

NIH Public Access

Author Manuscript

Endocrine. Author manuscript; available in PMC 2014 April 01.

Published in final edited form as:

Endocrine. 2013 April; 43(2): 293–302. doi:10.1007/s12020-012-9790-0.

Ectopic acromegaly due to growth hormone releasing hormone

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Keywords

acromegaly; paraganglioma; GHRH; ectopic

Introduction

Acromegaly is usually due to hypersecretion of growth hormone (GH) from a benign pituitary adenoma and is characterized by discordant tissue, skeletal and organ growth. The incidence of acromegaly is approximately 3 cases per 1 million persons per year and the prevalence is about 60 cases per million. In less than 1% of cases acromegaly may develop because of ectopic secretion of growth hormone releasing hormone (GHRH) [1–6] or more rarely GH, from a nonpituitary origin, mostly from a neuroendocrine tumor (NET) [7–9]. In 1959 Altman and Schutz reported an acromegaly patient who did not respond to pituitary irradiation but improved markedly after surgical resection of a lung carcinoid tumor [2]. Subsequent reports confirmed the association of acromegaly with carcinoid tumors, and ectopic GHRH secretion was definitively identified as the cause through isolation and extraction of GHRH from pancreatic tumor tissue in patients with acromegaly [10, 11]. Most cases of ectopic acromegaly result from GHRH derived from lung carcinoid tumors, leading to pituitary hyperplasia and GH hypersecretion [12-51]. Other reported cases include pancreatic cell tumors [28, 41, 44, 51–69], gastrointestinal tract [51, 70–75], thymus [76, 77], adrenal pheochromocytomas [78, 79], lung (adenoid cystic carcinoma) [80] and pituitary [81].

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Here, we comprehensively review all the published cases of ectopic acromegaly and also report a new cause i.e. acromegaly secondary to a GHRH secreting mediastinal paraganglioma with ectopic GHRH secretion.

Ectopic Acromegaly

Acromegaly secondary to ectopic GHRH secretion is rare. Since the initial reports of the association of bronchial carcinoid and acromegaly due to ectopic GHRH syndrome, 98 cases have been reported, of whom 2/3 were female. Mean age at diagnosis was 41 years, similar to pituitary acromegaly. Time from onset of symptoms to diagnosis, was between 1 to 22 years (8.3 ± 5.8 years). Mean duration of the disease to time of diagnosis, approximately 8 years, was also similar to the usual course of acromegaly, emphasizing the delay that often occurs in diagnosis due to gradual development of symptoms and small size of the corresponding tumors which can easily escape detection [18, 82].

Published causes of ectopic acromegaly

Ectopic acromegaly is almost always secondary to an NET originating in the lung and pancreas [44]. Association of acromegaly and pheochromocytoma has been reported in some patients (Table 1). Some of the cases were published before 1980 prior to isolation of GHRH and were labeled as part of MEN syndrome; hence their association with GHRH secretion cannot be proven. In a recent case, a 23 year old man who had ectopic acromegaly secondary to an incidentally discovered pheochromocytoma demonstrated improved symptoms after tumor resection, and positivity of the tumor tissue with GHRH immunostaining [79]. Rarely, ectopic acromegaly may be secondary to secretion of GH, as reported in a pancreatic islet cell tumor, a bronchial carcinoid tumor and lymphoma [7–9].

An additional cause--mediastinal paraganglioma

This cause was encountered by the authors on evaluation of a 56 year old woman for facial coarsening over the past 6 years. She complained of facial puffiness, increasing jaw underbite, widening of facial creases, and enlargement of nose, hands and feet. She complained of low back and bilateral knee pain but did not elicit galactorrhea. Past medical history was significant for menopause at age 52, lung surgery for bronchiectasis, hypertension, prediabetes, and a benign colon polyp. Physical examination was remarkable for the presence of multiple neck and chest skin tags. Her upper incisors were spread apart, soft tissue swelling of hands and feet and deep facial creases and coarse features were noted.

Endocrine testing demonstrated elevated serum IGF-1 levels of 780, 525 and 616 ng/ml (normal range 94–284 ng/ml). The diagnosis of acromegaly was confirmed after a 75 gram oral glucose tolerance test (OGTT) with nadir GH level of 2.2 ng/ml. Plasma GHRH levels were unable to be obtained. FSH and LH levels were in the menopausal range and thyroid function tests, morning cortisol and serum prolactin were within normal limits. Fasting blood glucose was 105 and 126 mg/dl, serum calcium was 9.6 mg/dl and 24 hour urine calcium was 304 mg and routine laboratory tests were within normal limits.

Pituitary MRI was performed twice and no pituitary adenoma or pituitary hyperplasia was visualized (Fig. 1). Computed tomography (CT) scan of the chest revealed a 4.5 by 3.5 cm lobulated soft tissue mass in the subcarinal region, posterior to the left atrium (Fig. 2). Abdominal CT scan showed a round lesion in the right lobe of the liver consistent with either a metastasis or a hemangioma (Fig. 2). Whole body Octreotide scan showed uptake concordant with the mediastinal mass but not with the liver lesion (Fig. 3).

An 18 gram mediastinal tumor was resected. Pathological analysis in Teheran University of Medical Sciences demonstrated the tumor to be highly positive for synaptophysin and chromogranin and scattered positivity for EMA, GH and S100 and negative for cytokeratin. Tumor Ki-67 was estimated at 7%. Subsequent pathological analysis at Cedars-Sinai Medical Center in Los Angeles was obtained and showed a monomorphous population of tumor cells arranged in nests and solid sheets. Tumor cells had moderate amounts of pale eosinophilic cytoplasm and uniform round nuclei with salt and pepper-like nuclear chromatin. Nucleoli were inconspicuous. Rare mitotic figures were noted (1/10 HPF) and there was no evidence of necrosis. Rare sustentacular cells were highlighted by S100 stain. Immunostaining confirmed that the tumor was immunopositive for GHRH (Fig. 4). Immunostaining for TTf-1 and for GH were both negative. There was strong and diffuse immunoreactivity for chromogranin and no staining for keratin (AE1/AE3). Final pathologic diagnosis was that of paraganglioma.

Despite postoperative symptom improvement, subsequent biochemical evaluation showed elevated basal serum GH and IGF-1 levels, 2.9 and 506 ng/ml respectively. The patient underwent a CT guided biopsy of the hepatic lesion, and histopathologic evaluation revealed that the tissue biopsy had morphologic characteristics similar to the original tumor. The patient was then started on monthly injection of long-acting octreotide 20 mg per month which resulted in normalization of serum IGF-1 (132 ng/ml at 3 months and 196 ng/ml at one year) and lowering of basal GH levels to 0.7 and 0.9 ng/ml, at 3 and 12 months, respectively.

Paragangliomas

The above case is the first published report of acromegaly caused by ectopic GHRH secretion secondary to a mediastinal paraganglioma.

Paragangliomas originate from extra-adrenal chromaffin cells located outside adrenal glands and comprise 10-18% of all catecholamine secreting tumors of the body. The tumors are located along the sympathetic chain from neck to the pelvis with the majority in the abdominal and pelvic region. While parasympathetic paragangliomas are primarily localized to neck or base of skull, sympathetic paragangliomas are mainly in the abdominal and pelvic region. Paragangliomas may manifest with symptoms of catecholamine hypersecretion, mass effect or may be incidentally discovered during imaging. Mediastinal paragangliomas are very rare and comprise only 2% of extra-adrenal catecholamine secreting tumors. Secretion of ectopic hormones from these tumors is very unusual. Most reported tumors secrete norepinephrine presenting with hypertension. Presence and bioactivity of GHRH has been demonstrated in pheochromocytomas, but not in paragangliomas [60, 83, 84]. In one series of 14 cases of mediastinal paragangliomas all patients had hypertension and one case was a nonfunctional paraganglioma [85-87]. Approximately one fourth to one third of paragangliomas are hereditary and can be seen in the settings of familial paragangliomas, neurofibromatosis type 1, Von Hippel-Lindau disease, Carney triad and rarely MEN 2. Mutations in succinate dehydrogenase (SDH) gene or mitochondrial complex 2 gene are responsible for most cases of hereditary paragangliomas.

Evaluation of patients for paragangliomas includes assessment of functional status through measurement of plasma or urinary fractionated metanephrines. Computed tomographic scanning (CT) can detect extra adrenal paragangliomas larger than 2 cm with a sensitivity of 95% and a specificity of 70%. Many tumors harbor extensive areas of necrosis. MRI has a sensitivity of 90% and a specificity of 70% in extra adrenal paragangliomas with brightness in T2 weighted images - a characteristic finding of pheochromocytoma and paragangliomas [82]. Somatostatin receptor scintigraphy (SRS) has also been used in localization of

paraganglioma though In-Octreotide may be more sensitive and specific in detecting head and neck paragangliomas [88].

Metaiodo-benzylguanidine (MIBG) using iodine-123 has an overall sensitivity of 83–100% and specificity of 95–100% [89]. However, in metastatic paragangliomas and head and neck paragangliomas, the sensitivity decreases to as low as 65% [90, 91]. In addition, Gallium-68 somatostatin receptor PET or PET/CT have a 93% sensitivity and 91% specificity for thoracic and gastroenteropancreatic NETs [92] and may eventually become first line tests in localization [93].

On light microscopy, paragangliomas are composed of cells arranged in nests or trabecular (zellballen pattern), separated with a thin vascular stroma. On electron microscopy, large dense core catecholamine-secreting granules measuring 150–250 nm in diameter can be detected [82].

In the current case, there were no specific symptoms of catecholamine excess as hypertension was mild and well controlled. Indeed, the patient did not present with features of a paraganglioma but rather presented functionally with acromegaly.

Clinical presentation of ectopic acromegaly

Signs and symptoms of ectopic acromegaly are similar to pituitary dependent acromegaly. In some cases, patients may present with symptoms specific to the corresponding tumor such as jaundice, cough, unilateral wheezing or other endocrinopathies [57, 67]. There may be an association of ectopic acromegaly with MEN1 as reported in 19 patients and presumptively in 5 additional cases [28, 40, 44, 51, 54, 55, 57–63, 66, 68, 69, 77]. Accordingly common genetic pathways may underlie the pathogenesis of the 2 disorders. Secretory ability of the tumor can change over time, as in a case of a GHRH producing tumor which changed to an insulin producing tumor [73]. Several cases of co-secretion of GHRH with other hormones such as ACTH have been reported [51, 76, 80].

Diagnostic evaluation of ectopic acromegaly

Distinction between pituitary and ectopic acromegaly is essential as pituitary hyperplasia secondary to GHRH production in ectopic acromegaly may be misdiagnosed as pituitary tumor leading to unnecessary pituitary surgery. Appropriate diagnosis of the source of acromegaly is of utmost significance as treatment strategies differ. In most cases in the literature, the preliminary diagnosis was pituitary acromegaly and diagnosis of ectopic acromegaly was confirmed either during the diagnostic work up or after unsuccessful pituitary surgery. There was a delay of 2 to 18 years between pituitary surgery and establishment of correct diagnosis [29, 45, 66]. Most patients had pituitary hyperplasia visible on imaging with some characteristics indistinguishable from pituitary adenomas, leading often to unnecessary transsphenoidal surgery (TSS) [14, 21, 22, 28, 29, 34, 39-41, 43, 44, 51, 53, 56, 62, 63, 66, 67, 69, 70, 74, 78, 79, 81, 94, 95]. Of the 98 reported cases, 30 patients underwent transphenoidal resection for presumed GH secreting pituitary adenoma. Key findings leading to correct diagnosis of ectopic acromegaly are summarized in Table 2. Pituitary surgery was indicated in one case where pituitary macro-adenoma was the origin of ectopic GHRH secretion [81] and in other cases where the tumor had metastasized to the pituitary gland [95, 96].

In most patients, corresponding tumors were detected by CT or MRI. The size of the tumors ranged from 1 to 25 cm (6.3 ± 4.0 , mean \pm SD). Pituitary enlargement was present in about 75% of cases and 1/3 were diagnosed with a pituitary adenoma. In some cases, normal appearance of the pituitary gland prompted workup for ectopic acromegaly (Table 4).

Positive Octreotide scan in 19 out of 21 cases shows a high sensitivity for this modality in the diagnosis of GHRH secreting tumors.

Serum levels of GH and IGF-1 are invariably elevated in patients with ectopic acromegaly, with nadir GH on OGTT > 1 mcg/L and IGF-1 levels above the age and gender matched normal range [97]. More sensitive assays lower the nadir GH cutoff to > 0.3 mcg/L [98]. Mean values of GH after OGTT may show a paradoxical increase.

Elevation of serum GH after thyrotropin releasing hormone (TRH) stimulation has been shown in some cases [53], but other laboratory tests are not useful in distinguishing pituitary from ectopic acromegaly. Measurement of plasma GHRH is the simplest and most accurate for diagnosis provided the laboratory facilities are available. The values for GHRH which are usually undetectable in pituitary acromegaly are elevated in patients with ectopic acromegaly reaching values hundreds or thousands-fold normal values [36, 44, 67]. Laboratory results including basal and post GTT values, serum IGF-1 and plasma GHRH for reported cases are summarized in Table 3. Thyrotropin releasing hormone (TRH) stimulation test was performed in 31 cases.

Diagnostic criteria for ectopic acromegaly include elevated plasma GHRH, complete recovery after resection of the corresponding tumor, positive GHRH immunostaining, positive bioassay (the ability of cultured rat pituitary cells to produce GH in response to the tumor extract) [15, 53, 70], arterio-venous gradient of GHRH across the tumor [31], detection of GHRH mRNA [64], or GHRH extraction from the tumor tissue [22, 31, 53, 56].

Pathologic evaluation of GHRH secreting tumors

Since 1985, immunohistochemistry (IHC) has been used to pathologically characterize GHRH secreting tumors. Positive immunoexpression for GHRH is seen in most tumors with ectopic acromegaly, though GHRH immunopositivity may not translate to biochemical and clinical manifestations of acromegaly [99]. The discordance between immunoexpression and clinical manifestations has been attributed to low concentration of GHRH in the vesicles, intravesicular degradation of GHRH or defects in secretory capacity of GHRH [99].

Histopathologic evaluation of reported GHRH secreting tumors confirmed that all except three were of neuroendocrine origin. One of these was an adenoid cystic carcinoma of the lung [80] and the other was a pituitary adenoma secreting both GH and GHRH [81]; the third one was a case of acromegaly with elevated GHRH and normal pituitary MRI in whom no tumor was identified despite extensive work-up [51]. Eighty four were bronchial carcinoid or endocrine pancreas tumors (Table 1). In 55 tumors, GHRH immunostaining was positive. In one case, tissue GHRH was measured using HPLC [22]. Bioassay was employed as confirmation of GHRH bioactivity in 3 cases [15, 53, 70].

General neuroendocrine markers such as chromogranin A, synaptophysin and NSE were positive by IHC in most cases. Results of pituitary histopathology from 29 cases are shown in Table 4.

In this report, plasma GHRH measurement was unavailable. However, negative pituitary imaging, positive IHC expression for GHRH and persistently elevated IGF-1 levels in the presence of hepatic metastasis are suggestive of a diagnosis of ectopic acromegaly secondary to GHRH secretion.

Management of ectopic acromegaly

Management of acromegaly from ectopic GHRH secretion consists of surgical resection of the primary tumor and biochemical normalization. Surgery is the treatment of choice and may be curative. In patients with extensive metastases or with unresectable tumor, somatostatin analogues may be used [22, 45, 47, 51]. Treatment modalities of the reported cases consisted of tumor resection in 55 cases, somatostatin analogue therapy in 13 patients, and adjuvant chemotherapy and radiotherapy in 2 patients. Of 90 cases, cure was documented in 44%, partial improvement in 31%, no improvement in 5%, and deaths in 10%. Lymph node or distant metastases were documented in 39 cases [15, 19, 21–24, 28–30, 32, 33, 43–45, 51, 55, 57, 60, 65, 70, 72–76, 80]. Outcome data was ascertained in 90 cases with mean follow-up of 5.3 ± 5.5 years. Although there is limited follow up of patients, GHRH producing NETs have a favorable prognosis if diagnosed early and surgically resected, as other forms of NET, even in the presence of metastases [100–102]. Anti-angiogenic medications and tyrosine kinase inhibitors may potentially have a role in inhibiting tumor growth of NETs [103, 104].

In the index case reported here, a mediastinal tumor was completely resected, but biochemical acromegaly persisted postoperatively due to a functional liver metastasis. Surgical resection of the liver metastasis would be the preferred treatment but the patient opted for therapy with monthly injections of Octreotide LAR which has controlled GH levels.

Conclusion

We report acromegaly secondary to ectopic GHRH secretion from a mediastinal paraganglioma in the context of ectopic acromegaly. Criteria consistent with this diagnosis include: clinical signs and symptoms of acromegaly, biochemically confirmed acromegaly, normal pituitary imaging, a pathologically confirmed mediastinal paraganglioma, and positive immunostaining for GHRH. The case highlights that paragangliomas may be functional and though very rare, can lead to ectopic GHRH secretion and consequent acromegaly.

Methods

Hormonal analysis

Plasma GH, IGF1, FSH, LH, TSH, T₄, T₃, cortisol and prolactin concentrations were measured by commercial kits.

Histopathologic examination

Resected specimens were fixed in formalin and embedded in paraffin. Paraffin sections were used for Hematoxylin-Eosin staining. Immunocytochemistry was performed using 0.03% H_2O_2 (DAKO solution). Immunohistochemical staining was performed with antibodies against chromogranin, synaptophysin, EMA, GH, S 100 and cytokeratin. Ki-67 nuclear antigen immunostaining was also performed. Rabbit anti-GHRH antibody was used at dilution 1:1000 and kindly provided by Dr. Lawrence Frohman (Univ. of Illinois at Chicago).

Imaging studies

High resolution computerized tomography (HRCT) scanning was used for chest imaging and T1-weighted sagittal and coronal pituitary MRI views were used. Whole body Somatostatin receptor scintigraphy was performed using ^{99m}Tc-pentetreotide. Also ^{99m}Tc-

Ectopic acromegaly cases in the literature

A comprehensive search of the English literature revealed that 98 cases of acromegaly secondary to ectopic secretion of GHRH were published between 1974 and 2011. Most were single case reports, and there were limited reviews on the topic. The results were prepared and analyzed using SPSS version 16. Data of the present case is also included in the analysis.

Acknowledgments

This work was supported by National Institutes of Health Grant K23DK085148 (O.C.) and Grant CA75979 (S.M.).

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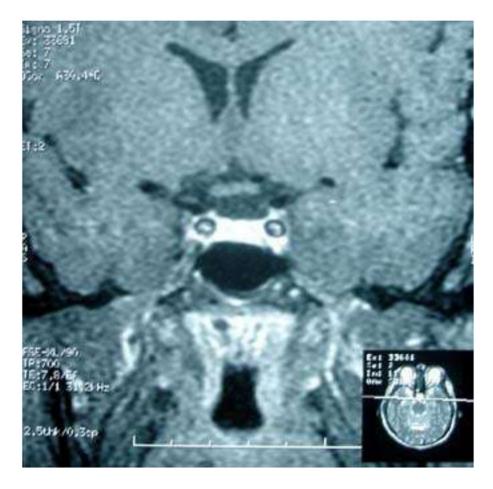


Figure 1.

MRI of pituitary, T1 weighted, coronal image demonstrates normal sized pituitary gland without an adenoma.



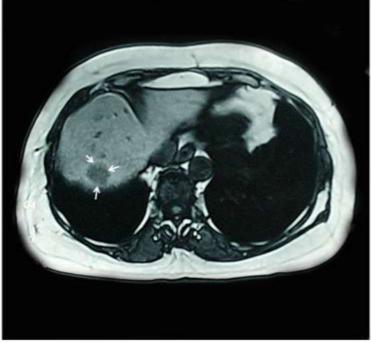


Figure 2.

a. CT scan of the chest, coronal image demonstrates a 4.5×3.5 cm mediastinal mass. b. CT scan of the abdomen demonstrates a hepatic mass.

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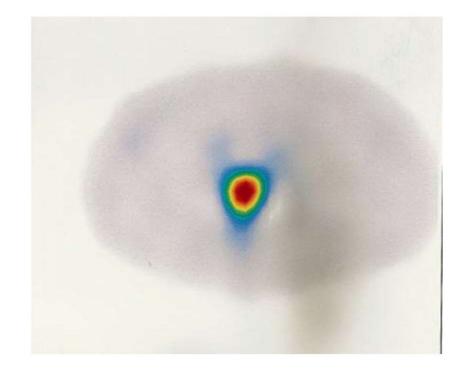


Figure 3.

Octreotide scan shows positive uptake corresponding to the mediastinal mass but not to the hepatic lesion.

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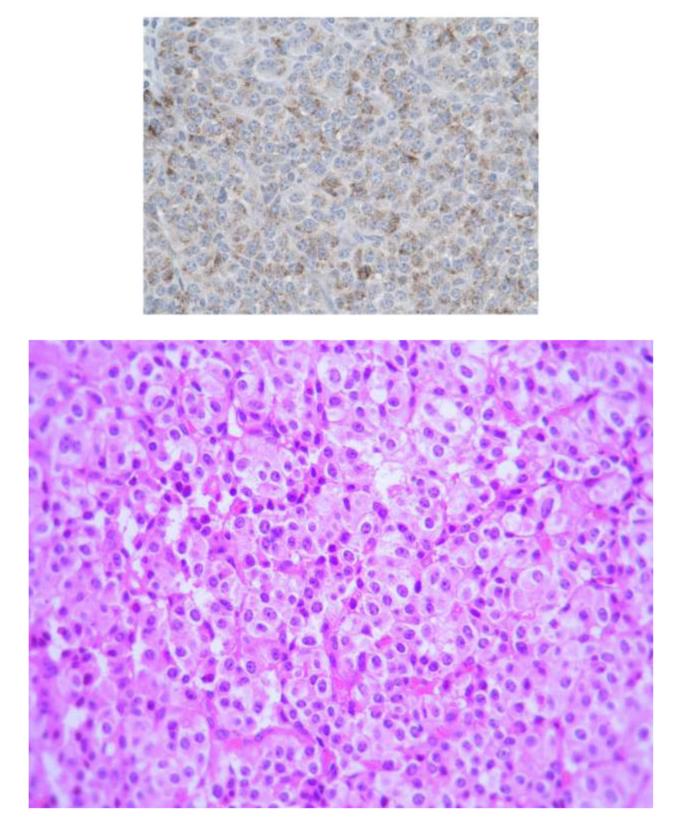


Figure 4.

a. H&E staining of mediastinal paraganglioma. b. tumor is positive for GHRH staining.

Table 1

Clinical features of 74 patients with ectopic acromegaly

n	%
64	64.6%
35	35.4%
Mean ± SD	range
40.4 ± 14.1	14–77
8.3 ± 5.8	1-22
6.3 ± 4.0	1–25
n	%
51	51.5
34	34.3
7	7.1
2	2
1	1
2	2
1	1
1	1
99	100
	$ \begin{array}{c} 64\\ 35\\ Mean \pm SD\\ 40.4 \pm 14.1\\ 8.3 \pm 5.8\\ 6.3 \pm 4.0\\ \mathbf{n}\\ 51\\ 34\\ 7\\ 2\\ 1\\ 2\\ 1\\ 1\\ 1 \end{array} $

* references 12-81

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Table 2

Findings which led to diagnosis of ectopic acromegaly

	n	%
Acromegaly in the setting of a known non-pituitary tumor	41	45.6
Persistence of acromegaly after transsphenoidal surgery	23	25.6
Incidentally discovered	12	13.3
Normal pituitary imaging	11	12.2
Routine GHRH measurement	3	3.3

Table 3

Laboratory results of patients with ectopic acromegaly

		(n)	Mean ± SD
	random	(70)	60 ± 81
Serum GH (ng/ml)			
	baseline GTT	(66)	35 ± 40
	post GTT	(55)	53 ± 77
	post TRH	(28)	187 ± 212
IGF1 [*]		(69)	2.56 ± 1.06
GHRH (ng/ml)		(73)	7693 ± 20276

 * IGF 1 values are expressed as patient's IGF 1/upper normal limit of age matched controls' IGF 1

Table 4

Pituitary imaging and histopathology in patients with ectopic acromegaly

		n	%
	Enlarged	41	46
Pituitary imaging			
	Adenoma	27	30
	Empty sella	2	2
	Normal	18	20
	Microcystic Lesion	2	2
Pituitary pathology	Hyperplasia	16	55
	Adenoma	6	21
	Others	6	21
	Normal	1	3