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Alternatively spliced tissue factor pathway inhibitor: Functional implications

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Abstract

Tissue factor pathway inhibitor (TFPI) is a factor Xa dependent inhibitor of tissue factor initiated blood coagulation. In recent years several alternatively spliced forms of TFPI have been identified. These alternatively spliced forms have different C-terminal regions and have different mechanisms for association with cell surfaces. They are differentially expressed in human and mouse tissues and may have distinct physiological functions.

Keywords

Tissue factor pathway inhibitor; alternative splicing; factor VIIa; factor Xa; Review

2. INTRODUCTION

Tissue factor pathway inhibitor (TFPI) is an anticoagulant protein found primarily on the surface of endothelium¹, but also within platelets^{2, 3} and circulating in plasma⁴. TFPI directly inhibits the tissue factor-factor VIIa (TF-fVIIa) catalytic complex that initiates blood coagulation. The *in vivo* importance of TFPI as an inhibitor of TF-fVIIa activity has been demonstrated through studies of mice lacking TF, fVIIa and TFPI. TFPI null mice die during embryogenesis from consumptive coagulopathy⁵. This embryonic lethal phenotype is rescued by breeding TFPI null mice into mice with low amounts of TF⁶ or into mice lacking fVIIa⁷, thereby demonstrating that TFPI directly counterbalances TF-fVIIa activity. The initial structural studies of TFPI described a protein containing an acidic N-terminal region, three Kunitz-type serine protease inhibitor domains and a highly basic C-terminal region.⁸ Functional studies identified the second Kunitz domain as a direct inhibitor of factor Xa (fXa) and the first Kunitz domain as a fXa dependent inhibitor of TF-fVIIa⁹. An inhibitory function for the third Kunitz domain has not yet been identified. Since its initial characterization, alternatively spliced forms of TFPI have been identified that are differentially expressed during mouse development and may have distinct physiological functions^{10–13}.

3. ALTERNATIVELY SPLICED FORMS OF TFPI

TFPI is produced in four alternatively spliced isoforms¹⁴ (Figures 1–4). Each isoform has the acidic N-terminal region and the first two Kunitz domains that are responsible for TFPI anticoagulant activity. However, they differ in the domain structure of their C-terminal regions and their mechanism of association with cell surfaces.

TFPIalpha is the full-length form of TFPI. It has all three Kunitz domains and a basic amino acid rich C-terminal region. The C-terminal region of human TFPIalpha is more basic than mouse TFPIalpha, with human having 14 basic amino acids, while mice have only nine. The third Kunitz domain and/or the basic C-terminal region may be important in mediating binding of TFPIalpha to endothelium¹⁵. TFPIalpha associates with the endothelium surface in two ways. About 90% is indirectly bound through an, as yet, unidentified GPI-anchored protein^{16, 17}. The association of TFPIalpha with endothelium through a GPI-anchored protein localizes TFPI to caveolae, where it may interact with caveolin-1 which increases its surface expression and anticoagulant activity¹⁸. The remaining 10% of TFPIalpha is nonspecifically bound to cell surface glycosaminoglycans¹⁷. In humans, the glycosaminoglycan bound TFPI alpha is released into plasma following heparin infusion causing the plasma TFPI concentration to increase 2- to 4-fold^{19, 20}.

TFPIbeta is a C-terminally truncated form of TFPI¹⁰. It has the first two Kunitz domains. Alternative splicing occurs just prior to the third Kunitz domain and produces a sequence encoding a GPI-anchor attachment site and, consequently, TFPIbeta directly associates with endothelium^{10, 11}. After processing of the C-terminal region to attach the GPI-anchor, human TFPIbeta protein has 12 amino acids not present in TFPIalpha, while mouse TFPIbeta has eight. Thus far, attempts to make high affinity antibodies that recognize the unique C-terminal region of TFPIbeta have not been successful making it difficult to directly identify TFPIbeta in tissues.

TFPIgamma is a C-terminally truncated form of TFPI found only in mice¹². It has the first two Kunitz domains. Alternatively splicing occurs at the same 5' splice acceptor site as TFPIbeta, just prior to the third Kunitz domain, but the 3' splice acceptor site is 276 nucleotides past the TFPIbeta stop codon within the TFPIbeta 3' untranslated region¹². This produces a protein sequence with 18 amino acids not present in TFPIalpha or TFPIbeta. As with TFPIbeta, attempts to make high affinity antibodies that recognize the unique C-terminal region of TFPIgamma have not been successful making it difficult to directly identify TFPIgamma in tissues. The C-terminal region of TFPIgamma does not encode a predicted GPI-anchor attachment sequence and following transfection into CHO cells, it is processed as a secreted protein¹².

TFPIDelta is a C-terminally truncated form of TFPI. Sequences are present within the NCBI GenBank database, but no other information about this isoform has been published to date. Alternative splicing occurs immediately following the second Kunitz domain and a sequence encoding a new C-terminal region of 12 amino acids is present.

4. EVOLUTION OF ALTERNATIVELY SPLICED ISOFORMS OF TFPI

TFPIalpha is well conserved from man to zebra fish who last shared a common ancestor 430 million years ago. The third Kunitz domain has maintained a similar high degree of sequence identity as the first and second Kunitz domains suggesting it has an important biological function¹³. In contrast, TFPIbeta-specific sequence has been identified in humans, other primates, mice, and rats¹³. This is not surprising because TFPIbeta sequence conservation between man and mouse is fairly poor (43%) with the exception of the 7 amino acid region proximal to the predicted GPI-anchor modification site. Analysis of zebrafish genomic sequence between the exons encoding K2 and K3 revealed no potential alternative exon that might encode the TFPIbeta specific or the TFPIgamma specific sequence. This suggests that TFPIbeta represents a 'recent' evolutionary adaptation whereas TFPIalpha existed prior to the divergence of the boney fish over 430 million years ago.

5. TISSUE EXPRESSION OF ALTERNATIVELY SPLICED FORMS OF TFPI

Examination of the expression of TFPIalpha and TFPIbeta mRNA in human and mouse tissues and endothelial cell lines using real time PCR revealed that each tissue has more message for TFPIalpha than TFPIbeta. This ranges from 4- to 50-fold with message for TFPIalpha on average 10-fold more abundant than message for TFPIbeta^{11, 12}. At the level of protein production, it appears that TFPIalpha is the major TFPI isoform produced in humans. TFPIalpha protein has been identified in human platelets³ and placenta¹⁷. As mentioned, human plasma contains TFPIalpha and the heparin-releasable form of TFPI is TFPIalpha²⁰. TFPI beta protein has not been identified in humans. The lack of a high affinity antibody that recognizes TFPIbeta but not TFPIalpha makes it difficult to differentiate TFPIbeta from partially degraded forms of TFPIalpha. Further studies are needed to definitively define the alternatively spliced isoforms of TFPI produced in different human vascular beds.

In mice, the placenta and embryo produce both TFPIalpha and TFPIbeta protein with placenta producing much more TFPIalpha than TFPIbeta and embryo producing approximately equal amounts of each¹³. However, as mice mature, the production of TFPIalpha decreases and adult mice produce predominantly TFPIbeta protein in all major vascular beds¹³. Mouse plasma contains essentially only TFPIbeta and heparin infusion has only minimal effect on the plasma TFPI concentration¹³. This finding is consistent with the structure of TFPIbeta, which lacks the basic C-terminal region responsible for association of TFPIalpha with glycosaminoglycans in human vasculature. Interestingly, it appears that mouse platelets make only TFPIalpha (Maroney SA et al., ATVB 2011 In Press). Thus, adult mice produce TFPIbeta in all tissues except for platelets that make TFPIalpha. TFPIgamma and TFPIdelta protein have thus far not been identified *in vivo*¹². Since TFPIalpha message is more abundant than TFPIbeta message in adult mouse tissues, control of TFPI protein production in mice occurs during mRNA translation.

6. PERSPECTIVE: POTENTIAL FOR DIFFERENTIAL FUNCTION OF ALTERNATIVELY SPLICED ISOFORMS OF TFPI

There is accumulating evidence that alternative splicing of TFPI produces physiologically relevant changes in TFPI activity. While all TFPI isoforms have the first two Kunitz domains and are capable of inhibiting TF-fVIIa and fXa catalytic activity, it is unclear whether they have relative equal inhibitory activity *in vivo* or whether they differentially inhibit TF procoagulant and pro-inflammatory activities. The evolutionary conservation of TFPIalpha and its conserved production by mouse embryo, placenta and platelets suggests that the third Kunitz domain and basic C-terminal region may have specific functions during vascular development and/or wound healing not performed by the other isoforms. The third Kunitz domain and C-terminal region of soluble TFPIalpha directly interact with fXa²¹, perhaps partially explaining the enhanced anticoagulant activity of TFPIalpha compared to truncated forms of TFPI in solution phase assays²²⁻²⁴. The anticoagulant activity of TFPIalpha is enhanced through interactions between the third Kunitz domain and protein S that produces enhanced inhibition of fXa by the second Kunitz domain^{25, 26}, but not of TF-fVIIa by the first Kunitz domain in solution phase assays²⁷. Further studies are needed to determine how protein S may enhance the activity of surface associated TFPIalpha. This enhancing effect of protein S on inhibition of fXa was not observed in mouse plasma, a finding that is consistent with TFPIbeta as the major alternatively spliced form of TFPI in adult mice.

Several studies using *in vitro* and *in vivo* assays have reported physiological activities of peptides corresponding to the basic C-terminal region of TFPI that occur independent of

TFPI anticoagulant activity further demonstrating that TFPI α may have unique functions not performed by the other isoforms. These include studies demonstrating that the TFPI α C-terminal peptide inhibits endothelial cell proliferation *in vitro* as well as primary and metastatic tumor growth *in vivo*²⁸. The C-terminal peptide has also been shown to have complement dependent antibacterial activity^{29, 30}. It is important to note that these studies have been performed using soluble forms of TFPI. Yet, most TFPI within the body is associated with cell surfaces which can greatly alter its activity. Further studies of the biological activity of the different alternatively spliced isoforms associated with cell surfaces are needed to understand their physiological functions.

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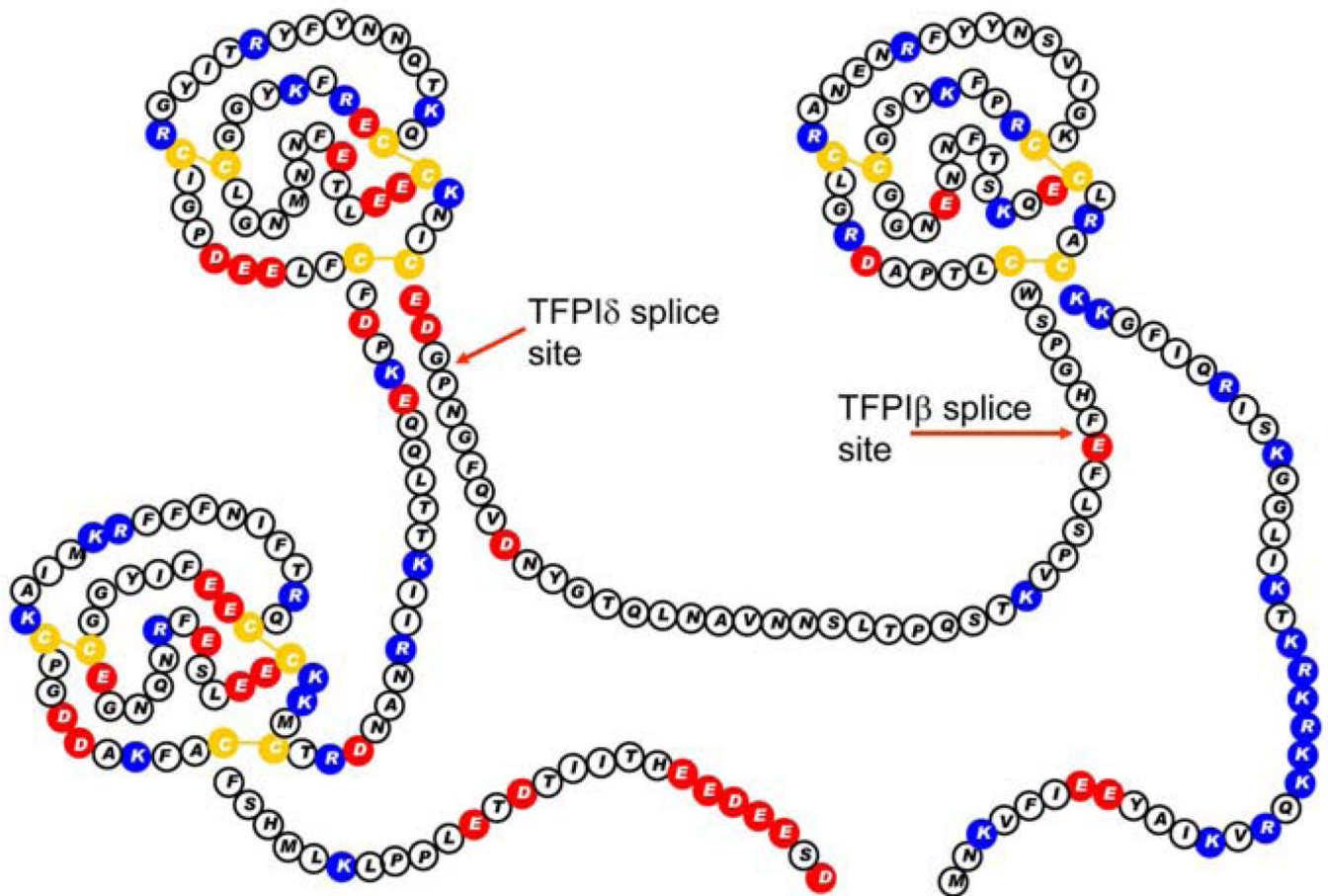
Human TFPI α 

Figure 1.
The amino acid sequence and Kunitz domain structure of human TFPI alpha. Red circles indicate acidic amino acids. Blue circles indicate basic amino acids. Yellow circles are cysteine residues.

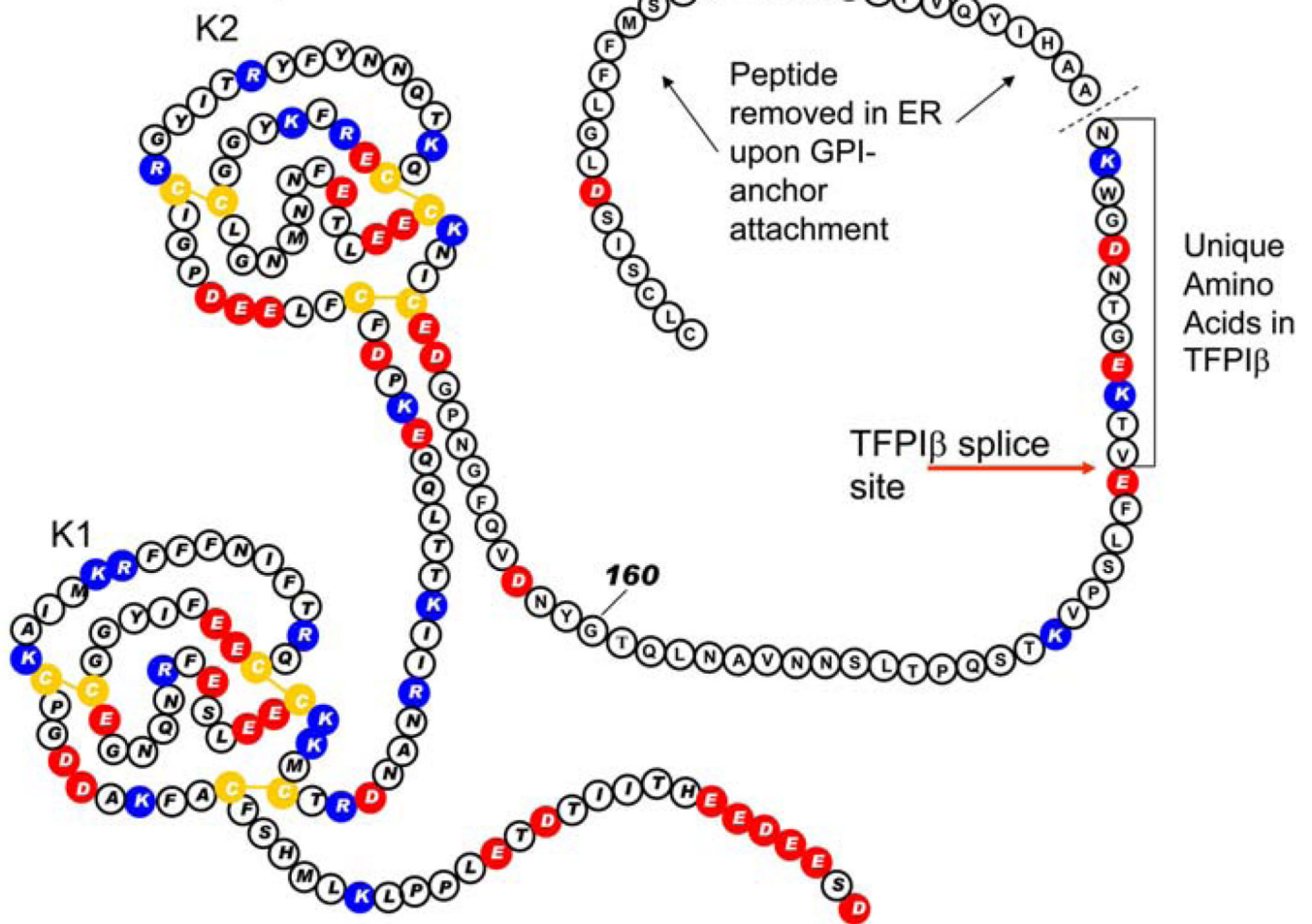
Human TFPI β 

Figure 2.

The amino acid sequence and Kunitz domain structure of human TFPI beta. Red circles indicate acidic amino acids. Blue circles indicate basic amino acids. Yellow circles are cysteine residues.

Human TFPI δ

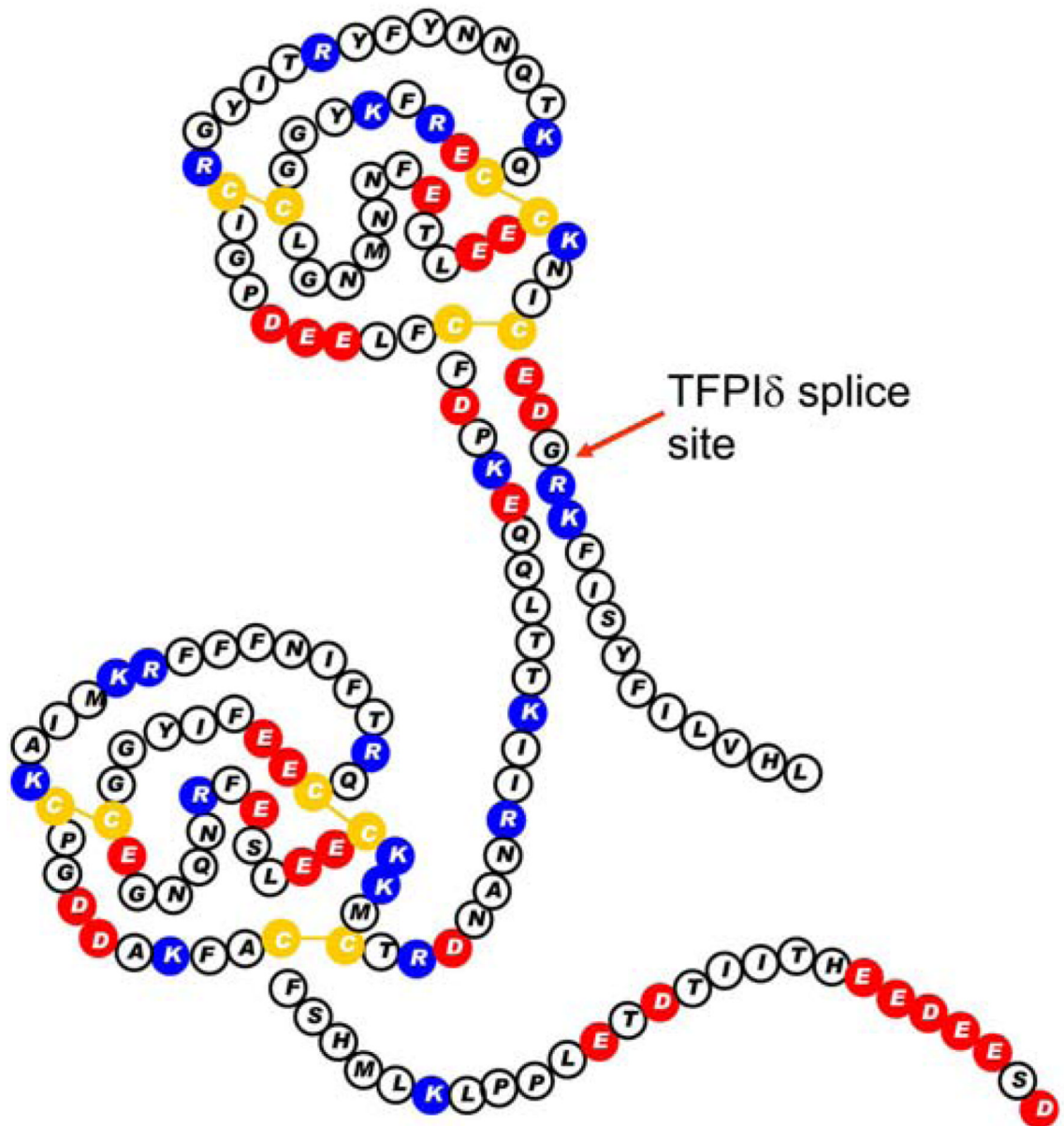


Figure 3.

The amino acid sequence and Kunitz domain structure of human TFPI gamma. Red circles indicate acidic amino acids. Blue circles indicate basic amino acids. Yellow circles are cysteine residues.

Mouse TFPI_γ

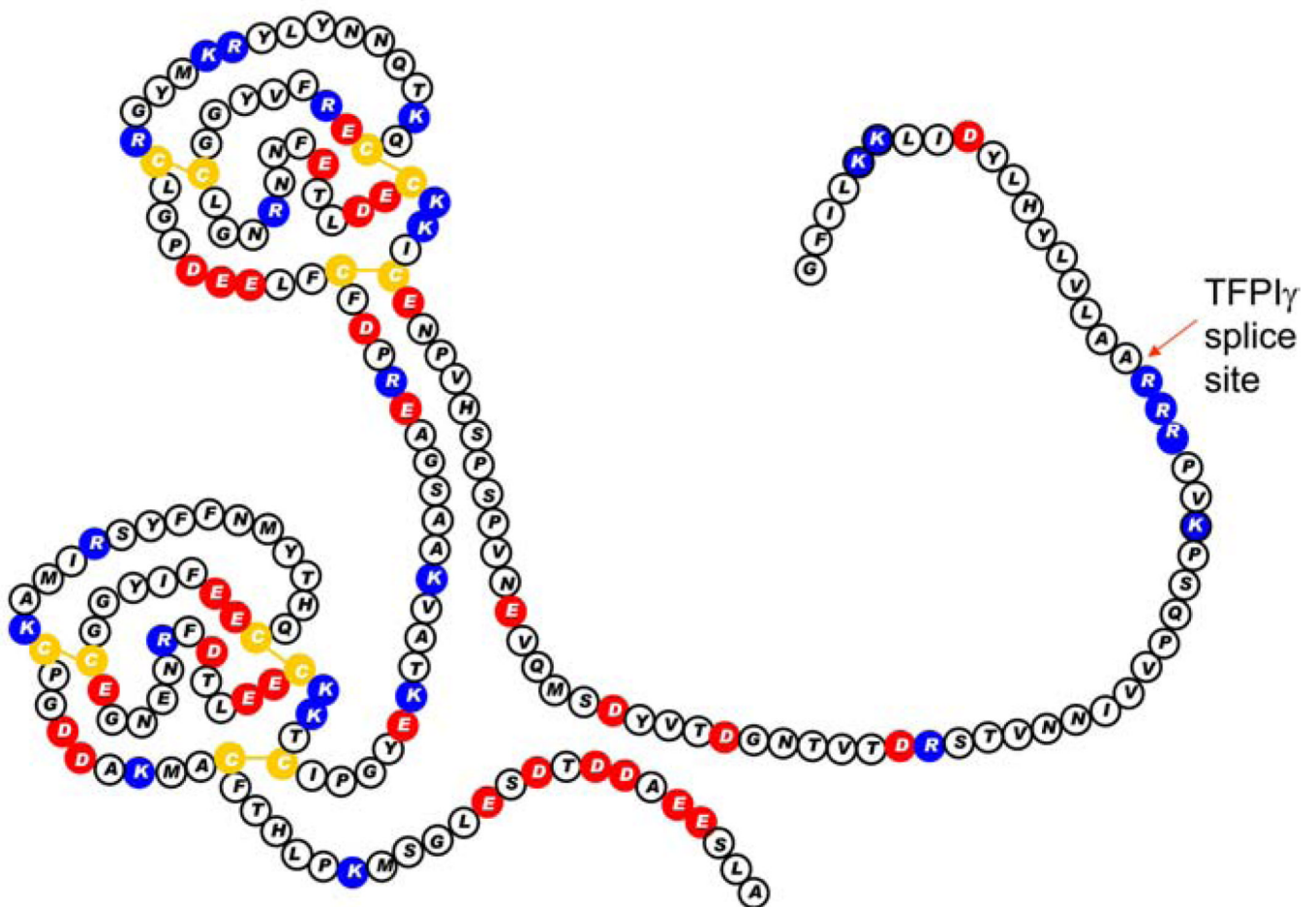


Figure 4.

The amino acid sequence and Kunitz domain structure of mouse TFPI delta. Red circles indicate acidic amino acids. Blue circles indicate basic amino acids. Yellow circles are cysteine residues.