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Reductions in glucose among postmenopausal women who use and do not use estrogen therapy

Catherine Kim, M.D., M.P.H., University of Michigan, Ann Arbor, MI

Shengchun Kong, M.S., University of Michigan, Ann Arbor, MI

Gail A. Laughlin, Ph.D., University of California, San Diego

Sherita H. Golden, M.D., M.H.S., Johns Hopkins University, Baltimore, MD

Kieren J. Mather, M.D., Indiana University, Indianapolis, IN

Bin Nan, Ph.D., University of Michigan, Ann Arbor, MI

John F. Randolph Jr., M.D., University of Michigan, Ann Arbor, MI

Sharon L. Edelstein, Sc.M., George Washington University, Rockville, MD

Fernand Labrie, M.D., Ph.D., Laval University, Quebec City, Canada and Al Imam Mohammed Ibn Saud Islamic University, Riyadh, Saudi Arabia

Elizabeth Buschur, M.D., and University of Michigan, Ann Arbor, MI

Elizabeth Barrett-Connor, M.D. University of California, San Diego

for The Diabetes Prevention Program Research Group

Abstract

Objective—Among postmenopausal women who do not use estrogen hormone therapy (HT) we have previously reported that intensive lifestyle intervention (ILS) leads to increases in sex hormone binding globulin (SHBG), and such increases were associated with reductions in fasting plasma glucose (FPG) and 2-hour post-challenge glucose (2HG). Oral HT decreases FPG and increases 2HG, while increasing both SHBG and estradiol (E2). It is unknown if ILS reduces

Address for Correspondence: Catherine Kim, M.D. M.P.H. 300 North Ingalls Building, Room 7C13, Mailstop 5429, Ann Arbor, MI 48109-5429, Telephone: (734) 936-5216, Fax: (734) 936-8944, cathkim@umich.edu.

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glucose among HT users, if changes in SHBG and E2 might mediate any glucose decreases in HT users, and if these patterns differ from non-HT users.

Methods—We conducted a secondary analysis of postmenopausal women in the Diabetes Prevention Program who used HT at baseline and 1 year follow-up (n=324) and who did not use HT at either time point (n=382). Participants were randomized to ILS, metformin, or placebo administered 850 mg twice a day.

Results—HT users were younger, more often white, and more likely to have had bilateral oophorectomy than non-HT users. Among HT users, ILS reduced FPG (p<0.01) and 2HG (p<0.01), and metformin reduced FPG (p<0.01) but not 2HG (p=0.56), compared to placebo. Associations between SHBG and total E2 with FPG and 2HG were not significant among women randomized to ILS or to metformin. These patterns differed from those observed among women who did not use HT.

Conclusions—We conclude that among glucose intolerant HT users, interventions to reduce glucose are effective but possibly mediated through different pathways than among women who did not use HT.

Keywords

menopause; estrogen; progestogen; hormone therapy; glucose

Introduction

Endogenous estradiol (E2) and sex hormone binding globulin (SHBG) have been associated with alterations in glucose levels.^{1–4} In the Diabetes Prevention Program (DPP),⁵ intensive lifestyle modification (ILS) and metformin reduced fasting plasma glucose (FPG) and 2-hour post-challenge glucose (2HG) among glucose-intolerant adults.⁵ Among postmenopausal DPP participants not using estrogen, ILS led to increases in sex hormone binding globulin (SHBG) which were associated with decreases in both FPG and 2HG.¹ Neither ILS nor metformin led to significant changes in E2 levels compared to placebo.¹

It has not been reported whether ILS and metformin reduce glucose levels among estrogen hormone therapy (HT) users, and whether SHBG or E2 are associated with glucose in HT users. The impact of ILS and metformin and mechanisms of action might differ between HT users and non-users for several reasons. The protective effects of endogenous SHBG and the detrimental effects of endogenous E2 might be obscured by HT use, which increases SHBG but also increases E2.⁶ Also, randomization to HT compared to placebo has been associated with both decreases in FPG as well as increases in 2HG.^{7–12} The mechanisms through which HT use might affect glucose levels are not known. In studies of HT use, the decreases in FPG persisted after adjustment for adiposity, suggesting that exogenous oral HT reduced FPG through other means, such as hepatic gluconeogenesis or hepatic insulin resistance.^{7, 12} HT use could increase postprandial glucose through reductions in whole-body insulin sensitivity,¹³ but at least one other study showed no association between HT use and insulin sensitivity.¹⁴

While HT use has declined after the publication of trials demonstrating an increased risk of morbidity,¹⁵ oral estrogens are still commonly prescribed for relief of menopausal symptoms. Therefore, the metabolic effects of HT remain important to examine, and potential mechanisms are relevant for understanding progression of glucose intolerance in both HT and non-HT users. In the DPP, participants randomized to ILS and metformin had maximal weight loss and reductions in glucose at 1-year after randomization.⁵ Changes in glucose were associated with changes in weight and insulin sensitivity.¹⁶ We examined whether postmenopausal women using oral HT at baseline and 1-year follow-up had

declines in glucose, and whether DPP treatments induced changes in E2 and SHBG that were associated with these changes in glucose. We also examined whether these patterns differed among women who did not use oral HT at either baseline or 1-year follow-up (Figure 1). We hypothesized that interventions would lead to reductions in both FPG and 2HG among HT users, but unlike in non-users, changes in SHBG and E2 would not be associated with these reductions. Finally, we examined whether associations between sex hormone changes and glucose changes persisted after consideration of changes in weight and insulin sensitivity.

Methods

Characteristics of DPP participants have been reported.⁵ Briefly, the DPP inclusion criteria included age 25 years, FPG of 95–125 mg/dl and 2HG of 140–199 mg/dl following a 75-glucose load, and body mass index (BMI) 24 kg/m² (22 kg/m² for Asian Americans). Written informed consent was obtained from all participants before screening, consistent with the guidelines of each participating center's institutional review board. Eligible participants recruited between 1996 and 1999 were randomly assigned to one of three interventions: 850 mg metformin twice daily, placebo twice daily, or ILS. The goals of ILS were to achieve and maintain a weight reduction of at least 7% of initial body weight through consumption of a low-calorie, low-fat diet, and to engage in moderate physical activity for at least 150 min/week.⁵ Weight and waist circumference were measured semiannually, and participants underwent an annual oral glucose tolerance test and semiannual FPG test. At the time of randomization, all women completed a questionnaire about their menses, gynecological history including surgeries, and exogenous HT use. Medication use was assessed every 6 months.

Women were classified as being postmenopausal if they met any of the following criteria: bilateral oophorectomy, lack of menses for at least one year without gynecologic surgery, cessation of menses prior to hysterectomy, cessation of menses within the past year and age

55 years, and cessation of menses with hysterectomy and age 55 years. For this report, we included women who were postmenopausal at randomization, had an available stored serum sample for E2 and SHBG measurement, and could be categorized as oral HT users both at randomization as well as at 1 year follow-up (n=324), or as non-HT users both at randomization and 1 year follow-up (n=382). Women who used injection, implant, transdermal, or transvaginal HT were excluded, as were women who used any HT at baseline but not at follow-up and vice-versa. We did not include women enrolled at Native American centers, as these women were not originally consented to participate in ancillary studies.

Baseline and year 1 assays were performed using the same batched assays. Glucose and insulin were measured as previously reported.¹⁶ Briefly, women were instructed to consume a usual diet and an oral glucose tolerance test was performed between 7 a.m. and 11 a.m. after an overnight fast. Blood was sampled from a vein before (fasting) and 2 hours after a 75-gram oral glucose load (Trutol 75; Custom Laboratories, Baltimore, MD). Plasma glucose was measured fasting and 2-hours, and plasma insulin was measured fasting. Insulin sensitivity was assessed using inverse fasting insulin levels.¹⁶ Insulin measurements were performed by a radioimmunoassay method using an anti–guinea pig antibody that measures total immunoreactive insulin. The assay is a 48-h polyethylene glycol–accelerated method with coefficients of variation (CVs) of 4.5% for high-concentration quality control samples and 6.9% for low-concentration quality control samples. The CV for masked split duplicates in this assay was <8.5%. SHBG, follicle stimulating hormone (FSH), and total E2 were measured as previously reported.¹ SHBG was measured at Endoceutics (Quebec City, Canada) using ELISA(Bioline) with interassay coefficients of variation of 7.8 and 5.0 at

18.2 and 63.1 nmol/l, respectively. FSH was measured at Endoceutics using ELISA (Bioline) with interassay coefficients of variation of 3.6 and 4.4 at 27.1 and 72.9 mIU/ml, respectively. E2 was analyzed using gas chromatography/mass spectrometry at Endoceutics.¹⁷ The limit of detection was 3.0 pg/ml for total E2 with an interassay coefficient of variation of 17.5 at 4.7 pg/ml. For measurements below the detection limit, values were extrapolated below the lower limit of quantitation using Mass Hunter Workstation software (Agilent, Santa Clara, CA).¹ We also performed sensitivity analyses where we assigned these values the equivalent of the lower limit of detection and found a similar pattern of results (not shown). Bioavailable E2 was calculated according to the method described by Sodergard and colleagues (macro courtesy of Frank Stanczyk, University of Southern California) taking the concentrationsof total E2 and SHBG into account and assuming a fixed albumin concentration of 4.0 g/dl.¹⁸

Statistical Analysis

Women who used HT at baseline and follow-up were analyzed separately from women who did not use HT at baseline and follow-up. For HT users and non-users, baseline characteristics were described using percentages for categorical variables and means (SD) for normally distributed quantitative variables. Skewed variables, including insulin levels, FSH, SHBG, total and bioavailable E2, were log-transformed before comparison. Changes in glucose, SHBG, total E2, and bioavailable E2 were calculated as year 1 level – baseline level. In order to assess the association between randomization assignment and change in FPG and 2HG as well as change in SHBG and E2 levels between baseline and year 1 follow-up, we used t-tests to compare levels of change between randomization arms. We also compared log-transformed SHBG and E2 levels and found a similar pattern of results. Models substituting the homeostasis model assessment of insulin resistance (HOMA-IR) for 1/fasting insulin had similar results (data not shown).

To determine whether changes in SHBG, bioavailable E2, or total E2 were associated with changes in glucose apart from changes in adiposity, we created a series of multiple linear regression models, with coefficients obtained by using ordinary least squares. Changes in the aforementioned sex hormones and associations with changes in FPG were examined after adjustment for baseline characteristics (age, race/ethnicity, baseline FSH, and baseline glucose levels), as well as changes in waist circumference and changes in 1/fasting insulin levels. We chose to adjust for weight and not physical activity, because in the DPP, reductions in body mass were the strongest predictor of reductions in glucose levels.¹⁹ While level of physical activity was associated with body mass, physical activity was not associated with reductions in glucose after adjustment for body mass.¹⁹ In other words, changes in weight mediated physical activity effects on glucose. We also chose to adjust for 1/fasting insulin due to previous reports suggesting that associations between sex hormones and glucose were mediated by fasting insulin.² In the DPP, weight and waist circumference were highly correlated, and we used waist circumference as a proxy for visceral adiposity.²⁰ Similar models were created that examined associations between changes in sex hormones and changes in 2HG. In sensitivity analyses, we examined only women using oral estrogen alone, without progestin. We observed similar results compared to women using oral estrogen and progestin (data not shown). We also examined models that did not include FSH or baseline glucose levels and observed similar results (data not shown). The SAS analysis system was used for all analyses (SAS Institute, Cary, NC).

Results

Baseline characteristics of the postmenopausal cohort of HT users and non-HT users are shown in Table 1. Reflecting DPP recruitment criteria, all women were overweight or obese at baseline and had elevated glucose levels. HT users were slightly younger and more often

Among HT users, there were no significant differences between women randomized to ILS (n=107), metformin (n=122), and placebo (n=95), with the exception that there were slightly more African-American women in the metformin arm than the placebo arm (9% vs. 5%, p<0.05). Of the oral estrogens that women reported using at baseline and follow-up, the most common was conjugated equine estrogen (Premarin), followed by 17- β -estradiol (Estrace), followed by esterified estrogen (Estratab, Menest) and estropipate (Ogen, Ortho-Est). Of the 211 women who had a hysterectomy, none were using progestin at baseline or follow-up. Of the 113 women who did not report a hysterectomy, all were using a progestin at baseline and follow-up. Among non-users, there were no significant differences between women randomized to ILS (n=133), metformin (n=122), or placebo (n=129) exception that women randomized to metformin had slightly lower FSH levels than women randomized to placebo (51.5 IU/1 vs. 59.3 IU/1, p<0.05).

Among both HT users and non-users, women randomized to ILS and metformin had declines in waist circumference compared to placebo (p<0.01 for all comparisons). The association between randomization arm and decreased waist circumference persisted after adjustment for age, race/ethnicity, and FSH levels (results not shown). Changes in glucose levels between baseline and year 1 by randomization arm are presented in Figure 2. Among HT users, women randomized to ILS had significant reductions in FPG (p<0.01) and 2HG (p<0.01) compared to placebo. Among HT users, women randomized to metformin had significant reductions in FPG (p<0.01) but not 2HG (p=0.56) compared to placebo. This pattern of results was similar across different types of estrogen (results not shown). Among non-users, women randomized to ILS had significant reductions in FPG (p<0.01) and 2HG (p=0.03) compared to placebo, but women randomized to metformin did not have significant reductions in FPG (p=0.20) or 2HG (p=0.50) compared to placebo. These associations between randomization arm and glucose levels persisted after adjustment for age, race/ ethnicity, and FSH levels (results not shown).

Changes in SHBG, total E2, and bioavailable E2 between baseline and year 1 by randomization arm are presented in Figure 3. Among HT users, women randomized to ILS or metformin did not have significant changes in SHBG (p=0.11 and p=0.16, respectively) or total E2 (p=0.46 and p=0.15, respectively) compared to placebo. However, women randomized to ILS (p=0.046) and metformin (p=0.042) did have significant reductions in bioavailable E2 compared to placebo. Among non-users, women randomized to ILS had significant increases in SHBG compared to placebo (p<0.01), but women randomized to metformin (p=0.58) did not have significant changes in SHBG. Among non-users, women randomized to ILS or metformin did not have significant changes in total E2 (p=0.36 and p=0.10, respectively) or bioavailable E2 (p=0.54 and p=0.11, respectively). This pattern of associations between randomization arm and SHBG and E2 levels remained similar after adjustment for age, race/ethnicity, and FSH levels (results not shown).

Table 2 shows the associations between changes in SHBG, total E2, and bioavailable E2 with changes in glucose. Among HT users who were randomized to ILS, there were no associations between changes in SHBG or E2 with changes in FPG or changes in 2HG.

Among HT users who were randomized to metformin, reductions in bioavailable E2 were associated with reductions in 2HG. This association persisted after further adjustment for changes in 1/fasting insulin (p-value=0.04), suggesting that the association between bioavailable E2 and 2HG was not mediated through adiposity or 1/fasting insulin. However, overall, significant reductions in 2HG among metformin users vs. placebo did not occur (Figure 2).

Among non-users (Table 2), increases in SHBG were associated with declines in FPG and 2HG after adjustment for waist circumference, and these associations persisted after further adjustments for change in 1/fasting insulin (p<0.05 for both associations). Among non-users, declines in total and bioavailable E2 were associated with declines in FPG, and this association persisted after further adjustment for change in 1/fasting insulin (p<0.05). However, overall, significant reductions in total E2 and bioavailable E2 among women randomized to interventions vs. placebo did not occur (Figure 3). Among non-users, excluding women who had undergone oophorectomy, ILS still led to declines in SHBG compared to placebo (p<0.01), and declines in SHBG were still associated with significant declines in FPG (p=0.01) although the association with declines in 2HG was attenuated (p=0.14).

Discussion

In a secondary analysis from the Diabetes Prevention Program, a randomized placebocontrolled trial, we observed that ILS led to reductions in both fasting and post-prandial glucose among HT users and non-users. However, ILS effects on SHBG and E2 differed between HT users and non-users, and SHBG and E2 had associations with glucose that differed by HT use. These findings suggest that the roles of SHBG and E2 in influencing glucose levels depend upon whether these hormones are exogenous or endogenous. Among HT users, ILS did not change SHBG, and SHBG was not associated with changes in FPG or 2HG. In contrast, among non-users, ILS led to increases in SHBG which were associated with decreases in glucose even after adjustment for adjposity and insulin sensitivity. Among HT users, ILS led to significant reductions in bioavailable E2 compared to placebo, although these changes were not associated with declines in glucose. Among non-users, ILS did not change E2 levels, although decreases in E2 were associated with decreases in FPG. Among HT users, the declines in FPG observed among metformin users did not appear to be associated with changes in SHBG, bioavailable E2, or E2. Among non-users, metformin had minimal impact upon glucose levels. In other words, the relationship between E2, SHBG and glucose were dissociated in HT users compared to non-users, even as interventions were still effective in reducing glucose in both HT users and non-users.

Our findings are notable for several reasons. First, we found that lifestyle modification can still reduce postprandial glucose levels in the context of HT use. In the Postmenopausal Estrogen/Progestin Intervention Study, randomization to HT was associated with significant reductions in FPG and fasting insulin, but with elevations in 2HG.⁷ The effects of exogenous estrogen on 2HG were not mitigated by lifestyle behaviors.⁷ Our results may have differed in that the DPP examined a glucose intolerant population, and any additional negative effects of HT on glucose tolerance may not have been as apparent. Also, women were randomized to lifestyle change which in turn resulted in significant reductions in weight and glucose levels in HT users, although SHBG was associated with reductions in both FPG and 2HG in non-users.¹ Among non-users, hepatic steatosis may be a common antecedent of SHBG and glucose, or SHBG may influence hepatic glucose production apart from steatosis.²¹ HT use itself was associated with almost a tripling of

SHBG levels, and it is possible that further increases in SHBG resulting from ILS had minimal effects on glucose levels. Third, we found that bioavailable E2 declined among HT users randomized to ILS and metformin compared to placebo, although we did not observe these changes among non-users. Baseline E2 levels were lower among non-users compared to HT users, and it is possible that further reductions in bioavailable E2 were minimized by these low levels. However, among HT users, these reductions in bioavailable E2 were not significantly associated with reductions in glucose. Even though reductions in bioavailable E2 were did not have statistically significant declines in 2HG overall.

To our knowledge, no reports have examined the impact of metformin in postmenopausal women without polycystic ovarian syndrome. Among HT users in this report, metformin had favorable effects on FPG with less marked reductions in 2HG. Metformin effects upon FPG did not appear to be mediated by metformin effects upon E2 levels, since total E2 and bioavailable E2 were not associated with FPG among HT users. Thus, sex hormone effects do not appear to be an important pathway for metformin effects on glucose among postmenopausal women using exogenous estrogen. Among non-HT users, metformin effects upon FPG and 2HG were minimal, consistent with other DPP studies demonstrating reduced effectiveness of metformin in older populations.²² While metformin has decreased sex steroid levels in women with polycystic ovarian syndrome,²³ sex steroid levels including estradiol were higher in women with polycystic ovarian syndrome than the postmenopausal women in this report. Thus, for different reasons compared to HT users, sex hormones do not appear to be an important pathway for metformin effects on glucose among postmenopausal women not using estrogen.

Limitations of this report are that women were not randomized to HT use as well as to lifestyle intervention. To assess optimally the interactions between HT use and glucosereduction interventions, randomization to HT use as well as to ILS, metformin, and placebo would have been necessary, although such a study is unlikely to be conducted. As HT use was strongly associated with surgical menopause, particularly bilateral oophorectomy, this would have required additional adjustment for indications of surgical menopause, which would have been logistically challenging. Therefore, despite uniform inclusion criteria for weight and glucose elevations, HT users differed from non-users regarding demographic characteristics, anthropometrics, and metabolic measurements, and these characteristics as well as other unmeasured characteristics may have led to the different roles of E2 and SHBG in glucose changes. It is possible that residual confounding from adiposity or insulin sensitivity occurred as the proxy measures used were not direct measures. HT was assessed via recording of medications and may have over- or underestimated HT use. While overweight and obese women tend to have lower FSH levels than normal weight women in the postmenopause,²⁴ some of the non-estrogen users may have been misclassified as being postmenopausal as opposed to premenopausal, and in turn, this may have increased or decreased observed associations. Although the analyses did not differ when we examined estrogen users only vs. estrogen-progestin users, progestin type may have altered the hormonal milieu. Finally, the women studied were already overweight and glucose intolerant, and our results may not extend to healthier women. Strengths of our report include its randomized design for glucose-reducing interventions, interventions which successfully reduced waist circumference, and performance of post-challenge as well as fasting glucose.

Conclusions

Our hypothesis-generating report suggests that glucose-lowering interventions might act via different mechanisms in estrogen users and non-users, and specifically that glucose-

reduction interventions are effective among HT users, and that the role of E2 and SHBG in glucose change differs depending upon whether women use HT. Intervention-associated increases in SHBG may be less important among women who use HT in part due to significant elevations in SHBG associated with HT. While there were reductions in bioavailable E2 among HT users with lifestyle change, the clinical significance of this finding is not clear. Metformin may be less effective for reducing post-challenge glucose among postmenopausal women than ILS, but this effect is observed among non-users as well as HT users and is likely not due to adverse effects of HT.

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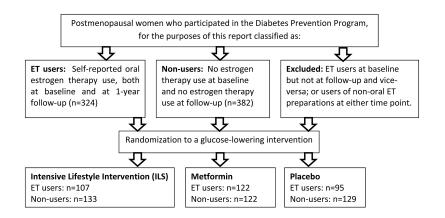


Figure 1.

Secondary analysis design. We conducted an analysis of the effectiveness of glucoselowering interventions in a randomized trial (the Diabetes Prevention Program), among postmenopausal women who were either 1) oral estrogen users at baseline and at 1-year follow-up or 2) non-estrogen users at baseline and at 1-year follow-up. Kim et al.

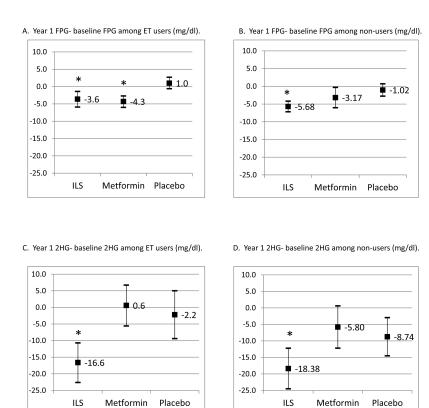
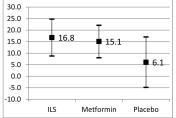


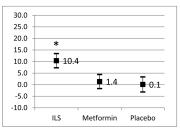
Figure 2.

Changes in fasting plasma glucose (FPG) and 2-hour post-challenge glucose (2HG) between baseline and year 1 follow-up, among estrogen therapy (ET) users and non-users. Means and 95% confidence intervals shown; * indicate significant differences from placebo arm.

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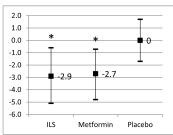


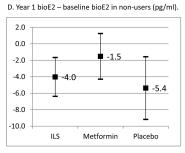




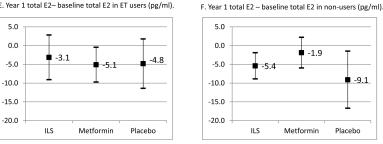
B. Year 1 SHBG - baseline SHBG in non-users (nmol/l).

C. Year 1 bioE2 – baseline bioE2 in ET users (pg/ml).





E. Year 1 total E2- baseline total E2 in ET users (pg/ml)



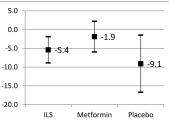


Figure 3.

Changes in select sex hormone measures (sex hormone binding globulin or SHBG, bioavailable estradiol or bioE2, total estradiol or total E2) between baseline and year 1 follow-up, among estrogen therapy (ET) users and non-users. Means and 95% confidence intervals shown; * indicate significant differences from placebo arm.

Table 1

Baseline characteristics of postmenopausal women by oral estrogen therapy (ET) use.

| | <u>Women with oral ET use at baseline</u> <u>and at follow-up</u> | <u>Women with no ET use at baseline and no</u> <u>ET use at follow-up</u> | |
|--|--|--|--|
| | n=324 | n=382 | |
| Age (years) * | 56.5 (7.6) | 58.7 (9.0) | |
| Race/ethnicity * | | | |
| Caucasian | 66 | 53 | |
| African-American | 16 | 28 | |
| Hispanic | 13 | 16 | |
| Asian | 4 | 3 | |
| Type of menopause * | | | |
| Bilateral oophorectomy | 42 | 20 | |
| Natural menopause | 39 | 67 | |
| Age 55 years and hysterectomy | 19 | 16 | |
| Years since final menstrual period | 14 (9) | 15 (10) | |
| Baseline weight (kg)* | 87.6 (17.8) | 91.0 (19.7) | |
| Baseline waist circumference (cm)* | 101 (14) | 104 (14) | |
| Baseline BMI (kg/m ²)* | 33.3 (6.5) | 34.6 (6.8) | |
| Baseline fasting insulin levels (IU/l)* | 23.4 (12.0) | 26.4 (16.0) | |
| Baseline fasting plasma glucose (mg/dl)* | 104 (7) | 107 (8) | |
| Baseline 2-hour glucose | 166 (18) | 164 (17) | |
| Baseline follicle stimulating hormone (IU/l)* | 34.8 (22.5) | 55.3 (26.6) | |
| Baseline sex hormone binding globulin $(nmol/l)^*$ | 85.3 (77.0) | 33.2 (18.3) | |
| Baseline total estradiol (pg/ml) * | 17.6 (14.8) | 8.5 (8.0) | |
| Baseline bioavailable estradiol (pg/ml)* | 8.6 (9.4) | 6.0 (6.4) | |

Means (SD) or percentages shown; medians (interquartile ranges or IQR) shown for sex hormones.

* indicates significant difference (p<0.05) between ET users and non-users.

Table 2

Associations between changes (Δ) in glucose (dependent variable) and sex hormone binding globulin (SHBG), bioavailable estradiol (E2), and total E2. ET indicates estrogen therapy, ILS indicates intensive lifestyle change, and FPG indicates fasting plasma glucose. Adjusted β coefficients (p-values) shown.^{*}

| | Women with oral ET use at baseline and at follow-up | | Women with no ET use at baseline and no ET use at follow-up | |
|--|---|--------------|---|--------------|
| | ILS | Metformin | ILS | Metformin |
| Association between \triangle SHBG and \triangle FPG | -0.01 (0.69) | -0.04 (0.12) | -0.12 (<0.01) | -0.05 (0.75) |
| Association between Δ bioavailable E2 and Δ FPG | 0.14 (0.51) | 0.08 (0.55) | 0.23 (0.02) | 0.14 (0.56) |
| Association between Δ total E2 and Δ FPG | 0.10 (0.36) | 0 (0.99) | 0.13 (0.048) | 0.08 (0.60) |
| Association between ∆ SHBG and ∆ 2-hour glucose | -0.07 (0.39) | -0.14 (0.11) | -0.39 (0.04) | -0.08 (0.82) |
| Association between ∆ bioavailable E2 and ∆ 2-hour glucose | 0.02 (0.96) | 1.1 (0.04) | 0.62 (0.14) | -0.04 (0.94) |
| Association between Δ total E2 and Δ 2-hour glucose | -0.13 (0.60) | 0.43 (0.06) | 0.37 (0.19) | -0.07 (0.83) |

Adjusted for age, race/ethnicity, baseline follicle stimulating hormone levels, baseline glucose levels, and changes in waist circumference.