

Published in final edited form as:

Lab Invest. 2012 October ; 92(10): 1472–1482. doi:10.1038/labinvest.2012.116.

Evidence questioning cromolyn’s effectiveness and selectivity as a “mast cell stabilizer” in mice

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Abstract

Cromolyn, widely characterized as a “mast cell stabilizer”, has been used in mice to investigate the biological roles of mast cells *in vivo*. However, it is not clear to what extent cromolyn can either limit the function of mouse mast cells or influence biological processes in mice independently of effects on mast cells. We confirmed that cromolyn (at 10 mg/kg *in vivo* or 10 – 100 μ M *in vitro*) can inhibit IgE-dependent mast cell activation in rats *in vivo* (measuring Evans blue extravasation in passive cutaneous anaphylaxis and increases in plasma histamine in passive systemic anaphylaxis) and *in vitro* (measuring peritoneal mast cell β -hexosaminidase release and prostaglandin D₂ synthesis). However, under the conditions tested, cromolyn did not inhibit those mast cell-dependent responses in mice. In mice, cromolyn also failed to inhibit the ear swelling or leukocyte infiltration at sites of passive cutaneous anaphylaxis. Nor did cromolyn inhibit IgE-independent degranulation of mouse peritoneal mast cells induced by various stimulators *in vitro*. At 100 mg/kg, a concentration ten times higher than that which inhibited passive systemic anaphylaxis in rats, cromolyn significantly inhibited the increases in plasma concentrations of mouse mast cell protease-1 (but not of histamine) during passive systemic anaphylaxis, but had no effect on the reduction in body temperature in this setting. Moreover, this concentration of cromolyn (100 mg/kg) also inhibited LPS-induced TNF production in genetically mast cell-deficient C57BL/6-*Kit^{W-sh/W-sh}* mice *in vivo*. These results question cromolyn’s effectiveness and selectivity as an inhibitor of mast cell activation and mediator release in the mouse.

Keywords

Anaphylaxis; cromolyn; disodium cromoglycate; DSCG; mast cell; mouse; rat

Mast cells have long been regarded as exceptionally efficient initiators and amplifiers of certain innate and acquired immune responses, especially IgE-dependent acute responses to challenge with specific antigen.^{1–5} In addition to their well-known functions in IgE-dependent responses, mast cells also can secrete mediators in response to a variety of other signals including products of pathogens, components of animal venoms,^{6–7} products of complement activation, neurotransmitters,⁸ vascular factors,⁹ and stem cell factor (SCF), suggesting that mast cells have a much larger spectrum of potential roles in health and disease than was originally thought.^{1–5}

One approach for understanding the roles of mast cells in health and disease would be to employ drugs that can effectively and selectively inhibit the function of this cell *in vivo*. Cromolyn (cromolyn sodium, sodium cromoglycate, SCG, disodium cromoglycate, DSCG)

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DISCLOSURE/DUALITY OF INTEREST

The authors state no conflict of interest.

is often used as such an agent. Although its mechanism of action has not been clarified, cromolyn has been used clinically as an anti-asthma drug for more than 30 years.¹⁰ Clinical studies showed that inhalation of cromolyn by subjects with allergic asthma blocked allergen-induced bronchospasm, presumably by inhibiting the release of mediators from mast cells,¹¹ an idea that was supported by the drug's ability to inhibit the degranulation of rat peritoneal mast cells in response to challenge with IgE and specific antigen *in vitro*.^{12–15} For this reason, cromolyn is often characterized clinically as a “mast cell stabilizer”,¹⁶ and this drug is now used for the treatment of other diseases thought to involve mast cell activation, including allergic rhinitis, allergic conjunctivitis, and mastocytosis.¹⁷

Although the early studies of cromolyn were performed predominantly in rats and humans, many groups are now employing this agent to “stabilize” mast cells or to suppress mast cell functions in mice. Indeed, treatment of mice with cromolyn *in vivo* has become commonplace as part of efforts to investigate roles of mast cells in mouse models of host defense or disease that are thought to be independent of IgE.^{18–29} However, surprisingly, there is little published information supporting the conclusion that cromolyn is either an effective or selective “stabilizer” (i.e., inhibitor) of the activation of mast cell populations in the mouse.

On the contrary, one group reported that cromolyn (at 200 mg/kg) did not inhibit Evans blue extravasation associated with IgE-dependent PCA reactions in mice, one of the most common assays used to measure mast cell function *in vivo*.³⁰ Moreover, other groups reported that treatment with cromolyn (at 1 – 100 μM ³¹ or 1 – 100 $\mu\text{g/mL}$,¹⁵ respectively) at the time of antigen challenge did not inhibit IgE plus antigen-induced degranulation³¹ or cysteinyl leukotriene release¹⁵ in mouse bone-marrow-derived cultured mast cells (BMCMCs) *in vitro*. However, Marquardt *et al.* reported that long term culture of mouse BMCMCs with cromolyn, as opposed to short-term treatment with the drug, resulted in a significant inhibition of IgE- and antigen-induced mediator release by BMCMCs, but the basis for the difference in the response of these cells to short-term versus long-term treatment with cromolyn was not defined.³¹ In addition, Forbes *et al.* reported that repeated treatment of mice with cromolyn inhibited the increases in blood concentrations of the mast cell-associated mediator, mouse mast cell protease-1 (mMCP-1), a product of mucosal mast cells, in mice that genetically overexpressed IL-9.³² Taken together, such studies raise the possibility that cromolyn might have inhibitory effects on some mast cell functions (such as the release of MCP-1 by mucosal mast cells³²) but not others (e.g., IgE-dependent activation of skin mast cells³⁰), and that the effects of the drug on even the same type of mast cells may depend on the conditions of cromolyn exposure.^{15,31}

However, there are very few reports analyzing the effects of cromolyn on mouse mast cells, and we are aware of no reports on the effects of cromolyn on mouse peritoneal mast cells (PMCs) – the mast cell type most thoroughly investigated in studies of the effects of cromolyn on mast cell activation in the rat.^{12–15} Accordingly, we think that it is difficult to draw any conclusion about the effectiveness or selectivity of cromolyn as a mast cell stabilizer in mice other than more studies of this topic are needed. By contrast, there have been many reports showing that cromolyn can inhibit rat PMC degranulation^{12–15} or prostaglandin D₂ (PGD₂) synthesis *in vitro*,³³ as well as reduce the extent of IgE-dependent degranulation of rat skin mast cells in passive cutaneous anaphylaxis (PCA) reactions *in vivo*.^{13,15,30}

In the present study, we used rats and rat PMCs as positive controls for experiments designed to investigate in detail the effectiveness of cromolyn as an inhibitor of mouse mast cell activation *in vivo* and *in vitro*. These studies included testing the drug's ability to interfere with mouse PMC activation by IgE and specific antigen or by multiple different

agents which can elicit IgE-independent mast cell activation *in vitro*. Because it has been well-established that pre-treatment of rat PMCs with cromolyn before stimulating the cells to degranulate substantially reduced the inhibitory effect of cromolyn on PMC degranulation (an effect of the drug called “tachyphylaxis”^{12,13}), we focused primarily on the effects of cromolyn treatment observed when the agent is administered at the time of mast cell activation. We also used genetically mast cell-deficient mice to assess whether the actions of this agent are selective for mast cells *in vivo*.

MATERIALS AND METHODS

Animals

Kit^{W-sh/+} mice generously provided by Peter Besmer (Memorial Sloan-Kettering Cancer Center) were backcrossed with C57BL/6J mice (Jackson Laboratories) for more than 11 generations to produce mast cell-deficient C57BL/6-*Kit^{W-sh/W-sh}* (herein: *Kit^{W-sh/W-sh}*) mice.³⁴ *Cpa3-Cre; Mcl-1^{+/+}* and mast cell- and basophil-deficient *Cpa3-Cre; Mcl-1^{fl/fl}* mice were bred in our laboratory.³⁵ C57BL/6J mice or Wistar rats were purchased from Jackson Laboratories or Charles River, respectively. All animal care and experimentation was conducted in accordance with the “Guide for the Care and Use of Laboratory Animals” prepared by the Institute of Laboratory Animal Resources, National Research Council, and published by the National Academy Press (revised, 1996) and with the approval of the Stanford University Committee on Animal Welfare.

Passive cutaneous anaphylaxis (PCA) reaction

Rats (female, 8 weeks) and mice (male or female, 8 weeks) were sensitized by intradermal (i.d.) injection of anti-DNP IgE (α DNP clone e26, generously provided by Dr. Fu-Tong Liu, UC Davis; 2.5 ng for rat dorsal skin; 10 ng for mouse ear pinnae) in saline (0.9% sodium chloride; 50 μ L for rats; 10 μ L for mice). Twenty four h later, DNP-HSA (Sigma; 1 mg/kg for rats; 10 mg/kg for mice) was injected i.v. with or without cromolyn sodium salt (MP Biomedicals). Mouse ear thickness was measured with a dial thickness gauge (G-1A, Ozaki, Tokyo, Japan). Mouse ear pinnae collected from mice killed by exposure to CO₂ 6 h after challenge were fixed in 10% formalin for preparation of paraffin sections stained with hematoxylin and eosin, and leukocyte numbers were quantified by light microscopy as per mm of horizontal field length of the ear (by an observer not aware of the identity of the individual sections). For measuring Evans blue extravasation, Evans blue (Sigma; 10 mg/kg for rats; 100 mg/kg for mice) was injected i.v. with DNP-HSA.³⁶ Thirty min later, skin areas were photographed and cut out. Evans blue dye was extracted by incubating the skin samples in DMSO (1 mL for rat samples; 0.5 mL for mouse samples) for 24 h at 37°C, and then O.D. 650 nm was measured.

Passive systemic anaphylaxis (PSA) reaction

Rats (female, 8 weeks) and mice (male, 8 weeks) were sensitized i.v. with anti-DNP IgE (1 μ g/kg for rats; 100 μ g/kg for mice³⁷). Twenty four h later, DNP-HSA (1 mg/kg for rats; 10 mg/kg for mice³⁷) was injected i.v. with or without cromolyn. Ninety sec later, blood was collected and the plasma histamine or mouse mast cell protease-1 (mMCP-1) concentrations were measured using histamine or mMCP-1 ELISA kits (Beckman coulter or eBioscience, respectively). Body temperature was measured with a rectal thermometer (Physitemp Instrument, Inc., NJ).

Preparation of PMCs

Whole peritoneal cells were collected in RPMI medium (GIBCO; with 1 mg/mL BSA and 10 units/mL heparin) from rat (female, 12 weeks) or mouse (female, retired breeder)

peritoneal cavities. The cells were mounted on 0.235 g/mL histodenz (Sigma), and were centrifuged at 400 $\times g$ (7 min for rats; 15 min for mice). The cells at the bottom of the tube were collected. More than 90% cells were PMCs by toluidine blue metachromatic staining. For stimulation with antigen (DNP-HSA), PMCs were cultured for 24 h with anti-DNP IgE in Opti-MEM (GIBCO; with 10% FBS) prior to stimulation with DNP-HSA. For other stimuli (recombinant mouse SCF was purchased from Peprotech, all other materials from Sigma), the PMCs were used immediately after isolation.

Measurement of β -hexosaminidase release

β -Hexosaminidase release was measured as described previously.^{6,36} The PMCs were resuspended (5×10^5 cells/mL) in Tyrode's buffer⁶ then treated with the stimuli in the absence or presence (added simultaneously) of cromolyn at 37°C. The reactions were stopped on ice 3 min later. The percentage of degranulation was calculated, taking the O.D. 405 nm of 0.5% triton X-100-treated cell sample as 100%.

Measurement of PGD₂ production

PMCs sensitized with IgE were resuspended (5×10^5 cells/mL) in Tyrode's buffer (without BSA), then treated with 100 ng/mL DNP-HSA in the absence or presence (added simultaneously [except Supplementary Figure S6]) of cromolyn at 37°C. The reactions were stopped on ice 10 min later and PGD₂ concentrations in the supernatants were measured by a PGD₂ EIA kit (Cayman chemical company).

LPS challenge *in vivo* and measurement of TNF

Vehicle (sterile, non-pyrogenic 0.9% sodium chloride, Hospira) or cromolyn (100 mg/kg in vehicle) was administered to mice (male, 8 weeks) by i.p. injection 30 min before challenge i.p. with 0.1 mg/kg LPS (LPS from *E. coli*, Serotype EH100, Enzo Life Sciences). Blood samples were collected 90 min after challenge and plasma was collected by centrifugation. TNF concentrations in plasma were measured using a mouse TNF ELISA kit (BD Biosciences).

LPS challenge *in vitro*

Spleen cells were collected from the spleens of mice (male, 8 weeks), treated with ACK lysing buffer for 5 min, and then passed through 70 μ m cell strainers. Peritoneal cells collected from mouse peritoneal cavities were washed twice and resuspended in RPMI (with 10% FBS). The spleen cells (5×10^6 cells/mL) or the peritoneal cells (5×10^5 cells/mL) were treated with or without cromolyn for 15 min and then stimulated with 1 ng/mL LPS for 3 h at 37°C. TNF concentrations in supernatants were measured by an ELISA as described above.

Statistical analyses

Unless noted otherwise, all results are expressed as mean + or \pm S.D. and were evaluated for statistical significance (defined as $p < 0.05$) by unpaired Student's *t*-test for comparisons between two groups (Supplementary Figure S1 and S3), by two-way ANOVA for comparisons between the time dependent curve of ear thickness (Figure 1c and S4c) and body temperature (Figure 2c), or by one-way ANOVA followed by Bonferroni test for comparisons between more than two groups (all other figures).

RESULTS

Cromolyn can effectively inhibit activation of rat, but not mouse, mast cells *in vivo*

First, we sought to confirm the well-established inhibitory effect of cromolyn on rat PCA reactions (Table 1).^{13,15,30} In accordance with previously published reports, we found that cromolyn (10 mg/kg) almost completely inhibited Evans blue extravasation induced by local IgE-dependent mast cell degranulation in rats (Figure 1a). We then tested PCA reactions in mice. The different concentrations of reagents (i.e., IgE, DNP-HSA, and Evans blue) and the different skin regions used to induce PCA reactions (i.e., dorsal skin or ear pinnae) that we used routinely for studies in rats or mice were chosen based on preliminary experiments that took into account species-specific variation in the amounts needed to elicit and assess robust reactions (Supplementary Figure S1). As expected, Evans blue extravasation was not observed when we tried to elicit PCA reactions in mast cell-deficient *Kit^{W-sh/W-sh}* mice (Figure 1b), confirming that this reaction is dependent on the presence of skin mast cells.³⁸ However, treatment with 10 mg/kg cromolyn did not inhibit the Evans blue extravasation at PCA reaction sites in wild type mice (Figure 1b). Even when administered at 100 mg/kg (a dose ten times higher than that used in rats), cromolyn did not significantly inhibit IgE-dependent PCA reactions in male mice (Figure 1b). Because we tested PCA in female rats, we also confirmed that there was no inhibitory effect of cromolyn (at 10 or 100 mg/kg) on PCA-associated Evans blue extravasation in female wild type mice (Supplementary Figure S2).

Ear swelling and numbers of dermal leukocytes also can be measured to evaluate PCA reactions in mice. In IgE-injected *Kit^{W-sh/W-sh}* mice, there were no significant changes in either ear thickness (Figure 1c) or skin leukocyte numbers (Figure 1d) after antigen challenge, confirming that both are largely or entirely dependent on the presence of skin mast cells. However, in support of our Evans blue extravasation results (Figure 1b), treatment with 10 or 100 mg/kg cromolyn failed to inhibit either the ear swelling (Figure 1c) or the local leukocyte response (Figure 1d) associated with PCA reactions in wild type mice.

We also measured plasma histamine concentrations 90 sec after inducing IgE-dependent PSA, again using higher concentrations of IgE and antigen in mice than rats in order to elicit robust reactions in each species (mice did not detectably respond to the concentrations of reagents that elicited a reaction in rats; see Supplementary Figure S3). When IgE-sensitized rats were challenged with antigen together with cromolyn (10 mg/kg), the antigen-induced increase in plasma histamine was completely inhibited (Figure 2a). Antigen challenge did not increase plasma histamine levels in IgE-sensitized *Kit^{W-sh/W-sh}* mice (Figure 2b), confirming that this finding is primarily dependent on mast cells. However, administration of 10 or 100 mg/kg cromolyn together with antigen failed to inhibit the increase in plasma histamine in IgE-sensitized wild type mice (Figure 2b). We also measured body temperature (Figure 2c) and plasma mMCP-1 concentration (Figure 2d) after inducing IgE-dependent PSA. Again, antigen challenge did not result in decreased body temperature (Figure 2c) or increased plasma mMCP-1 levels (Figure 2d) in IgE-sensitized *Kit^{W-sh/W-sh}* mice, confirming that these findings are primarily dependent on mast cells. However, administration of 10 or 100 mg/kg cromolyn together with antigen failed to inhibit the decrease in body temperature in wild type mice (Figure 2c). On the other hand, at the higher dose tested (100 mg/kg), cromolyn did significantly inhibit the increase in plasma mMCP-1 associated with IgE-dependent PSA in wild type mice (Figure 2d).

In some previous studies, cromolyn was administered repeatedly to mice.^{23,27,32} Therefore, we tested the effect of repeated administration of cromolyn (Supplementary Figure S4a) on IgE-dependent Evans blue dye extravasation (Supplementary Figure S4b) and ear swelling (Supplementary Figure S4c) in PCA reactions, and on increases in plasma histamine

(Supplementary Figure S4d) and mMCP-1 (Supplementary Figure S4e) in PSA. In support of our finding in Figure 2d, the repeated administration of cromolyn inhibited the increases in plasma mMCP-1 concentrations associated with PSA (Supplementary Figure S4e). However, repeated treatment with cromolyn at 100 mg/kg failed to inhibit other features of the IgE-dependent reactions (Supplementary Figure S4b – d), including the increases in plasma histamine associated with IgE-dependent PSA (Supplementary Figure S4d).

Cromolyn can effectively inhibit activation of rat, but not mouse, mast cells *in vitro*

To study the effects of cromolyn on rat or mouse mast cell activation more directly, we assessed mouse PMC *in vitro*. In preliminary experiments, we confirmed that a higher concentration of IgE was needed to induce high levels of β -hexosaminidase release in mouse PMCs (Supplementary Figure S5b) than in rat PMCs (Supplementary Figure S5a) *in vitro*. Therefore, in tests of mouse PMCs, we used both low and high concentrations of IgE (i.e., 50 ng/mL [the same concentration as for rat PMC experiments] and 5000 ng/mL [in order to induce more robust activation of mouse PMCs]). First, we tried to confirm the well-established inhibitory effect of cromolyn on IgE- and antigen-induced degranulation in rat PMCs (Table 1).^{12–15} In accordance with previously published reports,^{12,13} we found that pretreating rat PMCs with cromolyn before adding DNP-HSA reduced the inhibitory effect of cromolyn on PMC degranulation (so called “tachyphylaxis”; Supplementary Figure S6). In order to avoid this phenomenon, we added cromolyn simultaneously with DNP-HSA challenge. The simultaneous addition of cromolyn (10 – 100 μ M) inhibited antigen-induced IgE-dependent degranulation of rat PMCs in a concentration-dependent manner (Figure 3a). Cromolyn at 100 μ M also inhibited β -hexosaminidase release induced by different antigen concentrations in rat PMCs (Figure 3d). However, treatment of mouse PMCs with cromolyn (at 1 – 100 μ M) did not inhibit antigen- and IgE-induced degranulation (Figure 3b, c, e and f). Similarly, cromolyn (10 – 100 μ M) inhibited PGD₂ synthesis in rat (Figure 3g), but not mouse (Figure 3h and i), PMCs after challenge with IgE and antigen. We also tried to confirm the lack of effect of cromolyn on mouse BMCMC degranulation when the agent was administered at the time of antigen challenge. In accord with a prior report,³¹ treatment with cromolyn (at 1– 100 μ M) at the time of antigen challenge did not inhibit antigen- and IgE-induced degranulation in mouse BMCMCs (Supplementary Figure S7). Moreover, in our hands, maintaining BMCMCs with cromolyn (1 μ M to 1 mM) for up to 8 days also did not result in any inhibition of the cells’ degranulation in response to IgE and antigen (data not shown).

Although cromolyn did not inhibit IgE-dependent degranulation in mouse PMCs (Figure 3), the possibility remained that cromolyn might be able to inhibit IgE-independent degranulation induced by other activators. To investigate this possibility, we tested the ability of cromolyn to inhibit β -hexosaminidase release following stimulation with a variety of agents known to activate PMCs *in vitro*. We tested thapsigargin (Figure 4a and e), ionomycin (Figure 4b and f), substance P (Figure 4c and g), compound 48/80 (Figure 4d and h), phorbol 12-myristate 13-acetate (PMA) with ionomycin (Figure 4i), endothelin-1 (Figure 4j), adenosine (Figure 4k), and mouse SCF (Figure 4l). All agents induced β -hexosaminidase release from both rat and mouse PMCs in a dose dependent manner (Figure 4a – l). In rat PMCs (Figure 4a – d), despite differences in cromolyn sensitivity at different concentrations of certain stimuli (e.g., compound 48/80), the simultaneous addition of 100 μ M cromolyn inhibited the degranulation induced by each of the stimuli tested. However, we observed no significant inhibitory effects of 100 μ M cromolyn on the stimulus-induced degranulation responses of mouse PMCs to any of the agents we tested (Figure 4e – l).

Cromolyn can inhibit LPS-induced TNF production in both wild type and mast cell deficient mice

Finally, we investigated whether cromolyn might have detectable effects in genetically mast cell-deficient mice. Because *Kit^{W-sh/W-sh}* mice virtually lack mast cells, any effects of cromolyn observed in these mice would indicate an effect of the agent on other cell types. In wild type mice, increased levels of plasma TNF were detectable 90 min after injection of the mice with 0.1 mg/kg LPS (Figure 5a). Administration of 100 mg/kg cromolyn significantly inhibited the increase in plasma TNF levels in wild type mice (Figure 5a). In *Kit^{W-sh/W-sh}* mice, injection of LPS induced increases in plasma TNF to almost the same levels observed in wild type mice (Figure 5b), indicating that such LPS-induced TNF production can occur independently of mast cells. Moreover, administration of 100 mg/kg cromolyn significantly inhibited the increase in plasma TNF in *Kit^{W-sh/W-sh}* mice (Figure 5b). These results indicate that cromolyn can inhibit LPS-induced TNF production in mice in a mast cell-independent manner *in vivo*.

We next investigated the effects of cromolyn on LPS-induced TNF production *in vitro*. TNF was detectable in the supernatants of wild type spleen cells (Figure 5c) and peritoneal cells (Figure 5e) 3 h after stimulation with 1 ng/mL LPS, and the addition of cromolyn (1 – 100 μ M) inhibited such TNF production in a dose-dependent manner (Figure 5c and e). Treatment of spleen cells (Figure 5d) and peritoneal cells (Figure 5f) from *Kit^{W-sh/W-sh}* mice with 1 ng/mL LPS also induced TNF production. Moreover, cromolyn (1 – 100 μ M) inhibited LPS-induced TNF production by *Kit^{W-sh/W-sh}* spleen or peritoneal cells in a dose-dependent manner (Figure 5d and f). We also tested the effect of cromolyn on these reactions *in vitro* using *Kit*-independent mast cell- (and basophil-) deficient *Cpa3-Cre; Mcl-1^{fl/fl}* mice³⁵ (Supplementary Figure S8). In accordance with our data from *Kit^{W-sh/W-sh}* mice, we found that cromolyn (1 – 100 μ M) inhibited LPS-induced TNF production by spleen or peritoneal cells from both *Cpa3-Cre; Mcl-1^{+/+}* (littermate control) mice and *Cpa3-Cre; Mcl-1^{fl/fl}* (mast cell- and basophil-deficient) mice in a dose-dependent manner. Taken together, our findings indicate that cromolyn can inhibit LPS-induced TNF production in mice in a mast cell-independent manner.

DISCUSSION

We confirmed the well-established inhibitory effect of cromolyn on IgE-dependent PCA reactions in rats *in vivo*^{13,15,30} (Figure 1, Table 1) and on IgE- and antigen-dependent degranulation and mediator release in rat PMCs *in vitro*¹²⁻¹⁵ (Figure 3, Table 1). In contrast to rat PMCs, it was reported that cromolyn added 5 min before or at the time of antigen challenge did not inhibit IgE- and antigen-induced degranulation of either rat BMCMCs³⁹ or mucosal type mast cells isolated from rat intestine¹⁴ *in vitro* (Table 1). These findings suggest that, in rats, different mast cell populations may exhibit different sensitivity to cromolyn. In addition, we also confirmed the previously reported phenomenon that cromolyn pretreatment induces “tachyphylaxis” in rat PMCs^{12,13} (Supplementary Figure S6). This phenomenon shows that, even in rats, cromolyn can inhibit mast cell degranulation only under certain conditions of administration.

In mice, it has been reported that cromolyn treatment immediately before³⁰ or at the time of^{15,31} antigen challenge did not inhibit either IgE-dependent PCA reactions *in vivo*³⁰ or the activation of mouse BMCMCs *in vitro*^{15,31} (Table 1). Since, as noted above, different mast cell populations might exhibit different sensitivity to cromolyn in rats, it is possible that the drug also might vary in its ability to inhibit the activity of different populations of mouse mast cells *in vivo* or *in vitro*. We detected no inhibitory effects of cromolyn (either given once at the time of antigen challenge or administered every 12 hours starting 2.5 days before antigen challenge) on the increases in vascular permeability, tissue swelling, or leukocyte

infiltration at sites of IgE-dependent PCA reactions (Figure 1b – d, Supplementary Figure S4b and c) or on the increases in plasma histamine (Figure 2b and Supplementary Figure S4d) or decreases in body temperature (Figure 2c) associated with IgE-dependent PSA reactions in mice *in vivo*. However, we found that the increases in plasma levels of mMCP-1 associated with IgE-dependent PSA reactions were significantly inhibited by treatment of mice with a high dose (100 mg/kg) of cromolyn, either given once at the time of antigen challenge (Figure 2d) or administered repeatedly starting 2.5 days before antigen challenge (Supplementary Figure S4e). Since mMCP-1 is a serine protease which is thought to be stored and secreted in a tissue-specific manner by mucosal mast cells,⁴⁰ this result suggests that, under the conditions tested, cromolyn can inhibit the activation of mucosal type mast cells in mice. Forbes *et al.* reported that cromolyn inhibited the increases in blood concentrations of mMCP-1 in transgenic mice that overexpressed IL-9.³² Taken together, our results and those of Forbes *et al.*,³² suggest that, in mice, as well as in rats, different mast cell populations may exhibit different sensitivity to cromolyn.

Given the potency of cromolyn as an inhibitor of rat PMC degranulation,^{12–15} and given our inability to find any prior reports on the effects of cromolyn on degranulation of mouse PMCs (Table 1), we were especially interested to test the effects of this agent on mouse PMCs. We found that cromolyn did not effectively inhibit mouse PMC degranulation in response to IgE and specific antigen or to any of the various other activation stimuli we tested (Figures 3 and 4). Given that both PMCs and skin mast cells are considered connective tissue type mast cells, our results in experiments with mouse PMCs *in vitro* (Figures 3 and 4) and analyzing IgE-dependent PCA reactions in mice *in vivo* (Figure 1) support the conclusion that cromolyn is not a potent inhibitor of connective tissue mast cell activation in the mouse.

Studies with cromolyn illustrate two limitations that apply to many pharmacological approaches. First, there appear to be clear species-dependent differences in sensitivity to the drug. We confirmed that treatment with cromolyn (at 10 mg/kg *in vivo* or at 10 – 100 μ M *in vitro*) at the time of antigen challenge significantly inhibited IgE-dependent rat mast cell activation (Figures 1 – 4, Table 1). In mice, we found that the increase in plasma concentrations of mMCP-1 associated with IgE-dependent PSA was significantly inhibited by 100 mg/kg cromolyn administered at the time of antigen challenge *in vivo* (Figure 2d), but we did not detect inhibition by cromolyn of any of the other mast cell responses we tested, using the drug at 10 or 100 mg/kg *in vivo* and at 1 – 100 μ M *in vitro* (Figures 1 – 4, Table 1). It also has been reported that cromolyn, at 0.1 – 100 mg/kg *in vivo* and 1 – 100 μ g/mL *in vitro*, failed to inhibit guinea pig PCA reactions *in vivo* or histamine release from chopped guinea pig lung *in vitro*.¹⁵ In humans, a high concentration (1 mM) of cromolyn was required to achieve even modest inhibition of mediator release *in vitro* from mast cells isolated from lung,⁴¹ tonsils or intestines,⁴² or from cultured human mast cells derived from umbilical cord blood cells.⁴³ Moreover, human skin mast cells⁴² or cultured human mast cells derived from blood buffy coats⁴⁴ were unresponsive to this agent *in vitro*.

Second, cromolyn's mechanism of action still is not understood, including the basis for the phenomenon of "tachyphylaxis" illustrated in Supplementary Figure S6 or the explanation for the different effects of short-term versus long-term exposure to cromolyn on IgE- and antigen-dependent mediator release by BMCs that have been reported by Marquardt *et al.*³¹ However, several lines of evidence indicate that the effects of cromolyn are not restricted specifically to mast cells. It has been reported that cromolyn can bind directly to some ubiquitous proteins, for instance, heat-shock protein 90 (Hsp90),⁴⁵ S100 proteins,^{46,47} or G-protein-coupled receptor 35 (GPR35).⁴⁸ Considering that Hsp90 can interact with variety of proteins including those involved in hormone signaling, intracellular signal transduction, and cell cycle control,^{45,49} that S100 proteins are expressed in pancreatic,^{46,47}

breast,⁵⁰ lung⁵¹ and colon⁵² cancer cells and that GPR35 is expressed in leukocytes, monocytes, neutrophils and T cells,^{48,53} the cellular targets of cromolyn are unlikely to be restricted to mast cells.

Indeed, by testing genetically mast cell-deficient *Kit^{W-sh/W-sh}* or *Cpa3-Cre; Mcl-1^{fl/fl}* mice, we found that cromolyn (at 100 mg/kg *in vivo* and 10 – 100 μ M *in vitro*) can inhibit LPS-induced TNF production in a mast cell-independent manner *in vivo* and *in vitro* (Figure 5 and Supplementary Figure S8). Notably, acute treatment with the same high dose of cromolyn (100 mg/kg) which inhibited increases in plasma mMCP-1 in IgE-dependent PSA (Figure 2d) also resulted in a mast cell-independent inhibition of TNF production in mast cell-deficient *Kit^{W-sh/W-sh}* mice. In addition to our findings, there have been at least two other reports showing that cromolyn can inhibit certain immune responses in mast cell-deficient mice *in vivo*. In those reports, which employed WBB6F₁-*Kit^{W/W^v}* mice rather than *Kit^{W-sh/W-sh}* or *Cpa3-Cre; Mcl-1^{fl/fl}* mice, cromolyn inhibited both the neutrophil infiltration into the gastric mucosa that was observed after active anaphylaxis⁵⁴ and capsaicin-induced infiltration of eosinophils into the conjunctiva⁵⁵. Those results, and our findings that cromolyn can inhibit LPS-induced TNF release by genetically mast cell-deficient mice *in vivo* and by the spleen or peritoneal cells of such mice *in vitro*, are consistent with observations indicating that cromolyn can dose-dependently inhibit the activation of several different hematopoietic cell types. For example, cromolyn tested *in vitro* at concentrations as low as 10⁻⁷ M markedly reduced the formyl-methionyl-leucyl-phenylalanine-dependent activation of human peripheral blood neutrophils, eosinophils and monocytes, as well as the ability of highly purified human neutrophils or eosinophils to kill *Schistosoma mansoni* schistosomula,⁵⁶ and also markedly reduced *S μ* to *S ϵ* deletional switch recombination and IgE synthesis in highly purified human B cells.⁵⁷

In conclusion, based on our results and those of other groups, we think that there are extensive species-dependent differences in the responsiveness of mast cell populations to the inhibitory effects of cromolyn and that skin and peritoneal mast cells in mice are substantially less responsive to this drug than are the corresponding mast cell populations in the rat. Moreover, cromolyn can inhibit biological processes *in vivo* in mast cell-deficient mice and, when tested *in vitro*, can inhibit the activation of multiple hematopoietic cell types other than mast cells. Because neither the critical molecular targets of cromolyn that explain its ability to inhibit rat mast cell degranulation have been clearly defined nor has its mechanism(s) of action been identified, we think it might be very difficult to clarify why this drug inhibits the degranulation of some types of mast cells but not others, to explain why the effects of the drug on some mast cell populations can differ based on the duration of exposure to the agent, or to define the mechanism(s) by which it can inhibit the activation of other cell types in mast cell-deficient mice. Also, we obviously have not tested cromolyn in all strains of mice, in all biological responses thought to require mast cells, or with all agents capable of inducing mast cell activation. Therefore, it is possible that short or long-term treatment with cromolyn might be able to inhibit functions of mouse mast cells observed in settings other than those studied in this report. Indeed, our data, like those reported by Forbes *et al.*,³² suggest that treatment with high doses of cromolyn can inhibit the activation of mouse mucosal mast cells. However, given the results presented in this report, including the finding that cromolyn can inhibit TNF release in mast cell-deficient mice *in vivo*, we think that one should not assume that the effects of cromolyn treatment of mice *in vivo* necessarily reflect actions of this drug on mast cells.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This study was supported by grants from the National Institutes of Health AI070813, AI023990, and CA072074 to S. J. Galli.

We thank Mariola Liebersbach for support with mouse breeding and Chen Liu for processing slides for histological analysis.

ABBREVIATIONS

BMCMCs	bone-marrow-derived cultured mast cells
DNP-HSA	2,4-dinitrophenyl human serum albumin
GPR35	G-protein-coupled receptor 35
Hsp90	heat-shock protein 90
<i>Kit^{W-sh/W-sh}</i>	C57BL/6- <i>Kit^{W-sh/W-sh}</i>
mMCP-1	mouse mast cell protease-1
PCA	passive cutaneous anaphylaxis
PGD₂	prostaglandin D ₂
PMA	phorbol 12-myristate 13-acetate
PMCs	peritoneal mast cells
PSA	passive systemic anaphylaxis
SCF	stem cell factor

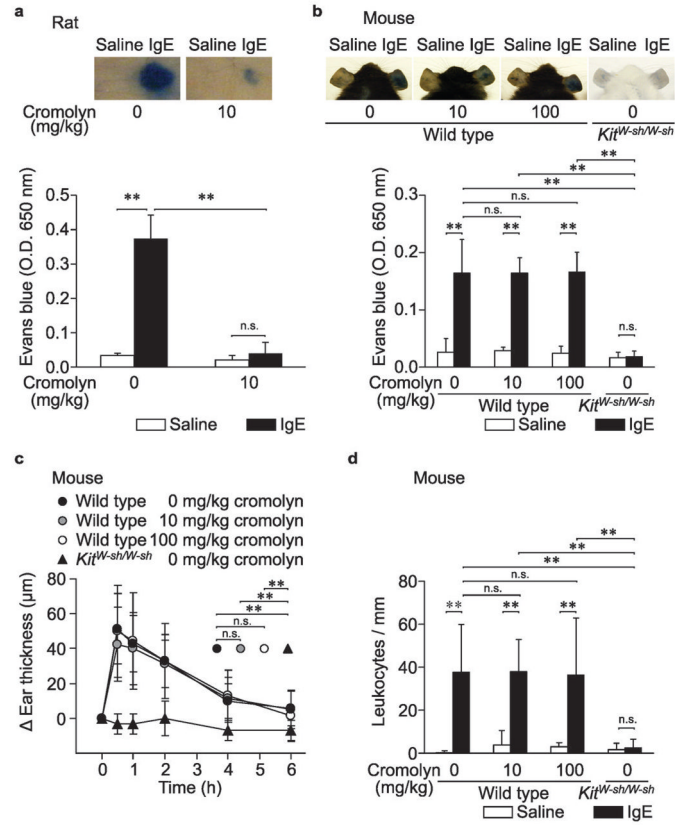
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**Figure 1.**

Effects of cromolyn on PCA reactions *in vivo*. Female rats (a) or male mice (b – d) injected i.d. with vehicle (saline) or anti-DNP IgE (IgE) were challenged i.v. with DNP-HSA with or without cromolyn (injected simultaneously); we then measured Evans blue extravasation (a and b), ear swelling (c) and leukocyte migration (d). N = 4 rats (a), 5 mice (b) and 7 – 8 mice (c and d) per group from at least 2 independent experiments. ** $p < 0.01$, * $p < 0.05$; n.s.: $p > 0.05$.

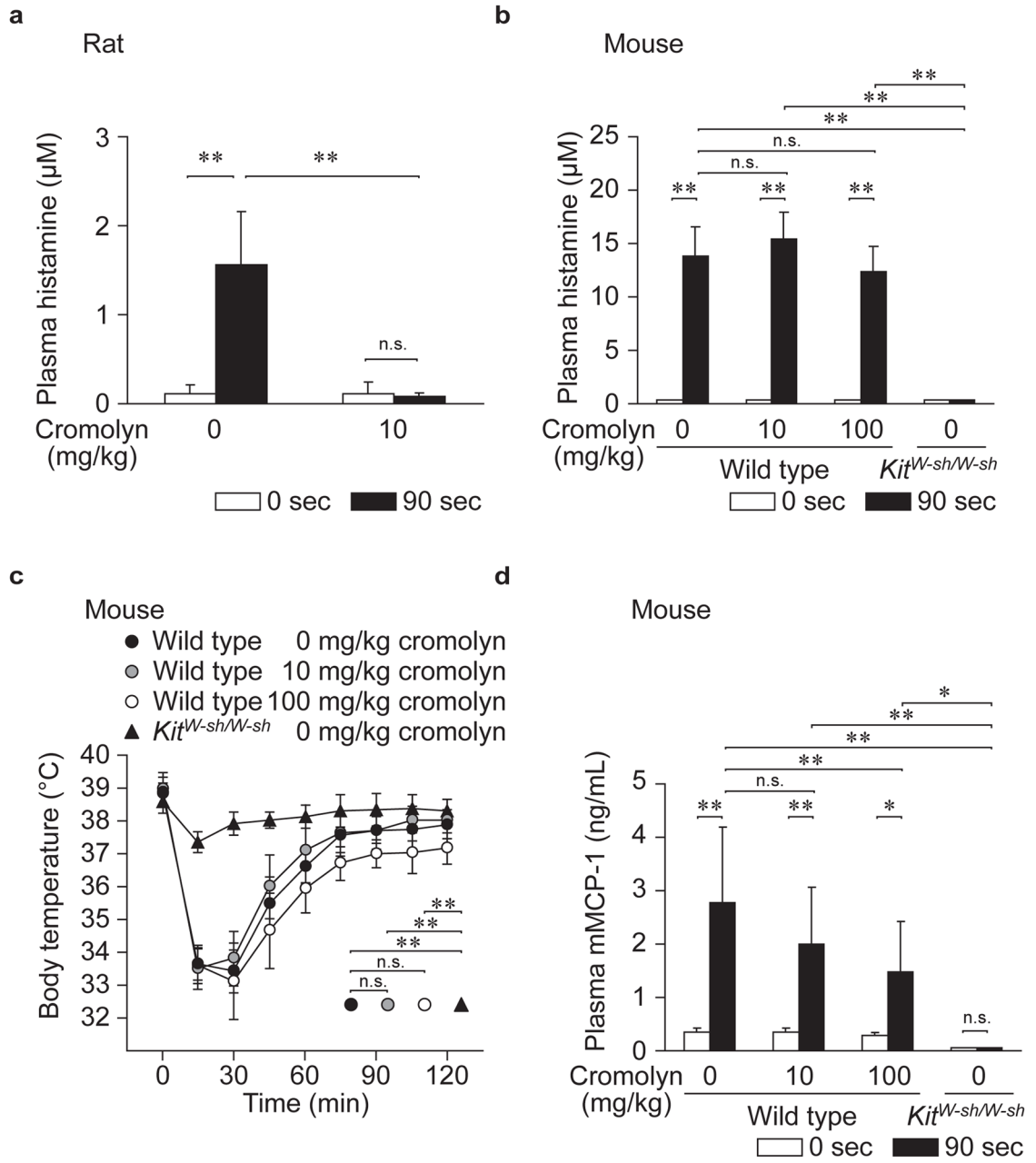


Figure 2.

Effect of cromolyn on PSA reactions *in vivo*. Female rats (a) or male mice (b – d) sensitized i.v. with anti-DNP IgE were challenged i.v. with DNP-HSA with or without cromolyn (injected simultaneously); we then measured plasma histamine (a and b), body temperature (c) and plasma mMCP-1 (d). N = 4 rats (a), and 5 – 10 mice (b – d) per group from at least 2 independent experiments. ** $p < 0.01$, * $p < 0.05$; n.s.: $p > 0.05$.

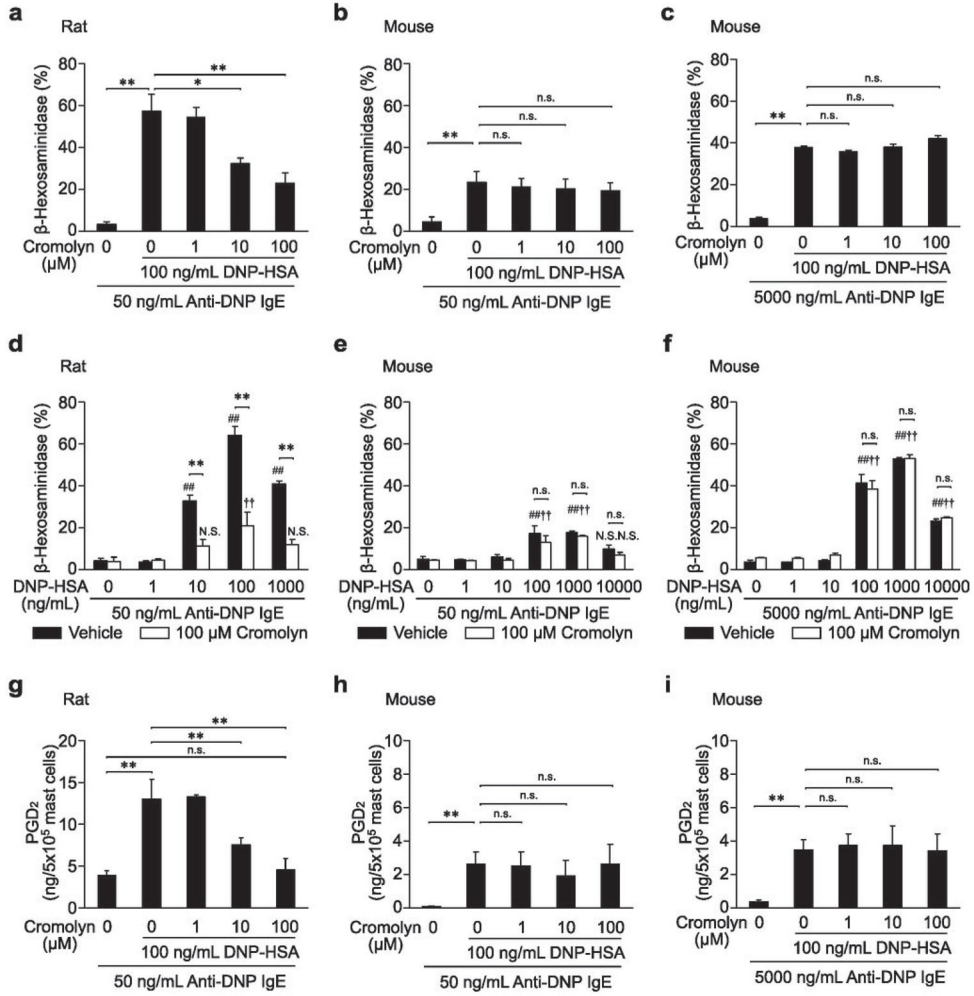


Figure 3. Effects of cromolyn on IgE-dependent PMC degranulation (a – f) or PGD₂ synthesis (g – i) *in vitro*. IgE-sensitized PMCs from female rats (a, d and g) or mice (b, c, e, f, h and i) were stimulated with DNP-HSA with or without cromolyn (added simultaneously). N = 3 – 10 per group from 1 – 2 independent experiments. ## $p < 0.01$, # $p < 0.05$, †† $p < 0.01$, † $p < 0.05$; N.S.: not significantly different versus corresponding values (vehicle or 100 μM cromolyn) for groups not treated with stimuli. ** $p < 0.01$, * $p < 0.05$; n.s.: $p > 0.05$.

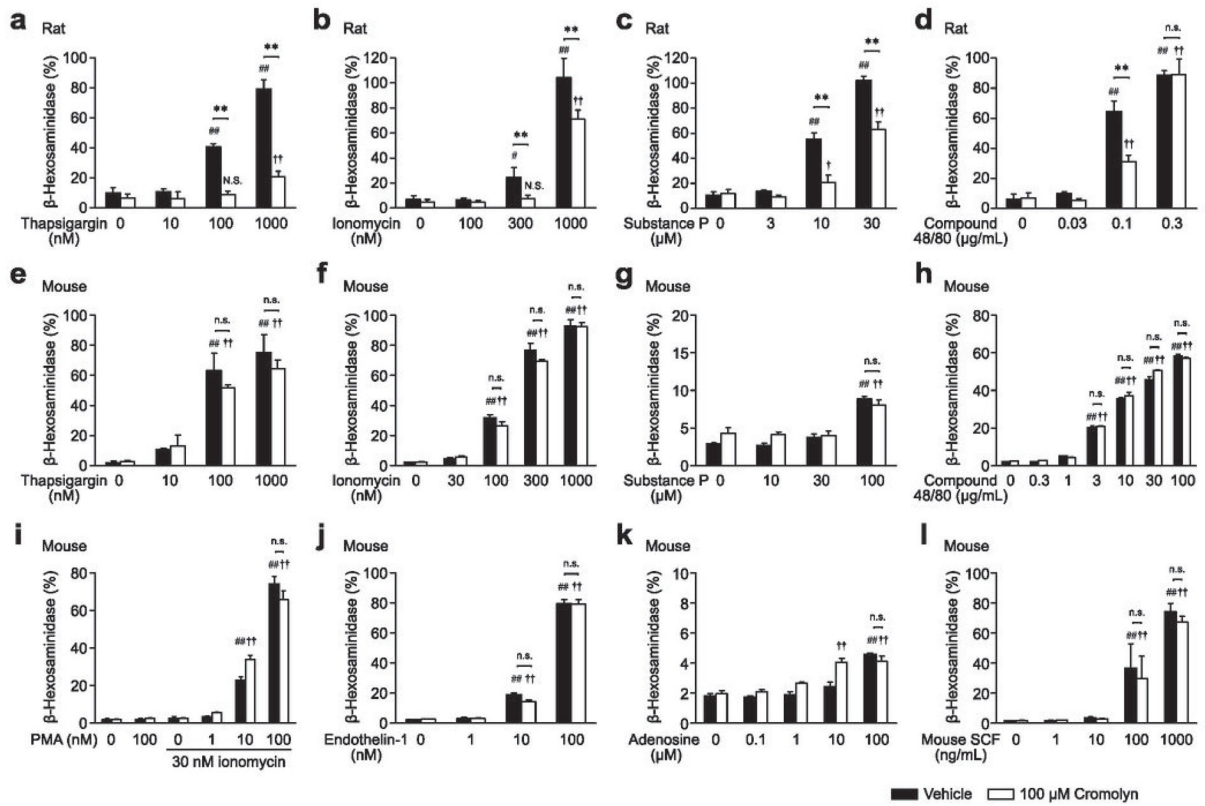


Figure 4.

Effect of cromolyn on IgE-independent degranulation of PMCs *in vitro*. PMCs from female rats (a – d) or mice (e – l) were stimulated with thapsigargin (a and e), ionomycin (b and f), substance P (c and g), compound 48/80 (d and h), PMA with ionomycin (i), endothelin-1 (j), adenosine (k), or mouse SCF (l), with or without cromolyn (added simultaneously). $N = 3 - 6$ per group from 1 – 2 independent experiments. ## $p < 0.01$, # $p < 0.05$, †† $p < 0.01$, † $p < 0.05$; N.S. not significantly different versus corresponding values (vehicle or 100 μ M cromolyn) for groups not treated with stimuli. ** $p < 0.01$, * $p < 0.05$; n.s.: $p > 0.05$.

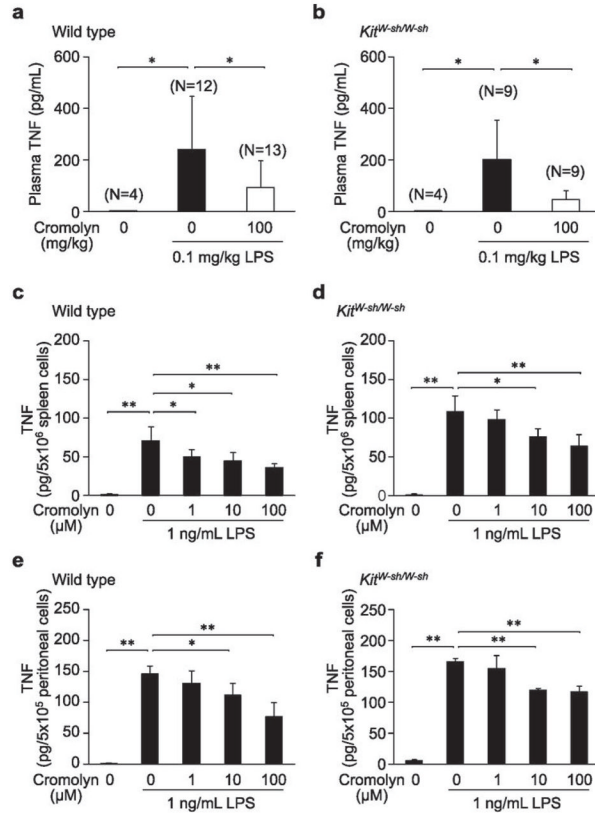


Figure 5.

Effects of cromolyn on LPS-induced TNF production in C57BL/6J wild type or mast cell-deficient *Kit^{W-sh/W-sh}* mice *in vivo* (a and b) or *in vitro* (c – f). Male wild type (a) or *Kit^{W-sh/W-sh}* (b) mice were injected i.p. with vehicle or 100 mg/kg cromolyn 30 min before i.p. challenge with LPS. Spleen cells (c and d) or peritoneal cells (e and f) from wild type (c and e) or *Kit^{W-sh/W-sh}* (d and f) mice incubated for 15 min with or without cromolyn were stimulated with LPS. N = 4 – 13 mice per group from 3 (a) or 2 (b) independent experiments or 4 per group from one of 2 – 3 independent experiments, each of which gave similar results (c – f). ** $p < 0.01$, * $p < 0.05$.

Table 1

Ability of acute exposure to cromolyn to inhibit features of PCA or PSA responses *in vivo* or mast cell degranulation *in vitro* in rats and mice. The table shows results obtained when cromolyn was injected either simultaneously with^{13,15} or immediately before^{13,30} antigen challenge *in vivo*, or added either simultaneously with^{12-15,31} or 5 min before^{12,39} antigen challenge *in vitro*. All of the results reported below from this report are from experiments in which cromolyn was administered simultaneously with antigen.

	Previous reports		This report	
	Rat	Mouse	Rat	Mouse
<i>In vivo</i> PCA	Yes ^{13,15,30}	No ³⁰	Yes (10 mg/kg)	No (10 – 100 mg/kg)
Ear swelling				No (10 – 100 mg/kg)
Leukocyte infiltration				No (10 – 100 mg/kg)
PSA Histamine release			Yes (10 mg/kg)	No (10 – 100 mg/kg)
Temperature decrease				No (10 – 100 mg/kg)
mMCP-1 release				Yes (only at 100 mg/kg)
<i>In vitro</i> PMCs	Yes ¹²⁻¹⁵		Yes (10 – 100 μM)	No (1 – 100 μM)
BMCs	No ³⁹	No ^{15,31}		No (1 – 100 μM)
Mucosal mast cells	No ¹⁴			