

Control of *Candida albicans* Metabolism and Biofilm Formation by *Pseudomonas aeruginosa* Phenazines

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ABSTRACT *Candida albicans* has developmental programs that govern transitions between yeast and filamentous morphologies and between unattached and biofilm lifestyles. Here, we report that filamentation, intercellular adherence, and biofilm development were inhibited during interactions between *Candida albicans* and *Pseudomonas aeruginosa* through the action of *P. aeruginosa*-produced phenazines. While phenazines are toxic to *C. albicans* at millimolar concentrations, we found that lower concentrations of any of three different phenazines (pyocyanin, phenazine methosulfate, and phenazine-1-carboxylate) allowed growth but affected the development of *C. albicans* wrinkled colony biofilms and inhibited the fungal yeast-to-filament transition. Phenazines impaired *C. albicans* growth on nonfermentable carbon sources and led to increased production of fermentation products (ethanol, glycerol, and acetate) in glucose-containing medium, leading us to propose that phenazines specifically inhibited respiration. Methylene blue, another inhibitor of respiration, also prevented the formation of structured colony biofilms. The inhibition of filamentation and colony wrinkling was not solely due to lowered extracellular pH induced by fermentation. Compared to smooth, unstructured colonies, wrinkled colony biofilms had higher oxygen concentrations within the colony, and wrinkled regions of these colonies had higher levels of respiration. Together, our data suggest that the structure of the fungal biofilm promotes access to oxygen and enhances respiratory metabolism and that the perturbation of respiration by bacterial molecules such as phenazines or compounds with similar activities disrupts these pathways. These findings may suggest new ways to limit fungal biofilms in the context of disease.

IMPORTANCE Many of the infections caused by *Candida albicans*, a major human opportunistic fungal pathogen, involve both morphological transitions and the formation of surface-associated biofilms. Through the study of *C. albicans* interactions with the bacterium *Pseudomonas aeruginosa*, which often coinfects with *C. albicans*, we have found that *P. aeruginosa*-produced phenazines modulate *C. albicans* metabolism and, through these metabolic effects, impact cellular morphology, cell-cell interactions, and biofilm formation. We suggest that the structure of *C. albicans* biofilms promotes access to oxygen and enhances respiratory metabolism and that the perturbation of respiration by phenazines inhibits biofilm development. Our findings not only provide insight into interactions between these species but also provide valuable insights into novel pathways that could lead to the development of new therapies to treat *C. albicans* infections.

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Candida albicans, a commensal resident of mucosal surfaces, can overgrow under favorable conditions, leading to a wide range of diseases, referred to as candidiasis (1). *C. albicans* infections often involve more than one cellular morphology; yeast, pseudohyphal, and hyphal growth forms have all been associated with virulence (2). It has been proposed that filaments are responsible for tissue penetration, whereas yeasts play an important role in early dissemination and in less invasive disease infections (3). *C. albicans* infections are also associated with its formation of biofilms, which are dense single-species and mixed-species populations adhering to one another through the action of surface adhesins and extracellular polymers. Biofilms on implanted medical devices can serve as a source for infection, and dense *C. albicans*

populations within the host may have properties in common with biofilms, such as high levels of drug resistance and resistance to host immune defenses (4, 5). Importantly, biofilm formation involves filamentation and adhesin-mediated aggregation of cells (6, 7).

Many environmental cues impact fungal morphology and biofilm formation. Many conditions, such as hypoxia, elevated extracellular pH, *N*-acetylglucosamine (GlcNAc), amino acids, low concentrations of glucose, body temperature, and elevated CO₂ have been associated with *C. albicans* filamentation (8, 9). There have been a number of studies that also indicate that metabolic status can influence the decision to form hyphae (10–16). There are still many questions regarding how the fungus integrates these

different signals and whether there are important links between known inducers of hyphal growth, biofilm formation, and metabolism.

C. albicans and bacteria can coexist within the host (17–19), where the nature of the interspecies interactions can determine the fate of the microbial populations (20) and thus probably the outcome of polymicrobial diseases. *C. albicans* and *Pseudomonas aeruginosa*, two species commonly found together in mixed-species, biofilm-related infections (21), interact in many different ways through physical association, killing by secreted factors and signaling events that modulate virulence properties (22–29). *P. aeruginosa* adheres to and forms biofilms on *C. albicans* filaments (23) and ultimately causes the death of the fungal hypha. A *P. aeruginosa* quorum-sensing molecule also induces a transition to yeast growth, and yeasts are more resistant to killing by *P. aeruginosa* (23). Redox-active phenazines produced by *P. aeruginosa* play a key role in controlling *C. albicans* when the two species are in close proximity (24, 29). We reported that *C. albicans* growth was antagonized by two *P. aeruginosa* phenazines, pyocyanin (PYO) and its biosynthetic precursor 5-methylphenazinium-1-carboxylate (5MPCA), as well as by a synthetic methylphenazinium analog phenazine methosulfate (PMS), which we have shown to be a surrogate for studying 5MPCA's antifungal activity (24, 29) (see Fig. S1 in the supplemental material for chemical structures). Phenazines play important roles in bacterial interactions with fungi in which high concentrations of phenazines inhibit fungal growth (30).

Here, we demonstrate a new role for bacterial phenazines as modulators of *C. albicans* metabolism and community behavior. In the presence of phenazine-producing *P. aeruginosa* or low concentrations of purified phenazines, the fungus grew but no longer developed wrinkled colonies or robust biofilms on plastic surfaces. Phenazines markedly inhibited the yeast-to-filament transition. Consistent with published data indicating that phenazines can impact mitochondrial activity (31, 32), their presence led to increased production of fermentation products and decreased respiratory activity at concentrations that are found in clinical samples (33). Methylene blue (MB), which can also interact with the electron transport chain (ETC) (34, 35), inhibited the development of wrinkled *C. albicans* colony biofilms. These studies also revealed a link between wrinkles in colony biofilms and respiratory activity, and this finding was supported by the detection of higher oxygen concentrations in wrinkled colonies than in smooth and flat colonies comprised of yeast. Together, our data revealed a previously unidentified role for phenazines as modulators of *C. albicans* metabolism and, through these effects, on cellular morphology, colony development, and biofilm formation on solid surfaces. Moreover, this work also provides new insights into the links between metabolism, morphogenesis, and community behaviors such as biofilm formation in *C. albicans*, and this may represent an emerging theme among the microbes.

RESULTS

***P. aeruginosa* phenazines repress *C. albicans* biofilm formation and hyphal growth.** When wild-type (WT) *P. aeruginosa* strain PA14 and WT *C. albicans* strain SC5314 were grown adjacent to one another, a striking change in *C. albicans* colony morphology occurred, with a smooth zone in the region adjacent to the bacterial colony and wrinkling in regions of the fungal streak that were more distant from the bacterial colony (Fig. 1A). In contrast, *C. al-*

bicans cocultured with a phenazine-deficient (Δphz) *P. aeruginosa* strain wrinkled robustly up to the edge of the bacterial colony, suggesting that the soluble excreted phenazines played a role in this effect on *C. albicans* colony morphology (Fig. 1A). When we cocultured *C. albicans* with *P. aeruginosa* mutants altered in the synthesis of individual phenazines (see Fig. S1 in the supplemental material for pathway), repression of colony wrinkling still occurred (see Fig. S2). Mutants unable to synthesize pyocyanin (PYO) (*phzS::TnM*) or 5MPCA (*phzM::TnM*) but capable of producing phenazine-1-carboxylate (PCA) still produced smooth zones.

To determine if phenazines were directly responsible for the change in *C. albicans* colony morphology, we spot inoculated *C. albicans* onto medium with and without purified phenazines. Control cultures without phenazines formed wrinkled colony structures, while colonies grown in the presence of low concentrations of any of three phenazines, PYO, PMS, or PCA, were smooth, flat, and undifferentiated (Fig. 1B; see also Fig. S3A in the supplemental material). The effects of different concentrations of PMS and PYO were tested, and complete repression of colony wrinkling was apparent at concentrations as low as 5 μM for PMS and 10 μM for PYO (see Fig. S3B), indicating that the concentrations required to inhibit wrinkled colony development were ~ 100 -fold below those shown previously to be toxic to *C. albicans* (24, 29, 36).

C. albicans hyphal growth has been associated with both wrinkled colony morphology and biofilm formation on plastic surfaces (5, 37). We found that wrinkled colonies, formed upon growth on medium with GlcNAc at 37°C, exhibited some biofilm-associated characteristics, including cells in hyphal, pseudohyphal, and yeast morphologies that remained associated with one another even upon suspension in liquid (Fig. 1B and data not shown). In contrast, smooth colonies formed in the presence of phenazines were comprised almost exclusively of yeast that dispersed easily upon transfer to liquid (Fig. 1B and data not shown). In addition to the change in cellular morphology, phenazines repressed expression of *HWPI*, a gene that is strongly correlated with hyphal growth and involved in biofilm formation (38, 39), as shown using a strain harboring a *lacZ* transcriptional reporter fused to the *HWPI* promoter. A strain expressing an actin transcriptional reporter (*ACT1::lacZ*) did not show differences between colonies grown in the presence or absence of phenazines (see Fig. S3C). Colonies grown at 30°C without GlcNAc (noninducing conditions) were smooth and contained only yeast (Fig. 1B); no phenotypic changes in colony morphology were observed upon inclusion of phenazines in the agar medium under these conditions (see Fig. S3D). The total numbers of CFUs in smooth colonies formed in the presence of either PMS or PYO were slightly but significantly ($P < 0.05$) lower than those in control colonies (Fig. 1C).

Consistent with the data that indicated that colony wrinkling and hypha formation were repressed by phenazines on agar surfaces, we found that these compounds inhibited *C. albicans* hyphal growth and biofilm formation in liquid. Planktonic control cultures grown under filament-inducing conditions contained long undulating filaments, whereas cultures exposed to either 5 μM PMS or 20 μM PMS contained small ovoid yeast cells (see Fig. S4A). Biofilm assays on polystyrene surfaces showed a significant reduction in biofilm formation in the presence of PMS or PYO in comparison to that of the untreated controls (see Fig. S4B).

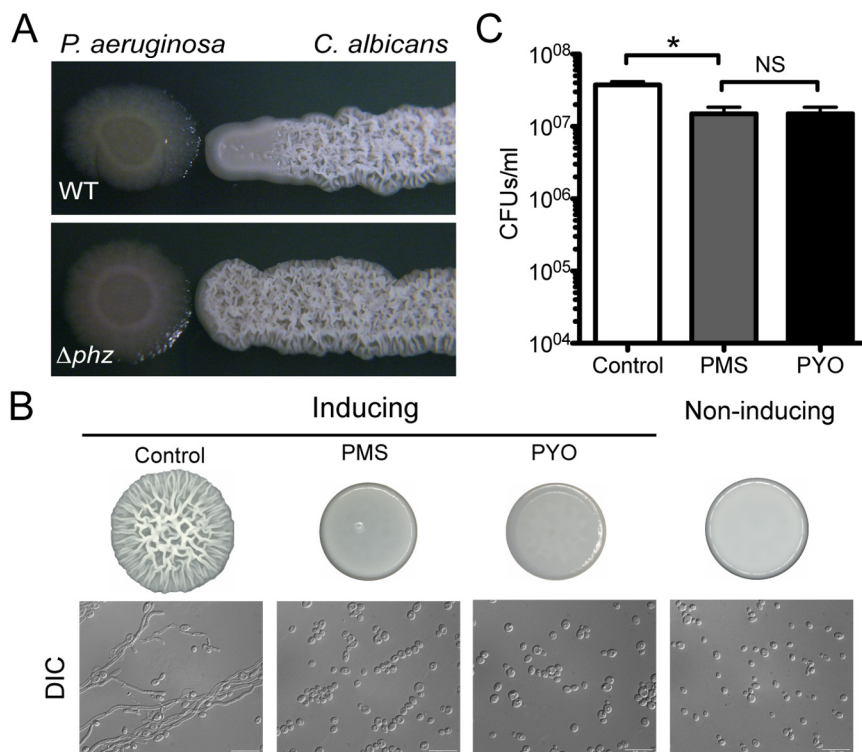


FIG 1 Effects of phenazines on *C. albicans* colony development and cellular morphology. (A) *C. albicans* SC5314 colony morphology when near *P. aeruginosa* WT or the Δphz strain. (B) Top, *C. albicans* colonies grown on Glu-AA-GlcNAc at 37°C (inducing conditions) or 30°C in the absence of GlcNAc (noninducing conditions) for 48 h in the absence or presence of 5 μ M PMS or 20 μ M PYO; bottom, microscopic view of cells from colonies, captured using differential interference contrast (DIC) microscopy. (C) Viable cell counts from *C. albicans* colonies exposed to phenazines. CFUs from colonies grown on Glu-AA at 30°C without or with 5 μ M PMS or 20 μ M PYO for 48 h (*, $P < 0.05$; $n = 4$).

Inhibition of extracellular alkalization by phenazines prevents *C. albicans* filamentation and wrinkling. Previous studies demonstrated links between *C. albicans* morphological transitions and extracellular pH, with yeast growth dominating under acidic conditions and filamentation promoted at neutral pH (40). Seminal studies by Vylkova and colleagues (41) showed that *C. albicans* amino acid catabolism can raise the extracellular pH in environments that are initially acidic. Since our culture medium was rich in amino acids and had an initial pH of 5.0, we determined the role of alkalization in the induction of filamentation and wrinkled colony morphology. As expected, alkalization of the medium was evident only in the presence of amino acids, while medium containing only glucose as a carbon source remained acidic (Fig. 2A). The addition of GlcNAc and incubation at 37°C (conditions that induce filamentation) promoted colony wrinkling on medium with amino acids that had increased pH (Fig. 2A; control). However, GlcNAc and 37°C were not sufficient to support wrinkled colony formation in the absence of amino acids (Fig. 2A). Concentrations of phenazines that inhibited colony wrinkling also had striking effects on the pH changes in the medium. While the medium surrounding control colonies became red by 48 h, indicating a pH greater near 7, the medium surrounding PMS- and PYO-grown colonies remained acidic (Fig. 2A). The addition of PMS or PYO did not change the pH of uninoculated media. These differences in extracellular pH led us to hypothesize that phenazines were impacting *C. albicans* metabolism.

Phenazine-induced fermentation correlates with *C. albicans* acidification of the extracellular milieu. In liquid culture condi-

tions, as observed on agar (Fig. 1C), PMS caused a small decrease in growth rate and final yield (Fig. 2B), and the pH in the extracellular medium of cultures grown with PMS was lower (Fig. 2C). Extracellular pH can increase upon ammonia release (41, 42), and acidic products can lower the pH. We first evaluated the effects of phenazines on ammonia release using an acid trap methodology (41). In control colonies, 138 ± 19 ppm NH_3 was detected in comparison to 290 ± 56 ppm NH_3 for colonies grown with PYO and 270 ± 69 ppm NH_3 for colonies grown with PMS. Thus, neither PMS nor PYO caused a decrease in ammonia release from colonies, suggesting that differences in amino acid catabolism were likely not the cause of the lower pH values in cultures with PMS or PYO.

To determine whether phenazines altered the production of acidic *C. albicans* products, we grew the fungus on filters floating on liquid medium with or without phenazines at 37°C (Fig. 3A) and analyzed the culture supernatants. As on agar, the pH of the supernatants decreased in the presence of phenazines, as visualized by bromocresol green addition (Fig. 3A, top). High-performance liquid chromatography (HPLC) analyses of the supernatants found 8- and 17-fold-higher levels of acetic acid produced per unit of optical density at 600 nm in the presence of PMS and PYO, respectively, at 24 h (Fig. 3B) despite the fact that slightly less glucose was consumed compared to control cultures (see Table S2). Other fermentation products, such as ethanol, were also found at higher relative levels upon phenazine exposure. There was 5.5- and 3.4-fold more ethanol (mg/ml/OD₆₀₀) in cultures grown with PMS and PYO, respectively, than in 24-h super-

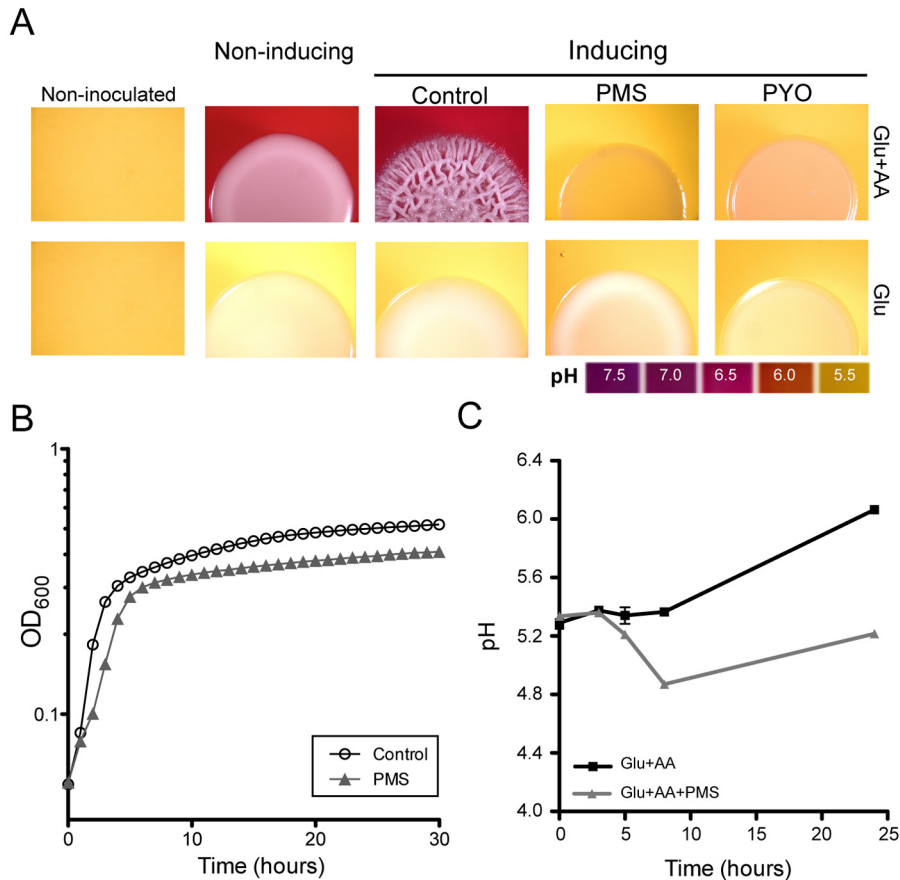


FIG 2 Phenazines modulate *C. albicans* extracellular alkalization and morphogenesis. (A) Fungal colonies grown on Glu+AA or Glu alone under filament-inducing and noninducing conditions without or with 5 μ M of PMS and 20 μ M PYO for 48 h; pH was monitored using bromocresol purple. The images were obtained by using a stereoscope (7.5 \times). (B and C) Growth (B) and pH (C) in liquid cultures grown with Glu-AA in the presence and absence of 5 μ M PMS at 37°C.

natants from untreated cultures of *C. albicans* colonies (Fig. 3B). HPLC analyses of *C. albicans* supernatants in planktonic cultures at 8 h also showed higher levels of ethanol as well as glycerol, another *C. albicans* fermentation product, in PMS-containing cultures than in controls (data not shown). Together, these data suggest a shift toward fermentation in the presence of phenazines.

Phenazines inhibit *C. albicans* growth on nonfermentable carbon sources. *C. albicans* can either ferment or respire glucose, whereas other substrates, like amino acids, alcohols, or organic acids, can be used only for energy generation via respiration (43). In light of the finding that phenazines increased production of *C. albicans* fermentation products (Fig. 3B), we tested the hypothesis that phenazines inhibited respiration by assessing their effects on growth on nonfermentable carbon sources. Indeed, while growth on glucose was only slightly slower in the presence of PMS (Fig. 4A), this phenazine caused a marked decrease in both growth rates and yield in cultures grown on amino acids (Fig. 4B) or succinate (see Fig. S5).

Phenazine-mediated *C. albicans* colony morphology changes are not due solely to altered extracellular pH. Phenazines repressed biofilm behaviors both on agar and in liquid medium with glucose (Fig. 1; see also Fig. S4B in the supplemental material) and caused acidification of the medium (Fig. 2A and C). Because of the known role of extracellular pH in *C. albicans* mor-

phological regulation (40, 41), we tested whether acidification of the extracellular milieu was required for phenazine-mediated inhibition of *C. albicans* filamentous growth and colony development. The fungus was spot inoculated on medium containing glucose and amino acids that was buffered at either pH 5 or pH 7 (Fig. 5A). As previously described (44), a stable acidic extracellular milieu (pH 5.0) completely inhibited *C. albicans* filamentation and wrinkled colony formation (Fig. 5A). Colonies formed on medium that was buffered at pH 7.0 were wrinkled in the absence of PMS or PYO. Interestingly, PYO and PMS still repressed filamentation and colony wrinkling on this buffered medium (Fig. 5A). Therefore, our results demonstrate that while phenazine-mediated decreases in pH were sufficient to repress hyphal growth, this change in extracellular pH was not required to prevent inhibition of wrinkled colony morphology formation and filamentation.

Phenazines alter *C. albicans* respiratory activity. Our data led us to propose that phenazines altered *C. albicans* respiratory activity. To assess whether perturbation of respiratory activity was sufficient to repress colony biofilm development, we determined if methylene blue (MB), another well-known electron acceptor capable of altering mitochondrial activity and with structural similarity to phenazines (34, 35), also repressed colony wrinkling. Similar to the effects of PMS, PCA, and PYO (Fig. 1B; see also

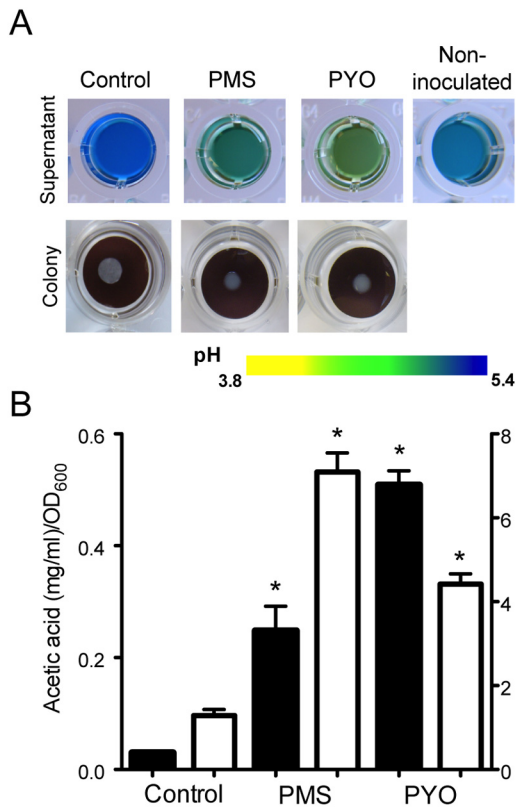


FIG 3 Phenazines promote glucose fermentation. (A) Top, extracellular changes in pH were visualized by addition of bromocresol green; bottom, *C. albicans* colonies grown for 24 h at 37°C on polycarbonate filters suspended on Glu-AA liquid medium in the presence or absence of 20 μM PYO and 5 μM PMS without bromocresol green. (B) Quantification of acetic acid (black bars) and ethanol (white bars) in supernatants of colonies grown on filters ($n = 3$; *, $P < 0.05$).

Fig. S3A in the supplemental material), micromolar concentrations of MB also caused *C. albicans* formation of smooth colonies on unbuffered medium (Fig. 5B), and the effects were dose dependent, with completely smooth colonies observed at 5 μM (see

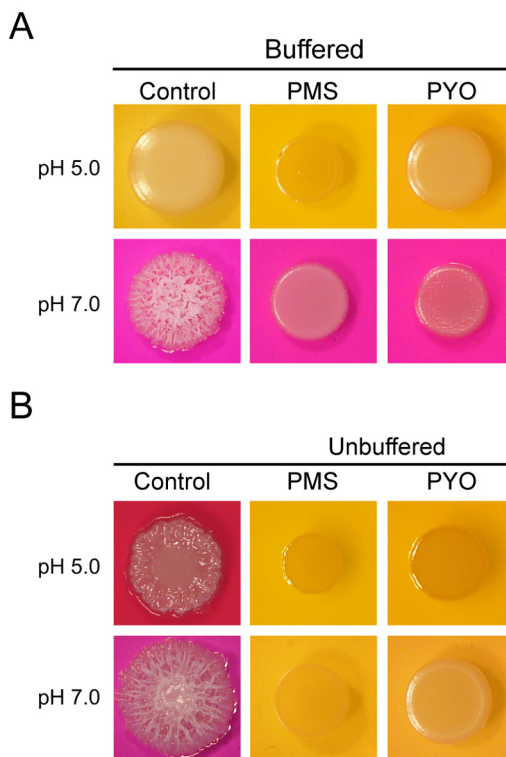
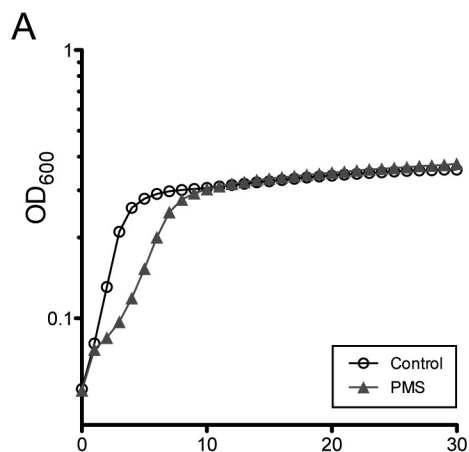


FIG 5 Phenazine-modulated inhibition of *C. albicans* wrinkled colony morphology is not solely due to decreased extracellular pH. (A and B) *C. albicans* colonies grown on Glu-AA-GlcNAc with the pH adjusted to 7.0 or 5.0 prior to inoculation. A total of 5 μM of PMS, PYO, or methylene blue (MB) was added to the medium when specified, and the pH indicator bromocresol purple, which yields a red-purple color under alkaline conditions and a yellow-orange color under acidic conditions, was included. In panel A, the medium was buffered with 40 mM MOPS (pH 7) or 100 mM citrate (pH 5) as indicated. Colonies were incubated for 48 h at 37°C.

Fig. S6). Unlike phenazines, which promoted extracellular acidification (Fig. 2A and C and 3A), cultures with MB alkalinized the medium when it was at an initial pH of 5 and did not acidify the extracellular milieu when the initial pH was adjusted to 7

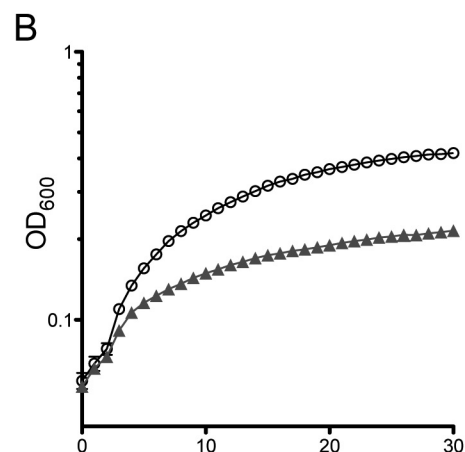


FIG 4 Phenazines alter *C. albicans* growth in fermentable and nonfermentable carbon sources. Growth curves of liquid cultures on media containing only glucose (A) or amino acids (B) in the presence (triangles) or absence (open circles) of 5 μM PMS at 37°C. The optical density (OD₆₀₀) of each culture was monitored every hour.

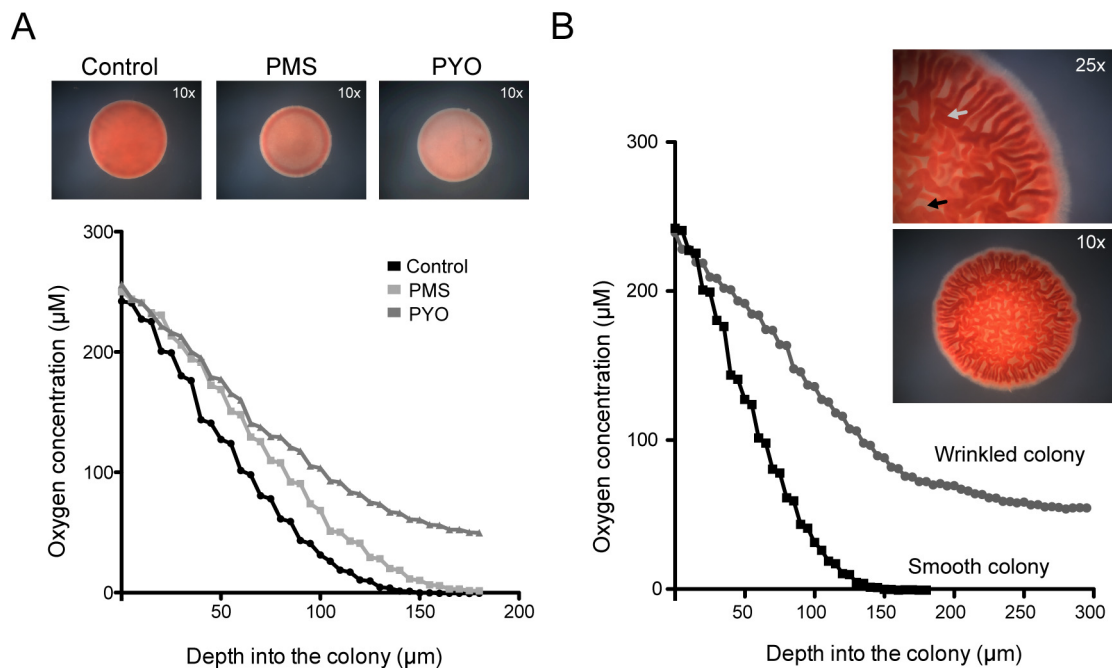


FIG 6 Effects of phenazines on respiratory metabolism and oxygen consumption. (A) *C. albicans* colonies grown in Glu-AA medium alone or with 5 μM PMS or 20 μM PYO under noninducing conditions. The respiratory activity of colonies was assessed using TTC, and images were captured using a stereoscope at 10 \times (top). Internal oxygen measurements in similarly grown colonies were made from the top to the bottom of control colonies (black), with PMS (light gray) or PYO (dark gray). (B) Oxygen gradients in colonies grown under noninducing (smooth) and inducing conditions (wrinkled) were measured at room temperature. The respiratory activity in wrinkled colonies was greater in wrinkles (gray arrow) than in valleys (black arrow) (inset).

(Fig. 5B). These data highlight that compounds that impact respiration can inhibit colony wrinkling and that extracellular pH changes were not required for the repression of hyphal growth and the development of wrinkled, structured colonies.

We next used TTC, a dye indicative of mitochondrial activity, in order to test the idea that phenazines could impair respiration directly. Reduction of TTC by the electron transport chain (ETC) leads to the formation of a red product that accumulates within cells (45). Colony biofilms formed without phenazines exhibited a high level of TTC reduction as evidenced by the strong red pigmentation (Fig. 6A and B). However, PMS- or PYO-grown colonies showed decreased TTC conversion in this assay and, thus, were much less metabolically active (Fig. 6A). As observed with phenazine-exposed *C. albicans*, colonies grown on agar with MB also demonstrated a reduced red pigmentation (see Fig. S6B).

To complement these studies, we used an electrode probe to measure the oxygen gradients as a function of depth from the top of the colony as an indicator of oxygen utilization within the colony interior. Under noninducing conditions, smooth colonies grown with PMS or PYO demonstrated higher concentrations of oxygen than control colonies when measured at comparable depths from the surface (Fig. 6A). These data are consistent with oxygen utilization being repressed by phenazines. Together, these findings indicate that phenazines cause a disruption of the normal respiratory patterns in *C. albicans* cells.

The wrinkles of *C. albicans* colonies have elevated oxygen concentrations and exhibit a high respiratory metabolism. When TTC was added to *C. albicans* wrinkled colonies formed in inducing conditions in the absence of phenazines, stronger red pigmentation was observed than in colonies grown under nonin-

ducing conditions (Fig. 6A and B), suggesting that wrinkled colony biofilms exhibited a more active respiratory metabolism than colonies with a smooth morphology. Consistent with this observation, phenazine-treated colonies grown under inducing conditions also showed decreased TTC-derived coloration than untreated wrinkled controls (data not shown). Strikingly, while there was even, red pigmentation from TTC conversion in smooth colonies (Fig. 6A), wrinkled colonies had higher levels of pigmentation in the wrinkled areas (higher regions of the colony). The “valleys” appeared to have less respiratory activity (Fig. 6B, inset). Based on these observations and in light of our previous results demonstrating that *C. albicans* wrinkled colony formation was repressed by phenazines and MB (Fig. 1B and 5B), we proposed that wrinkling was linked to respiration.

To determine whether the topology of the wrinkles promoted access to oxygen within dense colonies, we used an electrode to quantify the oxygen concentration within the wrinkles of colonies grown under inducing conditions compared to noninducing conditions at similar distances from the colony edge. Wrinkled colony areas had shallow oxygen gradients, and moderately high oxygen remained detectable throughout the structure (Fig. 6B). In sharp contrast, the gradients for smooth yeast colonies formed in noninducing conditions were much steeper and showed significantly lower oxygen concentrations at all levels within the colony. Thus, our results suggest a model in which wrinkling promotes aerobic respiration as part of fungal colony biofilm development.

DISCUSSION

Our work reveals a new role for *P. aeruginosa* phenazines as modulators of *C. albicans* metabolism and biofilm development. We

found that low, nonlethal concentrations of bacterial phenazines inhibited fungal morphogenesis and prevented the development of wrinkled colony biofilms, and subsequent analyses indicated that these effects were due to alteration of *C. albicans* respiratory activity. Several lines of evidence indicated that phenazines inhibited normal fungal respiratory activity: (i) fermentation products were at higher concentrations in the presence of phenazines (Fig. 3B), (ii) growth on nonfermentable carbon sources was severely impaired with only slight growth inhibition on glucose (Fig. 4B; see also Fig. S5 in the supplemental material), (iii) metabolic activity was lower in the presence of phenazines (Fig. 6A), and (iv) oxygen concentrations were higher in yeast colonies grown with phenazines than in yeast colonies grown in their absence (Fig. 6A). The increased secretion of acetic acid (Fig. 3B) (46) in the presence of phenazines likely reduced the extracellular pH (Fig. 2A and C); low pH was not the only mechanism responsible for the phenazine-mediated effects since buffering the medium at pH 7 did not ameliorate the effects of phenazines (Fig. 5A). Importantly, MB, another compound that alters the cellular respiratory activity (34, 35), also repressed hyphal growth and colony biofilm formation but did not lead to a pH change, suggesting that a different spectrum of fermentation products may have been produced (Fig. 5B). The different effects of MB and phenazines with respect to altering the extracellular pH may be explained by their respective redox properties (47), which could lead to different interactions with the ETC, or due to differences in other targets within the cell. Our data suggest that decreased ability to transfer electrons to oxygen led to decreased colony wrinkling and filamentation, and this model is supported by the fact that *C. albicans* wrinkled colony formation was inhibited (see Fig. S7) and biofilm formation on plastic in liquid medium does not occur in anoxic environments despite the fact that cells form hyphae (48, 49) (see Fig. S7).

Our observations that colony wrinkling was repressed under conditions that limit respiration suggest that these structures may allow cells to increase their access to oxygen in dense populations. Several findings support this model. First, while the absence of oxygen inhibited wrinkled colony formation (see Fig. S7), hypoxic conditions strongly promote colony wrinkling (10) (data not shown). Second, we found that wrinkled regions had higher rates of respiration than nonwrinkled regions of the colony (Fig. 6B) and that wrinkled colonies had more respiratory activity than colonies that lacked wrinkling (Fig. 6). Lastly, mutants locked in a highly respiratory state, such as strains lacking Ace2 and Tye2, have increased colony wrinkling (10) and filamentation within biofilms (50). We speculate that the *C. albicans* wrinkled phenotype is related to biofilm phenotypes on submerged, plastic surfaces, as both structures involve spatially distributed filamentation and cells that adhere tightly to one another. In multiple bacterial species, the study of colony wrinkling has led to the discovery of genes crucial for biofilm formation on solid surfaces (51–53).

Links between biofilm formation and oxygen availability may emerge as a common theme in diverse microbial species, including both fungi and bacteria. For example, wrinkled *P. aeruginosa* colonies have increased oxygen concentrations within the colony compared to smooth counterparts, and it has been hypothesized that biofilm structure enables access to oxygen by increasing surface area (54). In *P. aeruginosa*, endogenously produced phenazines serve as alternate electron acceptors, thereby facilitating *P. aeruginosa* survival in low-oxygen conditions (55, 56).

Phenazines, likely in part through modulation of intracellular redox, led to decreased surface-to-volume ratios in both flow cell and colony biofilms (57, 58) in a manner that is reminiscent to what we observed with *C. albicans*. In addition, *C. albicans* biofilm development may also be influenced by ethanol, a metabolite that has been shown to inhibit submerged biofilm formation in this fungus (59).

Previous studies have also shown links between *C. albicans* respiratory metabolism and cellular and colony morphology. For instance, *C. albicans* strains that lack the mitochondrial NADH dehydrogenase were unable to filament (13), and other ETC inhibitors such as rotenone (13) and thenoyl trifluoro acetone (16) repressed *C. albicans* filamentation. However, it is important to note that antimycin A, another respiratory chain inhibitor, promoted filamentation (10), suggesting that the inhibition of filamentation in response to altered ETC activity is likely due to more than decreased energy generation, such as a response to a specific metabolic signal (10). Biochemical studies that have examined the interactions between phenazines and respiratory electron transport systems in eukaryotic and bacterial cells have found that these compounds and related molecules can either inhibit or stimulate the flux of electrons through the ETC (31, 60, 61). Future work will determine what signal, such as ratios of intracellular electron carriers, the state of the ETC components, intracellular pH, or ATP levels, is sensed by the cell and which regulators respond to these important intracellular signals. Furthermore, additional studies will be required to explain the differences in respiration seen in the “folds” of wrinkled colonies compared to smooth regions. Possible explanations include regulated differences in metabolism or differences in cell density due to morphology or matrix production. While phenazines are likely too toxic for use as therapeutic agents, our findings with MB may have implications for antifungal therapy. MB is nontoxic in mammals and has been used as an antimalarial drug, a neurotoxin antidote, and a potentiating factor in photodynamic therapy (34, 35). Thus, there is potential for this or similar compounds to be used to inhibit *C. albicans* biofilm infections *in vivo*.

This work highlights that phenazines have different biological effects at different concentrations. Kerr and colleagues (27, 36) and our laboratory (24, 29) have demonstrated the anti-*Candida* properties of *P. aeruginosa* phenazines at high concentrations (>500 μ M). Here, we show that respiration is inhibited at concentrations that are 25- to 200-fold lower than those required to stop growth (29, 36) (Fig. 1B). In the lungs of cystic fibrosis (CF) patients, where *P. aeruginosa* and *C. albicans* are often found together (21, 62), PYO and PCA were found to be between 5 and 80 μ M (33). We show here that these concentrations repressed *C. albicans* filamentation and biofilm development but did not kill the fungus. A 1989 study by Bhargava et al. (63) that assessed fungal morphology in the CF lung suggests that this fungus exists as yeast in CF airways. Our findings also suggest the possibility that by releasing phenazines, *P. aeruginosa* causes *C. albicans* to secrete more fermentation products that are readily used by *P. aeruginosa* to enhance its own growth and survival.

MATERIALS AND METHODS

Strains, media, and culture conditions. See Table S1 for strain descriptions. *C. albicans* and *P. aeruginosa* were inoculated from overnight cultures onto yeast nitrogen base synthetic medium (YNB salts with ammonium sulfate that was supplemented with 10 mM glucose [Glu], 0.2%

[wt/vol] amino acids from yeast synthetic dropout medium supplement without tryptophan [AA], both Glu and AA, or succinate [10 mM]). Where stated, 40 mM MOPS (morpholinepropanesulfonic acid; pH 7) or 100 mM citrate buffer (pH 5) was added. Noninducing conditions that promoted growth as yeast and the formation of smooth colonies were defined as Glu or Glu-AA and incubation at 30°C. In inducing conditions that cause colony wrinkling and filamentation, 5 mM GlcNAc was added and cultures were incubated at 37°C. When indicated, media were supplemented with phenazines or methylene blue as described in the supplemental information. To monitor pH, bromocresol purple (0.01% [vol/vol]) was added to the agar medium. *C. albicans*-*P. aeruginosa* cocultures were performed on yeast extract-peptone-dextrose (YPD) agar.

Analysis of extracellular pH and ammonia production. See Methods in Text S1 in the supplemental material.

Respiratory activity analyses and oxygen tension analyses. To analyze the respiratory activity of colonies, we employed a TTC overlay technique (64) in which 2 ml of the TTC-agar solution was applied to 48-h-old spot-inoculated colonies (see Methods in Text S1 in the supplemental material). Oxygen tensions within the colonies were measured at room temperature using an automated micro Clark-type oxygen sensor (Unisense) in three areas per colony from the top to bottom after growth for 72 h.

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <http://mbio.asm.org/lookup/suppl/doi:10.1128/mBio.00526-12/-/DCSupplemental>.

Text S1, DOCX file, 0.1 MB.
Figure S1, TIF file, 0.3 MB.
Figure S2, TIF file, 0.8 MB.
Figure S3, TIF file, 1.2 MB.
Figure S4, TIF file, 0.9 MB.
Figure S5, TIF file, 0.1 MB.
Figure S6, TIF file, 0.5 MB.
Figure S7, TIF file, 0.2 MB.
Table S1, DOCX file, 0.1 MB.
Table S2, DOCX file, 0.1 MB.

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