

DEBATE

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Debate around infection-dependent hemophagocytic syndrome in paediatrics

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Abstract

Background: Hemophagocytic syndrome (HPS) is clinically defined as a combination of fever, liver dysfunction, coagulation abnormalities, pancytopenia, progressive macrophage proliferation throughout the reticuloendothelial system, and cytokine over-production, and may be primary or secondary to infectious, auto-immune, and tumoral diseases. The most consistent association is with viral infections but, as it is still debated whether any micro-organisms are involved in its pathogenesis, we critically appraised the literature concerning HPS and its relationship with infections.

Discussion: Infection-dependent HPS has been widely observed, but there are no data concerning its incidence in children. A better understanding of the pathophysiology of HPS may clarify the interactions between the immune system and the variously implicated potential infectious agents. Epstein-Barr virus (EBV) infection has been prominently associated with HPS, with clonal proliferation and the hyperactivation of EBV-infected T cells. However, a number of other viral, bacterial, fungal, and parasitic infections have been reported in association with HPS. In the case of low-risk HPS, corticosteroids and/or intravenous immunoglobulin or cyclosporine A may be sufficient to control the biological process, but etoposide is recommended as a means of reversing infection-dependent lymphohistiocytic dysregulation in high-risk cases.

Summary: HPS is a potential complication of various infections. A polymerase chain reaction search for infectious agents including EBV, cytomegalovirus and *Leishmania* is recommended in clinical settings characterised by non-remitting fever, organomegaly, cytopenia and hyperferritinemia.

Background

Hemophagocytic syndrome (HPS) is a potentially fatal condition due to dysregulated lymphocyte activation and proliferation, mainly characterised by impaired or inactive natural killer (NK) cells and cytotoxic T cells, which leads to macrophage hyperactivation and over-expression of cytokines [1]. The result of this process is uncontrolled and ineffective immune activation, multi-organ dysfunction, and hemophagocytosis throughout the reticuloendothelial system [2]. The pathognomonic characteristic of HPS is the activation of well-differentiated macrophages, phagocytosing erythrocytes, leukocytes and platelets in bone marrow, lymph nodes, spleen, liver and other organs, which can infiltrate almost anybody district and may

account for many of its systemic features [3]. HSP is still often under-diagnosed and sub-optimally managed in children [4], but the epidemiological data are fragmentary.

The syndrome was first described in 1939 as poorly-controlled histiocyte proliferation, but has since also been called hemophagocytic histiocytosis and macrophage activation syndrome [5-7]. It can be divided into a primary genetic form and a secondary reactive form (Table 1), a distinction that has historically been used to differentiate cases of often fatal infantile HPS from those caused by other etiologies that appear later in life and have a better prognosis. This difference may be artificially scholastic because primary forms can occur at any age (not only during infancy or early childhood) [8], and both primary and secondary forms can be precipitated by infections with a substantial risk of mortality [9]. Even secondary HPS occurs as an imbalance between insufficient host defense, obstinate hyperinflammation, and a heterogeneous triggering event, which can be of

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Table 1 Classification of hemophagocytic syndrome

Type of hemophagocytic syndrome	Subtype
Primary or genetic hemophagocytic syndrome	Familial hemophagocytic lymphohistiocytosis
	Immune deficiency syndromes (i.e., Chediak-Higashi syndrome, Griscelli syndrome, X-linked lymphoproliferative syndrome)
Secondary or reactive hemophagocytic syndrome	Infections (i.e., viral, bacterial, parasitic)
	Malignancies
	Auto-immune and auto-inflammatory diseases

infectious, rheumatic or neoplastic nature: therefore, the clinical disease results as the signature of a dysregulated immune activation, leading to macrophage proliferation and widespread hemophagocytosis in the reticuloendothelial system. The aim of this review is to make a critical appraisal of the literature concerning infection-related HPS in paediatrics.

Discussion

A multi-face disease with variable causes

Genetic HPS is heterogeneous and arises from defects in the critical regulatory pathways responsible for the natural termination of immune responses that lead to the failure of the homeostatic removal of cells that are superfluous or dangerous to the host. Since 1999, various genetic loci related to the activity of perforin and granzyme granules have been associated with genetic autosomal recessive HPS, thus explaining the impaired or absent function of NK cells and cytotoxic T cells [2,6]. The familial form, which was first described in 1952 [10], is an autosomal recessive disorder that is estimated to occur in 1/30,000-50,000 births, and usually appears during the first year of life [11,12]. Sporadic cases of HPS associated with multiple genetic mutations have also been described [13]. The different primary forms of HPS are associated with immune deficiencies, including Chediak-Higashi syndrome, Griscelli syndrome, X-linked lymphoproliferative syndrome, Wiskott-Aldrich syndrome, severe combined immunodeficiency, lysinuric protein intolerance, and Hermansky-Pudlak syndrome. Acquired HPS has also been associated with a variety of viral, bacterial and mycobacterial, fungal, and parasitic infections [14], auto-immune diseases [15], auto-inflammatory syndromes [16], and tumours, particularly T cell malignancies [17].

The constellation of signs and symptoms of HPS is not specific and none of the biochemical abnormalities is distinctive. The dramatic presentation of the syndrome includes unremitting fever, visceromegaly, thrombocytopenia, lethargy, seizures, skin rash, pulmonary failure, and cardiac and/or renal involvement, and the mortality rate is 8-22% [18]. The most common laboratory findings are due to liver dysfunction, and include low fibrinogen, and

high serum triglycerides and ferritin levels [18]. Two highly diagnostic clues are increased plasma concentrations of the alpha chain of interleukin-2 receptors (also known as sCD25) and impaired NK cell activity. As treatment can be life-saving and some of the clinical criteria occur late during the course of the disease, it is not necessary to satisfy all of the criteria before beginning therapy. The hallmark of HPS is the phagocytosis of blood cells and their precursors: bone marrow aspiration typically reveals the normal maturity of all cell lineages, and infiltration by activated macrophages “stuffed” with other blood cells [19].

Criteria for a diagnosis of HPS on the basis of clinical, laboratory, and histopathological findings are the following [3]:

- (1) Genetic diagnosis: genes known to cause the syndrome (*PRF1*, *UNC13D*, *STX11*, *STXBP2*, *RAB27A*)
- (2) Signs and symptoms (at least five of the following criteria):
 - a) Fever
 - b) Splenomegaly
 - c) cytopenias (minimum 2 cell lines reduced)
 - d) hypertriglyceridemia (≥ 265 mg/dL) and/or hypofibrinogenemia (≤ 150 g/dL)
 - e) hemophagocytosis in any involved organ
 - f) very little or no NK cell activity
 - g) increased ferritin ≥ 500 mg/L
 - h) increased soluble CD25 (serum interleukin-2 receptor alpha) ≥ 2.400 U/mL

The main pathophysiological abnormality in HPS is cytokine dysfunction, which leads to the uncontrolled accumulation and ectopic migration of activated T lymphocytes, antigen-presenting cells and histiocytes, and multi-system inflammation [20]. The pathophysiology of acquired HPS has not been fully defined, but deficient cytolytic activity leads to the persistent activation of lymphocytes and histiocytes, followed by the hypersecretion of pro-inflammatory cytokines and high soluble interleukin-2 receptor levels that correlate with the prognosis [21].

Infectious etiologies of HPS

The association between HPS and infections has been widely documented and both familial or sporadic cases are often precipitated by acute infections. It must also be pointed out that every form of HPS can mimic infectious diseases or overwhelming bacterial sepsis, thus hindering the diagnosis of a precipitating and treatable infectious illness. Virus-associated HPS was first described in 1979 by Risdall *et al.*, whose series consisted of 19 patients, most of whom were immunocompromised but without any confirmed genetic or acquired immunodeficiency, and all of whom showed serological signs of viral infection [22,23]. Since then, there have been reports of HPS associated with a host of infections [22].

HPS associated with viral infections

Epstein-Barr virus (EBV) is the most commonly reported trigger of HPS [24]. The epidemiology of EBV-related HPS is not well known, although a higher incidence has been observed in Asian countries, where it has been theorised there may be a more pathogenic viral strain that is genetically similar to the strains observed in nasopharyngeal carcinoma cell lines [25]. Two forms of EBV-related HPS have been described: the first occurring during primary infection and the second during a reactivation process [26]. During primary infection, EBV typically infects and replicates in B cells, whereas EBV-specific cytotoxic T cells are required to produce memory cells. In rare cases, EBV may infect T and NK cells and induce persistent EBV infection, which may lead to chronic active EBV infection, lymphoproliferative disorders and fulminant EBV-related HPS [27-29]. Serological testing can help determine whether EBV-associated HPS has occurred in the setting of acute infection or is the result of a reactivation process. In addition, the real-time polymerase chain reaction (PCR) detection and quantification of EBV nucleic acid is an important laboratory means of adequately reflecting viral replication and assessing EBV load in patients with EBV-related HPS [30]. The quantitative analysis of cell-free EBV genome copy numbers after four months of treatment can assess therapeutic responses and is prognostically significant [31].

The clonal expansion of EBV-infected T lymphocytes has been demonstrated in EBV-related HPS [32] and EBV-positive T cell lymphoma [33] on the basis of the presence of homogeneous viral terminal repeat sequences. The clonality of infected T lymphocytes is further suggested by the finding of monoclonal rearrangements of the T cell receptor-alpha gene in EBV-related HPS [34]. The distinction between the monoclonal proliferation of T lymphocytes seen in EBV-related HPS and EBV-positive T cell lymphomas may describe the extremes of a spectrum of disordered T lymphocyte proliferation following EBV infection. The inflammatory cytokine over-production seen

in patients with EBV-related HPS tends to be much more pronounced than that observed in patients with other forms of HPS [35]. Of all of the infections associated with HPS, EBV infection has the worst prognosis in the presence of underlying hereditary disorders, diffuse intravascular coagulation, neutropenia, or central nervous system involvement [36].

Treatment strategies vary significantly depending on the clinical features of the infection: mild cases of EBV-related HPS are treated conservatively as spontaneous regression has been reported, and antiviral therapy with acyclovir, ganciclovir or cidofovir has led to disappointing results [37]. In the case of severe EBV-related HPS, the introduction of immuno-chemotherapy and, if necessary, allogenic stem cell transplantation has radically changed the history and prognosis of the disease: in such cases, the optimal treatment strategy can be centred on immunosuppressive medications that inhibit overactive T and NK cell responses (i.e. corticosteroids, cyclosporine A, intravenous immunoglobulin, anti-thymocyte globulins, etoposide, rituximab, and plasma or blood exchange transfusions) [38,39]. Hematopoietic stem cell transplantation is the last treatment resort for refractory forms of EBV-related HPS, and in the case of EBV infection occurring in genetic forms of HPS [40].

The most frequent herpes viruses associated with HPS other than EBV are cytomegalovirus (CMV) and human herpes virus 8 (HHV8).

CMV up-regulates tumour necrosis factor gene expression and has been associated with HPS in otherwise healthy patients, patients with inflammatory bowel disease, rheumatological diseases and cancer, and transplant recipients [41]. HPS was observed in seven of a series of 171 patients undergoing hematopoietic stem cell transplantation, and was triggered by CMV in three cases [42]. Younger age may be associated with a worse prognosis [43]. A recent study has shown that the use of specific anti-CMV therapy, such as CMV immunoglobulin, foscarnet or ganciclovir, may be therapeutic [44].

HHV8 has been associated with HPS in 13 patients: most of these cases occurred in patients with a lymphoproliferative disorder [45] or immunocompromised patients [46], and rarely in immunocompetent hosts [47]. Treatment based on etoposide, ganciclovir, foscarnet or rituximab has led to successful results [48].

HPS can be associated with human immunodeficiency virus (HIV) infection, alone or with a wide variety of underlying disorders. It is likely that this condition is underestimated as HIV infection and HPS have many clinical and laboratory similarities. About 10% of bone marrow biopsies taken from HIV patients before the start of highly active antiretroviral therapy show active signs of hemophagocytosis [49]. HIV-related HPS can be observed in cases of acute or late HIV infection, and in

conjunction with immune reconstitution inflammatory syndrome, opportunistic infections, or malignancies [50]. HPS may even be the initial presentation of HIV infection [51], and it has been suggested that HIV itself may play a direct role in triggering the syndrome [52]. Other common viral triggers of HPS in HIV patients are EBV, CMV and HHV8, and EBV-related HPS seems to be more frequent in HIV-infected children [53].

Influenza-related HPS has been rarely reported in immunocompromised and otherwise healthy children [54-57]. One fatal case of HPS was observed among 32 children hospitalised with seasonal influenza in a prospective pediatric study [58], but reactive HPS has also been associated with avian and swine (non-pandemic) influenza [59,60]. In particular, patients with severe H5N1 (avian) influenza infection have symptoms and laboratory findings that are similar to those observed in patients with HPS, mainly encephalitis, organ dysfunction with hemophagocytosis, bone marrow failure, and pro-inflammatory cytokine over-production [61]. Clinical studies have found that mutations in some viral genes (NS1, PB2, HA and NA) are significantly related to cytokine release, and it has been demonstrated that recombinant hemagglutinin (H5) from H5N1 virus can suppress perforin expression and reduce the cytotoxicity of T cells, including their ability to kill H5-bearing cells [62]. Some authors have suggested treatment with a shorter course of etoposide and dexamethasone [63].

HPS has been reported in 28 cases of parvovirus B19 infection, most of whom had hereditary spherocytosis as the underlying disease: fewer than half were children [64-68]. Of these patients, 16 did not receive any treatment and 22 survived, thus suggesting that the prognosis of parvovirus-associated HPS is better than that of the other viral-mediated forms of HPS.

Fulminant viral hepatitis may mimic and even cause HPS, with hepatitis A virus being more frequently associated with HPS than the other hepatotrope viruses. Fifteen cases (including children) have been described, mainly in Asia: three of these patients also had a concurrent rheumatological disease (systemic juvenile idiopathic arthritis or Still's disease) and two also had hepatitis C. Their treatment consisted of corticosteroids, variously combined with intravenous immunoglobulin, but four patients received no specific treatment and 11 of the 15 experienced a favourable outcome [69-71].

Enterovirus-related HPS has been described in 12 pediatric cases: five occurred in infants aged <1 year, and the oldest patient was 18 years old. An underlying disease was found in four patients who experienced a fatal outcome (lymphoid neoplasms, lymphoblastic leukemia and juvenile idiopathic arthritis). Ten patients received intravenous immunoglobulin (six in combination with corticosteroids), but only seven patients survived [72].

Other viruses associated with HPS include adenovirus, paramyxovirus (leading to measles and mumps), rubella virus, human parainfluenza viruses, *Flavivirus* (leading to dengue fever) and hantavirus (leading to hemorrhagic fever and severe acute respiratory syndrome), all of which have been treated with varying courses of corticosteroids and intravenous immunoglobulin.

HPS associated with bacterial infections

Reactive HPS has frequently been associated with intracellular pathogens. The pathophysiology of HPS associated with non-viral agents may be related to the production of high levels of activating cytokines by host lymphocytes and monocytes. Although the pathophysiological response of the host immune system to the infectious agent is not fully understood, it is hypothesised that functional deficiencies in NK and cytotoxic T cells may occur during the illness [73].

HPS can be associated with disseminated *Mycobacterium tuberculosis* infection. Thirty-six cases (including infants and children) have so far been reported, approximately half of which were accompanied by comorbidities: eight patients had end-stage renal disease and were receiving hemodialysis or had undergone renal transplantation, four had a history of a malignancy, two had AIDS, and one had sarcoidosis. Fever was the most frequent clinical feature upon presentation, combined with visceromegaly and pancytopenia, and all of the patients underwent bone marrow aspirations that confirmed hemophagocytosis. Evidence of extra-pulmonary tuberculosis was found in 83% of cases. The concluding remarks of the report stated that tuberculosis-related HPS has a poor prognosis, with a mortality rate of approximately 50%, although anti-tuberculous and immunomodulatory therapy (consisting of high-dose corticosteroids, intravenous immunoglobulin, anti-thymocyte globulin, cyclosporine A, epipodophyllotoxin or plasma exchange) may lead to a better outcome [74]. Early diagnostic confirmation and the timely administration of anti-tuberculous medication seem to be crucial in these patients. One reported case of HPS occurred after childhood vaccination with the bacillus Calmette-Guérin [75].

HPS has also been described in association with brucellosis, with *Brucella melitensis* being the most frequently isolated organism [76]. Leptospirosis can cause life-threatening HPS as a result of an insufficient or misdirected immunological response to *Leptospira* itself: antibiotic treatment alone is not sufficient in such cases, and treatment with corticosteroids, intravenous immunoglobulin or etoposide is required [77]. Rickettsial diseases, transmitted to humans by arthropod bites and usually controlled at an intracellular level by nitric oxide synthesis, hydrogen peroxide production, and tryptophan degradation have also been related to HPS:

overall, 15 cases of rickettsial disease confirmed serologically and complicated by HPS have been published in the period 1990–2010, with only 3 cases occurring in patients less than 15 years and a prognosis influenced by the specific *Rickettsia* species, patient's immunologic equipment, and delay in antibiotic therapy or corticosteroid therapy [78]. In 2009, sepsis caused by multidrug-resistant *Acinetobacter baumannii* following urinary tract infection was reported for the first time in a previously healthy 3-year-old child, who recovered after multiple doses of granulocyte colony stimulating factor and red blood cell/platelet transfusions without any cytotoxic treatment or immunotherapy [79].

HPS associated with parasitic and fungal infections

HPS can be associated with *Leishmania donovani* and *Leishmania infantum* infections, but leishmaniasis may also mimic the syndrome, as it is characteristically associated with organomegaly and pancytopenia. This is particularly important in non-endemic areas, where visceral leishmaniasis is unlikely to be included in the differential diagnosis, and repeated bone marrow smears are often required to identify *Leishmania* species by means of PCR with species-specific probes [80]. Specific anti-leishmania treatment with amphotericin B is usually sufficient to control HPS. Unfortunately, sporadic cases of undiagnosed leishmaniasis have been treated as HPS with fatal consequences [81].

Malaria (caused by *Plasmodium falciparum* and *Plasmodium vivax*), toxoplasmosis, babesiosis, and strongyloidiasis have been rarely identified in association with HPS: a history of travel from endemic countries may help to identify these triggering agents [82].

Yeast (*Candida* spp., *Cryptococcus* spp. and *Pneumocystis* spp.) and moulds (*Histoplasma* spp., *Aspergillus* spp. and *Fusarium* spp.) have been associated with the occurrence of HPS, most commonly during HIV infection, neoplastic diseases, protracted corticosteroid administration, and transplantation [83-85].

Disseminated *Penicillium marneffe* infection is common among HIV-infected patients in many regions in Southeast Asia: the first case of HPS associated with penicilliosis in a Thai HIV-infected child was reported in 2001, with complete recovery after antifungal and intravenous immunoglobulin therapy [86].

Conclusions

Many immunological, neoplastic and genetic disorders may underlie HPS, but infectious causes are the most prevalent and most frequently reported in association with this syndrome.

The specific clinical and laboratory tests for microbiological identification of HPS are the following:

- (1) Blood, mid-stream urine, cerebrospinal fluid and sputum cultures for bacteria and fungi; cultures/microscopy for mycobacteria; tuberculin skin test and/or interferon-release tests
- (2) Bone marrow aspirate
- (3) Serological tests for acute Epstein-Barr, cytomegalovirus and human immunodeficiency virus infections
- (4) Peripheral blood nucleic acid tests for evidence of replicating Epstein-Barr virus, cytomegalovirus, herpes simplex virus, human herpes virus-8, human immunodeficiency virus, adenovirus and parvovirus B19
- (5) Serological tests for evidence of primary toxoplasmosis or reactivation
- (6) *Leishmania* antigen test
- (7) Serum cryptococcal antigen and serum galactomannans

As a fatal outcome may occur when infection-related HPS is only treated supportively, a multidisciplinary approach by experienced clinicians and infectious disease specialists is required in order to ensure the appropriate management of the syndrome itself, and the precipitating or underlying infection. Pediatricians should be alert and aware of the risk of the syndrome, because an early diagnosis can change its natural history and it has been shown that prompt treatment improves the overall prognosis. A combination of high fever unresponsive to broad-spectrum antibiotics, hyperferritinemia, hypertriglyceridemia, hypofibrinogenemia, cytopenia, organomegaly and characteristic histological findings in the setting of an infectious process (particularly EBV infection, but also other viral and bacterial, parasitic and fungal infections) is the key diagnostic clue. A better understanding of the pathophysiology of HPS should clarify the interactions between immune system pathways and infections. Specific antimicrobial therapy can be beneficial in selected cases, whereas antiviral drugs do not seem to be curative. Severe cases of infection-dependent HPS require immunosuppressants or chemotherapeutic agents, while bone marrow transplantation is the ultimate choice for persistent refractory cases.

Summary

Hemophagocytic syndrome (HPS) can occur as a rare complication of various infections in children. Clonal proliferation of T lymphocytes with an excessive activation of macrophages can be triggered by different infectious agents, thus indicating that infection *per se* is involved in the pathogenetic mechanism of the process. A number of studies have demonstrated that HPS is frequently triggered by one of many different viral, bacterial, parasitic or fungal infections, with large differences in terms of

treatment responses and overall outcomes. All patients meeting the criteria for HPS should undergo initial tests to diagnose the underlying infecting organism, which should be guided by epidemiological data and the patient's medical history. A polymerase chain reaction search for infectious agents, including EBV, CMV and *Leishmania*, is recommended in a clinical scenario characterised by unremitting fever, organomegaly, cytopenia and hyperferritinemia. As HPS may be associated with many infectious diseases and immunological, neoplastic or genetic disorders, the close cooperation of pediatricians and infectious disease specialists is crucial in order to define any precipitating or underlying condition.

Abbreviations

CMV: Cytomegalovirus; EBV: Epstein-Barr virus; HHV8: Human herpes virus 8; HIV: Human immunodeficiency virus; H5: Hemagglutinin; HPS: Hemophagocytic syndrome; NK: Natural killer.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

VA and DR drafted the manuscript and contributed equally to its preparation; SE revised the draft and made a substantial scientific contribution. All of the authors read and approved the final version of the manuscript.

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References

1. Maakaroun NR, Moanna A, Jacob JT, Albrecht H: **Viral infection associated with haemophagocytic syndrome.** *Rev Med Virol* 2010, **20**:93–105.
2. Ravelli A: **Macrophage activation syndrome.** *Curr Opin Rheumatol* 2002, **14**:548–552.
3. Douka E, Economidou F, Nanas S: **Infections associated with the hemophagocytic syndrome.** *Hosp Chron* 2012, **7**:16–24.
4. Ishii E, Ohga S, Imashuku S, Yasukawa M, Tsuda H, Miura I, Yamamoto K, Horiuchi H, Takada K, Ohshima K, Nakamura S, Kinukawa N, Oshimi K, Kawa K: **Nationwide survey of hemophagocytic lymphohistiocytosis in Japan.** *Int J Hematol* 2007, **86**:58–65.
5. Scott R, Robb-Smith A: **Histiocytic medullary reticulosis.** *Lancet* 1939, **2**:194–198.
6. Henter JI, Elinder G, Ost A: **Diagnostic guidelines for hemophagocytic lymphohistiocytosis.** The FHL Study Group of the Histiocyte Society. *Semin Oncol* 1991, **18**:29–33.
7. Henter JI, Elinder G: **Familial hemophagocytic lymphohistiocytosis. Clinical review based on the findings in seven children.** *Acta Paediatr Scand* 1991, **80**:269–277.
8. Allen M, De Fusco C, Legrand F, Clementi R, Conter V, Danesino C, Janka G, Aricò M: **Familial hemophagocytic lymphohistiocytosis: how late can the onset be?** *Haematologica* 2001, **86**:499–503.
9. Henter JI, Ehrnst A, Andersson J, Elinder G: **Familial hemophagocytic lymphohistiocytosis and viral infections.** *Acta Paediatr* 1993, **82**:369–372.
10. Farquhar J, Claireaux A: **Familial haemophagocytic reticulosis.** *Arch Dis Child* 1952, **27**:519–525.
11. Henter JI, Elinder G, Soder O, Ost A: **Incidence in Sweden and clinical features of familial hemophagocytic lymphohistiocytosis.** *Acta Paediatr Scand* 1991, **80**:428–435.
12. Ishii E, Ohga S, Tanimura M, Imashuku S, Sako M, Mizutani S, Miyazaki S: **Clinical and epidemiologic studies of familial hemophagocytic lymphohistiocytosis in Japan.** Japan LCH Study Group. *Med Pediatr Oncol* 1998, **30**:276–283.
13. Verbsky JW, Grossman WJ: **Hemophagocytic lymphohistiocytosis: diagnosis, pathophysiology, treatment, and future perspectives.** *Ann Med* 2006, **38**:20–31.
14. Fisman DN: **Hemophagocytic syndromes and infection.** *Emerg Infect Dis* 2000, **6**:601–608.
15. Rigante D, De Rosa G, Bertoni B, Ansuini V, Pardeo M, La Torraca I, Gaspari S, Stabile A: **Large pericardial effusion requiring pericardiocentesis as cardinal sign of macrophage activation syndrome in systemic onset-juvenile idiopathic arthritis.** *Rheumatol Int* 2007, **27**:767–770.
16. Rigante D, Capoluongo E, Bertoni B, Ansuini V, Chiaretti A, Piastra M, Pulitano S, Genovese O, Compagnone A, Stabile A: **First report of macrophage activation syndrome in hyperimmunoglobulinemia D with periodic fever syndrome.** *Arthritis Rheum* 2007, **56**:658–661.
17. Janka G, Imashuku S, Elinder G, Schneider M, Henter JI: **Infection and malignancy associated hemophagocytic syndromes. Secondary hemophagocytic lymphohistiocytosis.** *Hematol Oncol Clin North Am* 1998, **12**:435–444.
18. Jabado N, McCusker C, de Saint Basile G: **Pediatric hemophagocytic syndromes: a diagnostic and therapeutic challenge.** *Allergy Asthma Clin Immunol* 2005, **1**:142–160.
19. Stabile A, Bertoni B, Ansuini V, La Torraca I, Salli A, Rigante D: **The clinical spectrum and treatment options of macrophage activation syndrome in the pediatric age.** *Eur Rev Med Pharmacol Sci* 2006, **10**:53–59.
20. Osugi Y, Hara J, Tagawa S, Takai K, Hosoi G, Matsuda Y, Ohta H, Fujisaki H, Kobayashi M, Sakata N, Kawa-Ha K, Okada S, Tawa A: **Cytokine production regulating Th1 and Th2 cytokines in hemophagocytic lymphohistiocytosis.** *Blood* 1997, **89**:4100–4103.
21. Imashuku S, Hibi S, Sako M, Ishida Y, Mugishima H, Chen J, Tsunematsu Y: **Soluble interleukin-2 receptor: a useful prognostic factor for patients with hemophagocytic lymphohistiocytosis.** *Blood* 1995, **86**:4706–4707.
22. Roupheal NG, Talati NJ, Vaughan C, Cunningham K, Moreira R, Gould C: **Infections associated with haemophagocytic syndrome.** *Lancet Infect Dis* 2007, **7**:814–822.
23. Risdall RJ, McKenna RW, Nesbit ME, Krivit W, Balfour HH Jr, Simmons RL, Brunning RD: **Virus-associated hemophagocytic syndrome: a benign histiocytic proliferation distinct from malignant histiocytosis.** *Cancer* 1979, **44**:993–1002.
24. Chen CJ, Huang YC, Jaing TH, Hung IJ, Yang CP, Chang LY, Lin TY: **Hemophagocytic syndrome: a review of 18 pediatric cases.** *J Microbiol Immunol Infect* 2004, **37**:157–163.
25. Tabata Y, Hibi S, Teramura T, Kuriyama K, Yagi T, Todo S, Sawada T, Imashuku S: **Molecular analysis of latent membrane protein 1 in patients with Epstein-Barr virus-associated hemophagocytic lymphohistiocytosis in Japan.** *Leuk Lymphoma* 2000, **38**:373–380.
26. Imashuku S: **Clinical features and treatment strategies of Epstein-Barr virus-associated hemophagocytic lymphohistiocytosis.** *Crit Rev Oncol Hematol* 2002, **44**:259–272.
27. Ohshima K, Suzumiya J, Sugihara M, Nagafuchi S, Ohga S, Kikuchi M: **Clinicopathological study of severe chronic active Epstein-Barr virus infection that developed in association with lymphoproliferative disorder and/or hemophagocytic syndrome.** *Pathol Int* 1998, **48**:934–943.
28. Su IJ, Wang CH, Cheng AL, Chen RL: **Hemophagocytic syndrome in Epstein-Barr virus-associated T lymphoproliferative disorders: disease spectrum, pathogenesis, and management.** *Leuk Lymphoma* 1995, **19**:401–406.
29. Lindemann TL, Greene JS: **Persistent cervical lymphadenopathy in an adolescent with Epstein-Barr induced hemophagocytic syndrome: manifestations of a rare but often fatal disease.** *Int J Pediatr Otorhinolaryngol* 2005, **69**:1011–1014.
30. Kimura H, Hoshino Y, Hara S, Nishikawa K, Sako M, Hirayama M, Komada Y, Morishima T: **Viral load in Epstein-Barr virus-associated hemophagocytic syndrome.** *Microbiol Immunol* 2002, **46**:579–582.
31. Teramura T, Tabata Y, Yagi T, Morimoto A, Hibi S, Imashuku S: **Quantitative analysis of cell-free Epstein-Barr virus genome copy number in patients**

- with EBV-associated hemophagocytic lymphohistiocytosis. *Leuk Lymphoma* 2002, **43**:173–179.
32. Chen JS, Tzeng CC, Tsao CJ, Su WC, Chen TY, Jung YC, Su IJ: **Clonal karyotype abnormalities in EBV associated hemophagocytic syndrome.** *Haematologica* 1997, **82**:572–576.
 33. Lay JD, Tsao CJ, Chen JY, Kadin ME, Su IJ: **Upregulation of tumor necrosis factor-alpha gene by Epstein-Barr virus and activation of macrophages in Epstein-Barr virus-infected T cells in the pathogenesis of hemophagocytic syndrome.** *J Clin Invest* 1997, **100**:1969–1979.
 34. Craig F, Clare N, Sklar J, Banks P: **T-cell lymphoma and the virus-associated hemophagocytic syndrome.** *Am J Clin Pathol* 1991, **97**:189–194.
 35. Imashuku S, Hibi S, Tabata Y, Sako M, Sekine Y, Hirayama K, Sakazaki H, Maeda N, Kito H, Shichino H, Mugishima H: **Biomarker and morphological characteristics of Epstein-Barr virus-related hemophagocytic lymphohistiocytosis.** *Med Pediatr Oncol* 1998, **31**:131–137.
 36. Imashuku S, Teramura T, Morimoto A, Hibi S: **Recent developments in the management of haemophagocytic lymphohistiocytosis.** *Expert Opin Pharmacother* 2001, **2**:1437–1448.
 37. Bakhshi S, Pautu JL: **EBV associated hemophagocytic lymphohistiocytosis with spontaneous regression.** *Indian Pediatr* 2005, **42**:1253–1255.
 38. Imashuku S, Tabata Y, Teramura T, Hibi S: **Treatment strategies for Epstein-Barr virus-associated hemophagocytic lymphohistiocytosis (EBV-HLH).** *Leuk Lymphoma* 2000, **39**:37–49.
 39. Balamuth NJ, Nichols KE, Paessler M, Teachey DT: **Use of rituximab in conjunction with immunosuppressive chemotherapy as a novel therapy for Epstein Barr virus-associated hemophagocytic lymphohistiocytosis.** *J Pediatr Hematol Oncol* 2007, **29**:569–573.
 40. Imashuku S, Hibi S, Todo S, Sako M, Inoue M, Kawa K, Koike K, Iwai A, Tsuchiya S, Akiyama Y, Kotani T, Kawamura Y, Hirose M, Hasegawa D, Kosaka Y, Yamaguchi H, Ishii E, Kato K, Ishii M, Kigasawa H: **Allogeneic hematopoietic stem cell transplantation for patients with hemophagocytic syndrome (HPS) in Japan.** *Bone Marrow Transplant* 1999, **23**:569–572.
 41. Danishi EH, Dahms BB, Kumar ML: **Cytomegalovirus-associated hemophagocytic syndrome.** *Pediatrics* 1985, **75**:280–283.
 42. Abdelkefi A, Ben Jamil W, Torjman L, Ladeb S, Ksouri H, Lakhal A, Ben Hassen A, Ben Abdeladhim A, Ben Othman T: **Hemophagocytic syndrome after hematopoietic stem cell transplantation: a prospective observational study.** *Int J Hematol* 2009, **89**:368–373.
 43. Imashuku S, Ueda I, Teramura T, Mori K, Morimoto A, Sako M, Ishii E: **Occurrence of haemophagocytic lymphohistiocytosis at less than 1 year of age: analysis of 96 patients.** *Eur J Pediatr* 2005, **164**:315–319.
 44. Amenomori M, Migita K, Miyashita T, Yoshida S, Ito M, Eguchi K, Ezaki H: **Cytomegalovirus-associated hemophagocytic syndrome in a patient with adult onset Still's disease.** *Clin Exp Rheumatol* 2005, **23**:100–102.
 45. Pastore RD, Chadburn A, Kripas C, Schattner EJ: **Novel association of haemophagocytic syndrome with Kaposi's sarcoma-associated herpesvirus-related primary effusion lymphoma.** *Br J Haematol* 2000, **111**:1112–1115.
 46. Fardet L, Blum L, Kerob D, Agbalika F, Galicier L, Dupuy A, Lafaurie M, Meignin V, Morel P, Lebbé C: **Human herpesvirus 8-associated hemophagocytic lymphohistiocytosis in human immunodeficiency virus-infected patients.** *Clin Infect Dis* 2003, **37**:285–291.
 47. Grossman WJ, Radhi M, Schauer D, Gerday E, Grose C, Goldman FD: **Development of hemophagocytic lymphohistiocytosis in triplets infected with HHV-8.** *Blood* 2005, **106**:1203–1206.
 48. Re A, Facchetti F, Borlenghi E, Cattaneo C, Capucci MA, Ungari M, Barozzi P, Vallerini D, Potenza L, Torelli G, Rossi G, Luppi M: **Fatal hemophagocytic syndrome related to active human herpesvirus-8/Kaposi sarcoma-associated herpesvirus infection in human immunodeficiency virus-negative, non-transplant patients without related malignancies.** *Eur J Haematol* 2007, **78**:361–364.
 49. Sailler L, Duchayne E, Marchou B, Brousset P, Pris J, Massip P, Corberand J, Arlet P: **Etiological aspects of reactive hemophagocytoses: retrospective study in 99 patients.** *Rev Med Interne* 1997, **18**:855–864.
 50. Albrecht H, Schafer H, Stellbrink HJ, Greten H: **Epstein-Barr virus-Associated hemophagocytic syndrome. A cause of fever of unknown origin in human immunodeficiency virus infection.** *Arch Pathol Lab Med* 1997, **121**:853–858.
 51. Sun HY, Chen MY, Fang CT, Hsieh SM, Hung CC, Chang SC: **Hemophagocytic lymphohistiocytosis: an unusual initial presentation of acute HIV infection.** *J Acquir Immune Defic Syndr* 2004, **37**:1539–1540.
 52. Bhatia S, Bauer F, Bilgrami SA: **Candidiasis associated hemophagocytic lymphohistiocytosis in a patient infected with human immunodeficiency virus.** *Clin Infect Dis* 2003, **37**:e161–e166.
 53. Preciado MV, De Matteo E, Fallo A, Chabay P, Drelichman G, Grinstein S: **EBV-associated Hodgkin's disease in an HIV-infected child presenting with a hemophagocytic syndrome.** *Leuk Lymphoma* 2001, **42**:231–234.
 54. Potter MN, Foot AB, Oakhill A: **Influenza A and the virus-associated haemophagocytic syndrome: cluster of three cases in children with acute leukaemia.** *J Clin Pathol* 1991, **44**:297–299.
 55. Ando M, Miyazaki E, Hiroshige S, Ashihara Y, Okubo T, Ueo M, Fukami T, Sugisaki K, Tsuda T, Ohishi K, Yoshitake S, Noguchi T, Kumamoto T: **Virus associated hemophagocytic syndrome accompanied by acute respiratory failure caused by influenza A (H3N2).** *Intern Med* 2006, **45**:1183–1186.
 56. Watanabe T, Okazaki E, Shibuya H: **Influenza A virus-associated encephalopathy with haemophagocytic syndrome.** *Eur J Pediatr* 2003, **162**:799–800.
 57. Mou SS, Nakagawa TA, Riemer EC, McLean TW, Hines MH, Shetty AK: **Hemophagocytic lymphohistiocytosis complicating influenza A infection.** *Pediatrics* 2006, **118**:e216–e219.
 58. Samransamruajkit R, Hiranrat T, Chieochansin T, Sritippayawan S, Deerojanawong J, Prapphal N, Poovorawan Y: **Prevalence, clinical presentations and complications among hospitalized children with influenza pneumonia.** *Jpn J Infect Dis* 2008, **61**:446–449.
 59. Zhang Z, Zhang J, Huang K, Li KS, Yuen KY, Guan Y, Chen H, Ng WF: **Systemic infection of avian influenza A virus H5N1 subtype in humans.** *Hum Pathol* 2009, **40**:735–739.
 60. Kimura K, Adlakhia A, Simon PM: **Fatal case of swine influenza virus in an immunocompetent host.** *Mayo Clin Proc* 1998, **73**:243–245.
 61. Cheung CY, Poon LL, Lau AS, Luk W, Lau YL, Shorridge KF, Gordon S, Guan Y, Peiris JS: **Induction of proinflammatory cytokines in human macrophages by influenza A (H5N1) viruses: a mechanism for the unusual severity of human disease?** *Lancet* 2002, **360**:1831–1837.
 62. Hsieh SM, Chang SC: **Insufficient perforin expression in CD8 β T cells in response to hemagglutinin from avian influenza (H5N1) virus.** *J Immunol* 2006, **176**:4530–4533.
 63. Henter JI, Chow CB, Leung CW, Lau YL: **Cytotoxic therapy for severe avian influenza A (H5N1) infection.** *Lancet* 2006, **367**:870–873.
 64. Kaya Z, Ozturk G, Gursel T, Bozdayi G: **Spontaneous resolution of hemophagocytic syndrome and disseminated intravascular coagulation associated with Parvovirus b19 infection in a previously healthy child.** *Jpn J Infect Dis* 2005, **58**:149–151.
 65. Yilmaz S, Oren H, Demircioglu F, Firinci F, Korkmaz A, Irken G: **Parvovirus B19: a cause for aplastic crisis and hemophagocytic lymphohistiocytosis.** *Pediatr Blood Canc* 2006, **47**:8610.
 66. Larroche C, Scieucx S, Honderlick P, Piette AM, Morinet F, Bletry O: **Spontaneous resolution of hemophagocytic syndrome associated with acute Parvovirus B19 infection and concomitant Epstein-Barr virus reactivation in an otherwise healthy adult.** *Eur J Clin Microbiol Infect Dis* 2002, **21**:739–742.
 67. Dutta U, Mittal S, Ratho RK, Das A: **Acute liver failure and severe hemophagocytosis secondary to Parvovirus B19 infection.** *Indian J Gastroenterol* 2005, **24**:118–119.
 68. Ardalan MR, Shoja MM, Tubbs RS, Esmaili H, Keyvani H: **Postrenal transplant hemophagocytic lymphohistiocytosis and thrombotic microangiopathy associated with Parvovirus B19 infection.** *Am J Transplant* 2008, **8**:1340–1344.
 69. Watanabe M, Shibuya A, Okuno J, Maeda T, Tamama S, Saigenji K: **Hepatitis A virus infection associated with hemophagocytic syndrome: report of two cases.** *Intern Med* 2002, **41**:1188–1192.
 70. Tuon FF, Gomes VS, Amato VS, Graf ME, Fonseca GH, Lazari C, Nicodemo AC: **Hemophagocytic syndrome associated with hepatitis A: case report and literature review.** *Rev Inst Med Trop Sao Paulo* 2008, **50**:123–127.
 71. Russo RA, Rosenzweig SD, Katsicas MM: **Hepatitis A-associated macrophage activation syndrome in children with systemic juvenile idiopathic arthritis: report of 2 cases.** *J Rheumatol* 2008, **35**:166–168.
 72. Katsibardi K, Moschovi MA, Theodoridou M, Spanakis N, Kalabalkis P, Tsakris A, Tzortzatos-Stathopoulou F: **Enterovirus-associated hemophagocytic syndrome in children with malignancy: report of three cases and review of the literature.** *Eur J Pediatr* 2008, **167**:97–102.
 73. Janka GE: **Familial and acquired hemophagocytic lymphohistiocytosis.** *Annu Rev Med* 2012, **63**:233–46.

74. Brastianos PK, Swanson JW, Torbenson M, Sperati J, Karakousis PC: **Tuberculosis-associated haemophagocytic syndrome.** *Lancet Infect Dis* 2006, **6**:447–454.
75. Rositto A, Molinaro L, Larralde M, Ranalletta M, Drut R: **Disseminated cutaneous eruption after BCG vaccination.** *Pediatr Dermatol* 1996, **13**:451–454.
76. Sari I, Altuntas F, Hacioglu S, Kocyigit I, Sevinc A, Sacar S, Deniz K, Alp E, Eser B, Yildiz O, Kaynar L, Unal A, Cetin M: **A multicenter retrospective study defining the clinical and haematological manifestations of brucellosis and pancytopenia in a large series: hematological malignancies, the unusual cause of pancytopenia in patients with brucellosis.** *Am J Hematol* 2008, **83**:334–339.
77. Niller HH: **Myelodysplastic syndrome (MDS) as a late stage of subclinical hemophagocytic lymphohistiocytosis (HLH): a putative role for *Leptospira* infection. A hypothesis.** *Acta Microbiol Immunol Hung* 2010, **57**:181–189.
78. Cascio A, Giordano S, Dones P, Venezia S, Iaria C, Ziino O: **Haemophagocytic syndrome and rickettsial diseases.** *J Med Microbiol* 2011, **60**(Pt 4):537–542.
79. Gosh JB, Roy M, Bala A: **Infection associated with hemophagocytic lymphohistiocytosis triggered by nosocomial infection.** *Oman Med J* 2009, **24**:223–225.
80. Cascio A, Pernice LM, Barberi G, Delfino D, Biondo C, Beninati C, Mancuso G, Rodriguez-Morales AJ, Iaria C: **Secondary hemophagocytic lymphohistiocytosis in zoonoses. A systematic review.** *Eur Rev Med Pharmacol Sci* 2012, **16**:1324–1337.
81. Gagnaire MH, Galambrun C, Stephan JL: **Hemophagocytic syndrome: a misleading complication of visceral leishmaniasis in children - A series of 12 cases.** *Pediatrics* 2000, **106**:E58.
82. Singh ZN, Rakheja D, Yadav TP, Shome DK: **Infection-associated haemophagocytosis: the tropical spectrum.** *Clin Lab Haematol* 2005, **27**:312–315.
83. Numata K, Tsutsumi H, Wakai S, Tachi N, Chiba S: **A child case of haemophagocytic syndrome associated with cryptococcal meningoencephalitis.** *J Infect* 1998, **36**:118–119.
84. Koduri PR, Chundi V, DeMarais P, Mizock BA, Patel AR, Weinstein RA: **Reactive hemophagocytic syndrome: a new presentation of disseminated histoplasmosis in patients with AIDS.** *Clin Infect Dis* 1995, **21**:1463–1465.
85. De Lavaissiere M, Manceron V, Bourée P, Garçon L, Bisaro F, Delfraissy JF, Lambotte O, Goujard C: **Reconstitution inflammatory syndrome related to histoplasmosis, with a hemophagocytic syndrome in HIV infection.** *J Infect* 2009, **58**:245–247.
86. Chokeyhaibulkit K, Veerakul G, Vanprapar N, Chairprasert A, Tanphaichitr V, Chearskul S: **Penicilliosis-associated hemophagocytic syndrome in a human immunodeficiency virus-infected child: the first case report in children.** *J Med Assoc Thai* 2001, **84**:426–429.

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