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## Beneficial Effect of Hyperbaric Oxygenation After Neonatal Germinal Matrix Hemorrhage

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### Abstract

**Background**—Germinal matrix hemorrhage (GMH) is a potentially devastating neurological disease of very low birth weight premature infants. This leads to post-hemorrhagic hydrocephalus, cerebral palsy, and mental retardation. Hyperbaric oxygen (HBO) treatment is a broad neuroprotectant after brain injury. This study investigated the therapeutic effect of HBO after neonatal GMH.

**Methods**—Neonatal rats underwent stereotaxic infusion of clostridial collagenase into the right germinal matrix (anterior caudate) brain region. Cognitive function was assessed at 3 weeks, and then sensorimotor, cerebral, cardiac, and splenic growths were measured 1 week thereafter.

**Results**—Hyperbaric oxygen (HBO) treatment markedly improved upon the mental retardation and cerebral palsy outcome measurements in rats at the juvenile developmental stage. The

administration of HBO early after neonatal GMH also normalized brain atrophy, splenomegaly, and cardiac hypertrophy 1 month after injury.

**Conclusion**—This study supports the role of hyperbaric oxygen (HBO) treatment in the early period after neonatal GMH. HBO is an effective strategy to help protect the infant's brain from the post-hemorrhagic consequences of brain atrophy, mental retardation, and cerebral palsy. Further studies are necessary to determine the mechanistic basis of these neuroprotective effects.

### Keywords

Hyperbaric oxygenation; Neurological deficits; Stroke, experimental

## Introduction

Germinal matrix hemorrhage (GMH) is a potentially devastating clinical condition with life-long consequences. This occurs when immature blood vessels rupture within the subventricular (anterior caudate) region during the first 7 days of life [1, 2]. Approximately 20–25% of very low birth weight (VLBW < 1,500 g) premature infants will be affected, accounting for around 3.5 per 1,000 births in the United States every year [3–6]. The long-term consequences of GMH are hydrocephalus (post-hemorrhagic ventricular dilation), developmental delay, cerebral palsy, and mental retardation [4, 7]. This is an important clinical problem for which experimental studies investigating therapeutic modalities are generally lacking [8].

Hyperbaric oxygen (HBO) treatment will induce multiple neuroprotective pathways across a wide domain of brain injury [9]. HBO has been shown in randomized controlled trials to improve cognition and overall functioning in autistic children [10], and may ameliorate outcomes after cerebral palsy as well [11]. Emerging evidence shows that both HBO and normobaric oxygen (NBO) treatments exert similar neuroprotective mechanisms to improve outcomes after adult ischemic stroke [12], and perhaps after neonatal hypoxia-ischemia as well [13].

In light of this evidence, we hypothesized that HBO therapy can be a therapeutic strategy to ameliorate brain injury mechanisms, to thereby improve cognitive and sensorimotor outcomes after germinal matrix hemorrhage in neonatal rats.

## Methods and Materials

### Animal Groups and General Procedures

This study was in accordance with the National Institutes of Health guidelines for the treatment of animals and was approved by the Institutional Animal Care and Use Committee at Loma Linda University. Timed pregnant Sprague-Dawley rats were housed with food and water available *ad libitum*. Treatment consisted of 1 h normobaric oxygen (NBO, 0 atm) or hyperbaric oxygen (HBO, 2.5 atm) administration beginning 1 h after collagenase infusion. Postnatal day 7 (P7) pups were blindly assigned to the following ( $n = 8/\text{group}$ ): sham (naive), needle (control), GMH (collagenase-infusion), GMH + NBO, and GMH + HBO. All groups were evenly divided within each litter.

### Experimental Model of GMH

Using aseptic technique, rat pups were gently anesthetized with 3% isoflurane (in mixed air and oxygen) while placed prone onto a stereotaxic frame. Betadine sterilized the surgical scalp area, which was incised in the longitudinal plane to expose the skull and reveal the bregma. The following stereotaxic coordinates were determined: 1 mm (anterior), 1.5 mm

(lateral), and 3.5 mm (ventral) from the bregma. A bore hole (1 mm) was drilled, into which a 27-gauge needle was inserted at a rate of 1 mm/min. A microinfusion pump (Harvard Apparatus, Holliston, MA) infused 0.3 units of clostridial collagenase VII-S (Sigma, St Louis, MO) through a Hamilton syringe. The needle remained in place for an additional 10 min after injection to prevent back-leakage. After needle removal, the burr hole was sealed with bone wax, the incision sutured closed, and the animals allowed to recover. The entire surgery took on average 20 min. Upon recovering from anesthesia, the animals were returned to their dams. Needle controls consisted of needle insertion alone without collagenase infusion, while naïve animals did not receive any surgery.

### Cognitive Measures

Higher order brain function was assessed during the third week after collagenase infusion. The T-maze assessed short-term (working) memory [14]. Rats were placed into the stem (40 cm × 10 cm) of a maze and allowed to explore until one arm (46 cm × 10 cm) was chosen. From the sequence of ten trials, of left and right arm choices, the rate of spontaneous alternation (0% = none and 100% = complete, alternations/trial) was calculated, as routinely performed [15, 16]. The Morris water maze assessed spatial learning and memory on four daily blocks, as described previously in detail [17, 18]. The apparatus consisted of a metal pool (110 cm diameter) filled to within 15 cm of the upper edge, with a platform (11 cm diameter) for the animal to escape onto, which changed location for each block (maximum = 60 s/trial), and data were digitally analyzed by Noldus Ethovision tracking software. Cued trials measured place learning with the escape platform visible above water. Spatial trials measured spatial learning with the platform submerged, and probe trials measured spatial memory once the platform was removed. For the locomotor activity, in an open field, the path length in open-topped plastic boxes (49 cm long, 35.5 cm wide, 44.5 cm tall) was digitally recorded for 30 min and analyzed by Noldus Ethovision tracking software [18].

### Sensorimotor Function

At 4 weeks after collagenase infusion, the animals were tested for functional ability. The neurodeficit was quantified using a summation of scores (maximum = 12) given for (1) postural reflex, (2) proprioceptive limb placing, (3) back pressure towards the edge, (4) lateral pressure towards the edge, (5) forelimb placement, and (6) lateral limb placement (2 = severe, 1 = moderate, 0 = none), as routinely performed [15]. For the rotarod, striatal ability was assessed using an apparatus consisting of a horizontal, accelerated (2 rpm/5 s), rotating cylinder (7 cm-diameter × 9.5 cm-wide), requiring continuous walking to avoid falling, recorded by photo-beam circuit (Columbus Instruments) [17, 18]. For foot fault, the number of complete limb missteps through the openings while exploring over an elevated wire (3 mm) grid (20 cm × 40 cm) floor was counted over 2 min [16].

### Assessment of Treatment upon Cerebral and Somatic Growth

At the completion of experiments, the brains were removed and hemispheres separated by midline incision (loss of brain weight has been used as the primary variable to estimate brain damage in juvenile animals after neonatal brain injury [19]). For organ weights, the spleen and heart were separated from surrounding tissue and vessels. The quantification was performed using an analytical microbalance (model AE 100; Mettler Instrument Co., Columbus, OH) capable of 1.0 µg precision.

### Statistical Analysis

Significance was considered at  $p < 0.05$ . Data were analyzed using analysis of variance (ANOVA) with repeated measures (RM-ANOVA) for long-term neurobehavior. Significant

interactions were explored with conservative Scheffé post hoc and Mann-Whitney rank sum when appropriate.

## Results

HBO and NBO treatment normalized T-maze deficits (Fig. 1a,  $p > 0.05$  compared to controls) and water maze (spatial) learning deficits (Fig. 1b,  $p < 0.05$  compared to GMH), without improving spatial memory (Fig. 1c,  $p > 0.05$ ). Oxygenation treatment also normalized ( $p < 0.05$ ) sensorimotor dysfunction (compared to juvenile GMH animals) in the neurodeficit score, the number of foot faults, and accelerating rotarod falling latency (Fig. 2a–c,  $p < 0.05$ ). This was confirmed with normalization of brain atrophy (Fig. 3a,  $p < 0.05$  compared to GMH), splenomegaly, and cardiomegaly (Fig. 3b, c,  $p > 0.05$  compared to controls).

## Discussion

These results indicate that hyperbaric oxygen (HBO) treatment after neonatal GMH can reduce long-term brain atrophy and return sensorimotor and cognitive functional deficits back to near-normal levels in juvenile animals. In support of the findings from others, this provides preliminary evidence about the importance of HBO-mediated mechanisms upon improvement of outcomes after neonatal injury [10–13].

Hyperbaric oxygen (HBO) treatment can induce multiple neuroprotective pathways across a wide domain of cerebral pathophysiology [9]. Our results support descriptive clinical studies using HBO to improve cognition and overall functioning in autistic and cerebral palsy children [10, 11]. In agreement with our findings, recent evidence shows that both HBO and normobaric oxygen (NBO) therapy can upregulate neuroprotective mechanisms and improve outcomes in adult ischemic stroke models [12]. Although hyper-oxygenation treatment has been shown to be protective after neonatal hypoxia-ischemia [13], our study demonstrates the first preliminary evidence, in this patient sub-population, advocating the use of oxygenation after hemorrhagic brain injury as well.

This study supports the notion that HBO treatment has no adverse effects in neonatal rats and can be applied as a strategy to improve functional outcomes after brain injury from hemorrhagic stroke in premature infants. Further investigation is needed to determine the mechanistic basis of these neuroprotective effects.

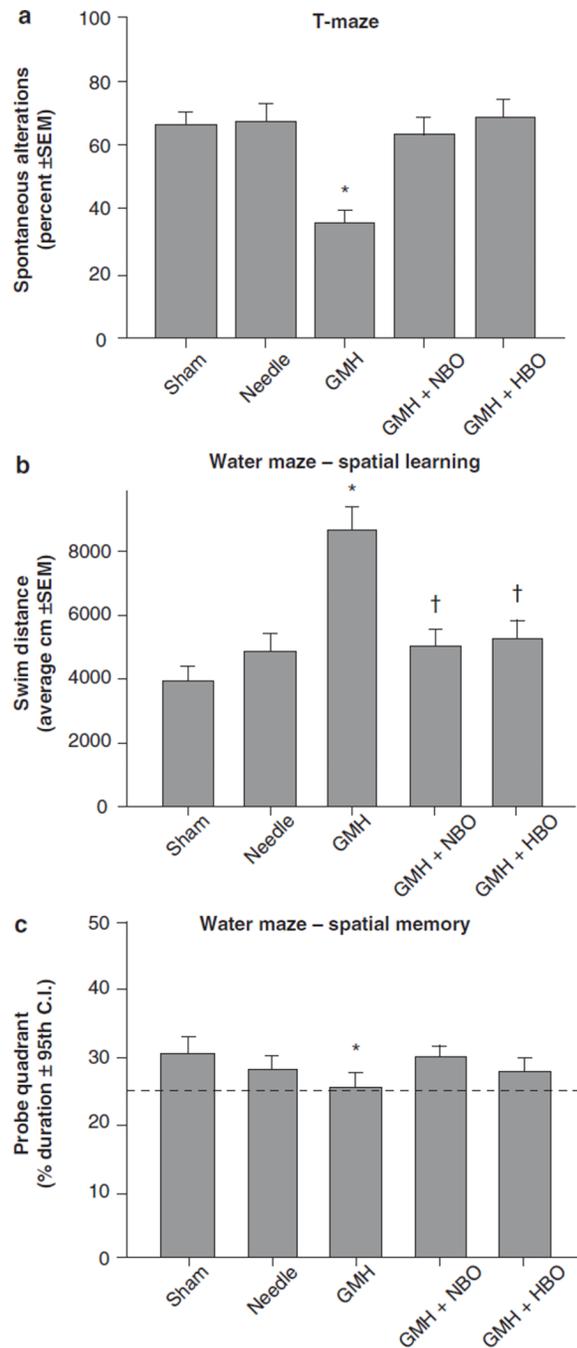
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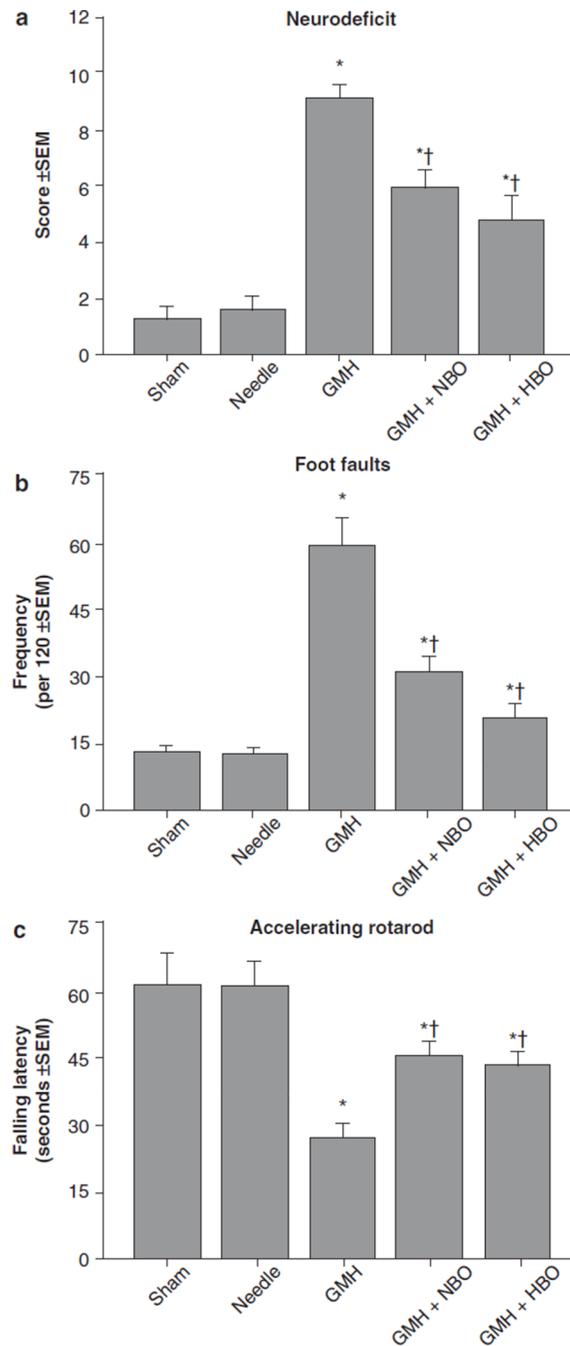
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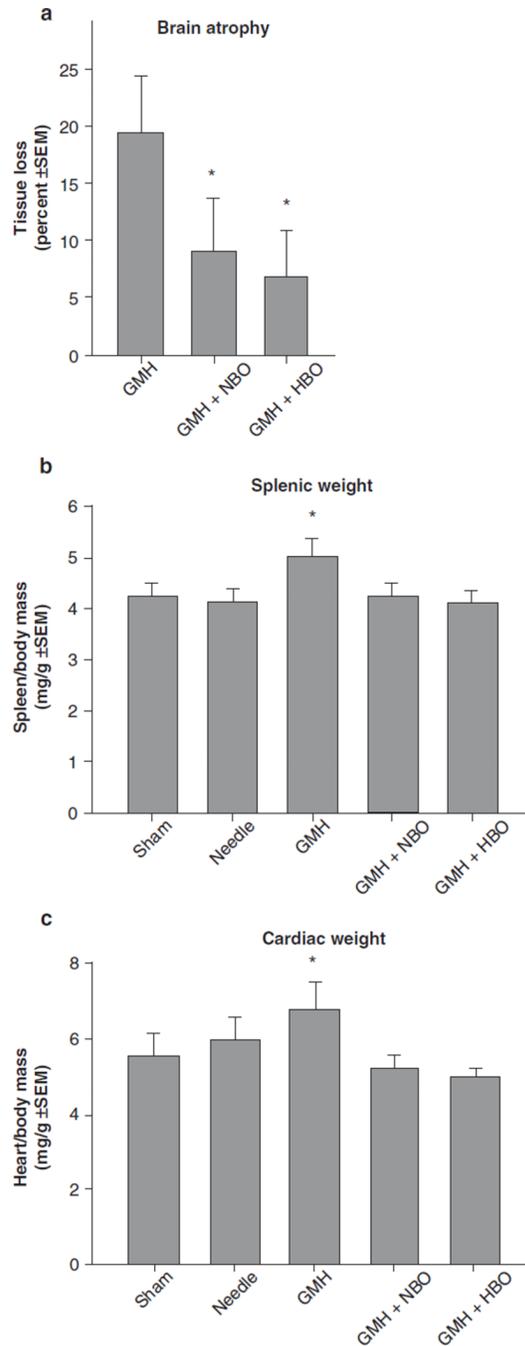
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**Fig. 1.** Cognitive function normalization in juvenile rats by HBO/ NBO after neonatal GMH. Higher order function was measured at the 3rd week after collagenase infusion: **(a)** T-maze, **(b)** spatial learning water maze, **(c)** spatial memory (probe) water maze. Values expressed as mean ± 95th CI (*probe quadrant*) or mean ± SEM (all others),  $n = 8$  (per group),  $*p < 0.05$  compared with controls (sham and needle trauma), and  $†p < 0.05$  compared with GMH



**Fig. 2.** Sensorimotor function normalization in juvenile rats by HBO/ NBO after neonatal GMH. Cerebral palsy measurements were performed in the juveniles at 1 month after collagenase infusion: **(a)** neurodeficit score, **(b)** foot faults and **(c)** rotarod. Values expressed as mean  $\pm$  SEM,  $n = 8$  (per group), \* $p < 0.05$  compared with controls (sham and needle trauma), and † $p < 0.05$  compared with GMH



**Fig. 3.** Cerebral and somatic growth normalization in juvenile rats by HBO/NBO after GMH. **(a)** Brain atrophy (percent tissue loss), **(b)** splenic weight, and **(c)** cardiac weight. Values expressed as mean  $\pm$  SEM,  $n = 8$  (per group), \* $p < 0.05$  compared with controls (sham and needle trauma)