- Stoop B, Lehner A, Iversen C, Fanning S, Stephan R. Development and evaluation of *rpoB* based PCR systems to differentiate the six proposed species within the genus *Cronobacter*. Int J Food Microbiol. 2009;136:165–8. http://dx.doi.org/10.1016/j.ijfoodmicro.2009.04.023
- Jarvis KG, Grim CJ, Franco AA, Gopinath G, Sathyamoorthy V, Hu L, et al. Molecular characterization of Cronobacter lipopolysaccharide O-antigen gene clusters and development of serotype-specific PCR assays. Appl Environ Microbiol. 2011;77:4017–26. http://dx.doi.org/10.1128/AEM.00162-11
- Hsueh PR, Teng LJ, Lee LN, Yu CJ, Yang PC, Ho SW, et al. Melioidosis: an emerging infection in Taiwan? Emerg Infect Dis. 2001;7:428–33.

Address for correspondence: Po-Ren Hsueh, Departments of Laboratory Medicine and Internal Medicine, National Taiwan University Hospital, #7, Chung-Shan South Rd, Taipei 100, Taiwan; email; hsporen@ntu.edu.tw

## MethicillinResistant Staphylococcus pseudintermedius in Rats

To the Editor: Staphylococcus pseudintermedius is a coagulasepositive species in the S. intermedius group. Previously misidentified as S. intermedius, S. pseudintermedius is now recognized as a leading cause of opportunistic infection in dogs (1) and a cause of sporadic infections in other species, including humans (1,2). Additionally, evidence of zoonotic transmission of S. pseudintermedius from dogs to humans has been reported (3,4). Although information regarding the pathogenic process of S. pseudintermedius is limited, the bacterium is known to possess virulence factors similar to those found in S. aureus, including a leukotoxin comparable to the Panton-Valentine leukocidase associated with community-acquired *S. aureus* infection (1).

Of concern is the emergence and widespread international recognition of methicillin-resistant S. pseudintermedius (MRSP) (I). One veterinary laboratory noted a 272% increase in MRSP cases from 2007–2008 through 2010–2011 (S). As with methicillin-resistant S. aureus, MRSP resistance is conferred by the mecA gene, making MRSP resistant to all  $\beta$ -lactam antimicrobial drugs and some other antimicrobial drug classes (I). Compared with methicillin-susceptible strains, MRSP seems better able to colonize humans (S).

The potential for zoonotic transmission and concerns that MRSP could be mistaken for other methicillin-resistant staphylococci (1,2) suggest the need for further investigation into the epidemiology of this pathogen. One question yet to be addressed is whether commensal pests, particularly rats (Rattus spp.), could serve as a source of MRSP because of their pervasiveness, their propensity toward close contact with humans, and the fact that they are the source of several other zoonotic diseases (6). We report MRSP carriage in wild Norway rats (R. norvegicus) in Vancouver, British Columbia, Canada.

During September–November 2011, Norway rats were trapped in a random sample of alleys in Vancouver's Downtown Eastside, an impoverished neighborhood with high levels of homelessness, intravenous drug use, and HIV infection. Immediately after the rats were euthanized, a sterile swab was used to sample the oropharynx and nares of each rat.

Swabs were placed in 2 mL of enrichment broth containing 10 g/L tryptone T, 75 g/L sodium chloride, 10 g/L mannitol, and 2.5 g/L yeast extract and incubated for 24 h at 35°C. Aliquots of 100  $\mu$ L were streaked onto mannitol salt agar with 2  $\mu$ g/mL oxacillin and incubated at 35°C for 48 h. Suspected staphylococcal isolates were subcul-

tured onto Columbia blood agar and identified according to colony morphologic appearance, Gram staining, and catalase reaction. Tube coagulasepositive isolates were speciated by using a multiplex PCR specific for the thermonuclease (nuc) gene (7). Methicillin resistance was confirmed by demonstrating penicillin-binding protein 2a antigen with the latex-agglutination test (Oxoid Ltd., Basingstoke, UK). Isolates were typed by sequencing of the *mec*-associated direct repeat unit (dru typing) (8). Antimicrobial drug susceptibility was evaluated by broth microdilution (Sensititre; Trek Diagnostics, Cleveland, OH, USA), according to Clinical and Laboratory Standards Institute guidelines (www. clsi.org/). The study was approved by the University of British Columbia Animal Care Committee.

MRSP was isolated from 5 (2.1%) of 237 rats trapped. However, lack of standardized screening methods for MRSP could have resulted in underestimation of MRSP prevalence. Of the 5 isolates, 3 were *dru* type dt11a, a strain commonly found in dogs (8), and the other 2 were a novel *dru* type (assigned dt7ac). All isolates tested demonstrated resistance to multiple antimicrobial drug classes (Table).

Carriage of MRSP has not been identified in wild rats; therefore, the epidemiologic and public health implications of these findings are difficult to determine. However, the isolation of a common dog-associated dru type from rats suggests that MRSP might be transmissible between dogs and rats. This possibility is not surprising given the potential for direct and indirect contact between these species. Indeed, rat-to-dog transmission of other bacterial pathogens has been recognized (9). Detection of a dru type not previously detected methicillin-resistant staphylococci suggests that these isolates might have evolved independently of methicillin-resistant staphylococci in other animal species.

Table. Sensitivity profiles for methicillin-resistant *Staphylococcus pseudintermedius* isolated from wild Norway rats, Vancouver, British Columbia, Canada\*

	Isolate no. (strain type)†			
Drug	1 (dt11a)	2 (dt11a)	3 (dt11a)	4 (dt7ac)
β-lactams	R	R	R	R
Clindamycin	R	R	R	R
Erythromycin	R	R	R	R
Daptomycin	S	S	S	S
Linezolid	S	S	S	S
Gentamicin	R	S	S	S
Tetracycline	S	S	S	R
Tigecycline	S	S	S	S
Trimethoprim-sulfamethoxazole	S	S	S	S
Quinupristin–dalfopristin	R	S	S	S
Rifampin	R	S	S	S
Chloramphenicol	S	S	S	S
Nitrofurantoin	S	S	S	S
Ciprofloxacin	R	S	S	S
Levofloxacin	l	S	S	S
Moxifloxacin	I	S	S	S
Vancomycin	S	S	S	S

\*Of 5 isolates, 1 was not available for antimicrobial susceptibility testing. R, resistant; S, sensitive; I, intermediate.

†Strain types identified through direct repeat unit typing

Rat carriage of MRSP does not prove that rats are capable of transmitting the bacterium to humans; however, rats are a source of other zoonotic pathogens (6). These pathogens are most commonly transmitted from rats to humans indirectly, through contamination of the environment or foodstuffs (6). Of note, MRSP has been shown to survive for extended periods in the environment (4), suggesting a mechanism through which the bacterium could be passed from rats to humans and dogs or vice versa.

In 2011, MRSA was detected in bedbugs from Vancouver's Downtown Eastside (10). We have identified another urban pest in this area as a potential source of a multidrug-resistant pathogen. As with bedbugs, rat infestations are most common in impoverished, inner-city neighborhoods, and residents of these neighborhoods are at greatest risk for rat-to-human pathogen transmission. These findings suggest that inner-city urban rats warrant further investigation as a potential source of MRSP and other contemporary zoonotic pathogens, including multidrug-resistant bacteria.

This study was supported by the Canadian Institutes of Health Research (MOP-119530 and CGV-104833), Murray

Wightman and Stuart McMillan, the British Columbia Centre for Disease Control, and the Urban Health Research Initiative. Field collection of rats was made possible by the assistance of the Vancouver Area Network of Drug Users.

## Chelsea Gardner Himsworth, David M. Patrick, Kirbee Parsons, Alice Feng, and J. Scott Weese

Author affiliations: University of British Columbia, Vancouver, British Columbia, Canada (C.G. Himsworth, D.M. Patrick, K. Parsons, A. Feng); British Columbia Ministry of Agriculture, Abbotsford, British Columbia, Canada (C.G.Himsworth); and University of Guelph, Guelph, Ontario, Canada (J.S. Weese)

DOI: http://dx.doi.org/10.3201/eid1901.120897

## References

- van Duijkeren E, Catry B, Greko C, Moreno MA, Constanca Pomba M, Pyorala S, et al. Review on methicillin-resistant Staphylococcus pseudintermedius. J Antimicrob Chemother. 2011;66:2705–14. http://dx.doi.org/10.1093/jac/dkr367
- Weese JS, van Duijkeren E. Methicillin-resistant Staphylococcus aureus and Staphylococcus pseudintermedius in veterinary medicine. Vet Microbiol. 2010;140:418–29. http://dx.doi. org/10.1016/j.vetmic.2009.01.039

- Paul NC, Moodley A, Ghibaudo G, Guardabassi L. Carriage of methicillin-resistant
   *Staphylococcus pseudintermedius* in small
   animal veterinarians: indirect evidence of
   zoonotic transmission. Zoonoses Public Health. 2011;58:533–9. http://dx.doi.
   org/10.1111/j.1863-2378.2011.01398.x
- Laarhoven LM, de Heus P, van Luijn J, Duim B, Wagenaar JA, van Duijkeren E. Longitudinal study on methicillin-resistant *Staphylococcus pseudintermedius* in households. PLoS ONE. 2011;6:e27788. http://dx.doi.org/10.1371/journal.pone.0027788
- Steen S. Meticillin-resistant strains of Staphylococcus pseudintermedius in companion animals. Vet Rec. 2011;169:53–4. http://dx.doi.org/10.1136/vr.d4247
- Meerburg BG, Singleton GR, Kijlstra A. Rodent-borne diseases and their risks for public health. Crit Rev Microbiol. 2009;35:221–70. http://dx.doi.org/10.1080/10408410902989837
- Sasaki T, Tsubakishita S, Tanaka Y, Sakusabe A, Ohtsuka M, Hirotaki S, et al. Multiplex-PCR method for species identification of coagulase-positive staphylococci.
   J Clin Microbiol. 2010;48:765–9. http://dx.doi.org/10.1128/JCM.01232-09
- 8. Kadlec K, Schwarz S, Weese JS. Evaluation of direct repeat unit (*dru*) typing for characterization of methicillin-resistant *Staphylococcus pseudintermedius*. In: Abstracts of the 2nd ASM-ESCMID Conference on Methicillin-Rresistant Staphylococci in Animals: Veterinary and Public Health Implications; Washington, DC, USA; 2011 Sept 8–11; Abstract 55. Washington (DC): American Society for Microbiology; 2011.
- Wouters EG, Ho HT, Lipman LJ, Gaastra W. Dogs as vectors of *Streptobacillus moniliformis* infection? Vet Microbiol. 2008;128:419–22. http://dx.doi.org/10.1016/j.yetmic.2007.10.019
- Lowe CF, Romney MG. Bedbugs as vectors for drug-resistant bacteria. Emerg Infect Dis. 2011;17:1132–4. http://dx.doi. org/10.3201/cid1706.101978

Address for correspondence: Chelsea G. Himsworth, 1767 Angus Campbell Rd, Abbotsford, British Columbia, V3G 2M3, Canada; email: Chelsea.Himsworth@gov.bc.ca

