508 macrophages nucleofected with siRNA against p62 or control siRNA for 48 h. Antibodies against p62 and BECN1 were used. *F*, Δ F508 macrophages were nucleofected with siRNA against p62 or control siRNA or siRNA against p62 and Atg7 together (*si-p62+si-Atg7*) for 48 h and then infected with *B. cepacia* for 2, 4, and 6 h. CFUs were enumerated. *G*, immunoblot analysis for Δ F508 macrophages nucleofected with control siRNA, siRNA against p62, siRNA against Atg7 (*si-Atg7*), or siRNA against p62 and Atg7 together for 48 h. Antibodies specific to p62 and Atg7 were used to detect the down-regulation. Data in *B*, *D*, and *F* are presented as means \pm S.D. of three different experiments. *B* and *D*, **, $p < 0.01$; ***, $p < 0.001$; significant differences between both types of macrophages at the designated time point.

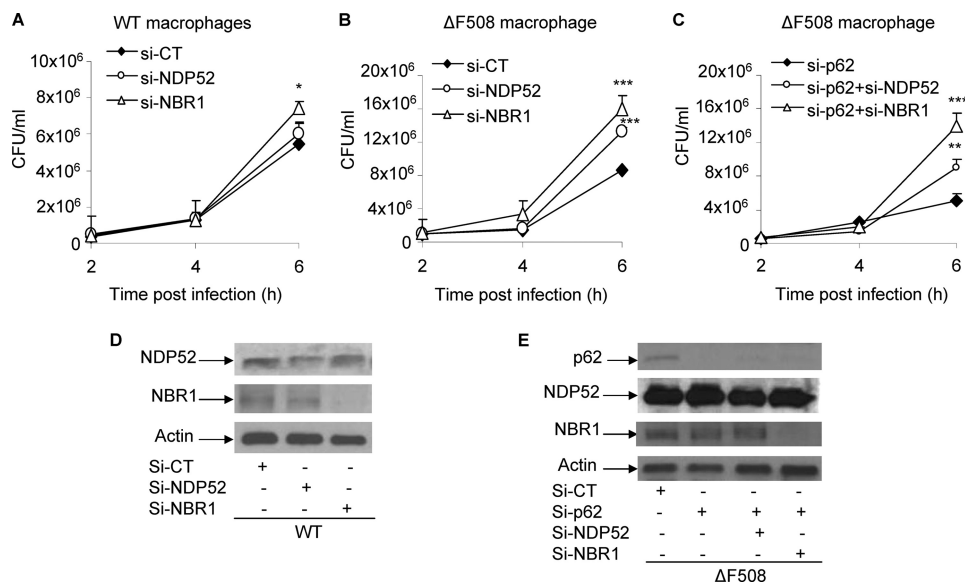


FIGURE 8. Depletion of p62 in Δ F508 macrophages improves clearance of *B. cepacia* by autophagosomes via NDP52 and NBR1. *A*, *B*, and *C*, WT (*A*) and Δ F508 (*B* and *C*) macrophages were nucleofected with siRNA against NDP52 (*si-NDP52*), NBR1 (*si-NBR1*), or control siRNA (*si-CT*) (*A* and *B*). *C*, Δ F508 macrophages nucleofected with siRNA against p62 (*si-p62*), *si-p62+si-NDP52*, or *si-p62+si-NBR1*. Macrophages in *A*, *B*, and *C* were then infected with *B. cepacia* for 2, 4, and 6 h. CFUs were quantified. *D* and *E*, Western blot analysis of macrophages treated as in *A* and *B*, respectively, with specific antibodies to NDP52, NBR1, p62, or actin. Data in *A*, *B*, and *C* are presented as the means \pm S.D. *A*, *C*, *D*, and *E*, *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$; significant differences at the respective time points.

because of reduced recycling in $\Delta F508$ macrophages as a consequence of compromised autophagosome formation and maturation. Alternatively, the accumulation of p62 could stimulate the formation of more $\Delta F508$ CFTR aggregates. This latter possibility agrees with the observation that depletion of p62 from $\Delta F508$ macrophages improves autophagy and decreases the BECN1-positive aggregates. Also, a previous study using CF epithelial cells showed that p62 promotes aggresome accumulation of misfolded or modified proteins (43, 45). Recently, it has reported that reducing the levels of p62 can rescue $\Delta F508$ -CFTR trafficking to the plasma membrane of CF airway epithelial cells (1, 2, 46).

The presence of intracellular bacteria such as *B. cepacia* increases the level of p62 expression in both WT and $\Delta F508$ macrophages. It is possible that p62 overexpression upon infection worsens the biology of $\Delta F508$ macrophages, providing an explanation for the deterioration of lung function and innate immune responses in the infected CF lung. There are several mechanisms by which *B. cepacia* may lead to the accumulation of p62. It is plausible that *B. cepacia* increases p62 accumulation by inhibiting autophagy in $\Delta F508$ macrophages, as we have published previously (37, 38). Notably, *B. cepacia* infection increases p62 mRNA. Regardless of the mechanism of p62 accumulation, the p62 aggregates sequester essential autophagy molecules such as BECN1, making them unavailable for efficient autophagosome formation (48).

The sequestration of BECN1 occurs via transglutaminase 2 (TG2)-mediated cross-linking in aggresomes because the BECN1 protein sequence contains QP and QXXP motifs, which are specific target sites for TG2 activity (48), and TG2 is an autophagy inhibitor in pancreatic adenocarcinoma cells (49). Increased reactive oxygen species in CF epithelia sustain high TG2 levels through TG2 SUMOylation (48). Thus, BECN1, and not all autophagy molecules, is specifically recruited to aggresomes in CF cells.

Examining the sequential acquisition of autophagy molecules by the *B. cepacia* vacuole revealed that although ubiquitination is efficient in both WT and $\Delta F508$ macrophages, BECN1 acquisition is defective only in $\Delta F508$ macrophages. BECN1, also known as autophagy-related gene product 6 (Atg6), and its binding partner class III PI3K (also named Vps34) are required for the initiation of the autophagosome formation (47). Thus, supplementation of p62 alone to $\Delta F508$ macrophages will not improve the targeting of *B. cepacia* to autophagosomes. This conclusion is supported by the overexpression experiment of p62 in $\Delta F508$ macrophages, which actually leads to more bacterial growth. Therefore, to correct the trafficking defect of *B. cepacia* in $\Delta F508$ macrophages, "free" BECN1 is required, which is achieved by depletion of p62.

p62 targets several pathogens, such as *S. typhimurium*, *Shigella*, and *Listeria* to the autophagosome (9, 10). Similarly, p62 associates with the *B. cepacia* vacuole in WT macrophages. However, depletion of p62 from $\Delta F508$ macrophages promotes *B. cepacia* uptake by autophagosomes and decreases the bacterial burden. It is possible that another adaptor molecule, such as NBR1, compensates for the loss of p62. The structure of NBR1 resembles that of p62. It can bind both LC3 and ubiquitinated proteins through the LC3 interaction region and ubiquitin-as-

sociated domain, respectively (11, 14). NDP52 is another cargo marker that drives certain bacteria to the autophagy machinery (9, 41). In this study, we found that NDP52 facilitates autophagy uptake of *B. cepacia* in $\Delta F508$ macrophages but not in WT cells. NBR1, however, appears to contribute to the delivery of *B. cepacia* to the autophagy machinery in both WT and $\Delta F508$ macrophages. To our knowledge, this is the first demonstration of a role for NBR1 in bacterial targeting by autophagy.

We showed previously (37) that autophagy stimulation by rapamycin can overcome the down-regulating effect of *B. cepacia* on the ATG genes and can control the *B. cepacia* infection in the $\Delta F508$ mouse model both *in vivo* and *in vitro*. In this work, we demonstrate that p62 depletion from $\Delta F508$ mouse macrophages is another approach to improve autophagic control on *B. cepacia* infection.

Together, these data provide a molecular framework to better understand the emerging complexity of diseases related to autophagic defect such as CF and the ability of macrophages to defend against the bacterial infection. This study also identifies p62 as a promising drug target for improving *B. cepacia* clearance in CF macrophages.

REFERENCES

- Luciani, A., Vilella, V. R., Esposito, S., Brunetti-Pierri, N., Medina, D., Settembre, C., Gavina, M., Pulze, L., Giardino, I., Pettoello-Mantovani, M., D'Apolito, M., Guido, S., Masliah, E., Spencer, B., Quarantino, S., Raia, V., Ballabio, A., and Maiuri, L. (2010) Defective CFTR induces aggresome formation and lung inflammation in cystic fibrosis through ROS-mediated autophagy inhibition. *Nat. Cell Biol.* **12**, 863–875
- Luciani, A., Vilella, V. R., Esposito, S., Brunetti-Pierri, N., Medina, D. L., Settembre, C., Gavina, M., Raia, V., Ballabio, A., and Maiuri, L. (2011) Cystic fibrosis. A disorder with defective autophagy. *Autophagy* **7**, 104–106
- Knorre, A., Wagner, M., Schaefer, H. E., Colledge, W. H., and Pahl, H. L. (2002) $\Delta F508$ -CFTR causes constitutive NF- κ B activation through an ER-overload response in cystic fibrosis lungs. *Biol. Chem.* **383**, 271–282
- Witko-Sarsat, V., Sermet-Gaudelus, I., Lenoir, G., and Descamps-Latscha, B. (1999) Inflammation and CFTR. Might neutrophils be the key in cystic fibrosis? *Mediators Inflamm.* **8**, 7–11
- Deretic, V. (2010) Autophagy in infection. *Curr. Opin. Cell Biol.* **22**, 252–262
- Amer, A. O., Byrne, B. G., and Swanson, M. S. (2005) Macrophages rapidly transfer pathogens from lipid raft vacuoles to autophagosomes. *Autophagy* **1**, 53–58
- Yang, Z., and Klionsky, D. J. (2010) Mammalian autophagy. Core molecular machinery and signaling regulation. *Curr. Opin. Cell Biol.* **22**, 124–131
- Mizushima, N., and Levine, B. (2010) Autophagy in mammalian development and differentiation. *Nat. Cell Biol.* **12**, 823–830
- Mostowy, S., Sancho-Shimizu, V., Hamon, M. A., Simeone, R., Brosch, R., Johansen, T., and Cossart, P. (2011) p62 and NDP52 proteins target intracytosolic *Shigella* and *Listeria* to different autophagy pathways. *J. Biol. Chem.* **286**, 26987–26995
- Zheng, Y. T., Shahnazari, S., Brech, A., Lamark, T., Johansen, T., and Brumell, J. H. (2009) The adaptor protein p62/SQSTM1 targets invading bacteria to the autophagy pathway. *J. Immunol.* **183**, 5909–5916
- Lamark, T., Kirkin, V., Dikic, I., and Johansen, T. (2009) NBR1 and p62 as cargo receptors for selective autophagy of ubiquitinated targets. *Cell Cycle* **8**, 1986–1990
- Kirkin, V., Lamark, T., Johansen, T., and Dikic, I. (2009) NBR1 cooperates with p62 in selective autophagy of ubiquitinated targets. *Autophagy* **5**, 732–733
- Kirkin, V., Lamark, T., Sou, Y. S., Bjørkøy, G., Nunn, J. L., Bruun, J. A., Shvets, E., McEwan, D. G., Clausen, T. H., Wild, P., Bilusic, I., Theurillat,

p62 Dictates the Fate of *B. cepacia* in Murine Macrophages

- J. P., Øvervatn, A., Ishii, T., Elazar, Z., Komatsu, M., Dikic, I., and Johansen, T. (2009) A role for NBR1 in autophagosomal degradation of ubiquitinated substrates. *Mol. Cell* **33**, 505–516
14. Kirkin, V., McEwan, D. G., Novak, I., and Dikic, I. (2009) A role for ubiquitin in selective autophagy. *Mol. Cell* **34**, 259–269
 15. Hocking, L. J., Lucas, G. J., Daroszewska, A., Mangion, J., Olavesen, M., Cundy, T., Nicholson, G. C., Ward, L., Bennett, S. T., Wuyts, W., Van Hul, W., and Ralston, S. H. (2002) Domain-specific mutations in sequestosome 1 (SQSTM1) cause familial and sporadic Paget's disease. *Hum. Mol. Genet.* **11**, 2735–2739
 16. Laurin, N., Brown, J. P., Morissette, J., and Raymond, V. (2002) Recurrent mutation of the gene encoding sequestosome 1 (SQSTM1/p62) in Paget disease of bone. *Am. J. Hum. Genet.* **70**, 1582–1588
 17. Komatsu, M., and Ichimura, Y. (2010) Physiological significance of selective degradation of p62 by autophagy. *FEBS Lett.* **584**, 1374–1378
 18. Komatsu, M., Waguri, S., Koike, M., Sou, Y. S., Ueno, T., Hara, T., Mizushima, N., Iwata, J., Ezaki, J., Murata, S., Hamazaki, J., Nishito, Y., Iemura, S., Natsume, T., Yanagawa, T., Uwayama, J., Warabi, E., Yoshida, H., Ishii, T., Kobayashi, A., Yamamoto, M., Yue, Z., Uchiyama, Y., Kominami, E., and Tanaka, K. (2007) Homeostatic levels of p62 control cytoplasmic inclusion body formation in autophagy-deficient mice. *Cell* **131**, 1149–1163
 19. Nezis, I. P., Simonsen, A., Sagona, A. P., Finley, K., Gaumer, S., Contamine, D., Rusten, T. E., Stenmark, H., and Brech, A. (2008) Ref(2)P, the *Drosophila melanogaster* homologue of mammalian p62, is required for the formation of protein aggregates in adult brain. *J. Cell Biol.* **180**, 1065–1071
 20. Kotrange, S., Kopp, B., Akhter, A., Abdelaziz, D., Abu Khweek, A., Caution, K., Abdulrahman, B., Wewers, M. D., McCoy, K., Marsh, C., Loutet, S. A., Ortega, X., Valvano, M. A., and Amer, A. O. (2011) *Burkholderia cenocepacia* O polysaccharide chain contributes to caspase-1-dependent IL-1 β production in macrophages. *J. Leukocyte Biol.* **89**, 481–488
 21. Orvedahl, A., and Levine, B. (2009) Eating the enemy within. Autophagy in infectious diseases. *Cell Death Differ.* **16**, 57–69
 22. Nakagawa, I., Amano, A., Mizushima, N., Yamamoto, A., Yamaguchi, H., Yoshimoto, T., Nara, A., Funao, J., Nakata, M., Tsuda, K., Hamada, S., and Yamimori, T. (2004) Autophagy defends cells against invading group A *Streptococcus*. *Science* **306**, 1037–1040
 23. Loutet, S. A., and Valvano, M. A. (2010) A decade of *Burkholderia cenocepacia* virulence determinant research. *Infect. Immun.* **78**, 4088–4100
 24. Saldías, M. S., and Valvano, M. A. (2009) Interactions of *Burkholderia cenocepacia* and other *Burkholderia cepacia* complex bacteria with epithelial and phagocytic cells. *Microbiology* **155**, 2809–2817
 25. Birmingham, C. L., Smith, A. C., Bakowski, M. A., Yoshimori, T., and Brumell, J. H. (2006) Autophagy controls *Salmonella* infection in response to damage to the *Salmonella*-containing vacuole. *J. Biol. Chem.* **281**, 11374–11383
 26. McCloskey, M., McCaughan, J., Redmond, A. O., and Elborn, J. S. (2001) Clinical outcome after acquisition of *Burkholderia cepacia* in patients with cystic fibrosis. *Ir. J. Med. Sci.* **170**, 28–31
 27. Tolman, J. S., and Valvano, M. A. (2012) Global changes in gene expression by the opportunistic pathogen *Burkholderia cenocepacia* in response to internalization by murine macrophages. *BMC Genomics* **13**, 63
 28. Rosales-Reyes, R., Skeldon, A. M., Aubert, D. F., and Valvano, M. A. (2012) The Type VI secretion system of *Burkholderia cenocepacia* affects multiple Rho family GTPases disrupting the actin cytoskeleton and the assembly of NADPH oxidase complex in macrophages. *Cell. Microbiol.* **14**, 255–273
 29. Hamad, M. A., Skeldon, A. M., and Valvano, M. A. (2010) Construction of aminoglycoside-sensitive *Burkholderia cenocepacia* strains for use in studies of intracellular bacteria with the gentamicin protection assay. *Appl. Environ. Microbiol.* **76**, 3170–3176
 30. Gavrilin, M. A., Bouakl, I. J., Knatz, N. L., Duncan, M. D., Hall, M. W., Gunn, J. S., and Wewers, M. D. (2006) Internalization and phagosome escape required for *Francisella* to induce human monocyte IL-1 β processing and release. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 141–146
 31. Gavrilin, M. A., Mitra, S., Seshadri, S., Nateri, J., Berhe, F., Hall, M. W., and Wewers, M. D. (2009) Pyrin critical to macrophage IL-1 β response to *Francisella* challenge. *J. Immunol.* **182**, 7982–7989
 32. Fan, W., Tang, Z., Chen, D., Moughon, D., Ding, X., Chen, S., Zhu, M., and Zhong, Q. (2010) Keap1 facilitates p62-mediated ubiquitin aggregate clearance via autophagy. *Autophagy* **6**, 416–421
 33. Fahy, R. J., Exline, M. C., Gavrilin, M. A., Bhatt, N. Y., Besecker, B. Y., Sarkar, A., Hollyfield, J. L., Duncan, M. D., Nagaraja, H. N., Knatz, N. L., Hall, M., and Wewers, M. D. (2008) Inflammasome mRNA expression in human monocytes during early septic shock. *Am. J. Respir. Crit. Care Med.* **177**, 983–988
 34. Amer, A., Franchi, L., Kanneganti, T. D., Body-Malapel, M., Ozören, N., Brady, G., Meshinchi, S., Jagirdar, R., Gewirtz, A., Akira, S., and Núñez, G. (2006) Regulation of *Legionella* phagosome maturation and infection through flagellin and host Ipaf. *J. Biol. Chem.* **281**, 35217–35223
 35. Akhter, A., Gavrilin, M. A., Frantz, L., Washington, S., Ditty, C., Limoli, D., Day, C., Sarkar, A., Newland, C., Butchar, J., Marsh, C. B., Wewers, M. D., Tridandapani, S., Kanneganti, T. D., and Amer, A. O. (2009) Caspase-7 activation by the Nlr4/Ipaf inflammasome restricts *Legionella pneumophila* infection. *PLoS Pathog.* **5**, e1000361
 36. Wang, C. W., and Klionsky, D. J. (2003) The molecular mechanism of autophagy. *Mol. Med.* **9**, 65–76
 37. Abdulrahman, B. A., Khweek, A. A., Akhter, A., Caution, K., Kotrange, S., Abdelaziz, D. H., Newland, C., Rosales-Reyes, R., Kopp, B., McCoy, K., Montione, R., Schlesinger, L. S., Gavrilin, M. A., Wewers, M. D., Valvano, M. A., and Amer, A. O. (2011) Autophagy stimulation by rapamycin suppresses lung inflammation and infection by *Burkholderia cenocepacia* in a model of cystic fibrosis. *Autophagy* **7**, 1359–1370
 38. Devenish, R. J. (2011) Autophagy and the evasion of host defense: a new variation on the theme for *Burkholderia cepacia*? *Autophagy* **7**, 1269–1270
 39. Vergne, I., and Deretic, V. (2010) The role of PI3P phosphatases in the regulation of autophagy. *FEBS Lett.* **584**, 1313–1318
 40. Zhang, J., Randall, M. S., Loyd, M. R., Dorsey, F. C., Kundu, M., Cleveland, J. L., and Ney, P. A. (2009) Mitochondrial clearance is regulated by Atg7-dependent and -independent mechanisms during reticulocyte maturation. *Blood* **114**, 157–164
 41. Cemba, M., Kim, P. K., and Brumell, J. H. (2011) The ubiquitin-binding adaptor proteins p62/SQSTM1 and NDP52 are recruited independently to bacteria-associated microdomains to target *Salmonella* to the autophagy pathway. *Autophagy* **7**, 341–345
 42. Johansen, T., and Lamark, T. (2011) Selective autophagy mediated by autophagic adapter proteins. *Autophagy* **7**, 279–296
 43. Mathew, R., Karp, C. M., Beaudoin, B., Vuong, N., Chen, G., Chen, H. Y., Bray, K., Reddy, A., Bhanot, G., Gelinas, C., Dipaola, R. S., Karantza-Wadsworth, V., and White, E. (2009) Autophagy suppresses tumorigenesis through elimination of p62. *Cell* **137**, 1062–1075
 44. Kuusisto, E., Salminen, A., and Alafuzoff, I. (2001) Ubiquitin-binding protein p62 is present in neuronal and glial inclusions in human tauopathies and synucleinopathies. *Neuroreport* **12**, 2085–2090
 45. Moscat, J., and Diaz-Meco, M. T. (2009) p62 at the crossroads of autophagy, apoptosis, and cancer. *Cell* **137**, 1001–1004
 46. Luciani, A., Vilella, V. R., Esposito, S., Gavina, M., Russo, I., Silano, M., Guido, S., Pettoello-Mantovani, M., Carnuccio, R., Scholte, B., De Matteis, A., Maiuri, M. C., Raia, V., Luini, A., Kroemer, G., and Maiuri, L. (2012) Targeting autophagy as a novel strategy for facilitating the therapeutic action of potentiators on $\Delta F508$ cystic fibrosis transmembrane conductance regulator. *Autophagy* **8**, 1–16
 47. Erlich, S., Mizrachy, L., Segev, O., Lindenboim, L., Zmira, O., Adi-Harel, S., Hirsch, J. A., Stein, R., and Pinkas-Kramarski, R. (2007) Differential interactions between Beclin 1 and Bcl-2 family members. *Autophagy* **3**, 561–568
 48. Lorand, L., and Graham, R. M. (2003) Transglutaminases. Crosslinking enzymes with pleiotropic functions. *Nat. Rev. Mol. Cell Biol.* **4**, 140–156
 49. Akar, U., Ozpolat, B., Mehta, K., Fok, J., Kondo, Y., and Lopez-Berestein, G. (2007) Tissue transglutaminase inhibits autophagy in pancreatic cancer cells. *Mol. Cancer Res.* **5**, 241–249
 50. Nezis, I. P., and Papassideri, I. (2008) Monitoring autophagy in insect eggs. *Methods Enzymol.* **451**, 669–683