

Spermidine promotes mating and fertilization efficiency in model organisms

Maria Anna Bauer,¹ Didac Carmona-Gutiérrez,¹ Christoph Ruckenstuhl,¹ Angela Reisenbichler,¹ Evgenia V. Megalou,² Tobias Eisenberg,¹ Christoph Magnes,^{3,4} Helmut Jungwirth,¹ Frank M. Sinner,^{3,4} Thomas R. Pieber,^{3,4} Kai-Uwe Fröhlich,¹ Guido Kroemer,^{5,6,7,8,9,*} Nektarios Tavernarakis² and Frank Madeo^{1,*}

¹Institute of Molecular Biosciences; Karl-Franzens University of Graz; Graz, Austria; ²Institute of Molecular Biology and Biotechnology; Foundation for Research and Technology-Hellas; Heraklion, Greece; ³Institute of Medical Technologies and Health Management; Joanneum Research; Graz, Austria; ⁴Department of Internal Medicine; Division of Diabetes and Metabolism; Medical University of Graz; Graz, Austria; ⁵INSERM; U848 and Institut Gustave Roussy; Villejuif, France; ⁶Metabolomics Platform; Institut Gustave Roussy; Villejuif, France; ⁷Centre de Recherche des Cordeliers; Paris, France; ⁸Pôle de Biologie; Hôpital Européen Georges Pompidou; Assistance Publique-Hôpitaux de Paris (AP-HP); Paris, France; ⁹Université Paris Descartes/Paris 5; Sorbonne Paris Cité; Paris, France

Keywords: *Caenorhabditis elegans*, spermidine, mating, fertilization, *Saccharomyces cerevisiae*, shmoo, autophagy, sexual reproduction

Spermidine is a naturally occurring polyamine involved in multiple biological processes, including DNA metabolism, autophagy and aging. Like other polyamines, spermidine is also indispensable for successful reproduction at several stages. However, a direct influence on the actual fertilization process, i.e., the fusion of an oocyte with a spermatocyte, remains uncertain. To explore this possibility, we established the mating process in the yeast *Saccharomyces cerevisiae* as a model for fertilization in higher eukaryotes. During human fertilization, the sperm capacitates and the acrosome reaction is necessary for penetration of the oocyte. Similarly, sexually active yeasts form a protrusion called “shmoo” as a prerequisite for mating. In this study, we demonstrate that pheromone-induced shmoo formation requires spermidine. In addition, we show that spermidine is essential for mating in yeast as well as for egg fertilization in the nematode *Caenorhabditis elegans*. In both cases, this occurs independently from autophagy. In synthesis, we identify spermidine as an important mating component in unicellular and multicellular model organisms, supporting an unprecedented evolutionary conservation of the mechanisms governing fertilization-related cellular fusion.

Introduction

The alarming decrease in fertility rates across industrialized countries is often interpreted to be rooted in a consciously decided decline of the desired number of children. This, in turn, is often thought to be caused by a change in the social and behavioral lifestyle. However, it seems premature to rely only on a sociopsychological explanation for this phenomenon, because prominent lifestyle changes as well as exposure to environmental hazards might have led to endocrine disruption as well.¹ Therefore, thorough understanding of the basic mechanisms and molecular players involved in mammalian reproduction is pivotal to tackle the ever-increasing problems related to human infertility.

Polyamines are known to be indispensable for successful mammalian reproduction at several stages, including spermatogenesis, oogenesis, embryogenesis, implantation or placentation.² Interestingly, some correlative data indicate a possible involvement in the actual fertilization process. For instance, the seminal plasma of infertile men contains markedly lower levels of the polyamines spermine and spermidine compared to normospermic controls.^{3,4} Also, the fertilization efficiency gradually declines with advanced paternal age, paralleling the decrease in

intracellular polyamine concentrations.^{5,6} Declining fertilization efficiency may be associated with reduced semen volume, sperm concentration as well as polyamine-dependent sperm motility and morphology.⁵ However, it remains to be elucidated if polyamines also determine the fusion efficiency of oocytes and spermatozoa, which would be crucial for the understanding and improvement of both in vivo and in vitro fertilization.

The budding yeast *Saccharomyces cerevisiae* has blossomed into a major, genetically tractable model organism for the analysis of basic molecular processes like the cell cycle, autophagy and protein-protein interactions.^{7,8} In addition, the last two decades have established yeast as a model for the study of cancer,^{9,10} aging^{11–13} and neurodegenerative disorders.^{14–16}

During the yeast mating process, two haploid cells of opposed mating types (“*MAT α* ” and “*MAT α* ”) combine to form a single diploid cell. This event is reminiscent of the human fertilization process, in which the fusion of two haploid cells, the spermatocyte and the oocyte, results in a diploid zygote. Here, we establish the budding yeast *S. cerevisiae* as a new model organism for the analysis of fertilization mechanisms in higher eukaryotes. Moreover, we unveil a conserved and specific role of spermidine in the positive regulation of this process.

*Correspondence to: Guido Kroemer and Frank Madeo; Email: kroemer@orange.fr and frank.madeo@uni-graz.at
Submitted: 10/11/12; Revised: 12/07/12; Accepted: 12/10/12
<http://dx.doi.org/10.4161/cc.23199>

Results

Disruption of spermidine production entails a severe mating defect in yeast. Polyamines are known to be indispensable for successful mammalian reproduction and have been implicated in both prenatal and postnatal processes, such as embryogenesis, placentation and lactation.² Interestingly, polyamines have been suggested to be involved in the modulation of membrane fusion events.¹⁷ However, their impact on the actual fertilization process remains undetermined, perhaps because of the technical difficulty of mechanistically exploring the fusion of spermatocytes and oocytes.¹⁸ To develop a method to model this process, we investigated sexual reproduction in the baker's yeast *S. cerevisiae*, during which two haploid cells of opposed gender or mating types ("MAT α " and "MAT α ") combine to generate one diploid yeast cell. To specifically explore the contribution of the polyamine spermidine, yeast mating was monitored between "MAT α " wild-type cells and "MAT α " deletion mutants lacking *SPE2* (Δ *spe2*), the gene coding for S-adenosyl-methionine decarboxylase, which is essential for spermidine biosynthesis. This disruption of spermidine production in one of the mating partners drastically reduced the overall mating efficiency as compared to the control mating scenario, involving "MAT α " and "MAT α " wild-type cells (Fig. 1A). Preceding addition of exogenous spermidine to cultures of "MAT α " Δ *spe2* cells restored the mating efficiency to control levels (Fig. 1A). In conclusion, spermidine is essential for efficient mating in *S. cerevisiae*.

Pheromone-induced shmoo formation depends on spermidine. An indispensable step during human fertilization is spermatocyte capacitation and the acrosome reaction, which is necessary for penetration of the egg by the spermatocyte.¹⁹⁻²¹ Similarly, sexually reproducing yeast cells must form a nodule called "shmoo" for mating.^{22,23} The shmoo is produced upon sensing the mating pheromone of the opposite sex, which is released by and signals the availability of suitable mating partners.^{23,24} Thus, to further characterize the impact of spermidine on yeast mating, we quantified shmoo formation upon external administration of 1 μ M α -pheromone to wild-type and Δ *spe2* "MAT α " cells. As expected, α -pheromone induced pronounced shmoo formation in viable wild-type cells as quantified by microscopic analysis (Fig. 1B and C). In contrast, pheromone-treated viable Δ *spe2* cells only occasionally developed shmoos (Fig. 1B and C). Again, external supplementation of 100 μ M spermidine reverted the deficient shmoo formation in *spe2* disruptants to wild-type control levels (Fig. 1B and C). Thus, spermidine is indispensable for shmoo formation, which, in turn, is required for successful mating in yeast. Of note, intracellular levels of spermidine as well as those of its precursor putrescine are significantly elevated upon treatment with mating pheromone (Fig. 1D).

Polyamine-mediated mating control in yeast is spermidine-specific. To evaluate the specificity of spermidine in accounting for the observed positive effect on yeast sexual reproduction, we performed mating experiments with cells lacking spermine synthase (*Spe4p*). This enzyme catalyzes the generation of the polyamine spermine from spermidine in the polyamine biosynthetic pathway. Intriguingly, deletion of *SPE4* did not compromise

mating efficiency, which showed similar levels as the wild-type control (Fig. 1E). Accordingly, Δ *spe4* cells displayed no defect in shmoo formation (Fig. 1F). Altogether, these results point towards a specific role for spermidine (rather than spermine) in the promotion of yeast mating.

Spermidine determines yeast mating in an autophagy-independent manner. We next asked whether the significance of spermidine during mating might involve autophagy, a process of cellular self-digestion that is potentially induced by spermidine.²⁵⁻³⁰ Deletion of the autophagy-related gene *ATG7*, which is essential for autophagic execution in *S. cerevisiae*, did not influence mating efficiency (Fig. 1G). Furthermore, the mating efficiency of Δ *atg7* yeast cells additionally deleted in *SPE2* could still be restored to wild-type levels via external spermidine supply (Fig. 1H). These data suggest that spermidine-controlled mating in yeast does not depend on autophagy.

Spermidine modulates fertilization in the nematode *Caenorhabditis elegans*. In order to establish whether our results might be relevant to fertilization in higher eukaryotes, we explored the impact of spermidine on fertilization in *C. elegans*. Disruption of spermidine biosynthesis through deletion of spermidine synthase (SPDS-1) led to a significant reduction of the total number of fertilized eggs laid by the worm (Fig. 2A). As in yeast, this deficiency could be largely prevented by external supply of spermidine to the culture medium (Fig. 2A). This spermidine-mediated fertilization control also seems to be autophagy-independent in worms, since RNAi-driven knockdown of *bec-1*, a gene encoding a cardinal autophagic regulator in *C. elegans*, did not influence egg fertilization rates (Fig. 2B). Thus, spermidine modulates fertilization in worms in a manner akin to yeast, suggesting that yeast mating constitutes a bona fide model for the fertilization process of oocytes by spermatocytes.

Discussion

Here, we demonstrate that sexually reproducing yeast may constitute a valuable model for studying basic mechanisms that govern the fertilization process in higher eukaryotes. Specifically, we identified the polyamine spermidine to be indispensable for successful mating in *S. cerevisiae*. Moreover, we observed that spermidine is equally crucial for effective fertilization in the nematode *C. elegans*. In both cases, the mechanisms underlying this pro-fertilizing effect are autophagy-independent. Altogether, we propose that, as it occurs in the sexual reproductive variant of baker's yeast, the fusion of two haploid yeast cells forming a diploid cell mirrors the fertilization process of higher eukaryotes, where two haploid gametes conjugate to generate a diploid zygote.

Our hypothesis arguing for a conserved regulation of such haploid fusion events during fertilization is supported by recent evidence suggesting that mating-associated yeast cell fusion may represent a useful model system for studying the mechanisms of cytogamy and karyogamy.³¹⁻³³ Intriguingly, a recent report has established structural similarities between proteins involved in sperm-egg recognition in mammals and fusion of haploid yeast cells.³⁴ Our results with yeast and nematodes now reinforce the

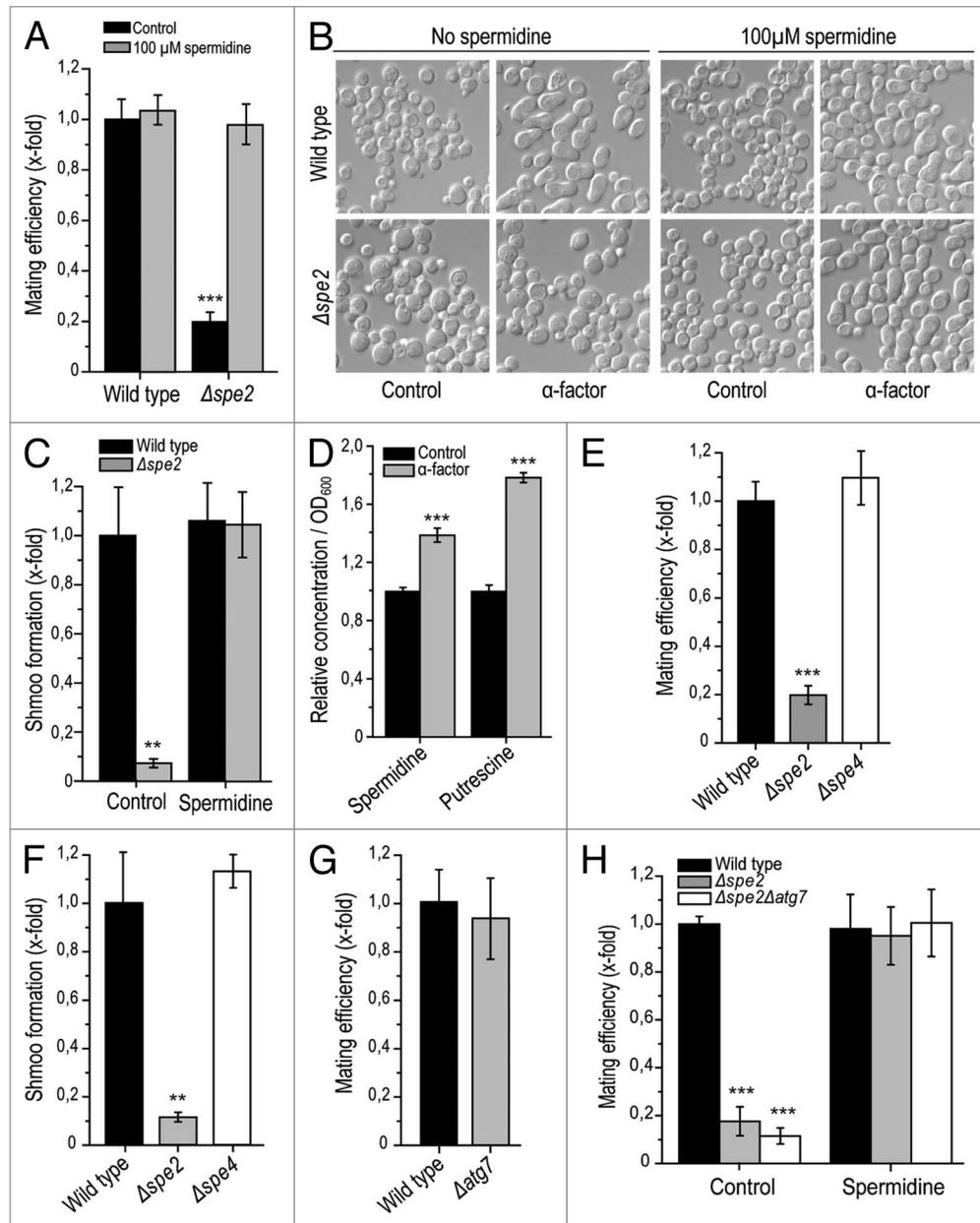


Figure 1. Yeast mating efficiency and shmoo formation depends on the availability of spermidine but is autophagy-independent. **(A)** Mating efficiency of wild-type and $\Delta spe2$ cells with or without external administration of 100 μM spermidine as determined via clonogenic mating assay. Mating efficiency of wild-type control was set at 1. Data represent mean \pm S.E.M. ($n = 3$; $***p < 0.001$). **(B)** Representative sample pictures of wild-type and $\Delta spe2$ cells (*MATa*) after treatment with 1 μM α -pheromone for 3 h with or without external administration of 100 μM spermidine. **(C)** Microscopic quantification of cells treated as described in **(B)** and displaying a shmoo. Only viable cells were considered, and at least 3,000 cells per strain and experiment were manually counted. The portion of cells displaying a shmoo was normalized to that of the untreated wild type control, which was set at 1. Data represent means \pm S.E.M. ($n = 4$; $**p < 0.01$). **(D)** Relative intracellular concentration of spermidine and its precursor putrescine per OD_{600} of wild-type cells (mating type a) either treated with 1 μM α -pheromone or left untreated (control), as determined using LC/MS/MS and normalized to the intracellular concentrations of the untreated control. Data represent means \pm S.E.M. ($n = 5$; $***p < 0.001$). **(E)** Mating efficiency of wild-type, $\Delta spe2$ and $\Delta spe4$ strains. Data were determined via clonogenic mating assay and represent means \pm S.E.M. ($n = 3$; $***p < 0.001$). **(F)** Shmoo-formation in wild-type, $\Delta spe2$ and $\Delta spe4$ cells (mating type a) after treatment with 1 μM α -pheromone for 3 h (compare to **Fig. 1B**). Data represent means \pm S.E.M. ($n = 4$; $**p < 0.01$). **(G)** Mating efficiency of wild-type and $\Delta atg7$ cells. Data were determined via clonogenic mating assay and represent means \pm S.E.M. ($n = 3$; $***p < 0.001$). Mating efficiency of wild-type control was set at 1. **(H)** Mating efficiency of wild-type, $\Delta spe2$ and $\Delta spe2\Delta atg7$ strains with or without external administration of 100 μM spermidine. Data was determined via clonogenic mating assays and represent means \pm S.E.M. ($n = 3$; $***p < 0.001$). Mating efficiency of wild-type control was set at 1.

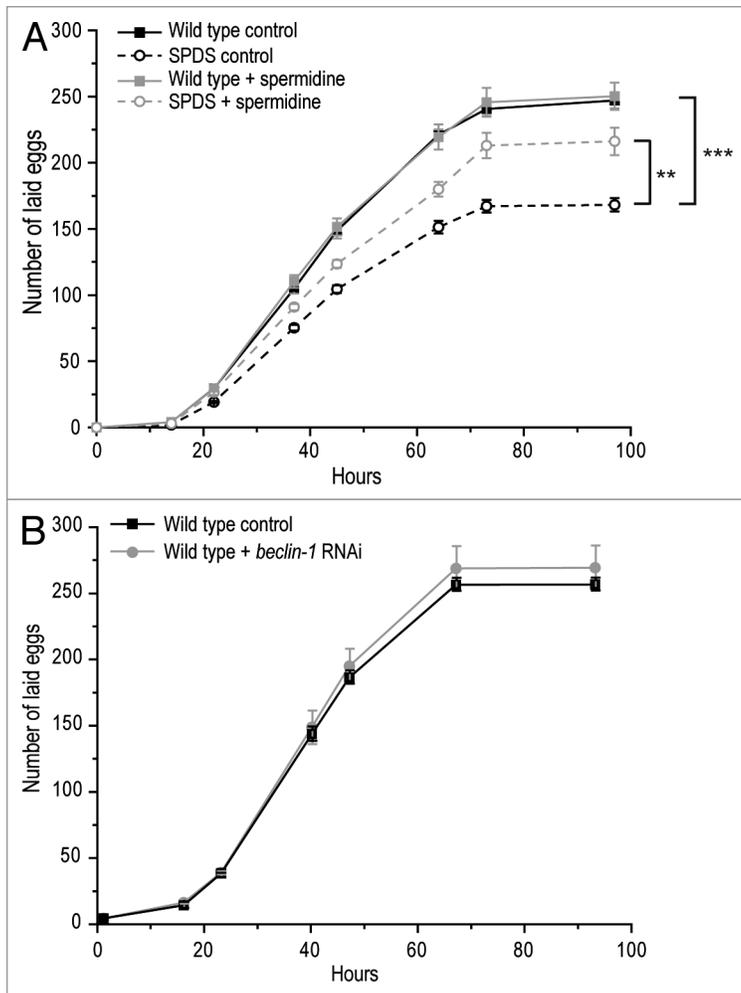


Figure 2. Fertilization in *C. elegans* is modulated by spermidine. (A) Total number of fertilized eggs laid by wild-type and spermidine synthase (SPDS-1)-deficient worms, fed with or without 50 μ M spermidine-supplemented food. Data represent mean \pm S.E.M. (n = 5; **p < 0.01, ***p < 0.001). (B) Total number of fertilized eggs laid by wild-type and *bec-1*-deficient worms. Data represent mean \pm S.E.M. (n = 5).

concept that the mechanisms accounting for the fusion among gametes are much more conserved than anticipated.

This study determines spermidine as a crucial regulator of mating/fertilization. Interestingly, spermidine levels have been shown to continuously decrease during yeast chronological aging^{11,30} and with increasing human age.^{5,6} This is, in turn, accompanied by decreased fertilization efficiency.⁵ We show that the intracellular concentrations of spermidine and its precursor putrescine are significantly elevated upon mating induction via treatment with the corresponding mating pheromone. However, such increase could also originate in other effects elicited through treatment with mating pheromone, which, e.g., also induces cell cycle arrest.²³ In fact, high spermidine concentrations have been previously linked to repression of cell division.³⁵⁻³⁷ Thus, a causal relationship between the herein identified elevation of intracellular spermidine levels and the induction of mating will need to be further elucidated in future studies.

Spermidine is required for shmooing, which is a prerequisite for successful mating and represents the polarized growth of the yeast cell towards the mating partner. Shmoo formation in yeast on the one hand and capacitation as well as the acrosome reaction in spermatocytes of higher eukaryotes on the other hand exhibit strong similarities: both manifest prior to the fusion of two haploid cells; both are indispensable for fusion efficiency, and both are morphologically similar to the extent that they involve protruding areas of the mating cells. In our hands, shmooing depends specifically on spermidine but does not seem to involve the generation of the closely related polyamine spermine. In contrast, recent *in vitro* data do not suggest such specificity for the acrosome reaction. In the ram spermatozoid, externally applied spermine does localize to the acrosome,³⁸ and low concentrations of spermine can stimulate the acrosome of bovine spermatozoa.³⁹ However, these *in vitro* studies did not compare spermine to spermidine and did not consider the possibility that spermine oxidase can convert spermine into spermidine. As a result, it is conceivable that such supposedly spermine-related effects might in fact be connected to the generation of effective spermidine levels. This will need clarification in future studies. Our conclusion that spermidine is specific for shmooing and mating control, however, is based on the observation that deletion of spermine synthase does not affect mating efficiency or shmooing. This does not exclude the possibility that spermine generation may be alternatively achieved through yet to be identified biosynthetic polyamine routes.⁴⁰

Spermidine potently induces autophagy in a range of model organisms including mice, worms, flies and yeast.³⁰ This had raised the question whether the positive effects on the sexual reproduction of yeast and nematodes might rely on its autophagy-promoting potential. Our data demonstrate that mating/fertilization is normal in yeast cells and nematodes lacking proteins that are essential for autophagy, like Atg7p in *S. cerevisiae* or beclin-1 in *C. elegans*. Moreover, external administration of spermidine restored the mating efficiency of spermidine biosynthesis-deficient yeast cells to wild-type levels, even in the context of deficient autophagy. Thus, autophagy is not involved in the positive effects of spermidine on mating/fertilization. Consequently, alternative pathways must be responsible for the herein observed effects and involve a different molecular target of spermidine. Possible candidates include calcium channels, which are regulated by spermidine⁴¹ and required for effective yeast mating as well as for capacitation and the acrosome reaction in humans.^{42,43} It is thus conceivable that a distortion of spermidine-tuned calcium flux regulation deriving from unbalanced spermidine levels might result in adverse mating/fertilization effects. Whether spermidine affects calcium channels that are crucial for haploid cell fusion remains to be explored.

Altogether, this study provides first *in vivo* evidence that the mechanisms underlying mating-related cell fusion of haploid yeast cells might parallel fertilization of higher eukaryotes. It

is tempting to speculate that yeast cell fusion might also model somatic fusion events as they occur in mammals during the formation of the syncytiotrophoblast, skeleton muscle fibers and osteoclasts.⁴⁴⁻⁴⁷ In fact, polyamines have been previously linked to the modulation of membrane fusion and aggregation.¹⁷ Yeast's enormous technical advantages, such as its inexpensiveness, convenient handling, rapid growth and uncomplicated DNA modification, have defined its value as a model for many areas of molecular biology. Thus, sexual reproduction and specifically fertilization might join as a further area to be explored in *S. cerevisiae* and thereby even more intimately link yeast and human.

Materials and Methods

Reagents. Spermidine (Sigma) was dissolved in sterilized water to a stock solution concentration of 100 mM and stored as single use aliquots at -20°C. Propidium iodide (PI, Sigma) was stored at -20°C as a stock water solution of 100 µg/ml.

Yeast strains and growth conditions. Yeast experiments were carried out in *Saccharomyces cerevisiae* BY4741 (*MATa his3Δ1 leu2Δ0 met15Δ0 ura3Δ0*) and the respective null mutants Δ *spe2* and Δ *spe4* as well as in BY4742 (*MATα his3Δ1 leu2Δ0 lys2Δ0 ura3Δ0*), which were all obtained from Euroscarf. All strains were grown on liquid SC medium containing 0.17% yeast nitrogen base (BD Diagnostics), 0.5% (NH₄)₂SO₄, 30mg/l of all amino acids (except 80mg/l histidine, 60 mg/l lysine and 200 mg/l leucine), 30 mg/l adenine, 320mg/l uracil and 2% glucose. For experiments, yeast strains from 25% glycerol stocks at -80°C were streaked on YPD plates and stored at 4°C for up to 10 d, from which overnight cultures were inoculated. Incubation of all liquid cultures was performed with 145 rpm at 28°C. Cell densities were determined using an automated cell counter (CASY1, Roche Innovatis).

Yeast-mating efficiency. To determine mating efficiency, BY4741 cultures were first inoculated into 100 ml quadruple-indented flasks at 10 ml cell culture each from fresh overnight cultures to a cell density of 1 × 10⁶ cells/ml and, depending on the experiment, supplemented or not with 100 µM spermidine. After 20–24 h of growth, 2 × 10⁶ cells of each flask were transferred to a new non-indented flask containing 10 ml SCD medium and mixed with 1 × 10⁷ cells BY4742 of a freshly made overnight culture. Of note, comparable viability of all cultures was verified by clonogenic survival plating on YPD as described before.^{12,13} Three hours after mixing, an aliquot was plated on two different agar plates, respectively: (1) 2,000 cells were plated on agar plates containing 0.17% yeast nitrogen base, 0.5% (NH₄)₂SO₄, 320 mg/l uracil, 200 mg/l leucine, 80 mg/l histidine and 30 mg/l methionine to determine the amount of both viable BY4741 (addition of methionine, see genotype) and diploid cells (arisen from successful mating of one BY4741 and one BY4742 cell) and (2) 2,000 cells were plated on agar plates containing 0.17% yeast nitrogen, 0.5% (NH₄)₂SO₄, 320 mg/l uracil, 200 mg/l leucine and 80 mg/l histidine to assess the number of viable diploid cells only. All plates were incubated at 28°C for 2 d, and resulting colony-forming units (CFU) were counted. The mating efficiency was determined as the number of diploid colonies per number

of colonies corresponding to BY4741 cells, i.e. CFU of plate 2/CFU of plate 1.

Yeast shmooing efficiency. For shmooing efficiency experiments, cultures were inoculated from fresh overnight cultures to a density with an absorbance of 0.1 as determined using a photometer; this absorbance correlated to approximately 1 × 10⁶ cells/ml. Depending on the experiment, the culture was supplemented or not with 100 µM spermidine and let grow for 20–24 h. Thereafter, the cultures were inoculated to a density with an absorbance of 0.3 in new non-indented flasks containing 10 ml fresh spermidine-free SCD medium that included 50 µM Na-succinate and was adjusted to pH 4.3 to retard pheromone proteolysis during subsequent pheromone treatment.⁴⁸ Cultures were then treated with 1 µM α-pheromone. After 3 h, cultures were centrifuged (3 min., 3,000 rpm) and diluted in PBS buffer containing propidium iodide (PI) at a concentration of 0.1 µg/ml. PI as an intercalating, fluorescent agent was used to evaluate cell viability and exclude inviable cells from subsequent microscopic quantification. Cells were incubated for 10 min and used for microscopy.

Microscopy. Microscopy was performed with a Zeiss Axioskop microscope using a Zeiss Plan-Neofluar objective lense with 40× magnification and 1.0 numerical aperture in oil (using Zeiss Immersol) at room temperature. Imaging medium was PBS. Fluorochrome was propidium iodide (PI). Fluorescence microscopic sample images were taken with a Diagnostic Instruments camera (Model: SPOT 9.0 Monochrome-6), acquired using the Metamorph software (version 6.2r4, Molecular Devices) and processed with IrfanView (version 3.97, Wiener Neustadt) software. Specifically, picture processing involved coloring of PI pictures, creation of corresponding DIC/PI overlays (used to determine inviable cells to exclude from quantification) with Metamorph and cropping of representative areas with IrfanView.

***C. elegans* strains and growth conditions.** Experiments were carried out using the *Caenorhabditis elegans* N2 wild-type strain. The following mutant strain was used: RB2481 *spds-1(ok3421) II* (referred to in the text as SPDS-1). We followed standard procedures for *C. elegans* strain maintenance.⁴⁹ Nematode rearing temperature was kept at 20°C.

***C. elegans* fertility determination.** Preparation of *Escherichia coli* (OP50) plates for worm feeding was performed as previously described.⁴⁹ Briefly, bacteria on seeded 7 ml NGM plates were killed by exposure to UV irradiation for 10 min (0.5 J/cm²) using a UV crosslinker (BIO-LINK-BLX-E365, Vilber Lourmat). Spermidine was prepared by dilutions in 50 µl sterilized water and applied to the top of the OP50 lawn on the agar medium (7 ml NGM plates). Identical solutions of drug-free water were used for the control plates. Plates were then allowed to dry overnight. N2 and SPDS-1 worms previously grown to L4 stage were transferred to fresh plates with or without spermidine. After 2 d, the progeny was transferred to corresponding fresh plates. Upon reaching L4 stage, single worms were transferred to plates without spermidine. One worm was transferred per plate. The day of this transfer was defined as t = 0. After t = 0, worms were transferred to fresh plates without spermidine every day in order to facilitate egg counting for each

day. The number of eggs laid by each single worm was monitored until egg laying ceased.

For fertility experiments involving RNAi-suppression of *bec-1*, a previously described^{50,51} plasmid, which directs the synthesis of a dsRNA corresponding to the *bec-1* gene in *E. coli*, was used. HT115(DE3) bacteria were transformed with either this *bec-1* RNAi plasmid or the pL4440 control vector. Before seeding, 2 mM IPTG were added to the bacterial culture and incubated for 15 min. The plates seeded with RNAi or pL4440 vector-containing bacteria were subsequently used as described above for the OP50-seeded plates in the fertility assay experiments. Each fertility assay was conducted at least twice, and figures represent typical assays.

Extraction and measurements of polyamines. Extraction and measurements of polyamines were essentially performed as described in Eisenberg et al.³⁰ For acid extraction of polyamines from yeast cells, culture equivalents of 2 OD₆₀₀ were washed with ddH₂O, resuspended in 1,000 μ l ice-cold 5% TCA with isotopically labelled internal standards for spermidine (¹³C₄-spermidine) and putrescine (²H₈-putrescine) and incubated on ice for 1h, with vortexing every 15min. [Calibration standards were prepared by spiking extraction buffer with specific concentrations of spermidine and putrescine and internal standards. Polyamines were derivatized to carbamyl-derivatives by adding 125 μ l of 1 M carbonate buffer (pH 9), 800 μ l water and 20 μ l of isobutyl chloroformate to 100 μ l of sample or calibration standard containing internal standards]. Supernatants were neutralized with 100 μ l of 2M K₂HPO₄ and stored at -80°C until polyamine measurements.

Polyamines were determined using LC/MS/MS. All analyses were carried out on an Ultimate 3000 System (Dionex, LCPackings) coupled to a Quantum TSQ Ultra AM (Thermo Scientific) using an electrospray ion source. The system was controlled by Xcalibur Software 2.0. The stationary phase was a Kinetex 2.6 μ m C18 100A 50 mm \times 2.1 mm column (Phenomenex). Two hundred and fifty microliters of the derivatized samples were loaded on to an online-SPE column (Strata

X, Phenomenex) using eluent A (flow rate 1.5 ml min). After 2 min, online SPE was switched to the analytical column and the polyamines were eluted and separated on the analytical column within 4 min using isocratic conditions (80% eluent B, flow rate 250 μ l/min). Polyamines were detected in MRM mode using following transitions: Spermidine (m/z 446 \rightarrow 298V), putrescine (m/z 289 \rightarrow 115), ¹³C₄-spermidine (m/z 450 \rightarrow 302), ²H₈-putrescine (m/z 297 \rightarrow 123).

Statistical analyses. Error bars (\pm S.E.M.) are shown for independent experiments. In cases when experiments were performed in parallel, a common ONC for each strain was used and (1) fresh medium in separate flasks was inoculated directly from the corresponding ONCs (knockout studies). The number of independent data points (n) is indicated in the figure legends of the corresponding graphs. Statistical analyses were carried out using the Microsoft Office 2003 Excel software package (Microsoft Corporation). Mean values were compared using unpaired t-tests.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

We thank Ulrike Potocnik and Angela Pasparaki for assistance. We are grateful to the Austrian Science Fund FWF (Austria) for grants P23490-B12 and P24381-B20. N.T. acknowledges support by grants from the European Research Council (ERC) and the European Commission 7th Framework Programme. This work is supported by grants to G.K. from the Ligue Nationale contre le Cancer (Equipe labellisée), Agence Nationale pour la Recherche (ANR), AXA Chair for Longevity Research, European Commission (ArtForce, ChemoRes), Fondation pour la Recherche Médicale (FRM), Institut National du Cancer (INCa), Cancéropôle Ile-de-France, Fondation Bettencourt-Schueller, the LabEx Onco-Immunology and the Paris Alliance of Cancer Research Institutes. The authors declare no conflict of interest.

References

1. Phillips KP, Tanphaichitr N. Human exposure to endocrine disrupters and semen quality. *J Toxicol Environ Health B Crit Rev* 2008; 11:188-220; PMID:18368553; <http://dx.doi.org/10.1080/10937400701873472>
2. Lefèvre PL, Palin MF, Murphy BD. Polyamines on the reproductive landscape. *Endocr Rev* 2011; 32:694-712; PMID:21791568; <http://dx.doi.org/10.1210/er.2011-0012>
3. Calandra RS, Rulli SB, Frungieri MB, Suescun MO, González-Calvar SI. Polyamines in the male reproductive system. *Acta Physiol Pharmacol Ther Latinoam* 1996; 46:209-22; PMID:9222386
4. Shohat B, Maayan R, Singer R, Sagiv M, Kaufman H, Zukerman Z. Immunosuppressive activity and polyamine levels of seminal plasma in azo-spermic, oligospermic, and normospermic men. *Arch Androl* 1990; 24:41-50; PMID:2327812; <http://dx.doi.org/10.3109/01485019008986857>
5. Stewart AF, Kim ED. Fertility concerns for the aging male. *Urology* 2011; 78:496-9; PMID:21884897; <http://dx.doi.org/10.1016/j.urolgy.2011.06.010>
6. Scalabrino G, Ferioli ME. Polyamines in mammalian ageing: an oncological problem, too? A review. *Mech Ageing Dev* 1984; 26:149-64; PMID:6384679; [http://dx.doi.org/10.1016/0047-6374\(84\)90090-3](http://dx.doi.org/10.1016/0047-6374(84)90090-3)
7. Pertschy B, Saveanu C, Zisser G, Lebreton A, Tengg M, Jacquier A, et al. Cytoplasmic recycling of 60S preribosomal factors depends on the AAA protein Drg1. *Mol Cell Biol* 2007; 27:6581-92; PMID:17646390; <http://dx.doi.org/10.1128/MCB.00668-07>
8. Hohmann S. The Yeast Systems Biology Network: mating communities. *Curr Opin Biotechnol* 2005; 16:356-60; PMID:15961037; <http://dx.doi.org/10.1016/j.copbio.2005.04.006>
9. Carmona-Gutiérrez D, Eisenberg T, Büttner S, Meisinger C, Kroemer G, Madeo F. Apoptosis in yeast: triggers, pathways, subroutines. *Cell Death Differ* 2010; 17:763-73; PMID:20075938; <http://dx.doi.org/10.1038/cdd.2009.219>
10. Ruckenstein C, Büttner S, Carmona-Gutiérrez D, Eisenberg T, Kroemer G, Sigrist SJ, et al. The Warburg effect suppresses oxidative stress induced apoptosis in a yeast model for cancer. *PLoS One* 2009; 4:e4592; PMID:19240798; <http://dx.doi.org/10.1371/journal.pone.0004592>
11. Carmona-Gutiérrez D, Bauer MA, Ring J, Knauer H, Eisenberg T, Büttner S, et al. The propeptide of yeast cathepsin D inhibits programmed necrosis. *Cell Death Dis* 2011; 2:e161; PMID:21593793; <http://dx.doi.org/10.1038/cddis.2011.43>
12. Herker E, Jungwirth H, Lehmann KA, Maldener C, Fröhlich KU, Wissing S, et al. Chronological aging leads to apoptosis in yeast. *J Cell Biol* 2004; 164:501-7; PMID:14970189; <http://dx.doi.org/10.1083/jcb.200310014>
13. Madeo F, Herker E, Maldener C, Wissing S, Lächelt S, Herlan M, et al. A caspase-related protease regulates apoptosis in yeast. *Mol Cell* 2002; 9:911-7; PMID:11983181; [http://dx.doi.org/10.1016/S1097-2765\(02\)00501-4](http://dx.doi.org/10.1016/S1097-2765(02)00501-4)
14. Krobitsch S, Lindquist S. Aggregation of huntingtin in yeast varies with the length of the polyglutamine expansion and the expression of chaperone proteins. *Proc Natl Acad Sci USA* 2000; 97:1589-94; PMID:10677504; <http://dx.doi.org/10.1073/pnas.97.4.1589>
15. Büttner S, Bitto A, Ring J, Augsten M, Zabrocki P, Eisenberg T, et al. Functional mitochondria are required for alpha-synuclein toxicity in aging yeast. *J Biol Chem* 2008; 283:7554-60; PMID:18192273; <http://dx.doi.org/10.1074/jbc.M708477200>

16. Braun RJ, Büttner S, Ring J, Kroemer G, Madeo F. Nervous yeast: modeling neurotoxic cell death. *Trends Biochem Sci* 2010; 35:135-44; PMID:19926288; <http://dx.doi.org/10.1016/j.tibs.2009.10.005>
17. Schuber F, Hong K, Düzgünes N, Papahadjopoulos D. Polyamines as modulators of membrane fusion: aggregation and fusion of liposomes. *Biochemistry* 1983; 22:6134-40; PMID:10627119; <http://dx.doi.org/10.1021/bi00295a015>
18. Leahy T, Gadella BM. Sperm surface changes and physiological consequences induced by sperm handling and storage. *Reproduction* 2011; 142:759-78; PMID:21964828; <http://dx.doi.org/10.1530/REP-11-0310>
19. Yanagimachi R. In: Neill EKJ, ed. *The Physiology of Reproduction*. New York: Raven Press, 1988:135-85
20. Yanagimachi R. In: Biggers LMJD, ed. *Fertilization and embryonic development in vitro*. New York: Plenum Press, 1981:82-182
21. Russell L, Peterson R, Freund M. Direct evidence for formation of hybrid vesicles by fusion of plasma and outer acrosomal membranes during the acrosome reaction in boar spermatozoa. *J Exp Zool* 1979; 208:41-56; PMID:381567; <http://dx.doi.org/10.1002/jez.1402080106>
22. Madden K, Snyder M. Cell polarity and morphogenesis in budding yeast. *Annu Rev Microbiol* 1998; 52:687-744; PMID:9891811; <http://dx.doi.org/10.1146/annurev.micro.52.1.687>
23. Bardwell L. A walk-through of the yeast mating pheromone response pathway. *Peptides* 2004; 25:1465-76; PMID:15374648; <http://dx.doi.org/10.1016/j.peptides.2003.10.022>
24. Cross F, Hartwell LH, Jackson C, Konopka JB. Conjugation in *Saccharomyces cerevisiae*. *Annu Rev Cell Biol* 1988; 4:429-57; PMID:2848554; <http://dx.doi.org/10.1146/annurev.cb.04.110188.002241>
25. Madeo F, Tavernarakis N, Kroemer G. Can autophagy promote longevity? *Nat Cell Biol* 2010; 12:842-6; PMID:20811357; <http://dx.doi.org/10.1038/ncb0910-842>
26. Minois N, Carmona-Gutierrez D, Madeo F. Polyamines in aging and disease. *Aging (Albany NY)* 2011; 3:716-32; PMID:21869457
27. Morselli E, Mariño G, Bennetzen MV, Eisenberg T, Megalou E, Schroeder S, et al. Spermidine and resveratrol induce autophagy by distinct pathways converging on the acetylproteome. *J Cell Biol* 2011; 192:615-29; PMID:21339330; <http://dx.doi.org/10.1083/jcb.201008167>
28. Minois N, Carmona-Gutierrez D, Bauer MA, Rockenfeller P, Eisenberg T, Brandhorst S, et al. Spermidine promotes stress resistance in *Drosophila melanogaster* through autophagy-dependent and -independent pathways. *Cell Death Dis* 2012; 3:e401; PMID:23059820; <http://dx.doi.org/10.1038/cddis.2012.139>
29. Madeo F, Eisenberg T, Büttner S, Ruckenstein C, Kroemer G. Spermidine: a novel autophagy inducer and longevity elixir. *Autophagy* 2010; 6:160-2; PMID:20110777; <http://dx.doi.org/10.4161/auto.6.1.10600>
30. Eisenberg T, Knauer H, Schauer A, Büttner S, Ruckenstein C, Carmona-Gutierrez D, et al. Induction of autophagy by spermidine promotes longevity. *Nat Cell Biol* 2009; 11:1305-14; PMID:19801973; <http://dx.doi.org/10.1038/ncb1975>
31. Ydenberg CA, Rose MD. Yeast mating: a model system for studying cell and nuclear fusion. *Methods Mol Biol* 2008; 475:3-20; PMID:18979235; http://dx.doi.org/10.1007/978-1-59745-250-2_1
32. Ydenberg CA, Stein RA, Rose MD. Cdc42p and Fus2p act together late in yeast cell fusion. *Mol Biol Cell* 2012; 23:1208-18; PMID:22323294; <http://dx.doi.org/10.1091/mbc.E11-08-0723>
33. Sens KL, Zhang S, Jin P, Duan R, Zhang G, Luo F, et al. An invasive podosome-like structure promotes fusion pore formation during myoblast fusion. *J Cell Biol* 2010; 191:1013-27; PMID:21098115; <http://dx.doi.org/10.1083/jcb.201006006>
34. Swanson WJ, Aagaard JE, Vacquier VD, Monné M, Sadat Al Hosseini H, Jovine L. The molecular basis of sex: linking yeast to human. *Mol Biol Evol* 2011; 28:1963-6; PMID:21282709; <http://dx.doi.org/10.1093/molbev/msr026>
35. Deeb F, van der Weele CM, Wolniak SM. Spermidine is a morphogenetic determinant for cell fate specification in the male gametophyte of the water fern *Marsilea vestita*. *Plant Cell* 2010; 22:3678-91; PMID:21097708; <http://dx.doi.org/10.1105/tpc.109.073254>
36. Gömürçen AN, Mutlu F, Bozcuk S. Effects of polyamines (putrescine, spermidine and spermine) on root tip mitosis and chromosomes in *Allium cepa* L. *Cytologia (Tokyo)* 2005; 70:217-24; <http://dx.doi.org/10.1508/cytologia.70.217>
37. Osborne HB, Cormier P, Lorillon O, Maniey D, Bellé R. An appraisal of the developmental importance of polyamine changes in early *Xenopus* embryos. *Int J Dev Biol* 1993; 37:615-8; PMID:8180006
38. Rubinstein S, Breitbart H. Cellular localization of polyamines: cytochemical and ultrastructural methods providing new clues to polyamine function in ram spermatozoa. *Biol Cell* 1994; 81:177-83; PMID:7849608; [http://dx.doi.org/10.1016/S0248-4900\(94\)80008-1](http://dx.doi.org/10.1016/S0248-4900(94)80008-1)
39. Rubinstein S, Lax Y, Shalev Y, Breitbart H. Dual effect of spermine on acrosomal exocytosis in capacitated bovine spermatozoa. *Biochim Biophys Acta* 1995; 1266:196-200; PMID:7742386; [http://dx.doi.org/10.1016/0167-4889\(95\)00007-F](http://dx.doi.org/10.1016/0167-4889(95)00007-F)
40. Michael AJ. Exploring polyamine biosynthetic diversity through comparative and functional genomics. *Methods Mol Biol* 2011; 720:39-50; PMID:21318865; http://dx.doi.org/10.1007/978-1-61779-034-8_2
41. Gomez M, Hellstrand P. Endogenous polyamines modulate Ca²⁺ channel activity in guinea-pig intestinal smooth muscle. *Pflügers Arch* 1999; 438:445-51; PMID:10519136; <http://dx.doi.org/10.1007/s004240051060>
42. Iida H, Nakamura H, Ono T, Okumura MS, Anraku Y. MID1, a novel *Saccharomyces cerevisiae* gene encoding a plasma membrane protein, is required for Ca²⁺ influx and mating. *Mol Cell Biol* 1994; 14:8259-71; PMID:7526155
43. Orta G, Ferreira G, José O, Treviño CL, Beltrán C, Darzón A. Human spermatozoa possess a calcium-dependent chloride channel that may participate in the acrosomal reaction. *J Physiol* 2012; 590:2659-75; PMID:22473777; <http://dx.doi.org/10.1113/jphysiol.2011.224485>
44. Vignery A. Osteoclasts and giant cells: macrophage-macrophage fusion mechanism. *Int J Exp Pathol* 2000; 81:291-304; PMID:11168677; <http://dx.doi.org/10.1046/j.1365-2613.2000.00164.x>
45. Pötgens AJ, Schmitz U, Bose P, Versmold A, Kaufmann P, Frank HG. Mechanisms of syncytial fusion: a review. *Placenta* 2002; 23(Suppl A):S107-13; PMID:11978067; <http://dx.doi.org/10.1053/plac.2002.0772>
46. Horsley V, Pavlath GK. Forming a multinucleated cell: molecules that regulate myoblast fusion. *Cells Tissues Organs* 2004; 176:67-78; PMID:14745236; <http://dx.doi.org/10.1159/000075028>
47. Chen EH, Olson EN. Unveiling the mechanisms of cell-cell fusion. *Science* 2005; 308:369-73; PMID:15831748; <http://dx.doi.org/10.1126/science.1104799>
48. Ciejek E, Thorner J. Recovery of *S. cerevisiae* a cells from G1 arrest by alpha factor pheromone requires endopeptidase action. *Cell* 1979; 18:623-35; PMID:391400; [http://dx.doi.org/10.1016/0092-8674\(79\)90117-X](http://dx.doi.org/10.1016/0092-8674(79)90117-X)
49. Brenner S. The genetics of *Caenorhabditis elegans*. *Genetics* 1974; 77:71-94; PMID:4366476
50. Samara C, Syntichaki P, Tavernarakis N. Autophagy is required for necrotic cell death in *Caenorhabditis elegans*. *Cell Death Differ* 2008; 15:105-12; PMID:17901876; <http://dx.doi.org/10.1038/sj.cdd.4402231>
51. Tavernarakis N, Pasparaki A, Tasdemir E, Maiuri MC, Kroemer G. The effects of p53 on whole organism longevity are mediated by autophagy. *Autophagy* 2008; 4:870-3; PMID:18728385