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Obesity and Nonalcoholic Fatty Liver Disease: Biochemical, Metabolic and Clinical Implications

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Abstract

Obesity is associated with an increased risk of nonalcoholic fatty liver disease (NAFLD). Steatosis, the hallmark feature of NAFLD, occurs when the rate of hepatic fatty acid uptake from plasma and *de novo* fatty acid synthesis is greater than the rate of fatty acid oxidation and export (as triglyceride within VLDL). Therefore, an excessive amount of intrahepatic triglyceride represents an imbalance between complex interactions of metabolic events. The presence of steatosis is associated with a constellation of adverse alterations in glucose, fatty acid and lipoprotein metabolism. It is likely that abnormalities in fatty acid metabolism, in conjunction with adipose tissue, hepatic, and systemic inflammation, are key factors involved in the development of insulin resistance, dyslipidemia and other cardiometabolic risk factors associated with NAFLD. However, it is not clear whether NAFLD causes metabolic dysfunction or whether metabolic dysfunction is responsible for IHTG accumulation, or possibly both. Understanding the precise factors involved in the pathogenesis and pathophysiology of NAFLD will provide important insights into the mechanisms responsible for the cardiometabolic complications of obesity.

Obesity is associated with a spectrum of liver abnormalities, known as nonalcoholic fatty liver disease (NAFLD), characterized by an increase in intrahepatic triglyceride (IHTG) content (i.e. steatosis) with or without inflammation and fibrosis (i.e. steatohepatitis). NAFLD has become an important public health problem because of its high prevalence, potential progression to severe liver disease, and association with serious cardiometabolic abnormalities, including type 2 diabetes mellitus (T2DM), the metabolic syndrome and coronary heart disease (CHD).¹ In addition, the presence of NAFLD is associated with a high risk of developing T2DM, dyslipidemia (high plasma TG and/or low plasma HDL-cholesterol concentrations), and hypertension.² The purpose of this review is to provide a comprehensive assessment of the complex clinical and physiological interactions among NAFLD, adiposity, and metabolic dysfunction.

DIAGNOSIS AND PREVALENCE

The hallmark feature of NAFLD is steatosis. Excessive intrahepatic triglyceride (IHTG), or steatosis, has been chemically defined as IHTG content >5% of liver volume or liver weight,³ or histologically defined when 5% or more of hepatocytes contain visible intracellular triglycerides (TG).⁴ Recently, data obtained from two studies, which evaluated IHTG content by using magnetic resonance spectroscopy (MRS) in large numbers of

subjects, provide additional insights into defining “normal” IHTG content.^{5,6} The results from one study, conducted in a cohort of Hispanic and non-Hispanic Caucasians and African American subjects, who were considered to be at low-risk for NAFLD (i.e. BMI < 25 kg/m², no diabetes, and normal fasting serum glucose and alanine aminotransferase concentrations), suggest the threshold for a normal amount of IHTG should be 5.6% of liver volume, because this value represented the 95th percentile for this “normal” population.⁶ Data from the second study, found the 95th percentile for IHTG content was 3% in lean, young adult, and Caucasian men and women who had normal oral glucose tolerance.⁵ However, none of the values proposed for diagnosing steatosis are based on the relationship between IHTG and a rigorous assessment of either metabolic or clinical outcome. In fact, the relationship between insulin sensitivity and IHTG content in obese subjects is monotonic, without evidence of an obvious threshold that can be used to define normality.⁷

The prevalence rate of NAFLD increases with increasing body mass index (BMI).⁸ An analysis of liver histology obtained from liver donors,⁹ automobile crash victims,¹⁰ autopsy findings,¹¹ and clinical liver biopsies¹² suggests that the prevalence rates of steatosis and steatohepatitis are approximately 15% and 3%, respectively, in non-obese persons, 65% and 20%, respectively, in persons with class I and II obesity (BMI 30.0–39.9 kg/m²), and 85% and 40%, respectively, in extremely obese patients (BMI ≥ 40 kg/m²). The relationship between BMI and NAFLD is influenced by racial/ethnic background and genetic variation in specific genes.^{5,13,14}

LIVER PHYSIOLOGY AND PATHOPHYSIOLOGY

The liver is a metabolic workhorse that performs a diverse array of biochemical functions necessary for whole-body metabolic homeostasis. The metabolic activities of the liver require a rich blood supply for delivery and export of substrates, hormones, and nutrients. The hepatic vascular network consists of a dual contribution from the hepatic artery, which delivers ~30%, and the portal vein, which delivers ~70%, of the blood reaching the liver.¹⁵ During basal conditions, 1.5 L of blood are transported to the liver every min, which deliver a large load of compounds that require metabolic processing. Excessive accumulation of IHTG is associated with alterations in glucose, fatty acid (FA) and lipoprotein metabolism and inflammation, which have adverse consequences on health. However, it is not clear whether NAFLD causes these abnormalities or whether these metabolic abnormalities cause IHTG accumulation. In addition, the relationship between NAFLD and metabolic complications is often confounded by concomitant increases in visceral adipose tissue and intramyocellular TG, which are also risk factors for metabolic dysfunction.^{7,16,17} Therefore, persons with increased IHTG often have increased ectopic fat accumulation in other organs and increased visceral fat mass.¹⁷

Hepatic Lipid Metabolism

Steatosis develops when the rate of FA input (uptake and synthesis with subsequent esterification to TG) is greater than the rate of FA output (oxidation and secretion). Therefore, the amount of TG present in hepatocytes represents a complex interaction among: 1) *hepatic fatty acid uptake*, derived from plasma free fatty acid (FFA) released from hydrolysis of adipose tissue TG and FFA released from hydrolysis of circulating TG, 2) *de novo* fatty acid synthesis (*de novo* lipogenesis [DNL]), 3) *fatty acid oxidation* (FAO), and 4) *fatty acid export within VLDL-TG* (Figure 1).

Fatty acid uptake—The rate of hepatic FFA uptake depends on the delivery of FFA to the liver and the liver’s capacity for FFA transport. During postabsorptive conditions, the major source of FFA delivered to the liver is derived from FFA released from subcutaneous adipose tissue, which enter the systemic circulation and are then transported to the liver by

the hepatic artery and portal vein, after passage through splanchnic tissues. Although lipolysis of visceral adipose tissue TG releases additional FFA directly into the portal system, the relative contribution of portal vein FFA derived from visceral fat is small compared with FFA derived from subcutaneous fat; only about 5% and 20% of portal vein FFA originate from visceral fat in lean and obese subjects, respectively.¹⁸ We have found that the rate of FA release into the systemic circulation increases directly with increasing fat mass in both men and women, so that the rate of FFA release in relationship to fat-free mass is greater in obese than lean subjects.¹⁹ In addition, gene expression of hepatic lipase and hepatic lipoprotein lipase (LPL) are higher in obese subjects with NAFLD than subjects without NAFLD, suggesting that FFA released from lipolysis of circulating TG also contribute to hepatocellular FFA accumulation and steatosis.^{20,21} It is possible that these increases in hepatic lipase and hepatic LPL, along with higher postprandial lipemia and FFA concentrations reported in subjects with NAFLD,²² are responsible for the increased postprandial incorporation of dietary fatty acids into IHTG observed in obese subjects with T2DM.²³ Membrane proteins that direct trafficking of FFA from plasma into tissues are also likely involved in increased hepatic FFA uptake. Gene expression and/or protein content of FAT/CD36, which is an important regulator of tissue FFA uptake from plasma, are increased in liver and skeletal muscle but decreased in adipose tissue in obese subjects with NAFLD compared with obese subjects who have normal IHTG content,^{24,25} suggesting that membrane fatty acid transport proteins redirect the uptake of plasma FFA from adipose tissue toward other tissues. Therefore, the summation of these data suggests that alterations in adipose tissue lipolytic activity, regional hepatic lipolysis of circulating TG, and tissue FFA transport proteins are involved in the pathogenesis of steatosis and ectopic fat accumulation (Figure 2).

De novo lipogenesis—The liver synthesizes fatty acids *de novo* through a complex cytosolic polymerization in which acetyl-CoA is converted to malonyl-CoA by acetyl-CoA carboxylase (ACC), and undergoes several cycles metabolic reactions to form one palmitate molecule. The rate of DNL is regulated by the fatty acid synthase (FAS) complex, ACC 1 and 2, diacylglycerol acyltransferase (DGAT) 1 and 2, stearoyl-CoA desaturase 1 (SCD1), and several nuclear transcription factors (SREBPs, ChREBP, liver X receptor α [LXR α], farnesoid X receptor [FXR], and peroxisome proliferator-activated receptors [PPARs]).²⁶ Hepatic DNL is regulated independently by insulin and glucose, through the activation of SREBP-1c²⁷ and ChREBP,²⁸ which transcriptionally activate nearly all genes involved in DNL. Data from studies conducted in mouse models demonstrate that hepatic overexpression of SREBP-1c or hyperinsulinemia stimulate lipogenesis and cause hepatic steatosis,^{29,30} whereas the levels of all the enzymes involved in DNL are reduced in ChREBP knockout mice.³¹ In humans, NAFLD is associated with increased hepatic expression of several genes involved in DNL.^{32,33}

The contribution of DNL to total IHTG production in normal subjects is small and accounts for less than 5% of fatty acids incorporated into secreted VLDL-TG (~1–2 g/d)³⁴. However, the contribution of DNL to total IHTG production in subjects with NAFLD is much higher and accounts for 15%–23% of the fatty acids within IHTG and secreted in VLDL-TG.^{34,35} Moreover, data from a study that used sophisticated magnetic resonance spectroscopy techniques to evaluate postprandial glucose metabolism *in vivo* suggest that the increase in DNL precedes the development of NAFLD.³⁶ Compared with insulin-sensitive subjects, consumption of a high-carbohydrate meal was associated with a much lower rate of muscle glycogen synthesis and a diversion of most of the ingested glucose toward hepatic DNL and IHTG synthesis in insulin-resistant subjects who had normal IHTG content. These data suggest that insulin resistance in skeletal muscle could promote IHTG accumulation by diverting ingested carbohydrate away from storage as muscle glycogen and toward *de novo* fatty acid synthesis.

Although hepatic DNL is a quantitatively minor pathway for TG synthesis, the rate of DNL might have important metabolic regulatory functions. For example, intrahepatic fatty acids that have been synthesized *de novo* activate PPAR α to maintain glucose and lipid homeostasis.³⁷ In addition, malonyl-CoA, the first intermediate of DNL, inhibits carnitine palmitoyltransferase 1 (CPT1) activity, thereby preventing the entry of FFA into the mitochondrion and inhibiting FAO.³⁸ The notion of potential allosteric inhibition of FAO by DNL is supported by data that found hepatic CPT-1 expression is decreased in subjects with NAFLD.³³

Fatty acid oxidation—The complex metabolic processes performed by the liver require a considerable amount of energy; the metabolic rate of liver tissue (~0.28 kcal/g of tissue per day) is similar to that of the brain, and is nearly 20 times greater than the metabolic rate of resting skeletal muscle and 50 times greater than the metabolic rate of adipose tissue.³⁹ Therefore, although the liver weighs only ~1.5 kg in adults, representing a small portion of total body weight (~2.5% in lean persons), it consumes ~450 kcal/d and accounts for ~20% of total resting energy expenditure.³⁹ The mix of fuels used by the liver *in vivo* is difficult to quantify accurately because of the complicated exchange of metabolites between multiple biochemical pathways and technical limitations. It is estimated that fatty acid and amino acid oxidation provide ~90% of the fuel for basal hepatic energy requirements, and that the use of FFA as a fuel decreases during the fed state.⁴⁰

The oxidation of intrahepatocellular fatty acid occurs primarily within mitochondria, and to a much lesser extent by peroxisomes and microsomes. Transport of FA inside the mitochondrial matrix is regulated by a carnitine-dependent enzyme shuttle, sequentially CPT1, carnitine translocase, and CPT2. Mitochondrial β -oxidation progressively shortens the fatty acyl-CoA by two carbon units at each cycle (released as acetyl-CoA), through a series of dehydrogenation, hydration, and cleavage reactions that involve a membrane-bound and soluble enzymes, which are transcriptionally regulated by PPAR- α .⁴¹ Acetyl-CoA derived from FAO can either enter the tricarboxylic acid cycle for complete oxidation and energy production for the liver, or can be condensed to form ketone bodies (acetoacetate and beta-hydroxybutyrate) which are exported to provide fuel for other tissues.³⁸

Data from studies conducted in rodent models demonstrate that inhibition or activation of intrahepatic FAO can influence IHTG content. Genetic or experimentally-induced deficiencies in mitochondrial oxidative enzymes lead to hepatic steatosis,^{42,43} whereas increasing the expression or activity of hepatic enzymes involved in FAO, reduces IHTG accumulation.^{44–47} However, it is not known whether FAO is defective in human subjects with NAFLD, because there are currently no reliable methods for measuring hepatic FAO *in vivo*. Indirect measures of hepatic mitochondrial FAO, assessed by plasma ketone body concentrations, suggest that hepatic FAO is either increased or normal in subjects with NAFLD.^{48–51} In addition, although CPT1 expression is decreased, gene expression of other hepatic fatty acid oxidative enzymes are generally greater in subject with NAFLD than in those with normal IHTG content.^{24,33} In contrast, subjects with NAFLD have evidence of hepatic mitochondrial structural and functional abnormalities, including loss of mitochondrial cristae and paracrystalline inclusions,^{49,52} a decrease in mitochondrial respiratory chain activity,⁵³ impaired ability to resynthesize ATP after a fructose challenge,⁵⁴ and increased hepatic uncoupling protein 2,³³ which affect energy production but not FAO. These abnormalities could represent an adaptive uncoupling of FAO and ATP production, which allows the liver to oxidize excessive FA substrates without generating unneeded ATP.

VLDL kinetics—Very-low-density lipoproteins are complex lipoprotein particles that are produced by the liver and secreted into the systemic circulation. The formation of VLDL

provides an important mechanism for converting water-insoluble TG into a water-soluble form that can be exported from the liver and delivered to peripheral tissues. Hepatic VLDL assembly involves the fusion of a newly synthesized apolipoprotein B-100 (apoB-100) molecule with a TG droplet through the action of microsomal triglyceride transfer protein (MTP); each VLDL particle contains a single molecule of apoB-100. The fatty acids that are esterified into TG and secreted as VLDL are derived from several sources. In subjects with normal IHTG content, ~70% of FA incorporated into VLDL-TG originate from systemic plasma and the remaining are derived from several non-systemic FA sources, including hepatic DNL, lipolysis of IHTG, and lipolysis of visceral adipose tissue.⁵⁵

Intrahepatocellular fatty acids that are not oxidized are esterified to TG, which can either be incorporated into VLDL and secreted into the circulation or stored within the liver. Therefore, the secretion of VLDL provides a mechanism for reducing IHTG content. In fact, an impairment in hepatic VLDL secretion, caused by genetic defects, such as familial hypobetalipoproteinemia,⁵⁶ or pharmacological agents that inhibit MTP,⁵⁷ are associated with an increase in IHTG content. However, data from most^{58,59} but not all³⁴ studies have found that VLDL-TG secretion rate is greater in subjects with NAFLD than in those with normal IHTG content. We found that the rate of VLDL-TG secretion was twice as great in non-diabetic obese subjects with NAFLD than in those with normal IHTG content, who were matched on BMI and percent body fat (Figure 3). The increase in VLDL-TG secretion was almost entirely accounted for by a marked increase in the contribution of non-systemic FA, presumably derived from lipolysis of intrahepatic and visceral fat and DNL, to VLDL-TG secretion.⁵⁹ In addition, the relationship between VLDL-TG secretion and IHTG content differed between the two groups; VLDL-TG secretion increased linearly with increasing IHTG content in subjects with normal IHTG, but appeared to reach a plateau in subjects with NAFLD, independent of IHTG content (Figure 4). Therefore, the increase in VLDL-TG secretion rate in subjects with NAFLD is not able to adequately compensate for the increased rate of IHTG production, so steatosis is maintained.

The mechanism responsible for the inadequate increase in hepatic TG export is not known, but might be related to physical limitations in the liver's ability to secrete large VLDL particles. In contrast to VLDL-TG kinetics, the secretion rate of VLDL-apoB-100 was not different between subjects with high and low IHTG content, so the molar ratio of VLDL-TG to VLDL-apoB-100 secretion rates, an index of the TG content of nascent VLDL, was more than two-fold greater in those with NAFLD.⁵⁹ Data from a study conducted in transgenic mice that overexpress SREBP-1a and develop massive steatosis found that very large VLDL particles cannot be secreted from the liver because they exceed the diameter of the sinusoidal endothelial pores, resulting in an accumulation of IHTG.⁶⁰ Therefore, the composite of these data suggest that the failure to upregulate VLDL-apoB secretion rate in obese subjects with NAFLD leads to the production of large VLDL particles, which cannot penetrate sinusoidal endothelial pores for export out of the liver.

Insulin sensitivity

Insulin has important metabolic effects in multiple organ systems. Although the term “insulin resistance” is usually used to describe impaired insulin-mediated glucose uptake in skeletal muscle, insulin resistance associated with obesity and NAFLD also involves the liver (impaired insulin-mediated suppression of glucose production) and adipose tissue (impaired insulin-mediated suppression of lipolysis). The presence of steatosis is an important marker of multi-organ insulin resistance, independent of BMI, percent body fat, and visceral fat mass,^{7,16,25,48,61} Moreover, insulin resistance in liver, adipose tissue and skeletal muscle is directly related to percent liver fat (Figure 5).^{7,48,49,61} However, it is not known whether NAFLD causes or is a consequence of insulin resistance, or possibly both.

Fatty acid metabolism—Whole-body lipolytic rates, expressed as the rate of FFA release per unit of fat-free mass, is usually greater in obese than lean persons and is directly related with body fat mass.¹⁹ The presence of NAFLD in obese persons is associated with adipose tissue insulin resistance and even greater rates of adipose tissue lipolysis than in obese persons without NAFLD.^{7,16,48,61} Excessive rates of release of FFA from adipose tissue into the circulation increases the delivery of FFA to the liver and skeletal muscle, which can simultaneously lead to an increase in IHTG and cause insulin resistance in liver and skeletal muscle.⁶² Skeletal muscle insulin resistance and hyperinsulinemia can further increase the accumulation of IHTG by stimulating hepatic DNL and TG synthesis.³⁶ An increase in IHTG content itself could be involved in the pathogenesis of hepatic insulin resistance by releasing FA into the cytoplasm, which can have adverse effects on insulin signaling.⁶²

The cellular mechanism(s) responsible for fatty-acid induced insulin resistance in muscle and liver is not completely clear. A large volume of data from studies conducted in animal models and human subjects suggest that excessive intracellular lipid intermediates generated by fatty acid metabolism, particularly diacylglycerol (DAG), long chain fatty acyl-CoA, ceramide, lysophosphatidic acid, and phosphatidic acid, can interfere with insulin action by activating protein kinase C and mTOR, and inhibiting Akt, which have direct adverse effects on insulin signaling, and by activating the nuclear factor kinase B (NFκB) system which can cause insulin resistance through activation of inflammatory pathways (Figure 6).^{63,64} However, these conclusions are based primarily on studies that have simply demonstrated an association between these lipid intermediates and impaired insulin action, and not a cause-and-effect relationship. Moreover, the results from some studies have found that an increase in these lipid intermediates is not associated with insulin resistance.^{65–67} The ability to identify the cellular mediators responsible for FA-induced insulin resistance is further complicated by the possibility that the mechanism might not be the same among all tissues. Transgenic mice that overexpress *muscle* diacylglycerol acyltransferase (DGAT2), which catalyzes the final step of TG synthesis by adding fatty acyl-CoA to DAG, have high intramyocellular levels of DAG, long chain fatty acyl-CoA, and ceramide and have abnormal hepatic insulin sensitivity, impaired insulin signaling and insulin-mediated glucose uptake.⁶⁸ In contrast, transgenic mice that overexpress *hepatic* DGAT2 have high intrahepatocellular levels of DAG, long chain fatty acyl-CoA, and ceramide, but do not have abnormal hepatic insulin sensitivity.⁶⁹

Adipose tissue inflammation—Adipose tissue contains several different cell types, including adipocytes and macrophages that produce cytokines (e.g. IL-6 and TNF-α), and chemokines (e.g. CCL2 [also known as monocyte chemoattractant protein 1]), which can cause inflammation and insulin resistance.⁷⁰ Adipose tissue macrophage content and production of cytokines and chemokines are greater in obese than lean subjects.⁷¹ Moreover, macrophage infiltration and inflammatory markers are greater in adipose tissue of subjects with NAFLD than BMI-matched subjects with normal IHTG content.⁷² Therefore, the secretion of adipose tissue inflammatory proteins in obese persons with NAFLD is likely involved in the pathogenesis of insulin resistance, but the relative contribution of adipose tissue inflammation in comparison with other potential factors that can cause insulin resistance is not known.

Intrahepatic inflammation—Diet-induced and genetically-induced obesity in rodent models cause steatosis, insulin resistance, and increased hepatic NF-κB activity.^{73,74} In addition, selective activation of hepatocellular NF-κB causes hepatic inflammation without steatosis, and results in both hepatic and skeletal muscle insulin resistance.⁷⁴ These animals have increased hepatocyte expression of IL-6 and plasma IL-6 concentrations, suppression of IL-6 activity by administering neutralizing IL-6 antibodies resulted in a decrease in both hepatic and peripheral insulin resistance. These data suggest that steatosis can cause both

hepatic and systemic insulin resistance by activating NF- κ B, which upregulates the production of proinflammatory cytokines that affect both local and systemic insulin action.

Adipocyte-derived hormones—Adipocytes produce a series of peptide hormones, which are associated with insulin action (e.g. resistin, retinol-binding protein 4, adiponectin, leptin). Among these proteins, adiponectin, which is the most abundant secretory protein produced by adipose tissue, is the most closely related with insulin action. Plasma adiponectin concentrations are inversely associated with hepatic steatosis,^{22,75} insulin resistance,⁷⁶ T2DM,⁷⁷ and the metabolic syndrome. Delivery of recombinant adiponectin into mice with liver steatosis markedly reduced hepatomegaly and IHTG content.⁴⁴

Endoplasmic reticulum stress—The endoplasmic reticulum (ER) is a critical intracellular organelle that coordinates synthesis, folding and trafficking of proteins. Transmembrane and secreted proteins are folded into the ER and then directed to cellular destinations. Unfolded or misfolded proteins are detected, removed from the ER and degraded by proteasome system.⁷⁸ Under stress conditions, such as hypoxia, alterations in energy and substrates, toxins, viral infections, unfolded proteins accumulate in the ER and initiate an adaptive response known as the unfolded protein response (UPR) in an effort to restore organelle function.⁷⁹ The UPR is initiated by three ER transmembrane sensors, PKR-like endoplasmic-reticulum kinase (PERK), inositol-requiring enzyme 1 (IRE-1), and activating transcription factor 6 (ATF6). These transmembrane sensors activate an adaptive response that results in cessation of protein synthesis, increase of protein-folding chaperones, and increase in ER-associated degradation genes. The UPR is also able to induce activation of the c-Jun NH2-terminal kinase (JNK) pathway and thereby inhibit insulin signaling through the subsequent phosphorylation and/or degradation of IRS1.⁸⁰ Recent data from experimental models indicate that ER stress is critical to the initiation and integration of pathways of inflammation and insulin action in obesity, T2DM, and NAFLD. ER stress response can be induced in the liver by saturated FA in rats,⁸¹ and this activation can lead to activation of JNK and insulin resistance.⁸⁰ Activation of ER stress in the liver has also been shown in human subjects with NAFLD, as documented by activation of PERK and an increase in the ER chaperone GRP78 mRNA.⁸² Data from a study conducted in extremely obese patients, found that ER stress is associated with NAFLD and improves with weight loss and resolution of steatosis. Bariatric surgery-induced weight loss increased insulin sensitivity in multiple organs and decreased IHTG content and both liver and adipose tissue activation of all three ER stress pathways.⁸³

Hepatic steatosis in the absence of insulin resistance—The complexity of the relationship between NAFLD and insulin resistance is underscored by the observation that steatosis is not always associated with insulin resistance. A dissociation between steatosis and insulin resistance has been reported in selected genetically-altered or pharmacologically-manipulated animal models and human subjects. Overexpression of hepatic DGAT,⁶⁹ blockade of hepatic VLDL secretion,⁶⁶ and pharmacological blockade of β -oxidation⁸⁴ in mice causes hepatic steatosis, but not hepatic or skeletal muscle insulin resistance, whereas inhibiting hepatocyte TG synthesis in obese mice decreases hepatic steatosis but does not improve insulin sensitivity.⁸⁵ In addition, hepatic steatosis caused by genetic deficiency of apoB synthesis and decreased VLDL hepatic secretion in patients with familial hypobetalipoproteinemia is not accompanied by hepatic or peripheral insulin resistance. (S. Klein unpublished observations). These data support the notion that hepatic accumulation of TG does not necessarily cause insulin resistance. In fact, it is possible that the esterification of excessive FA to inert TG molecules protects the hepatocyte by preventing the accumulation of potentially toxic intracellular fatty acids.⁸⁶ Inhibiting hepatocyte TG synthesis by treatment with DGAT2 antisense oligonucleotide in obese mice

decreased hepatic steatosis, but increased hepatic free FA, markers of lipid peroxidation/oxidant stress, lobular necroinflammation, and fibrosis.⁸⁷ The mechanism(s) responsible for the marked differences in the relationship between hepatic steatosis and insulin resistance across different studies is not known, but suggest that other factors associated with steatosis, such as inflammation, circulating adipokines, ER stress, or as yet unidentified lipid metabolites, affect insulin sensitivity but are not necessarily directly related with IHTG content. It is also possible that there is a temporal dissociation between steatosis and insulin resistance, so that IHTG accumulation is secondary to a primary defect in skeletal muscle insulin action, by diverting ingested carbohydrates away from muscle glycogen storage to DNL.³⁶

ENERGY BALANCE AND NAFLD

Calorie restriction and weight loss is an effective therapy for obese patients with NAFLD. A marked decrease in IHTG content and improvement in hepatic insulin sensitivity occurs very rapidly, within 48 h, of calorie restriction (~1100 kcal/d diet).⁸⁸ A comprehensive review of 14 studies that evaluated the effect of lifestyle weight loss therapy on NAFLD/NASH,⁸⁹ and data from recent prospective diet intervention studies,^{88,90} found that a 5–10% weight loss improved liver biochemistries, liver histology (steatosis and inflammation) and IHTG content, in conjunction with an increase in hepatic and skeletal muscle insulin sensitivity, and decrease in hepatic VLDL-TG secretion rate.⁹¹ Bariatric surgery is the most effective available weight loss therapy. There has been concern that the large and rapid weight loss, induced by bariatric surgery, can actually worsen NAFLD by increasing hepatic inflammation and fibrosis.⁹² However, data from more recent surgical series suggest that weight loss induced by bariatric surgery decreases steatosis, inflammation and fibrosis.^{93,94} In addition, bariatric surgery induced weight loss has considerable beneficial metabolic effects in the liver manifested by a decrease in: 1) hepatic glucose production, 2) hepatic VLDL-triglyceride secretion rate, and 3) hepatic gene expression of factors that regulate hepatic inflammation and fibrogenesis.⁹⁵ These data suggest that bariatric surgery-induced weight loss is an effective therapy for NAFLD in patients with morbid obesity, by normalizing the metabolic abnormalities involved in the pathogenesis and pathophysiology of NAFLD, and by preventing the progression of hepatic inflammation and fibrosis.

The effect of overfeeding on IHTG content and metabolic function has not been carefully studied in human subjects. Data from a study conducted in rodents suggest that overfeeding first has metabolic effect on the liver followed by an effect on muscle; insulin resistance in liver was observed after 3 d and in muscle after 7 days of overfeeding.⁹⁶ Four wks of overfeeding in lean men and women, designed to cause a 5%-15% increase in body weight, resulted in a significant increase in IHTG content (from 1.1% to 2.8%), a decline in insulin sensitivity, and an increase in serum transaminase concentrations.⁹⁷

CONCLUSIONS

Although obesity is associated with multiple metabolic risk factors for cardiovascular disease, including insulin resistance, diabetes, and dyslipidemia, about 30% of obese adults are “metabolically normal”, usually defined by some measure of insulin sensitivity or having 1 cardiometabolic abnormality.^{98–100} Excessive intrahepatic triglyceride (IHTG) content in obese persons is a robust marker of metabolic abnormalities (insulin resistance in liver, muscle and adipose tissue, alterations in FFA metabolism, and increased VLDL-TG secretion rate), independent of BMI, percent body fat, and visceral fat mass. Conversely, obese persons who have normal IHTG content appear to be resistant to developing obesity-related metabolic complications. However, it is not known whether NAFLD is a cause or a consequence of metabolic dysfunction. A better understanding of the mechanisms

responsible for the pathogenesis and pathophysiology of NAFLD will potentially identify both novel biomarkers for metabolic risk and unique targets for therapeutic intervention.

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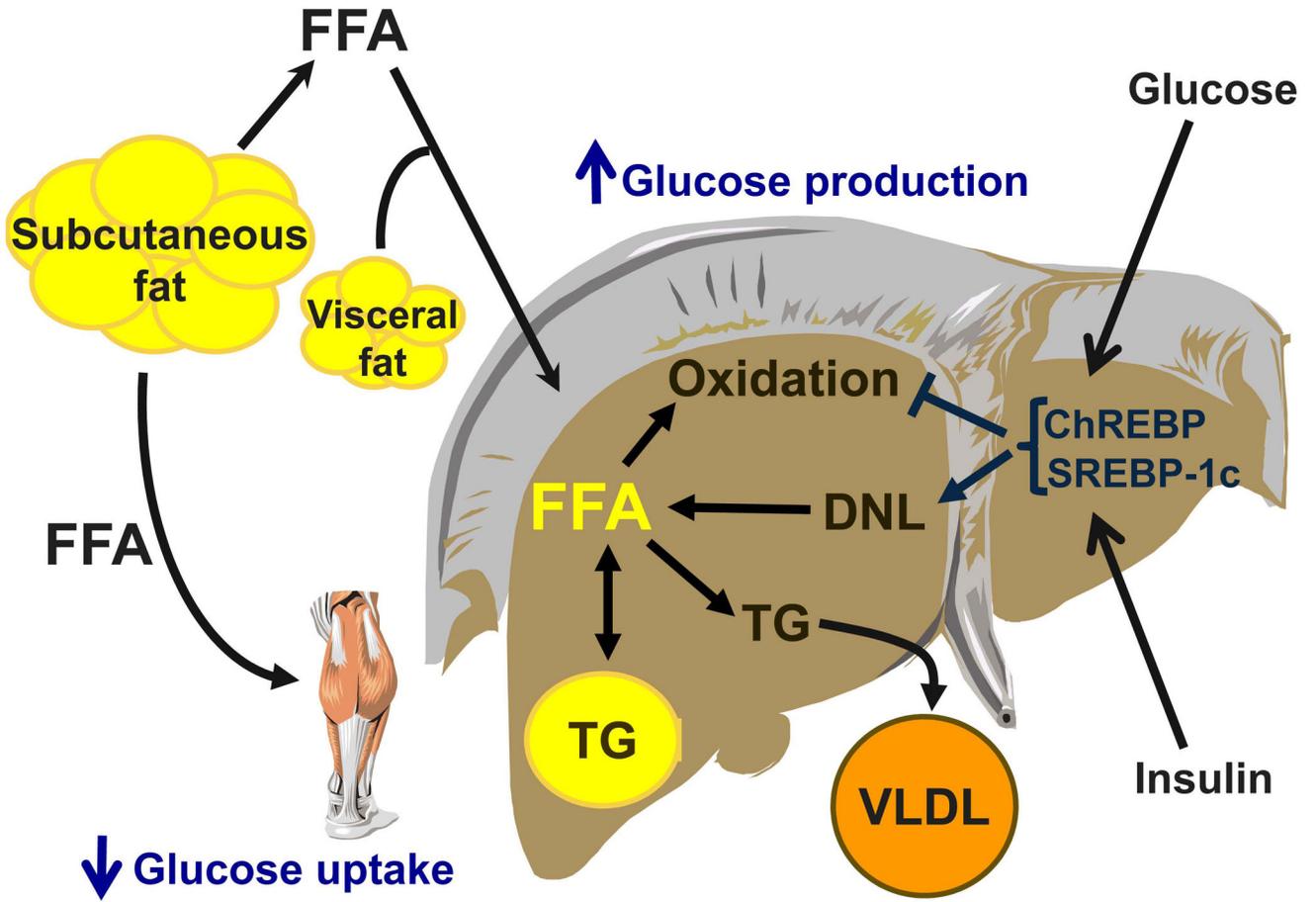


Figure 1. Physiological interrelationships among fatty acid metabolism, insulin resistance, dyslipidemia, and intrahepatic triglyceride content in nonalcoholic fatty liver disease (NAFLD). The rate of release of FFA from adipose tissue and delivery to the liver and skeletal muscle is increased in obese persons with NAFLD, which results in an increase in hepatic and muscle FFA uptake. In addition, intrahepatic *de novo* lipogenesis (DNL) of fatty acids is greater in subjects with NAFLD than in those with normal intrahepatic triglyceride (IHTG), which further contributes to the accumulation of intracellular fatty acids. The production and secretion of TG in VLDL is increased in subjects with NAFLD, which provides a mechanism for removing IHTG; however, the rate of secretion does not adequately compensate for the rate of TG production. Increased plasma glucose and insulin associated with NAFLD stimulate DNL and inhibit fatty acid oxidation, by affecting sterol regulatory element binding protein (SREBP-1c) and carbohydrate responsive element binding protein (ChREBP). These metabolic processes lead to an increase in intracellular fatty acids that are not oxidized or exported within VLDL-TG, and are esterified to TG and stored within lipid droplets. Certain lipid intermediates of fatty acid metabolism can impair insulin signaling and cause tissue insulin resistance. Therefore, this scheme illustrates how alterations in fatty acid metabolism can lead to an accumulation of intrahepatic (and intramuscular) TG, stimulate VLDL-TG secretion with subsequent hypertriglyceridemia, and cause insulin resistance in the liver and skeletal muscle.

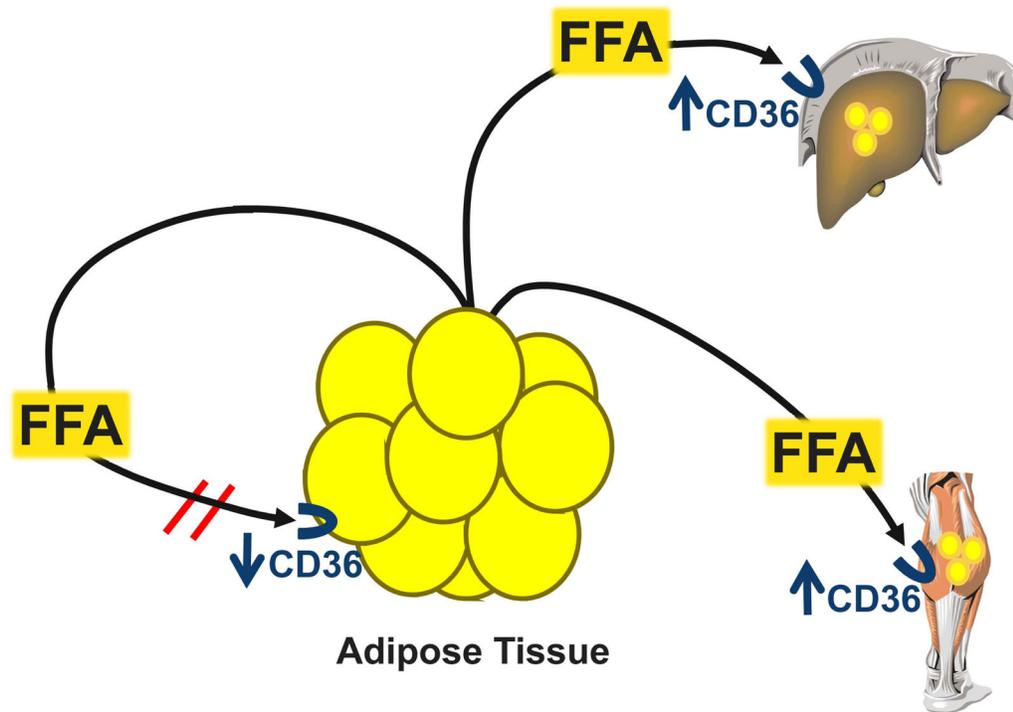


Figure 2.

Alterations in cellular fatty acid transport facilitate ectopic fat accumulation in the liver and skeletal muscle. The fatty acid translocase, CD36, regulates tissue FFA uptake from plasma. CD36 expression and protein content is decreased in adipose tissue, but increased in the liver and skeletal muscle of insulin-resistant animals and human subjects who have increased intrahepatic and intramyocellular triglyceride content. These findings suggest that alterations in tissue fatty acid transport could be involved in the pathogenesis of ectopic triglyceride accumulation by redirecting plasma fatty acid uptake from adipose tissue toward other tissues.

Hepatic VLDL-TG Secretion Rate

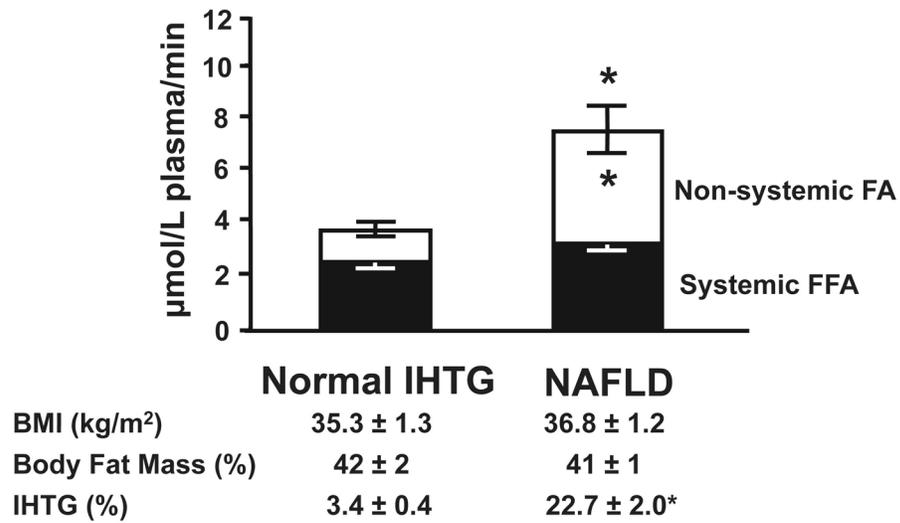


Figure 3.

Total VLDL-TG secretion rate (sum of grey and white bars) in subjects with normal and increased (nonalcoholic fatty liver disease [NAFLD]) intrahepatic triglyceride (IHTG) content, who were matched on BMI and percent body fat. White bars represent fatty acids in VLDL-TG that originated from systemic plasma free fatty acids, presumably derived primarily from lipolysis of subcutaneous fat, whereas black bars represent fatty acids in VLDL-TG that originated from non-systemic fatty acids, presumably derived primarily from lipolysis of intrahepatic and visceral fat and *de novo* lipogenesis. *Value significantly different from corresponding value in the Normal IHTG group, $P < 0.05$. (Adapted from: Fabbrini E, Mohammed BS, Magkos F, Korenblat KM, Patterson BW, Klein S. Alterations in adipose tissue and hepatic lipid kinetics in obese men and women with nonalcoholic fatty liver disease. *Gastroenterology* 2008;134:424–431).

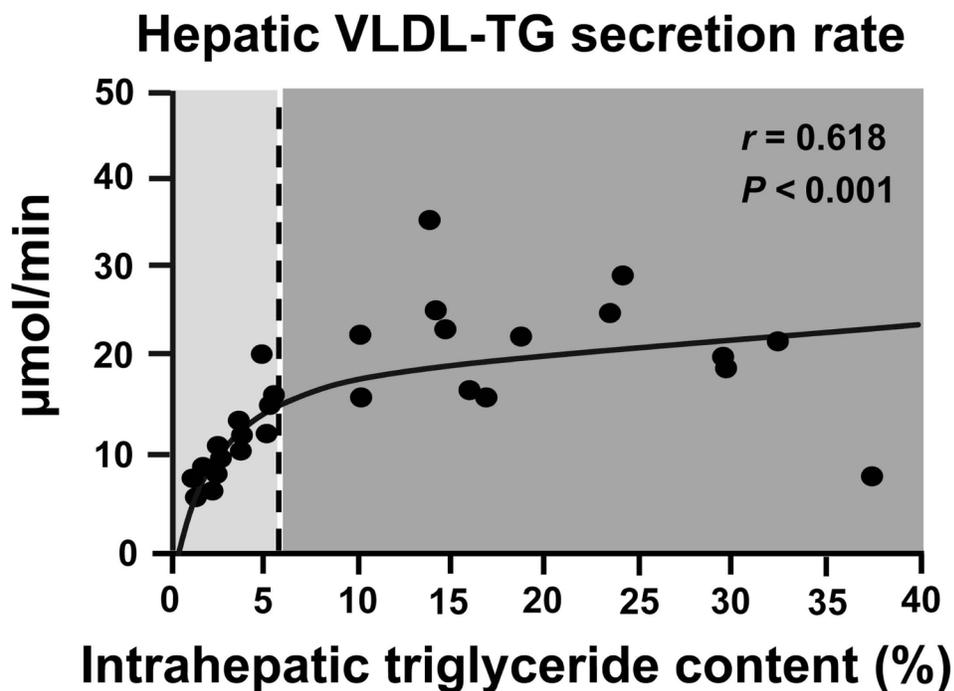


Figure 4. Relationship between VLDL-TG secretion rate and intrahepatic triglyceride content (IHTG) in subjects with normal IHTG (triglyceride content <5.6% of liver volume) and nonalcoholic fatty liver disease (NAFLD) (triglyceride content >10% of liver volume). (Adapted from: Fabbrini E, Mohammed BS, Magkos F, Korenblat KM, Patterson BW, Klein S. Alterations in adipose tissue and hepatic lipid kinetics in obese men and women with nonalcoholic fatty liver disease. *Gastroenterology* 2008;134:424–431).

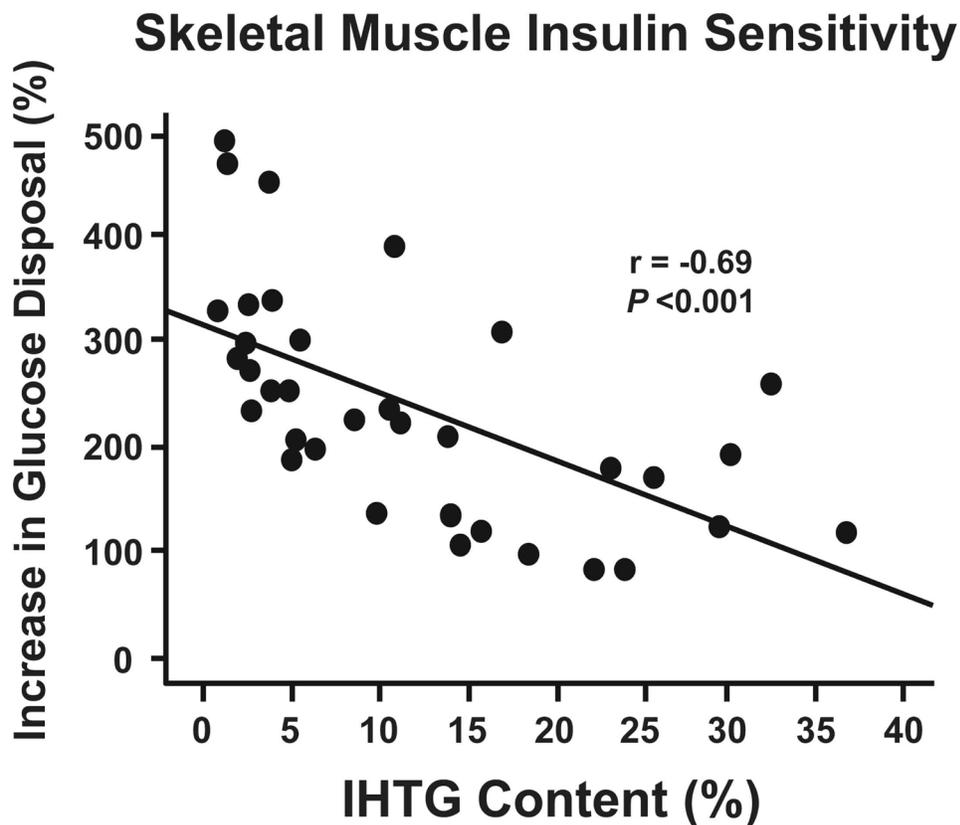


Figure 5. Relationship between intrahepatic triglyceride (IHTG) content and skeletal muscle insulin sensitivity, defined as the percent increase in the rate of glucose disposal in response to insulin infusion during a hyperinsulinemic-euglycemic clamp procedure. (Adapted from: Korenblat KM, Fabbrini E, Mohammed BS, Klein S. Liver, Muscle, and Adipose Tissue Insulin Action Is Directly Related to Intrahepatic Triglyceride Content in Obese Subjects. *Gastroenterology* 2008).

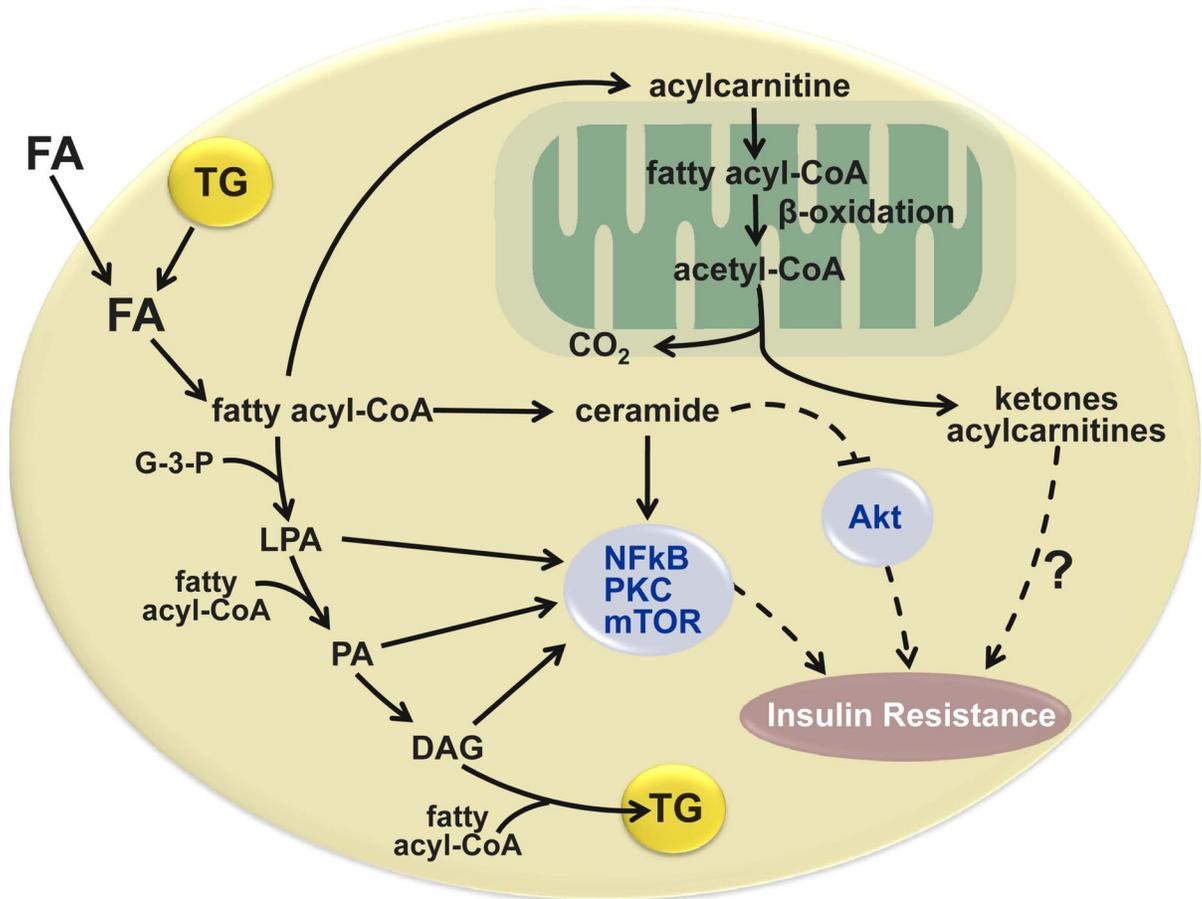


Figure 6.

Potential cellular mechanisms responsible for the relationship between fatty acid metabolism and insulin resistance in the liver and skeletal muscle. Obese persons with nonalcoholic fatty liver disease have increased rates of adipose tissue lipolysis and fatty acid (FA) release into plasma and increased intrahepatic and intramyocellular triglyceride (TG) content. Intracellular FA delivered from plasma or derived from lipolysis of intracellular triglyceride (TG) can be transported to the mitochondria for oxidation, esterified to TG or partially metabolized to several lipid intermediates, long chain fatty acyl-CoA, ceramide, lysophosphatidic acid (LPA), and phosphatidic acid (PA), and diacylglycerol (DAG). These lipid intermediates can interfere with insulin signaling by activating protein kinase C (PKC), mammalian target of rapamycin (mTOR), and nuclear factor kinase B (NFκB), and inhibiting Akt (also known as protein kinase B). The oxidation of intracellular FAs involve the conversion of long-chain fatty acyl-CoAs to acylcarnitines, which enter the mitochondria, and are progressively shortened by β-oxidation, which produces acetyl-CoA that can enter the tricarboxylic acid cycle for complete oxidation. The incomplete oxidation of fatty acyl-CoA generates ketone bodies and acylcarnitines, which might also have adverse effects on insulin action. (Adapted from: Schenk S, Saberi M, Olefsky JM. Insulin sensitivity: modulation by nutrients and inflammation. *J Clin Invest* 2008;118:2992–3002, and Nagle CA, Klett EL, Coleman RA. Hepatic triacylglycerol accumulation and insulin resistance. *J Lipid Res* 2009;50 Suppl:S74–79).