

MRI-based biodistribution assessment of holmium-166 poly (L-lactic acid) microspheres after radioembolisation

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Abstract

Objectives To demonstrate the feasibility of MRI-based assessment of the intrahepatic Ho-PLLA-MS biodistribution after radioembolisation in order to estimate the absorbed radiation dose.

Methods Fifteen patients were treated with holmium-166 (^{166}Ho) poly(L-lactic acid)-loaded microspheres (Ho-PLLA-MS, mean 484 mg; range 408–593 mg) in a phase I study. Multi-echo gradient-echo MR images were acquired from which R_2^* maps were constructed. The amount of Ho-PLLA-MS in the liver was determined by using the relaxivity r_2^* of the Ho-PLLA-MS and compared with the administered amount. Quantitative single photon emission computed tomography (SPECT) was used for comparison with MRI regarding the whole liver absorbed radiation dose.

Results R_2^* maps visualised the deposition of Ho-PLLA-MS with great detail. The mean total amount of Ho-PLLA-MS detected in the liver based on MRI was 431 mg (range 236–666 mg) or 89 ± 19 % of the delivered amount (correlation coefficient $r=0.7$; $P<0.01$). A good correlation was

found between the whole liver mean absorbed radiation dose as assessed by MRI and SPECT (correlation coefficient $r=0.927$; $P<0.001$).

Conclusion MRI-based dosimetry for holmium-166 radioembolisation is feasible. Biodistribution is visualised with great detail and quantitative measurements are possible.

Key Points

- Radioembolisation is increasingly used for treating unresectable primary or metastatic liver tumours.
- MRI-based intrahepatic microsphere biodistribution assessment is feasible after holmium-166 radioembolisation.
- MRI enables quantification of holmium-166 microspheres in liver in a short imaging time.
- MRI can estimate the whole liver absorbed radiation dose following holmium-166 radioembolisation.

Keywords Radioembolisation · Holmium · MRI · Dosimetry · Biodistribution

Introduction

Yttrium-90 (^{90}Y) radioembolisation (RE) is increasingly used for the treatment of patients with unresectable primary or metastatic liver tumours [1–6]. The two commercially available products, SIR-Spheres[®] (Sirtex Medical Limited, North Sydney, NSW, Australia) and TheraSphere[®] (Nordion, Ottawa, Canada), use empirical methods for dose calculation, based on body surface area and total liver weight respectively. However, apart from the total dose delivered to the liver, the efficacy of RE will largely depend on the intrahepatic dose distribution or, more specifically, on the ratio between the tumour absorbed dose and healthy tissue absorbed dose.

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Quantitative assessment of the post-administration intrahepatic microsphere distribution is therefore indispensable for evaluation of RE toxicity and efficacy and can potentially be used for the prediction of patient response and patient-specific therapeutic dose optimisation. Image-based approaches to assess microsphere distribution after RE have gained interest but are mostly hampered by the limited imaging possibilities of the isotope ^{90}Y . Quantitative ^{90}Y -SPECT imaging has limited spatial resolution because it is based on ^{90}Y Bremsstrahlung [7–10], whereas ^{90}Y -PET has better spatial resolution but low sensitivity [11, 12]. As a consequence, alternative methods of visualising the microspheres have been investigated, such as MR imaging of iron-labelled microspheres [13].

In the past decade, holmium-166 (^{166}Ho) poly(L-lactic acid)-loaded microspheres (Ho-PLLA-MS) have been developed as a potential radioembolisation particle [14, 15]. Neutron activated holmium-166 is a β -emitting ($E_{\text{max}}=1.77$ and 1.854 MeV; $I_{\beta}=48.7\%$ and 50 %, respectively; $T_{1/2}=26.83$ h) and γ -emitting ($E_{\gamma}=80.6$ keV) lanthanide that, embedded in microspheres of poly(L-lactic acid), yields a particle suitable for internal radiation therapy that can be visualised with a range of clinical imaging techniques, including SPECT and MRI. This multimodal imaging has previously been demonstrated in preclinical animal studies [16–18]. It was also shown that MRI combines high sensitivity with high spatial-temporal resolution and with superior soft tissue contrast [19] and thus can be used to cover a broad range of clinically interesting imaging parameters. More recently, it was shown for an ex vivo situation that MRI is able to provide a good measurement of the Ho-PLLA-MS radiation-absorbed dose by convolution of quantitative MRI data with a ^{166}Ho dose point kernel [20].

The feasibility of image-guided RE using Ho-PLLA-MS was tested in a first-time-in-man clinical phase I study. The aim of the work described in this article was to investigate the capability of MRI to measure the intrahepatic microsphere distribution in order to quantify the absorbed radiation dose in patients treated with Ho-PLLA-MS radioembolisation.

Materials and methods

Patients

To be eligible to enter the phase I study each patient needed to comply with the following inclusion criteria: written informed consent; age 18 years or older; presence of liver-dominant, unresectable, chemorefractory liver metastases of any primary tumour with at least one measurable lesion of ≥ 10 mm in the longest diameter on CT; an estimated life expectancy >3 months; World Health Organisation (WHO) performance status 0–2; and a negative pregnancy test for women. Furthermore, patients with contraindications for

MRI were excluded from treatment. Ethics approval for this study was obtained from the institutional review board; the study was performed in accordance with the Declaration of Helsinki and was registered with Clinicaltrials.gov, no. NCT01031784.

Treatment

Holmium-165 poly(L-lactic acid) microspheres with a mean diameter of 30 μm (range 20–50 μm) and a holmium content of 18.7 % (weight/weight) were prepared under Good Laboratory Practice (GLP) guidelines as previously described [14]. The amount of microspheres (600 mg) was weighed, packed in high-density polyethylene vials (Posthumus Plastics, Beverwijk, The Netherlands) and sent to the nuclear reactor (Delft University of Technology, Delft, The Netherlands) for neutron activation [21]. The calculation of the amount of radioactivity was based on liver weight, conforming to the clinically used approach for TheraSphere[®]. The amount of administered ^{166}Ho -radioactivity was calculated using the target liver absorbed dose in a dose-escalation fashion (20, 40, 60, or 80 Gy) as previously described [22]. All patients received whole liver treatment. The activity to be administered was calculated assuming homogeneous distribution of the activity in the liver using the following formula: $A_{\text{Ho166}} (\text{MBq}) = \text{Liver Dose} \times 63 (\text{MBq/J}) \times \text{LW}$ where A_{Ho166} is the administered activity, LW is the liver weight in kg, and Liver Dose is the intended liver absorbed radiation dose in Gy. While the radioactivity was different for each patient, the amount of administered Ho-PLLA-MS was kept constant at 600 mg. Before administration, the amount of radioactivity in the vials was measured using a dose calibrator (VDC-404, Veenstra Instrumenten B.V., Joure, The Netherlands), and the specific activity of the Ho-PLLA-MS was determined. After administration, activity measurements were performed on the administration system to determine the amount of microspheres that was actually delivered to the subjects retrospectively.

Radioembolisation procedures

Radioembolisation was performed according to standard recommendations [23, 24]. Relevant vessels in particular branches of the hepatic artery supplying organs other than the liver, e.g., the gastroduodenal artery (GDA) and right gastric artery (RGA), were coiled during a pre-treatment angiographic procedure to prevent extrahepatic deposition of activity. A scout dose of $^{99\text{m}}\text{Tc}$ -MAA (150 MBq, 0.8 mg, TechneScan LyoMaa[®], Mallinckrodt Medical B.V., Petten, The Netherlands) was injected into the hepatic artery followed by planar gamma camera imaging and SPECT to check for inadvertent extrahepatic deposition. The $^{99\text{m}}\text{Tc}$ -MAA lung shunt was measured by planar scintigraphic imaging, and

shunt fractions were determined by region of interest (ROI) analyses [25]. If the lung shunt fraction of ^{99m}Tc -MAA was <20 % and no other extrahepatic deposition of ^{99m}Tc -MAA was detected, treatment with Ho-PLLA-MS was performed within 2 weeks.

Data acquisition for dosimetry

For intrahepatic biodistribution assessment of the microspheres, abdominal SPECT and MRI were performed.

MRI

Magnetic resonance images were acquired 1–2 weeks before and 1 week after administration of the therapeutic dose of Ho-PLLA-MS. Since MRI utilises the paramagnetic nature of the microspheres rather than the radioactivity, the post-therapy imaging time point was of minor relevance, and post-therapy images were acquired during the patients' first weekly visit at the outpatient clinic after treatment. Imaging was performed using a 1.5-T whole body system (Achieva, Philips Healthcare, Best, The Netherlands) equipped with a 16-element torso coil. For detection and quantification of Ho-PLLA-MS, a multi-slice multi-gradient echo (MGE) sequence was used, acquiring 16 echoes during breath hold with an in-plane resolution of $2 \times 2 \text{ mm}^2$ and a slice thickness of 6 mm. Imaging parameters included: FOV: $288 \times 384 \text{ mm}^2$, number of slices: 45, TR/TE1/ Δ TE: 440 ms/1.33 ms/1.15 ms, flip angle: 50° . Sensitivity encoding (SENSE) with a factor of 2.5 was used for acceleration resulting in an imaging time of $3 \times 19 \text{ s}$ during breath hold. For anatomical information and segmentation purposes, T_2 -weighted turbo spin echo (TSE) images were acquired with identical FOV and voxel size. Imaging parameters included: TR/TE: 830 ms/80 ms; SENSE factor: 2; imaging time: $2 \times 19 \text{ s}$ during breath hold. In addition, T_1 -weighted turbo field echo (TFE) images were acquired using an in-plane resolution of $1.8 \times 3.6 \text{ mm}^2$; slice thickness: 10 mm; TR/TE: 8.5 ms/4.18 ms; total imaging time: $2 \times 14 \text{ s}$ during breath hold.

To determine the sensitivity of MRI for Ho-PLLA-MS, as expressed by the r_2^* relaxivity (relaxation rate per mg/ml Ho-PLLA-MS), a calibration phantom setup containing known microsphere concentrations (0, 2.1, 4.1, 6.1 and 8 mg/ml Ho-PLLA-MS in agarose gel) was imaged using the MGE sequence described above.

SPECT

Single photon emission computed tomography images were acquired 3–6 days after administration of Ho-PLLA-MS. The imaging time point did depend on the total activity that was administered to the patient and was based on the maximum tolerated count rate of the SPECT system to acquire

images without dead-time effects. The first nine patients underwent imaging using a FORTE™ dual-headed gamma camera (FORTE™, Philips Medical Systems, Milpitas, CA, USA) equipped with gadolinium-153 scanning line sources for transmission CT. The last six patients underwent imaging using a Siemens Symbia T16 SPECT/CT system (Siemens, Erlangen, Germany), which combines a dual-headed gamma camera with a 16-slice CT system. Medium energy collimators were used on both systems. Energy windows were set at 80.6 keV (15 % window width) for the holmium-166 photo peak and at 118 keV (12 % window width) for the correction of downscatter. One hundred twenty projections of 30 s were acquired in a 180° (FORTE) or 360° (Symbia T16) orbit around the liver. Quantitative image data were reconstructed to a 128^3 matrix size with an isotropic voxel size of 4.7 mm^3 (FORTE) or 4.8 mm^3 (Symbia T16), using an ordered subsets expectation maximisation algorithm including a hybrid scatter correction method [26], resulting in an absolute quantitative 3D activity distribution in MBq/voxel.

Data analyses

Magnetic resonance images were processed using software code written in MATLAB (MathWorks, Natick, MA, USA). From the MGE MRI patient data, R_2^* values were estimated voxelwise using a mono-exponential fitting algorithm weighting all signal amplitudes equally. To minimise the influence of noise, signal intensities lower than 3σ were excluded from the fitting procedure, with σ the standard deviation (SD) of the signal determined in a region with homogeneous signal intensity and free from Ho-PLLA-MS. R_2^* values were determined for data obtained both before and after RE. For further analysis, the liver was manually segmented based on the tissue contrast of the corresponding T_2 -weighted TSE images. The distribution of estimated liver R_2^* values was measured, and the mean R_2^* value together with the SD was calculated.

To determine the change in R_2^* (ΔR_2^*) after therapy, a baseline R_2^* value determined by the mean R_2^* of the liver before Ho-PLLA-MS administration was subtracted from the post-therapy R_2^* maps. Voxelwise concentrations of Ho-PLLA-MS were determined from the ΔR_2^* maps by the relationship $[\text{Ho-PLLA-MS}] = \Delta R_2^* / r_2^*$, with r_2^* measured from the calibration phantom setup. Using the voxel volume and the total volume included in the segmentation, the total amount of Ho-PLLA-MS in the liver was determined.

For all patients, the MRI-based amount was compared with the total amount that was assumed to be delivered to the liver. This amount was determined by correcting the delivered amount of Ho-PLLA-MS by the ^{99m}Tc -MAA lung shunt fraction and the amount of Ho-PLLA-MS left in the

administration system. Data from patients who had surgical clips implanted (as a consequence of former partial liver resection) were separated from the patients without clips, and means were also compared separately for these two groups. This separation was made because metallic surgical clips are known to distort gradient echo images [27–29] and hence were suspected to influence MRI-based quantification.

As a final step, the mean radiation absorbed dose was calculated on a whole liver level using MRI and SPECT dose maps that were constructed using the method previously described for an ex-vivo liver model [20]. In short, MRI concentration values were converted into MBq/voxel using the voxel size and the specific activity of the microspheres (MBq/mg) at the moment of injection. The SPECT reconstructions provided an absolute quantitative activity in MBq/voxel. Both MRI and SPECT activity maps were convolved with a ^{166}Ho dose point kernel that was calculated using the Monte Carlo code MCNPX (vs. 2.5.0; LANL, Los Alamos, NM, USA) according to the method described in Medical Internal Radiation Dose (MIRD) Pamphlet 17 [30], assuming a tissue density for liver tissue of 1.06 g/cm^3 according to International Commission on Radiation Units and Measurements (ICRU) Report 44 [31]. The voxel size of the dose point kernel was equal to the native resolution of the image technique ($2\times 2\times 6\text{ mm}$ for MRI, 4.7 mm isotropic for SPECT). After manual co-registration of SPECT maps to MRI and segmentation, mean liver absorbed dose values were calculated for both techniques.

Statistical analysis

Descriptive statistics of means, standard deviations and ranges were calculated for continuous variables. The paired Student's *t*-test was used for comparison of means (two-tailed 95 % confidence interval). Bivariate Pearson's correlation (one-tailed 95 % confidence interval) was used for correlation between Ho-PLLA-MS calculations based on MRI and the actual amount for the whole liver and the correlation between MRI and SPECT dosimetry on a whole liver level. SPSS software (SPSS for Windows, version 13.0; SPSS Inc., Chicago, IL, USA) was used for all analyses.

Results

Fifteen patients were included in this phase I dose-escalation study. Nine men and six women, median age 55 years (range 38–87 years), were treated with escalating whole liver doses of 20–80 Gy. The median number of liver tumours was 5 (range 1–21), with a median fractional liver involvement of 14 % (range 2–52 %), originating from

an ocular melanoma (7 patients), colorectal carcinoma (5 patients), cholangiocarcinoma (2 patients) or breast carcinoma (1 patient). Five patients had surgical clips implanted in the liver as a consequence of former partial liver resection (number of clips ranging from 1 to >20). MRI data were analysed for 14 patients. One patient was excluded from MRI analysis because of incomplete data. The patients received a mean of 523 mg Ho-PLLA-MS (range 438–640 mg). Corrected for shunting to the lungs (mean 7.4 %; range 3.0–13.0 %), the mean administered amount of Ho-PLLA-MS to the liver was 484 mg (range 408–593 mg; Table 1).

Ho-PLLA-MS caused enhanced MR signal decay after administration, translating into high R_2^* values (Fig. 1). Strongly increased R_2^* values were observed locally, corresponding to tumour lesions with high signal intensity on T_2 -weighted images. Histograms of the R_2^* values found in the liver showed an overall shift of R_2^* values toward higher values and a broadening of the R_2^* distribution after treatment (Fig. 2).

The mean R_2^* values, measured over the entire liver for all patients before treatment (mean 31.5 s^{-1} ; range 27.9 – 37.3 s^{-1}), significantly increased after administration of Ho-PLLA-MS (mean 52.7 s^{-1} ; range 41.6 – 78.2 s^{-1} ; $P<0.001$; Table 1).

From the phantom setup, an r_2^* relaxivity of $103\pm 4\text{ s}^{-1}\cdot\text{mg}^{-1}\cdot\text{ml}$ was measured for the used Ho-PLLA-MS at 1.5 T, which is in good agreement with the values in the literature [19, 20]. This value was subsequently used to calculate Ho-PLLA-MS concentration maps. The concentration maps were comparable to the R_2^* maps with relatively high concentrations found at tumour sites (Fig. 3). From the concentration maps and the volume of the liver, the amount of Ho-PLLA-MS was calculated. The mean total amount of Ho-PLLA-MS that was detected in the liver based on MRI was 431 mg (range 236–666 mg). For all patients this was $89\pm 19\%$ (mean \pm SD) of the amount delivered to the liver (correlation coefficient $r=0.7$; $P<0.01$). By excluding the data of the patients with surgical clips implanted in the liver, the mean MRI-based detected fraction increased to $96\pm 13\%$ (mean \pm SD; correlation coefficient $r=0.8$; $P<0.01$; Table 1). MRI showed that the presence of surgical clips in the liver led to undetermined areas (black spots) surrounded by (virtual) high concentration values (Fig. 4). This led to a poor correlation between Ho-PLLA-MS amounts calculated on MRI and the actual delivered amount of Ho-PLLA-MS (correlation coefficient $r=0.3$; $P=0.29$).

A good correlation was found between the whole liver mean absorbed radiation dose as assessed by MRI and SPECT (correlation coefficient $r=0.93$; $P<0.001$ including all patients, correlation coefficient $r=0.98$; $P<0.001$ excluding patients with surgical clips, correlation coefficient $r=0.81$; $P=0.10$ for only patients with clips).

Table 1 Acquired biodistribution data of all 15 patients included in the phase I dose-escalation study (20, 40 60 and 80 Gy). One patient was excluded from MRI analyses because of incomplete data. Data of patients with surgical clips present in the liver are shaded grey.

Administered amounts (admin Ho-PLLA-MS) were determined based on activity measurements of the administration system and lung shunt fraction. R_2^* , concentration and absorbed dose values are mean values over the entire liver

Detection of Ho-PLLA-MS in the whole liver										
Subject	Dose cohort (Gy)	No. of surgical clips	Admin. Ho-PLLA-MS (mg)	R_2^* pre \pm SD (s^{-1})	R_2^* post \pm SD (s^{-1})	Concentration Ho-PLLA-MS \pm SD (mg/ml)	MRI-based Ho-PLLA-MS in mg (% of admin)	MRI-based absorbed dose (Gy)	SPECT-based absorbed dose (Gy)	
1	20	-	490	excl	excl	excl	excl	excl	9.5	
2	20	-	408	28.4 \pm 12.4	43.3 \pm 21.8	0.14 \pm 0.21	467 (115)	12.4	9.1	
3	20	5-10	449	31.1 \pm 18.6	42.4 \pm 36.2	0.11 \pm 0.35	287 (64)	6.6	12.5	
4	20	-	465	31.8 \pm 16.4	45.4 \pm 21.2	0.13 \pm 0.21	452 (97)	16.4	14.1	
5	20	-	489	37.3 \pm 24.1	61.4 \pm 39.7	0.23 \pm 0.39	415 (85)	13.4	11.0	
6	20	-	461	30.6 \pm 14.8	59.2 \pm 23.6	0.28 \pm 0.23	447 (97)	15.9	11.9	
7	40	10-15	411	33.2 \pm 20.3	52.2 \pm 36.7	0.18 \pm 0.36	291 (71)	19.8	22.2	
8	40	-	472	33.9 \pm 19.7	52.7 \pm 36.2	0.19 \pm 0.35	351 (74)	19.4	20.9	
9	40	10-15	453	30.9 \pm 21.7	50.2 \pm 29.8	0.28 \pm 0.23	495 (109)	29.5	27.7	
10	60	-	501	27.9 \pm 15.7	42.2 \pm 21.4	0.14 \pm 0.21	436 (87)	32.4	36.6	
11	60	-	485	31.4 \pm 19.6	53.8 \pm 32.2	0.23 \pm 0.30	457 (94)	41.3	36.9	
12	60	>20	478	35.8 \pm 34.2	45.3 \pm 30.0	0.09 \pm 0.29	236 (49)	19.6	37.3	
13	80	-	593	29.2 \pm 13.8	69.4 \pm 56.4	0.39 \pm 0.55	597 (101)	59.3	53.1	
14	80	1	522	25.5 \pm 21.0	41.6 \pm 25.7	0.16 \pm 0.25	442 (85)	42.0	41.3	
15	80	-	591	33.7 \pm 16.8	78.2 \pm 54.8	0.43 \pm 0.53	666 (113)	68.8	55.2	

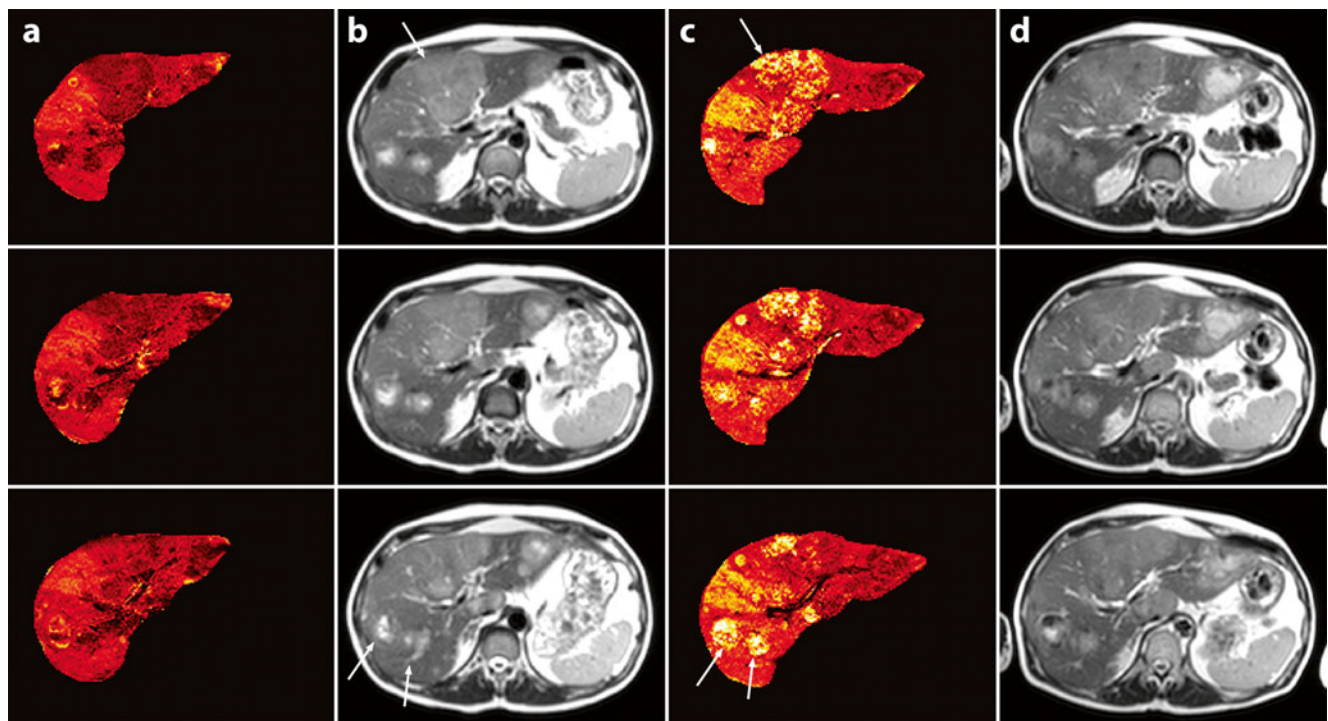


Fig. 1 R_2^* maps and SE images (TR/TE: 830 ms/80 ms, flip angle: 90°) before (a+b) and after (c+d) RE. Variations in R_2^* values before RE (a) correspond to variations on SE images (b). After RE, increased

R_2^* values were observed at locations with Ho-PLLA-MS deposition (c), corresponding to tumour lesions with high intensities on SE images (arrows) (d)

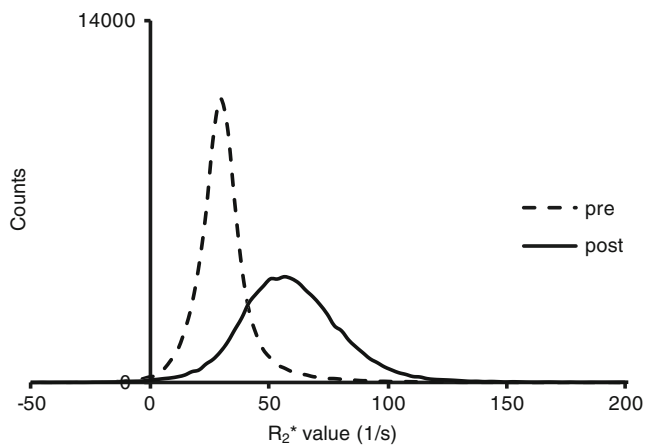


Fig. 2 Typical histogram of R_2^* values measured in the whole liver of a patient before (---) and after (—) administration of Ho-PLLA-MS. After administration the curves shifted toward higher values and a broader distribution of R_2^* values was observed

Magnetic resonance imaging displayed the local microsphere distribution in much more detail because of the relatively high in-plane resolution, for example revealing an absence of Ho-PLLA-MS in the central part of a furthermore highly targeted tumour in some cases. This situation was rather obscured in the corresponding SPECT images owing to the lower resolution and blurring effects (Fig. 5).

Discussion

In this article we presented the first *in vivo* quantitative MRI data of ^{166}Ho -poly(L-lactic acid) microspheres for image-guided radioembolisation using MRI.

Three-dimensional MRI-based assessment of the intrahepatic biodistribution of Ho-PLLA-MS was feasible. Ho-PLLA-MS concentration maps, which were constructed by using a time series of T_2^* -weighted images, clearly showed the ability of MRI to visualise the intrahepatic biodistribution of the microspheres. Furthermore, MRI provided both a quantitative measure of the Ho-PLLA-MS distribution and anatomical information, enabling direct localisation of the microsphere deposition within the tissue. MRI detected 89 % of the actual delivered Ho-PLLA-MS in the whole liver. The mean whole liver absorbed radiation dose calculated on MRI correlated very well with the absorbed radiation dose on SPECT. A more detailed intra-tumour biodistribution assessment was possible using MRI compared with SPECT because of the better spatial resolution of MRI. The quantitative nature of MRI-based biodistribution assessment of Ho-PLLA-MS allows for personalised dosimetry *in vivo* and may ultimately lead to optimisation of radioembolisation with regard to safety and efficacy.

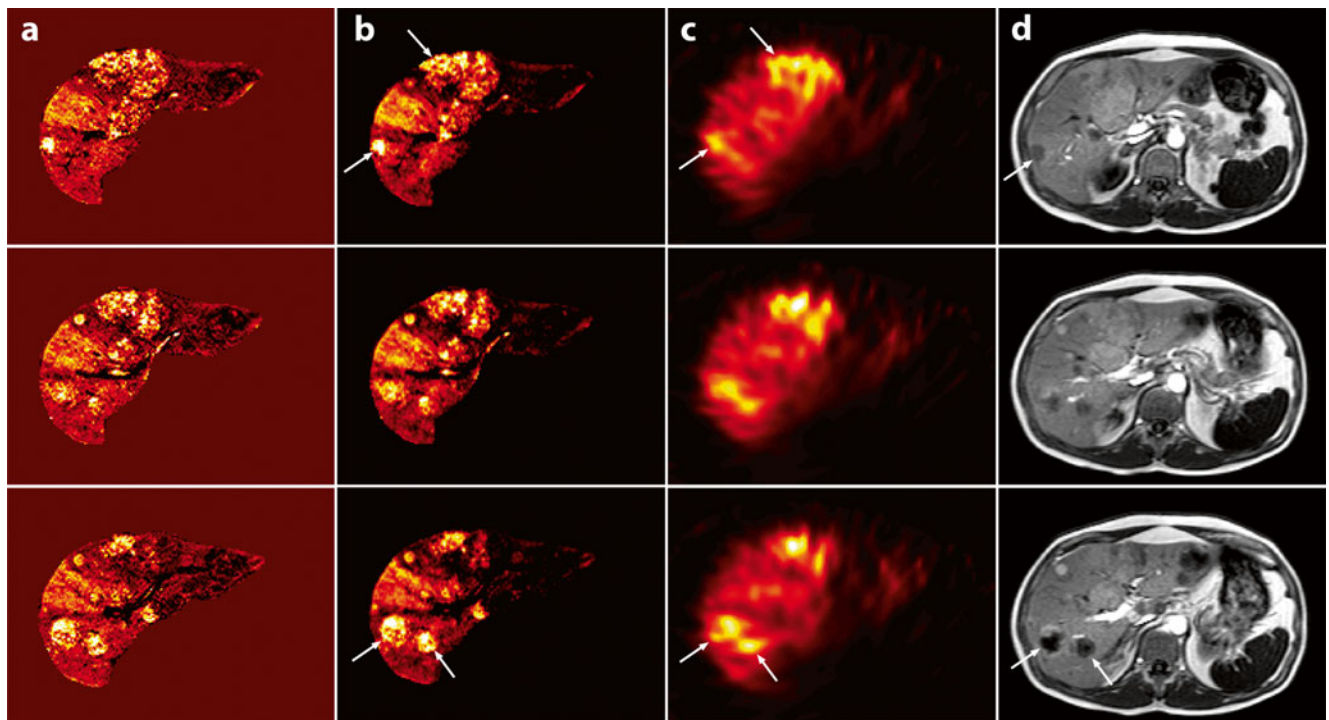
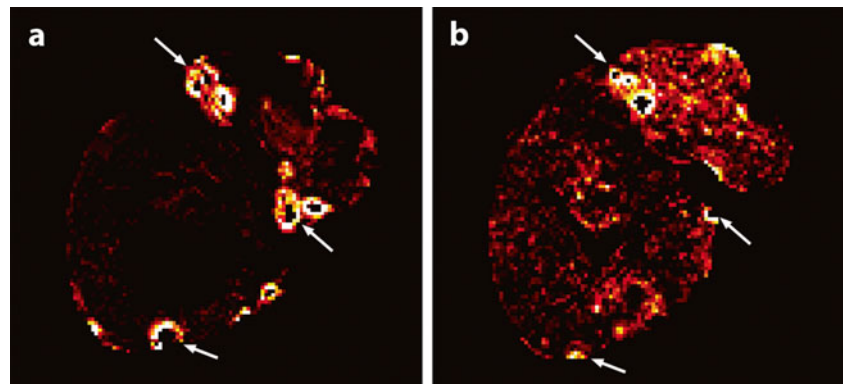


Fig. 3 MRI-based concentration maps (a) after Ho-PLLA-MS administration, MRI-based absorbed dose maps (b) together with their corresponding SPECT images (c) and T_1 -weighted images (d) (TR/TE: 8.5 ms/4.18 ms, flip angle: 10°). The concentration maps were constructed from normalised R_2^* maps by using the r_2^* relaxivity of

Ho-PLLA-MS. MRI-based dose maps were constructed by applying a dose point kernel after conversion of concentrations Ho-PLLA-MS to MBq/voxel. MRI-based concentration and absorbed dose correspond to the activity distribution on the SPECT images (c) and the tumour sites on T_1 -weighted images (d) (arrows)

Fig. 4 MRI-based Ho-PLLA-MS concentration maps before (a) and after (b) radioembolisation in a patient with multiple surgical clips after partial liver resection. At the location of the clips (arrows), quantification failed (black spots), while in the near vicinity extremely high concentration values were found (white rings)



Compared with the actual administered amount of Ho-PLLA-MS in the liver, determined based on the injected amount and taking into account the ^{99m}Tc -MAA-based lung shunt fraction, the MRI quantification method presented in this work showed on average an underestimation. As shown, this was in part due to the presence of surgical clips (Fig. 4). These clips induced field distortions, resulting in large MR signal voids surrounded by large R_2^* values. Therefore, locally the amount of Ho-PLLA-MS cannot be determined resulting in an underestimation of the microsphere amount in the vicinity of the clips. Although the presence of surgical clips does not necessarily lead to a contraindication for MRI-based dosimetry, biodistribution measurements in the vicinity of the clips should be interpreted with care. Another source of the discrepancy between the MRI-based amount and the amount delivered may be the use of ^{99m}Tc MAA as a predictor of the lung shunt fraction. Although used to correct for extrahepatic deposition in order to estimate the Ho-PLLA-MS liver amount, it is known that, because of a number of dissimilarities between ^{99m}Tc MAA

and Ho-PLLA-MS such as size, shape and density, ^{99m}Tc MAA is not the optimal microsphere analogy [6]. Incorrect assessment of the lung shunt fraction may have led to an erroneous estimation of the amount of Ho-PLLA-MS delivered in the liver. This explanation is supported by the good correlation that was found between the MRI-based mean liver absorbed dose and the absorbed dose based on the more conventional method of image-based dosimetry, SPECT.

Quantification errors may also arise from the need to correct for baseline R_2^* values in order to determine ΔR_2^* . ΔR_2^* values were calculated by subtracting the mean liver R_2^* value before administration from the post-treatment R_2^* maps. Because of the variance of R_2^* in the absence of microspheres as a result of tissue characteristics (Fig. 1), over- or underestimations will be introduced at the voxel level. The bias introduced in ΔR_2^* will depend on the local voxel R_2^* value with respect to the mean R_2^* over the whole VOI. In addition to these variations over space, changes may occur in tissue T_2 in the time interval between pre- and post-therapy imaging. Such changes may occur, for example, because of short-term therapy-induced oedema [32], which leads to increasing T_2 values, and thus decreasing R_2 values, owing to the higher water content as is generally reflected by hyperintense areas on T_2 -weighted images [33]. A decrease in tissue R_2 may therefore counteract the increase in R_2^* because of Ho-PLLA-MS leading to an underestimation of Ho-PLLA-MS concentration.

To minimise these quantification errors, voxel-based subtraction of post- and pre-therapy maps seems to be crucial. Manual or (semi-) automatic registration of these maps may partly solve the problem, but a more robust solution will be to integrate MR imaging as part of the RE procedure. Data needed to construct the concentration maps shown in this work were acquired within 1 min, and data processing can be performed within several minutes. Considering this short time scale, it is feasible to perform personalised MRI-based dosimetry during the actual RE procedure. This would enable fast verification and give the opportunity, if necessary, to immediately adjust the RE procedure. Moreover, MRI would enable monitoring of the administration of the Ho-PLLA-MS in real time. This has previously been demonstrated in a pig model [34].

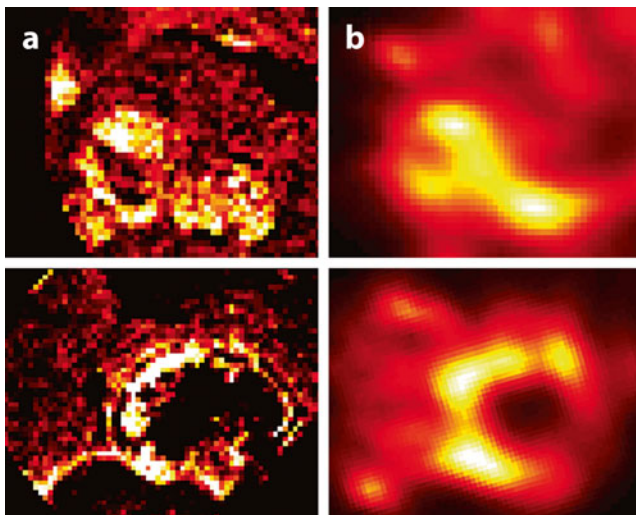


Fig. 5 Ho-PLLA-MS deposition around tumours. MRI-based concentration maps (a) show more detail (higher resolution) about the distribution of the microspheres than the SPECT-based activity maps. The Ho-PLLA-MS are primarily distributed in the peripheral parts of the tumour (a). This is less pronounced on the SPECT images (b)

The first MRI data of patients treated with ^{166}Ho -poly (L-lactic acid) microspheres show the feasibility of MRI-based dosimetry for radioembolisation. Intrahepatic microsphere biodistribution can be visualised with great detail, and a good quantitative measurement of the absorbed radiation dose can be obtained within several minutes of the radioembolisation procedure. Biodistribution assessment in the vicinity of surgical clips should be interpreted with care.

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References

- Salem R, Thurston KG (2006) Radioembolization with yttrium-90 microspheres: a state-of-the-art brachytherapy treatment for primary and secondary liver malignancies. *J Vasc Interv Radiol* 17: 1571–1594
- Kennedy AS, Coldwell D, Nutting C et al (2006) Resin 90Y-microsphere brachytherapy for unresectable colorectal liver metastases: modern USA experience. *Int J Radiat Oncol Biol Phys* 65:412–425
- Vente MAD, Hobbelink MGG, van het Schip AD, Zonnenberg BA, Nijsen JFW (2007) Radionuclide liver cancer therapies: from concept to current clinical status. *Anticancer Agents Med Chem* 7:441–459
- Gulec SA, Fong Y (2007) Yttrium 90 microsphere selective internal radiation treatment of hepatic colorectal metastases. *Arch Surg* 142:675–682
- Vente MAD, Wondergem M, van der Tweel I et al (2009) Yttrium-90 microsphere radioembolization for the treatment of liver malignancies: a structured meta-analysis. *Eur Radiol* 19:951–959
- Bult W, Vente MAD, Zonnenberg BA, Nijsen JFW (2009) Microsphere radioembolization of liver malignancies: current developments. *Q J Nucl Med Mol Imaging* 53:325–335
- Fabbri C, Sarti G, Cremonesi M et al (2009) Quantitative analysis of 90Y Bremsstrahlung SPECT-CT images for application to 3D patient-specific dosimetry. *Cancer Biother Radiopharm* 24:145–154
- Minarik D, Sjögreen-Gleisner K, Ljungberg M (2008) Evaluation of quantitative 90Y SPECT based on experimental phantom studies. *Phys Med Biol* 53:5689–5703
- Minarik D, Sjögreen-Gleisner K, Linden O et al (2010) 90Y Bremsstrahlung imaging for absorbed-dose assessment in high-dose radioimmunotherapy. *J Nucl Med* 51:1974–1978
- Elschot M, Nijsen JFW, Dam AJ, de Jong HWAM (2011) Quantitative evaluation of scintillation camera imaging characteristics of isotopes used in liver radioembolization. *PLoS One* 6:e26174
- Lhommel R, van Elmbt L, Goffette P et al (2010) Feasibility of 90Y TOF PET-based dosimetry in liver metastasis therapy using SIR-spheres. *Eur J Nucl Med Mol Imaging* 37:1654–1662
- Gates VL, Esmail AAH, Marshall K, Spies S, Salem R (2011) Internal pair production of 90Y permits hepatic localization of microspheres using routine PET: proof of concept. *J Nucl Med* 52:72–76
- Gupta T, Virmani S, Neidt TM et al (2008) MR Tracking of iron-labeled glass radioembolization microspheres during transcatheter delivery to rabbit VX2 liver tumors: feasibility study. *Radiol* 249:845–854
- Nijsen JFW, Zonnenberg BA, Woittiez JRW et al (1999) Holmium-166 poly lactic acid microspheres applicable for intra-arterial radionuclide therapy of hepatic malignancies: effects of preparation and neutron activation techniques. *Eur J Nucl Med* 26:699–704
- Nijsen JFW, van Steenberg MJ, Kooijman H et al (2001) Characterization of poly(L-lactic acid) microspheres loaded with holmium acetylacetonate. *Biomaterials* 22:3073–3081
- Nijsen JFW, Seppenwoolde JH, Havenith T, Bos C, Bakker CJG, van het Schip AD (2004) Liver tumors: MR imaging of radioactive holmium microspheres—phantom and rabbit study. *Radiol* 231:491–499
- Seppenwoolde JH, Nijsen JFW, Bartels LW, Zielhuis SW, van het Schip AD, Bakker CJG (2005) Internal radiation therapy of liver tumors: qualitative and quantitative magnetic resonance imaging of the biodistribution of holmium-loaded microspheres in animal models. *Magn Reson Med* 53:76–84
- Vente MAD, Nijsen JFW, de Wit TC et al (2008) Clinical effects of transcatheter hepatic arterial embolization with holmium-166 poly (L-lactic acid) microspheres in healthy pigs. *Eur J Nucl Med Mol Imaging* 7:1259–1271
- Seevinck PR, Seppenwoolde JH, de Wit TC et al (2007) Factors affecting the sensitivity and detection limits of MRI, CT, and SPECT for multimodal diagnostic and therapeutic agents. *Anticancer Agents Med Chem* 7:317–334
- Seevinck PR, van de Maat GH, de Wit TC, Vente MAD, Nijsen JFW, Bakker CJG (2012) Magnetic resonance imaging-based radiation-absorbed dose estimation of 166-Ho microspheres in liver radioembolization. *Int J Rad Oncol Biol Phys* 83:e437–e444
- Vente MA, Nijsen JF, de Roos R et al (2009) Neutron activation of holmium poly(L-lactic acid) microspheres for hepatic arterial radioembolization: a validation study. *Biomed Microdevices* 11:763–772
- Smits MLJ, Nijsen JFW, van den Bosch et al (2010) Holmium-166 radioembolization for the treatment of patients with liver metastases: design of the phase I HEPAR trial. *J Exp Clin Cancer Res* 29:70
- Coldwell D, Sangro B, Wasan H, Salem R, Kennedy A (2011) General selection criteria of patients for radioembolization of liver tumors: an international working group report. *Am J Clin Oncol* 34:337–341
- Kennedy A, Nag S, Salem R et al (2007) Recommendations for radioembolization of hepatic malignancies using yttrium-90 microsphere brachytherapy: a consensus panel report from the radioembolization brachytherapy oncology consortium. *Int J Radiat Oncol Biol Phys* 68:13–23

25. Leung WT, Lau WY, Ho SKW et al (1994) Measuring lung shunting in hepatocellular carcinoma with intrahepatic-arterial technetium-99 m macroaggregated albumin. *J Nucl Med* 35:70–73
26. de Wit T, Xiao J, Nijssen JF et al (2006) Hybrid scatter correction applied to quantitative holmium-166 SPECT. *Phys Med Biol* 51:4773–4787
27. Brown MA, Carden JA, Coleman RE, McKinney R Jr, Spicer LD (1987) Magnetic field effects on surgical ligation clips. *Magn Reson Imag* 5:443–453
28. Czervionke LF, Daniels DL, Wehrli FW et al (1988) Magnetic susceptibility artifacts in gradient-recalled echo MR imaging. *Am J Neuroradiol* 9:1149–1155
29. Port JD, Pomper MG (2000) Quantification and minimization of magnetic susceptibility artifacts on GRE images. *J Comput Assist Tomogr* 24:958–964
30. Bolch WE, Bouchet LG, Robertson JS et al (1999) MIRD pamphlet no. 17: the dosimetry of nonuniform activity distributions—radionuclide S values at the voxel level. Medical Internal Radiation Dose Committee. *J Nucl Med* 40:11S–36S
31. International Commission on Radiation Units and Measurements (1989) *Tissue Substitutes in Radiation Dosimetry and Measurement*. 44th edn. Bethesda, MD
32. Kamel IR, Bluemke DA (2002) Imaging evaluation of hepatocellular carcinoma. *J Vasc Interv Radiol* 13:S173–S183
33. Rummeny E, Weissleder R, Stark DD et al (1989) Primary liver tumours: diagnosis by MR imaging. *Am J Roentgenol* 152:63–72
34. Seppenwoolde JH, Bartels LW, van der Weide R, Nijssen JFW, van het Schip AD, Bakker CJG (2006) Fully MR-guided hepatic artery catheterization for selective drug delivery: a feasibility study in pigs. *J Magn Reson Imaging* 23:123–129