

Published in final edited form as:

*J Vasc Res.* 2012 ; 49(5): 375–389. doi:10.1159/000338747.

## CONTRIBUTION OF FLOW-DEPENDENT VASOMOTOR MECHANISM TO THE AUTOREGULATION OF CEREBRAL BLOOD FLOW

Akos Koller<sup>1,2</sup> and Peter Toth<sup>1,2</sup>

<sup>1</sup>Department of Pathophysiology and Gerontology, Medical School, University of Pécs, Pécs, Hungary

<sup>2</sup>Department of Physiology, New York Medical College, Valhalla, New York, USA

### Abstract

Regulation of cerebral blood flow (CBF) is the result of multilevel mechanisms to maintain blood supply for the brain, in the same time it has to comply with the limited space available in the cranium. This latter requirement is ensured by the autoregulation of CBF, in which the pressure-sensitive myogenic response is known to play a pivotal role. However, in vivo increases in pressure are accompanied by increases in flow; the effects of flow on the vasomotor tone of cerebral vessels are less known. Earlier studies, showing flow-sensitive dilation and/or constriction or both, thus no clear picture emerged. Recently an important role of flow sensitive mechanism(s) eliciting constriction of cerebral vessels has been demonstrated, thus this review focuses on the effect of hemodynamic forces (especially intraluminal flow) on the vasomotor tone of cerebral vessels and the underlying cellular and molecular mechanisms. A novel concept of autoregulation of CBF is proposed suggesting that (in certain areas of the cerebrovascular tree) pressure- and flow-induced constrictions together maintain an effective autoregulation, and alterations of these mechanisms may contribute to the development of cerebrovascular disorders. Future studies are warranted to explore the signals, details of signaling processes and in vivo importance of these mechanisms.

### Keywords

flow-induced response; myogenic response; 20-HETE; ROS; cerebral circulation

### Introduction

Regulation of cerebral blood flow (CBF) is of utmost importance to supply the myriad function of the brain. It has to ensure many roles: appropriate supply of nutrition and gas for cerebral tissue, fluid and gas exchange in the capillaries, maintenance of cerebral blood volume, thus intracranial volume and pressure in a very limited range. Obviously, such a complex function requires complex regulatory mechanisms ensured by the dynamic interaction of mechanotransduction, metabolic (for example adenosine), chemical (such as changes in pCO<sub>2</sub>, pH and pO<sub>2</sub>) and other factors, for example, recently there are emerging evidence regarding an important role for glial (astrocytes), pericytes and neural control of CBF [1–14].

---

**Correspondence:** Akos Koller M.D., Ph.D., Department of Pathophysiology and Gerontology Medical School University of Pécs, H-7624 Pécs, Szigei Str. 12., Hungary, akos.koller@aok.pte.hu, Tel: +36-30-338-4496.

Because all of these mechanisms have to comply with limited space in the closed cranium [15], maintenance of a relatively constant cerebral blood flow despite of the variations in pressure and flow historically has been always in the center of investigation. Although, autoregulation of CBF is used to be explained by the pressure sensitive myogenic mechanism (here we do not include metabolic and other factors in the notion of autoregulation, we define autoregulation as the sum of the mechanisms activated in response to changes in blood pressure) [4–7, 16–19], careful analysis of earlier data and recent findings question the sole role of myogenic mechanism eliciting autoregulation of CBF [20–24].

Thus, in this review we focus on the potential flow-dependent autoregulatory mechanisms and provide an explanation of the different results of previous studies regarding flow-induced diameter changes of cerebral vessels.

## I. Autoregulation of CBF

### 1) General considerations

Total cerebral blood flow has to be relatively constant in order to allow a stable and continuous supply of cerebral tissue and maintain intracranial volume and pressure constant. On the basis of Hagen-Poiseuille law it can be assumed that CBF is related to the 4<sup>th</sup> power of vessel radius, thus an increase in the diameter of vessels elicits an *exponential* increase in blood flow. Therefore in the closed cranium, a general vasodilatation would lead to substantial increase in CBF and cerebral blood volume (CBV) and would lead to elevation of intracranial pressure (and vice versa) [19, 25]. Thus, tight control of CBF and CBV is essential for the organism. Indeed, in a wide range (from ~ 60 to 140 mmHg) of perfusion pressure CBF increases only slightly in a *linear* manner measured by different in vivo techniques [5, 18]. At this point it has to be noted, that however in mathematical models gain = 1 is used to indicate so called perfect autoregulation [22, 23], as also depicted in Figure 2, it is likely that such perfect horizontal relationship does not exist in vivo and it would not be beneficial regarding appropriate supply of brain tissue with blood, either. Rather, as Rosenblum suggested it is likely that the slope increases linearly as pressure and flow increases [5, 18, 26, 27]. The linear and not exponential (!) increase of CBF in the face of increasing blood pressure is achieved by the mechanisms of cerebral autoregulation. In this review we do not include metabolic factors into the term of autoregulation, however by some authors metabolic factors/mechanisms are also considered to contribute to “autoregulation” of CBF. This interpretation of “autoregulation” means adjusting CBF to the metabolic demands and to function of neural tissues. Although metabolic regulation of CBF [2, 5, 13, 14, 28] is obviously an important issue, in this review we define autoregulation to be vasomotor responses to changes in hemodynamic forces achieved by mechanisms intrinsic to the vascular wall, rendering the cerebral blood perfusion independent from changes in systemic blood pressure, and without activating metabolic, chemical, glial, neural and other (for example capillary blood flow regulation by pericytes) regulatory mechanisms [1, 2, 5–11, 18, 19, 29]. Because changes in pressure are accompanied by changes in flow, in vivo responses of cerebral vessels to changes in hemodynamics are most likely a combination of pressure and flow-induced mechanisms [30–33]. Thus one can hypothesize that changes in flow contributes to autoregulation of CBF. In other words, in vivo when systemic pressure changes diameter responses of cerebral vessels thus changes of CBF are determined by the combined effect of pressure and flow.

### 2) Regional and segmental differences in autoregulation of CBF

Autoregulation of cerebral blood flow was documented in a variety of animal species and human using in vivo techniques, such as clearance of diffusible indicators, observation of

pial vessels, measurements of arterial inflow and venous outflow, autoradiography, distribution of radioactive microspheres, and recently by various magnetic resonance imaging, ultrasound and other techniques [18, 34–42].

Importantly, in the cerebral circulation large arteries represent a significant part ( $\approx 40\%$ ) of total cerebrovascular resistance [9, 18, 43–45]. In the late 1970's, Kontos et al. investigating the cat pial circulation found that large surface vessels from the circle of Willis to pial arterioles up to 200 micrometer contribute  $\approx 30\%$  of cerebrovascular resistance at 120 mmHg intraluminal pressure and proposed that these vessels are exclusively responsible for autoregulation between 120 and 160 mmHg intraluminal pressure [18]. Similar findings were observed in the rat cerebral circulation [44]. These results showing that in the cerebral circulation both large arteries and small arterioles take part in regulation of CBF indicated that there might have been a hierarchy in vascular responsiveness, a basis for more complicated regulation. This also raised the possibility of spatial characteristics of cerebral autoregulation. Indeed, regional differences were shown in cerebral autoregulatory capacity: hypertension exceeded autoregulation in the cerebrum of the cat (supplied by the internal carotid system), but not in brain stem (supplied by the vertebro-basilar system) demonstrating that autoregulation of flow is more effective in the vasculature of the brain stem [41, 46, 47]. In human cerebral circulation regional differences have also been shown regarding autoregulatory function [48, 49].

However, different magnitude of response in cerebral arteries and arterioles to elevation in pressure was already shown by Kontos et al. [18], later it was proposed again then proved by Faraci and Heistad et al. that different segmental responses of cerebral vessels to changes in intraluminal pressure underlies the heterogeneity of autoregulation. They demonstrated that pial arteriolar pressure is greater in brain stem than in the cerebrum implying that pressure drop is greater on larger arteries in the cerebrum than in the brain stem. Also they found that resistance of larger arteries increase in the cerebrum and decrease in the brain stem during moderate increase in blood pressure. They concluded that autoregulation depended on arterioles in the brain stem, but in the cerebrum larger vessels play an important role in autoregulation [50, 51]. These findings are in line with the findings of Kontos et al. that in the cerebrum larger arteries play a significant part in autoregulation.

### 3) Physiological role

Collectively, autoregulation ensures that CBF remains relatively constant in a wide range despite increase or decrease of systemic blood pressure. In vivo, autoregulation of CBF shows regional and segmental differences. In the brain stem (supplied by the vertebro-basilar system) resistance of larger arteries decreases whereas that of arterioles increases in response to elevation in pressure. In contrast, in the cerebrum (supplied by the internal carotid artery system) larger arteries increase their resistance to elevation in pressure greater than the arterioles, playing primary role in autoregulation, and protecting downstream brain circulation from pressure and volume overload [43, 50, 52].

It is important to note however, that in these in vivo studies of autoregulation of CBF and cerebrovascular response to pressure the effects of pressure and flow could not be separated and the effect of flow on the diameter of vessels was not even considered [18, 34, 36–43, 50, 53].

## II. Pressure-induced myogenic responses of cerebral vessels

### 1) Earlier findings and conclusions

Until very recently, cerebral autoregulation (relative independence of CBF from changes in systemic blood pressure, especially at higher pressure values) is primarily explained by the

pressure-induced myogenic response [52]: the inherent property of vascular smooth muscle to dilate to decrease and to constrict to increase in intraluminal pressure. Since its first description by Bayliss [35], early in the 20<sup>th</sup> century the myogenic response of different arteries has been widely investigated. In the cerebral circulation it was first showed by Fog, Forbes and Wolff that pial arterioles of the cat actively dilate and constrict to changes of blood pressure [8, 54, 55]. These findings were confirmed later in studies mentioned above investigating the regulation of CBF, measuring also the simultaneous changes of blood flow, arteriolar pressure etc.

First in the 1980's and since that time, presence of myogenic response has been also shown in vitro in isolated cerebral arteries [22, 56]. In these in vitro studies investigating the myogenic response only pressure was changed, flow was kept constant (this could be done due to the development of novel technics in which by cannulation of vessel on both ends the intraluminal pressure could be raised by increasing both the inflow and outflow pressure or by closing the outflow end of the vessel and increasing the inflow pressure. In both cases the intraluminal flow remains unchanged) [57–59]. Therefore the observed diameter responses were due to changes in pressure alone and were not influenced by changes in flow. Interestingly, in many of these studies cerebral vessels only maintained a constant diameter between 60–140 mmHg intraluminal pressure [20–22, 24]. This response is referred as the 2<sup>nd</sup> phase of in vitro arterial myogenic behavior proposed by Osol et al [21]. If however, one extrapolated this finding to in vivo conditions, because of the constant diameter, increasing pressure would result in an increasing blood flow velocity thus increasing CBF. In contrast, in vivo measurements of CBF did show that CBF remained relatively constant while intraluminal pressure (and flow velocity) increased! These observations should have prompted the investigators to hypothesize the existence of a flow sensitive mechanism, as well, resulting in further constriction.

However, in many vascular beds the myogenic constriction alone likely prevents increases in blood flow during increases in pressure, for example in the cremaster muscle circulation [60]. Although, the direct comparison of cerebral vessels to peripheral vessels has to be carried out carefully, for instance, Bohlen and Harper [61] and Meininger et al. [62] found that magnitude of myogenic constriction in arterioles of the cremaster muscle is greater than in the comparable sized arterioles of the rat cerebral cortex.

Also, the strength of the myogenic constriction or dilation can be modulated by other effects. For example, hypertension and exercise can enhance the myogenic response leading to increased constriction to increase in intraluminal pressure [22, 61, 63, 64]. Also, myogenic dilation can increase significantly under certain conditions. For instance, arterioles in the cat sartorius muscle dilate significantly more to decrease in intraluminal pressure during sympathetic nerve stimulation, enhancing myogenic dilation and providing increasing flow [65]. Although, in case of increasing blood pressure in cerebral vessels the myogenic constriction is considered to be the most important mechanisms maintaining constant or near constant blood flow [52, 63, 66]. In contrast, as we mentioned above, in most cerebral vessels the myogenic response does not reduce the diameter of vessels beyond the basal level in response to elevation of intraluminal pressure [20–24]. It cannot be excluded however, that magnitude of the myogenic constriction have regional and size differences, and that these vessels possess strong and efficient myogenic response. At present however the investigation of myogenic response of microvessels deeply seeded in the brain is technically challenging and the presence of confounding mechanisms in vivo makes it difficult to interpret the findings. The ratio of the magnitude of pressure (and flow-induced, see later) responses in different brain regions and their contribution to cerebral autoregulation should be clarified in the future.

## 2) Molecular mechanisms of the myogenic response of cerebral vessels

Although there are several excellent reviews [67–71] of molecular mechanisms of myogenic response, we felt that it would be still appropriate to give a brief overview here.

It is now widely accepted that the primary stimulus to trigger the myogenic response is the pressure-elicited stretch of vascular wall, leading to increased wall tension [72]. However, the sensors are still in question: among others stretch-activated cation channels, Gq-coupled receptors, and interactions between matrix metalloproteinases, extracellular matrix, integrins and the cytoskeleton were proposed [73–76].

The first event in the mechanotransduction is the depolarization of smooth muscle membrane [77]. Although, there is no consensus regarding the initiators of membrane depolarization, stretch-activated cation channels (TRPC6, TRPM4), calcium-activated potassium channels and chloride channels are probably involved [78–83]. Activation of voltage-gated potassium channels limits depolarization as a negative-feedback mechanism [84].

Depolarization of the membrane then leads to the opening of voltage-gated  $\text{Ca}^{2+}$  channels (which are also capable to respond to stretch directly [85]) and thus elevated inward  $\text{Ca}^{2+}$  current [86, 87].  $\text{Ca}^{2+}$  can be released from sarcoplasmic stores, as well; however their role in myogenic response seems to be minor [85, 88]. The increased  $\text{Ca}^{2+}$  concentration via  $\text{Ca}^{2+}$ -calmodulin complex leads to myosin light-chain kinase (MLCK) activation. MLCK phosphorylates myosin light chain ( $\text{MLC}_{20}$ ) leading to increased actin-myosin interaction and consequent shortening of smooth muscle cell [89–91]. Because of the circumferential orientation of the smooth muscle cells shortening is translated into constriction of the vessel.

Interestingly, after the initial elevation the intracellular  $\text{Ca}^{2+}$  concentration does not increase, but the vessel is still constricting [21]. Among others this observation resulted in the discovery of novel  $\text{Ca}^{2+}$ -independent mechanisms of the myogenic response which can sensitize the constrictor apparatus to  $\text{Ca}^{2+}$ . As it was mentioned an increased actin-myosin interaction and consequent shortening of smooth muscle cell depends on the phosphorylated state of  $\text{MLC}_{20}$ , which is governed by myosin light-chain kinase (MLCK) and myosin light-chain phosphatase (MLCP) [71, 92].  $\text{Ca}^{2+}$ -independent mechanisms altering  $\text{Ca}^{2+}$  sensitivity of smooth muscle cell act through regulation of MLCP. PKC activation, production of diacylglycerol, RhoA/Rho kinase are involved in these processes [71, 92–94].

As a second messenger, 20-hydroxyeicosatetraenoic acid (20-HETE) was also found to play a role in myogenic response (also in other vascular beds) [3, 95–97]. 20-HETE is capable to inhibit large conductance  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels and to increase influx of  $\text{Ca}^{2+}$  via L-type  $\text{Ca}^{2+}$  channels [3, 96, 98]. Also, one of the major molecular targets of 20-HETE is PKC [3, 96, 98, 99].

Collectively, there are two critical mechanisms ( $\text{Ca}^{2+}$ -dependent and independent) contributing to myogenic constriction: 1) increase in pressure via increase in wall tension and smooth muscle cell stretch leads to membrane depolarization,  $\text{Ca}^{2+}$  influx and constriction via MLCK and phosphorylation of  $\text{MLC}_{20}$ . 2) In the same time,  $\text{Ca}^{2+}$ -independent mechanisms involving PKC, diacylglycerol, RhoA/Rho kinase and 20-HETE regulate the activity of MLCP determining the phosphorylated state of  $\text{MLC}_{20}$  thus sensitizing actin-myosin to  $\text{Ca}^{2+}$ .

## III. Flow-induced responses of cerebral vessels

As we have mentioned above, changes in pressure are accompanied by changes in flow [30–33], and based on theoretical considerations flow-induced mechanisms may play a role in

cerebral autoregulation. Interestingly, flow-induced responses of cerebral vessels varied between species, vessel types and methods used. In theory, flow-induced dilation would reduce the magnitude of myogenic constriction of cerebral vessels, which would reduce the gain of autoregulation of CBF, whereas if flow elicited constriction, it could contribute to a more efficient autoregulation of CBF. In the following we summarize the results of studies regarding flow-induced responses of cerebral vessels.

### 1) Diameter responses of isolated cerebral vessels differ to increases in flow

**a) Dilation of cerebral vessels to flow**—First Fujii et al. showed that basilar artery of rat dilated *in vivo* when intraluminal flow was increased [100, 101]. They used craniotomy to visualize basilar artery. Metabolic parameters (blood O<sub>2</sub> and CO<sub>2</sub>, pH) were maintained at a constant physiological level, and blood pressure was kept constant by controlled bleeding of the animal. Intraluminal flow was elevated in the basilar artery by bilateral carotid artery occlusion. It is of note, that among *in vivo* methods this approach is able to separate the effect of flow and pressure on vessel diameter, while other factors (i.e. metabolic) are controlled. However it cannot be entirely excluded that occlusion of the carotid arteries, which decreases blood flow in the carotid system would generate a signal propagating to basilar artery to elicit dilation. The group of Sobey confirmed dilation of basilar artery using the same method [102]. In 1993, Gaw and Bevan found also dilation of rabbit cerebral vessels using a wire myograph [103], in which however flow and pressure cannot be controlled properly. Recently, Drouin et al. demonstrated flow-induced dilation in cerebral arteries. They studied isolated posterior and anterior cerebral arteries of mice in a pressure-flow chamber in a well-controlled manner [104, 105]. In recent *in vitro* studies (unpublished observation) we confirmed that in isolated rat basilar arteries increases in flow elicit dilation. An original record of such response is depicted on Figure 1, showing substantial dilation of an isolated basilar artery of rat to increases in flow.

**b) Biphasic response of cerebral vessels to flow**—Interestingly, there have been studies, in which biphasic response, dilation and constriction of cerebral vessels to increases in flow was observed. Garcia Roldan and Bevan showed that isolated rabbit pial arteries dilated to 20  $\mu$ l/min flow at 30 mmHg and they constricted to the same flow at 90 mmHg of intraluminal pressure [106, 107]. They used a Halpern perfusion system, in which at a constant pressure, flow was generated by pressure difference due to changing the perfusion by the inflow and outflow pumps.

Thorin-Trescases et al. confirmed these findings also in rabbit cerebral vessels. They found in secondary and tertiary branches of posterior cerebral arteries that flow induced dilation at 40 mmHg, dilation and a small constriction at 60 mmHg, and a small dilation followed by constriction at 80 mmHg intraluminal pressures [108]. Ward et al. observed similar pressure dependency of flow-induced response of cerebral vessels: in arteriolar branches ( $\approx$  80  $\mu$ m) of the rat posterior cerebral artery flow elicited dilation at 60 mmHg, and constriction at 120 mmHg intraluminal pressure [109].

Ngai and Winn studied arteriolar branches of rat middle cerebral arteries (MCA), approximately 35–88  $\mu$ m in diameter. They achieved increasing flow at a constant (60 mmHg) pressure by changing the inflow and outflow pressure to an equal degree, but opposite direction (the same method used by us when constrictions were observed in rat MCA and human intracerebral arteries to increases in flow). They also observed that isolated cerebral arterioles, branches of MCA dilated to flow up to 10  $\mu$ l/min, then restored their diameter at higher flow rates to the initial value [110].

Similar flow-rate dependency was found by Shimoda et al. Anterior and middle cerebral arteries of the neonatal pig constricted to flow between 0.077–0.212 ml/min, and dilated



when flow was raised further (up to 1.6 ml/min). There were no differences in the flow-induced responses at 20 and 60 mmHg intraluminal pressures. In these studies flow was initiated by a syringe pump, and pressure was measured in both the inflow and outflow cannulas by transducers. When flow was raised a micromanipulator decreased the outflow pressure adjacent to the increase in pressure due to flow by sensing the pressure difference between the transducers. The luminal pressure was also measured by inserting a cannula into the lumen of the vessel [111].

In a study Garcia Roldan and Bevan found the mixture of pressure and flow-rate dependency of flow-induced response of cerebral vessels. In isolated rabbit pial arterioles they demonstrated that vessels constricted in response to increase in intraluminal flow from 0 to 20  $\mu$ l/min at 90 mmHg, but did not at the presence of 60 mmHg of intraluminal pressure. Increasing flow up to 100  $\mu$ l/min constricted vessels at both pressure values [107]. It has to be clear, that biphasic response of diameter to flow does not mean that vessels exhibit opposite responses in a temporal manner, instead it reflects different responses at different pressure values or flow rates.

**c) Constriction of cerebral vessels to increases flow**—The variation of observed diameter responses of cerebral vessels in response to increases in flow prompted further investigations. Madden at al. studied isolated cat middle cerebral arteries. They applied 1–4 ml/min flow at 70 and 100 mmHg intraluminal pressures, and found that increases in flow led to decrease in diameter and depolarization of smooth muscle cell membrane. Arteries constricted by flow dilated when PCO<sub>2</sub> was increased, implying that metabolic/chemical signals can override flow-induced constriction. Also, when flow was stopped vessels dilated, and when flow was suddenly increased to the maximal values constriction occurred. Endothelium denudation did not affect flow response, suggesting that flow-induced constriction is endothelium-independent [112].

Sipkema at al. found flow-induced constriction in isolated basilar artery of the Rhesus monkey and in order to make sure that the conditions did not influence the response in the same set up they used isolated femoral arteries as well in which they observed dilation to flow. However, in many of these earlier studies changes in pressure during changes in flow could not be excluded with great certainty. Therefore constriction in these studies to flow could be elicited by the activation of myogenic mechanism, whereas dilation of femoral artery to flow might be caused by flow-induced mechanism, and/or passive dilation (femoral arteries did not develop spontaneous tone). This conclusion is supported by the finding that the active pressure-diameter curves of these vessels did not differ from the passive diameter obtained in calcium free conditions [113]. However, to our best knowledge this is the only study investigating isolated cerebral arteries of monkeys, thus it cannot be excluded that constriction of the basilar artery to flow may be due to species characteristic.

In 2001 Bryan at al. aimed to clarify the controversial findings (dilation, constriction and biphasic responses) regarding flow-induced responses of cerebral vessels. They proposed that controversy and confusion in previous studies were due to different species, different vessel types, different techniques and technical problems causing artifacts, namely the different pH of the extraluminal and intraluminal bath (cerebral vessels are highly sensitive to pH), and the pressure-change accompanying the increase in flow. Therefore in the same experimental conditions they investigated isolated middle cerebral arteries (MCA), penetrating cerebral arterioles (PA) ( $\approx$  70  $\mu$ m) and cremaster muscle arterioles (CMA) (which are known to dilate to flow) of rats. They used a syringe pump to generate flow, and decreased the outflow pressure according to the elevation in pressure due to flow both counted by an algorithm based on the resistance of the tubing and micropipettes and by measuring directly the intraluminal pressure. They made sure that the flow was laminar

adjusting the necessary length of the vessel. Maximal flow for PA was 40  $\mu\text{l}/\text{min}$ , for MCA 300–500  $\mu\text{l}/\text{min}$ . They allowed to equilibrate the perfusate before entering the vessel lumen passing it through gas permeable tubing in the extraluminal bath in order to avoid pH differences. In this experimental setup CMA dilated, MCA and PA constricted to increases in flow. The diameter decreased as a function of calculated shear stress, as well (discussed later) [114].

Recently, we have also investigated isolated middle cerebral arteries of the rat [23]. The sizes of glass pipettes used in this study were matched to both each other and the diameter of the vessels in order to achieve equal resistance. In addition, inflow and outflow reservoirs and position of the chamber were built in a symmetrical manner providing equal pressures or generating flow in the presence of constant pressure in the midsection of vessels. We raised pressure, then flow, then pressure and flow together and measured the diameter changes. We found that rat middle cerebral arteries constricted to increases in flow (also see at Figure 1), and flow-induced constriction enhanced the only pressure-induced constriction. Importantly, in line with findings of Bryan et al. [114] flow-induced constriction was observed in smaller vessels, as well. We have found that not only large arteries (i.e. middle cerebral arteries) but smaller side branches of MCA (about 50 micrometer in diameter) also constricted to increases in flow (unpublished observation). Also, very recently Filosa's group demonstrated flow-induced constriction of rat cerebral arterioles in brain slices [115].

It is important to note that isolated middle cerebral arteries maintain a stable steady state diameter in the presence of a constant flow rate. Upon increase in flow they constrict and in response to decrease in flow they dilate and maintain the new diameter [23]. These findings suggest that in vitro steady state flow rates activate vasomotor mechanisms that are able to provide a stable diameter. It is likely however that in vivo if such diameter is not sufficient to maintain adequate CBF other mechanisms are activated to provide the sufficient flow to brain tissues. For example, metabolic dilator adenosine inhibited functionally the flow-induced constriction, proposing that metabolic factors can override the flow-induced responses (similar to the case of increase in  $\text{CO}_2$ , as shown by Madden et al.; see earlier in this section) [23].

**Human cerebral arteries constrict to flow, as well:** Importantly, we also found that isolated human intracerebral arteries (from the internal carotid circulatory area) constricted to increases in flow, and the flow-induced constriction was mediated by 20-HETE (See later the mechanisms of flow-induced responses) [23]. It has to be noted that the active internal diameters of these vessels were approximately 200–250  $\mu\text{m}$ , equal with the studied rat middle cerebral arteries. These size of vessels correspond to large arteries of the circle of Willis in the rat, but small arteries in the human brain. Also, the human vessels were isolated from the fronto-temporal cortex of patients, thus different responses of human cerebral vessels to flow from different brain regions cannot be excluded.

In conclusion, studies using a well-controlled methodological approach found dilation to flow in rat and mice in the vertebro-basilar circulatory area; constriction was found in cat, rat and importantly, in human isolated cerebral arteries from the internal carotid circulatory area; biphasic responses were observed in rabbit and rat cerebral arterioles showing pressure and flow-rate dependency (dilated at lower and constricted at higher pressure and flow-rates).

## 2) What are the potential signals of flow-induced responses of cerebral vessels?

Changes in flow are accompanied by changes of many factors, among others wall shear stress. It is widely established that in most vascular beds changes of flow are sensed by the endothelium by sensing wall shear stress (WSS) elicited by increases in flow. The



consequent flow-induced dilation reduces WSS, thus WSS is regulated in a negative feedback manner [57–59]. This can be described by the following equation:

$$WSS=4\eta \times Q/\pi \times r^3,$$

where WSS is wall shear stress,  $\eta$  is viscosity, Q is flow and r is vessel radius [59]. It is believed that the overall physiological "purpose" of this mechanism is to optimize the circulatory energy loss. This mechanism is likely to play role in the development of functional hyperemia, for example during exercise (in skeletal and cardiac muscle) [57]. In case of cerebral vessels, in which flow elicits dilations - for example in the basilar artery – such WSS-sensitive mechanism can be at operation.

It is more difficult to envision the *mechanotransduction of flow-induced constriction*. What could be the signal that associated with increasing flow and sensed and converted to mechanical responses? Many previous studies were very careful to exclude any changes in intraluminal pressure during the investigation and observation of flow-induced constriction by keeping pressure constant [112, 114], thus increase in pressure cannot be the possible trigger.

The easiest hypothesis would be that in some cerebral vessels increasing WSS is converted to constriction instead of dilation by an unknown mechanism. For example, 20-HETE, the mediator of flow-induced constriction in MCA can cause dilation in basilar arteries known to dilate to flow, because it is converted into - as of yet unknown - dilator metabolites [116]. In this case constriction can be the function of WSS as calculated by Bryan at al. in rat middle cerebral arteries and in penetrating arterioles elevated by either increasing flow or viscosity of the perfusate solution [114, 117]. We have also found similar results in MCA, i.e. the magnitude of constriction could be plotted as a function of WSS (unpublished observation). However, there are theoretical considerations that argue against the idea that WSS acting on the vascular endothelium is the signal in flow-induced constriction of cerebral vessels:

- a. If WSS is sensed, how does it relate to changes in CBF and cerebral blood volume (CBV)? How can a positive feed-back mechanism "serve" a negative feed-back mechanism i.e. autoregulation? Because constriction increases WSS further, it should result in further constriction of cerebral vessels, then further increase in WSS (positive feed-back manner), while CBF and CBV would decrease continuously. Such changes in CBF and CBV have not been observed, however.
- b. Bryan at al. and other previous studies found that constriction of cerebral vessels to flow is independent of the presence of a functioning endothelial layer [112, 114, 117], and mechanisms responsible for the constriction are localized into the smooth muscle layer of cerebral arteries. This finding also argues against that WSS is the signal. The intriguing question is: how can WSS be sensed by the smooth muscle if the flowing blood is in contact with the intraluminal side of endothelium and not with the smooth muscle? Bryan at al. proposed, that the endothelium plays a role in attenuating the shear stress-induced constrictions by an unknown dilating factor, by a dilating process, or by attenuating the mechanical force of the WSS as it is transmitted to the abluminal side of the vessel [117]. This dilator mechanism may be present, but it needs further experimental support. This could be NO, since inhibition of NOS in zero flow conditions reduced the diameter of cerebral vessels [117, 118].

- c. One can assume that WSS is sensed by the smooth muscle through interstitial flow driven by the transvascular pressure difference, which was shown to be capable to cause smooth muscle constriction [119]. This idea is supported by the findings that integrins play a role (as sensors) in interstitial flow-related mechanotransduction and blocking integrins inhibits the flow-induced constriction of cerebral arteries [112, 114, 119]. This hypothesis could also explain why intact endothelium attenuates flow-induced constriction, because in case of endothelial injury or denudation the magnitude of transvascular flow is much greater than in intact arteries [119].

It is of note however, that significant amount of fluid cannot leave the vessels in the closed cranium. Although, it is unlikely that larger amount of fluid would leave the wall of larger vessels due to the presence of well developed internal elastic lamina and several layers of smooth muscle cells [52, 120]. Moreover, the amount of transmural fluid movement is likely to be minimal, because of the tight connections between endothelial cells [121]. Thus it is unlikely to lead to “leakiness” or increased filtration of plasma into the brain parenchyma. Nevertheless, the exact nature of the “transmural fluid signal” proposed by Tarbell et al. [119] still remains obscure.

- d. Flow velocity increases as diameter decreases if flow (and pressure) constant. Flow velocity itself could be a candidate for the trigger parameter, as it is related to changes in CBF and CBV, but it is difficult to envision a sensor for flow velocity, since velocity is not a force that can be sensed by cells.
- e. Another possibility is that increasing flow (mass transport) washes in or out vasoactive substances providing a tonic effect on vascular smooth muscle. For instance, NO has been proposed to perform a tonic dilator effect on cerebral vessels [29, 117, 118]. We have also shown a significant reduction in basal diameter of cerebral vessels when NOS was inhibited. It is also known that NO can inhibit the production of 20-HETE [122], which was shown to mediate flow-induced constriction of cerebral arteries [23]. Thus, hypothetically a decreased effect of NO could lead to an enhanced constrictor effect of 20-HETE. Reactive oxygen species were also shown to play a role in flow-induced constriction in rat middle cerebral arteries [23], and they can be also potential candidate to decrease the bioavailability of NO leading to enhanced 20-HETE production. Endothelial-derived dilator arachidonic acid metabolites by lipoxygenase, epoxyeicosatrienoic acids (EETs) are also known to be produced by the endothelium in response to shear stress [123], and also they are capable to inhibit TXA<sub>2</sub> receptor [124] which was shown to mediate the constriction to flow by 20-HETE [23]. Thus, they could also play a role in the “wash-out” theory. However, presently existing data question these possibilities, showing that flow-induced constriction of cerebral vessels still exists in the presence of NO synthase inhibitor L-NAME and after denudation of endothelium [114, 117] and such constriction was not observed in basilar arteries instead of dilations. Also, at a constant flow rate increases in the viscosity of the perfusing solution could constrict the vessels while increased wall shear stress, as well [114].

These ideas and other controversial observations need to be addressed by future studies. Such “physiological” investigations would be very important since precise regulation of CBF has important clinical consequences to prevent and to treat headache of vascular origin, TIA, stroke, vascular cognitive impairment (and mixed vascular dementia with Alzheimer disease), edema formation after brain injury or associated with tumor formation and other fatal cerebral pathologies. Also, without knowing the exact nature of mechanical, vasomotor response only “signaling” studies would not yield valuable information.

### 3) Cellular and molecular signaling of flow-induced responses of cerebral vessels

In this section we briefly summarize the molecular mechanisms that are shown to play role in the flow-induced responses of cerebral vessels (Figure 3).

**a) Flow-induced dilation of cerebral vessels**—There are only a few data in the literature regarding the mechanisms of flow-induced dilation of cerebral vessels. In the rat basilar artery Paravicini et al. found that the dilation is mediated by  $H_2O_2$  and NO generated by NADPH oxidase and eNOS, and PI3-K activation is important in activation of NADPH oxidase [102]. Recently Drouin et al. demonstrated, similar to Paravicini et al. that  $H_2O_2$  mediates flow-induced dilation, but it derives from NO synthase, which is activated by an Akt-dependent pathway (Figure 3) [104, 105].

**b) Biphasic response of cerebral vessels to flow**—In line with the observations of flow-induced dilation, in some of the studies founding biphasic response to flow the dilation was endothelium-dependent and was blocked by inhibition of NO synthase (L-NAME) [108, 110]. Ward et al. also found that indomethacin blocked the response and proposed a role for cyclooxygenases-derived metabolites [109]. The constrictor response was found to be dependent on increases in intracellular (smooth muscle)  $Ca^{2+}$  concentration [106, 107].

**c) Flow-induced constriction of cerebral vessels**—Madden et al. found that administration of integrin-binding peptides, scavenging reactive oxygen species by superoxide dismutase (SOD) and inhibition of tyrosine kinase blocked the constriction suggesting that integrin signaling, free radicals and tyrosine kinase play a role in mediation of flow-induced constriction in an endothelium-independent manner [112]. Similarly Bryan et al. observed that an integrin blocker specific for  $\beta_3$ -integrin and SOD abolished the flow-induced constriction. They also measured  $Ca^{2+}$  concentration in the vessel wall and demonstrated that flow-induced constriction is accompanied by increases in  $[Ca^{2+}]_i$ . Denudation of the endothelium did not affect flow-induced constriction [114, 117].

Recently we have found that flow-induced constriction of cerebral vessels was blocked by inhibition of 20-HETE synthesis,  $TXA_2$  (TP) receptor, COX activity and scavenging ROS. Therefore we proposed that increases in flow activate arachidonic acid cascade, and the metabolites are further metabolized by cytochrome P450 4A enzymes (CYP450 4A) into 20-hydroxieicosatetraenoic acid (20-HETE). It is known that CYP450 4A also produces reactive oxygen species (ROS), which contribute to the constriction. Thus it seems that flow-induced constriction of MCA and that of human cerebral arteries is mediated by 20-HETE via thromboxane  $A_2$ /prostaglandin  $H_2$  (TP) receptors and requires COX activity (Figure 3) [23].

### 4) Physiological significance of flow induced responses of cerebral vessels of different sizes and regions. Implications for heterogeneity of autoregulation

From the observed responses one can estimate the change in CBF by using diameter values induced by only changes in pressure and then simultaneous changes in pressure and flow using the Hagen–Poiseuille equation [23]. Also, a “gain factor (G)” can be calculated indicating the strength or efficacy of the autoregulation of blood flow ( $G = 1$  indicates perfect autoregulation, whereas  $G < 1$  means inefficient autoregulation, when CBF increases as a function of intraluminal pressure) [22, 23]. In case of flow-induced constriction we found that simultaneous increase of pressure+flow enhanced the only pressure-induced decrease in diameter. Also, the gain of autoregulation (G) calculated using diameters induced by pressure alone was below 1, whereas G was  $\approx 1$  when pressure+flow were increased simultaneously indicating a more efficient autoregulation. When only pressure was increased estimated CBF showed a linear increase, but when pressure+flow increased

simultaneously estimated CBF decreased significantly and remained relatively constant [23]. Based on this we propose that in the circulatory areas where vessels constrict to flow simultaneous operation of pressure- and flow-induced constrictions is necessary to explain an effective autoregulation of CBF (Figure 2).

In the cerebrum, in the internal carotid circulatory area resistance is profoundly determined by larger arteries [18, 43, 44, 50]. In line with this, larger arteries (i.e MCA) constrict to increases in flow, flow-induced constriction enhances the pressure-induced tone of cerebral vessels leading to a more efficient autoregulation of CBF, and via this flow-induced constriction of cerebral arteries plays a role in regulating cerebral blood volume and intracranial pressure. Therefore, in addition to myogenic response, flow-induced constriction (or dilation) may also participate in the development of segmental resistance of the cerebral circulation, because both large arteries and arterioles respond to changes in flow with either constriction or dilation [23, 100, 105–107, 110, 114, 125].

This segmental vascular resistance is thought to provide constant blood flow when blood flow is altered locally (for example due to metabolic factors), and to protect downstream brain circulation from pressure and volume overload [43, 50, 52]. Regarding flow-induced responses the importance of segmental function of cerebrovascular resistance and the role of large arteries are underlined by the findings of Fujii et al. and Garcia-Roldan and Bevan [106, 110] showing that isolated cerebral arterioles constricted when pressure or flow rate is high and dilated when they are low.

Also, the vasomotor tone “set” by the two hemodynamic forces can be modulated or overridden by other factors sensitive to the needs of neural tissues. Whereas flow-induced constriction may play an important role (together with the pressure sensitive myogenic tone) in regulation and maintenance of cerebral volume and intracranial pressure, local neural needs can increase cerebral blood flow regionally via neural, glial and other regulatory mechanisms, which can also be propagated to upstream vessels [1, 17]. This concept is in line with the suggestion of studies showing that metabolic dilation could overcome the constrictor effect of pressure or flow [23, 112, 126, 127]. Conversely, in the brain stem (vertebro-basilar system) arterioles are the major site of resistance [50, 51], thus larger arteries, such as the basilar artery “can” dilate to flow participating in reactive hyperemia [125]. The heterogeneity of autoregulation is likely due to the different efficacy of pressure and flow sensitive vascular mechanisms and/or contribution of metabolic mechanisms and neurovascular coupling. The proposed concept should be further clarified in the future by in vitro investigation and comparison of pressure- and flow-induced responses of cerebral arteries and arterioles from different regions of cerebrovascular tree [128]. More importantly, in vivo imaging of CBF in experimental animals and humans could be performed by means of MRI, laser speckle imager and other novel techniques in order to clarify the mentioned concept of the role of hemodynamic forces in regulation of CBF, in which pharmacological intervention of signaling mechanisms may reveal further detail of autoregulation of CBF.

All in all, regulation of cerebral blood flow (CBF) is the result of multilevel, interacting complex mechanisms to maintain an appropriate blood flow providing nutritional and gas supply for the brain tissue. To simplify, the tone of cerebral vessels is affected by both intraluminal pressure-induced constriction and flow-induced dilation or constriction, “setting a basal vasomotor tone”, which is then modulated by metabolic, astrocytic-glial, neural signals providing the complex multilevel regulation of CBF. The sum of these mechanisms is capable to match the requirement of maintaining an appropriate blood flow for the brain tissue and in the same time complying with the limited space in the closed cranium.

## 5) Pathophysiological and clinical relevance of pressure- and flow-induced autoregulatory mechanisms of CBF

**Changes in hypertension**—Previous studies described that myogenic responses of cerebral vessels is enhanced in hypertension [22, 63, 129, 130]. Logically it is considered to be an adaptation extending the range of CBF autoregulation to higher systemic blood pressure. Based on our recent concept described in this review (Fig 2) enhanced flow-dependent constriction may also contribute to the functional adaptation of cerebral vessels to hypertension in order to “protect” brain tissue and intracranial space from high pressure and volume. Indeed, flow-induced constriction seems to be enhanced in hypertension [131], and mediated by upregulation of 20-HETE signaling (Toth et al 2012, manuscript in preparation). This could be due to the chronic elevation of blood flow due to the greater pressure drop across the cerebral circulation in hypertension.

**Possible contribution to stroke and other cerebrovascular disorders**—As mentioned above, myogenic response is enhanced in hypertension protecting the cerebral circulation from higher blood pressure. The importance of this mechanism was demonstrated by Smeda et al showing that the enhanced myogenic response of rat cerebral arteries is attenuated before and lost after the onset of cerebral hemorrhage [132, 133]. Parallel with the myogenic response we found that enhanced flow-induced constriction in hypertension is attenuated before and lost after the onset of hemorrhagic stroke in stroke prone spontaneously hypertensive rats (unpublished observation, Toth P et al., 2011). On the other hand enhanced constriction to flow in hypertension can cause decreased capability to adapt to decrease in blood pressure probably contributing to ischemic damage of the brain. This idea is supported by the findings that inhibition of 20-HETE production reduces significantly the infarct size after middle cerebral artery occlusion [134–136]. However, all of these ideas need to be investigated in the future.

### Summary

Although autoregulation of cerebral blood flow at higher pressures has been explained solely by the pressure sensitive myogenic mechanism, more and more data show that it is insufficient to do so. Because during changes in pressure flow changes as well, it is logical to assume that changes in flow also has a role. Probably this was the reason for conducting many studies to elucidate the nature, the underlying mechanisms and physiological importance of flow-induced responses of cerebral vessels in the last 30 years. These studies showed that increases in intraluminal flow is sensed by the vascular tissues of cerebral vessels and converted into changes in diameter: constriction, dilation or biphasic response. It seems that the nature of response depends on the region of the brain, or vascular segment, size or localization. Also, species and gender differences cannot be ruled out. These differences are likely to have physiological role by modulating myogenic mechanism and regulation of CBF and CBV and intracranial pressure, and at the same time, providing an appropriate amount of blood flow to the brain tissue according to the needs.

Although the precise signaling of mechanotransduction of flow signal is not yet known, arachidonic metabolites, especially 20-HETE, nitric oxide, potassium and TRP channels seem to play important role. In addition reactive oxygen species produced by these and other enzymes, such as NAD(P)H oxidase may also contribute to the modulation of vasomotor tone.

It is clear however, that there are still many controversial observations, unexplained signaling and unknown mechanisms that could all contribute to the diameter response of cerebral vessels of different size and origin to changes in hemodynamic forces and thus regulation of cerebral blood flow. Because cerebrovascular diseases (stroke, vascular

dementia, cognitive impairment, aging etc.) associated with the impaired regulation of CBF are still leading causes of human morbidity and mortality it is imperative to further investigate the responses and the underlying mechanism of cerebral vessels to hemodynamic/rheological forces.

## Acknowledgments

**Sources of Support:** American Heart Association, Founders Affiliate, 0855910D, NIH PO-1 HL-43023, Hungarian National Science Research Fund (OTKA) K71591 and K67984, MHT 2011.

## References

1. Attwell D, Buchan AM, Charpak S, Lauritzen M, Macvicar BA, Newman EA. Glial and neuronal control of brain blood flow. *Nature*. 2010; 468:232–243. [PubMed: 21068832]
2. Betz E. Cerebral blood flow: Its measurement and regulation. *Physiol Rev*. 1972; 52:595–630. [PubMed: 4555515]
3. Harder DR, Gebremedhin D, Narayanan J, Jefcoat C, Falck JR, Campbell WB, Roman R. Formation and action of a p-450 4a metabolite of arachidonic acid in cat cerebral microvessels. *Am J Physiol*. 1994; 266:H2098–H2107. [PubMed: 8203608]
4. Harper AM. Autoregulation of cerebral blood flow: Influence of the arterial blood pressure on the blood flow through the cerebral cortex. *J Neurol Neurosurg Psychiatry*. 1966; 29:398–403. [PubMed: 5926462]
5. Kontos HA. Regulation of the cerebral circulation. *Annu Rev Physiol*. 1981; 43:397–407. [PubMed: 6782941]
6. Lassen NA. Cerebral blood flow and oxygen consumption in man. *Physiol Rev*. 1959; 39:183–238. [PubMed: 13645234]
7. Lassen NA. Control of cerebral circulation in health and disease. *Circ Res*. 1974; 34:749–760. [PubMed: 4598993]
8. McHedlishvili G. Physiological mechanisms controlling cerebral blood flow. *Stroke*. 1980; 11:240–248. [PubMed: 6771898]
9. McHedlishvili GI, Mitagvaria NP, Ormotsadze LG. Vascular mechanisms controlling a constant blood supply to the brain ("autoregulation"). *Stroke*. 1973; 4:742–750. [PubMed: 4751085]
10. van Beek AH, Claassen JA, Rikkert MG, Jansen RW. Cerebral autoregulation: An overview of current concepts and methodology with special focus on the elderly. *J Cereb Blood Flow Metab*. 2008; 28:1071–1085. [PubMed: 18349877]
11. Rosenblum WI, Shimizu T, Nelson GH. Endothelium-dependent effects of substance p and calcitonin gene-related peptide on mouse pial arterioles. *Stroke*. 1993; 24:1043–1047. discussion 1047–1048. [PubMed: 7686695]
12. Rosenblum WI, Wei EP, Kontos HA. Dilation of rat brain arterioles by hypercapnia in vivo can occur even after blockade of guanylate cyclase by odq. *Eur J Pharmacol*. 2002; 448:201–206. [PubMed: 12144942]
13. Kovach AG, Dora E, Szedlaczek S, Koller A. Effect of the organic calcium antagonist d-600 on cerebrocortical vascular and redox responses evoked by adenosine, anoxia, and epilepsy. *J Cereb Blood Flow Metab*. 1983; 3:51–61. [PubMed: 6822618]
14. Dora E, Koller A, Kovach AG. Effect of topical adenosine deaminase treatment on the functional hyperemic and hypoxic responses of cerebrocortical microcirculation. *J Cereb Blood Flow Metab*. 1984; 4:447–457. [PubMed: 6470059]
15. Schaller B, Graf R. Different compartments of intracranial pressure and its relationship to cerebral blood flow. *J Trauma*. 2005; 59:1521–1531. [PubMed: 16394936]
16. Faraci FM, Baumbach GL, Heistad DD. Myogenic mechanisms in the cerebral circulation. *J Hypertens Suppl*. 1989; 7:S61–S64. discussion S65. [PubMed: 2681598]
17. Iadecola C, Yang G, Ebner TJ, Chen G. Local and propagated vascular responses evoked by focal synaptic activity in cerebellar cortex. *J Neurophysiol*. 1997; 78:651–659. [PubMed: 9307102]



18. Kontos HA, Wei EP, Navari RM, Levasseur JE, Rosenblum WI, Patterson JL Jr. Responses of cerebral arteries and arterioles to acute hypotension and hypertension. *Am J Physiol.* 1978; 234:H371–H383. [PubMed: 645875]
19. Lassen NA, Christensen MS. Physiology of cerebral blood flow. *Br J Anaesth.* 1976; 48:719–734. [PubMed: 7284]
20. Cipolla MJ, Gokina NI, Osol G. Pressure-induced actin polymerization in vascular smooth muscle as a mechanism underlying myogenic behavior. *Faseb J.* 2002; 16:72–76. [PubMed: 11772938]
21. Osol G, Brekke JF, McElroy-Yaggy K, Gokina NI. Myogenic tone, reactivity, and forced dilatation: A three-phase model of in vitro arterial myogenic behavior. *Am J Physiol Heart Circ Physiol.* 2002; 283:H2260–H2267. [PubMed: 12388265]
22. Osol G, Halpern W. Myogenic properties of cerebral blood vessels from normotensive and hypertensive rats. *Am J Physiol.* 1985; 249:H914–H921. [PubMed: 4061668]
23. Toth P, Rozsa B, Springo Z, Doczi T, Koller A. Isolated human and rat cerebral arteries constrict to increases in flow: Role of 20-hete and tp receptors. *J Cereb Blood Flow Metab.* 2011; 31:2096–2105. [PubMed: 21610722]
24. Wallis SJ, Firth J, Dunn WR. Pressure-induced myogenic responses in human isolated cerebral resistance arteries. *Stroke.* 1996; 27:2287–2290. discussion 2291. [PubMed: 8969795]
25. Schaller B, Graf R. Different compartments of intracranial pressure and its relationship to cerebral blood flow. *J Trauma.* 2005; 59:1521–1531. [PubMed: 16394936]
26. Lucas SJ, Tzeng YC, Galvin SD, Thomas KN, Ogoh S, Ainslie PN. Influence of changes in blood pressure on cerebral perfusion and oxygenation. *Hypertension.* 2010; 55:698–705. [PubMed: 20083726]
27. Rosenblum WI. Autoregulatory plateau: Does it exist? *J Cereb Blood Flow Metab.* 1995; 15:174–177. [PubMed: 7798336]
28. Peterson EC, Wang Z, Britz G. Regulation of cerebral blood flow. *Int J Vasc Med.* 2011; 2011:823525.
29. Rosenblum WI. Is the edrf in the cerebral circulation no? Its release by shear and the dangers in interpreting the effects of nos inhibitors. *Keio J Med.* 1998; 47:142–149. [PubMed: 9785759]
30. Pohl U, de Wit C. A unique role of no in the control of blood flow. *News Physiol Sci.* 1999; 14:74–80. [PubMed: 11390824]
31. Pohl U, Herlan K, Huang A, Bassenge E. Edrf-mediated shear-induced dilation opposes myogenic vasoconstriction in small rabbit arteries. *Am J Physiol.* 1991; 261:H2016–H2023. [PubMed: 1721502]
32. Sun D, Huang A, Koller A, Kaley G. Flow-dependent dilation and myogenic constriction interact to establish the resistance of skeletal muscle arterioles. *Microcirculation.* 1995; 2:289–295. [PubMed: 8748953]
33. Ungvari Z, Koller A. Selected contribution: No released to flow reduces myogenic tone of skeletal muscle arterioles by decreasing smooth muscle ca(2+) sensitivity. *J Appl Physiol.* 2001; 91:522–527. discussion 504–525. [PubMed: 11408472]
34. Ayata C, Dunn AK, Gursoy OY, Huang Z, Boas DA, Moskowitz MA. Laser speckle flowmetry for the study of cerebrovascular physiology in normal and ischemic mouse cortex. *J Cereb Blood Flow Metab.* 2004; 24:744–755. [PubMed: 15241182]
35. Bayliss WM. On the local reactions of the arterial wall to changes of internal pressure. *J Physiol.* 1902; 28:220–231. [PubMed: 16992618]
36. Bellapart J, Fraser JF. Transcranial doppler assessment of cerebral autoregulation. *Ultrasound Med Biol.* 2009; 35:883–893. [PubMed: 19329245]
37. Lunn V, Fog M. The reaction of the pial arteries to some cholin-like and adrenalin-like substances. *J Neurol Psychiatry.* 1939; 2:223–230. [PubMed: 21610953]
38. MacKenzie ET, Farrar JK, Fitch W, Graham DI, Gregory PC, Harper AM. Effects of hemorrhagic hypotension on the cerebral circulation. I. Cerebral blood flow and pial arteriolar caliber. *Stroke.* 1979; 10:711–718. [PubMed: 524412]
39. MacKenzie ET, Strandgaard S, Graham DI, Jones JV, Harper AM, Farrar JK. Effects of acutely induced hypertension in cats on pial arteriolar caliber, local cerebral blood flow, and the blood-brain barrier. *Circ Res.* 1976; 39:33–41. [PubMed: 1277403]

40. Magun JG. The effect of pharmacologically increased blood pressure on brain circulation. Angiographic investigation of arterial diameter and blood flow in patients with normal and pathological angiograms. *Z Neurol.* 1973; 204:107–134. [PubMed: 4121466]
41. Mueller SM, Heistad DD, Marcus ML. Total and regional cerebral blood flow during hypotension, hypertension, and hypocapnia. Effect of sympathetic denervation in dogs. *Circ Res.* 1977; 41:350–356. [PubMed: 890889]
42. Zaharchuk G, Mandeville JB, Bogdanov AA Jr, Weissleder R, Rosen BR, Marota JJ. Cerebrovascular dynamics of autoregulation and hypoperfusion. An mri study of cbf and changes in total and microvascular cerebral blood volume during hemorrhagic hypotension. *Stroke.* 1999; 30:2197–2204. discussion 2204-2195. [PubMed: 10512929]
43. Faraci FM, Heistad DD. Regulation of large cerebral arteries and cerebral microvascular pressure. *Circ Res.* 1990; 66:8–17. [PubMed: 2403863]
44. Harper SL, Bohlen HG, Rubin MJ. Arterial and microvascular contributions to cerebral cortical autoregulation in rats. *Am J Physiol.* 1984; 246:H17–H24. [PubMed: 6696087]
45. Stromberg DD, Fox JR. Pressures in the pial arterial microcirculation of the cat during changes in systemic arterial blood pressure. *Circ Res.* 1972; 31:229–239. [PubMed: 5049738]
46. Baumbach GL, Heistad DD. Regional, segmental, and temporal heterogeneity of cerebral vascular autoregulation. *Ann Biomed Eng.* 1985; 13:303–310. [PubMed: 3898928]
47. Baumbach GL, Heistad DD. Heterogeneity of brain blood flow and permeability during acute hypertension. *Am J Physiol.* 1985; 249:H629–H637. [PubMed: 3929626]
48. Rozet I, Vavilala MS, Lindley AM, Visco E, Treggiari M, Lam AM. Cerebral autoregulation and co2 reactivity in anterior and posterior cerebral circulation during sevoflurane anesthesia. *Anesth Analg.* 2006; 102:560–564. [PubMed: 16428561]
49. Willie CK, Colino FL, Bailey DM, Tzeng YC, Binsted G, Jones LW, Haykowsky MJ, Bellapart J, Ogoh S, Smith KJ, Smirl JD, Day TA, Lucas SJ, Eller LK, Ainslie PN. Utility of transcranial doppler ultrasound for the integrative assessment of cerebrovascular function. *J Neurosci Methods.* 2011; 196:221–237. [PubMed: 21276818]
50. Faraci FM, Mayhan WG, Heistad DD. Segmental vascular responses to acute hypertension in cerebrum and brain stem. *Am J Physiol.* 1987; 252:H738–H742. [PubMed: 3565591]
51. Mayhan WG, Faraci FM, Heistad DD. Disruption of the blood-brain barrier in cerebrum and brain stem during acute hypertension. *Am J Physiol.* 1986; 251:H1171–H1175. [PubMed: 3098113]
52. Cipolla, MJ. *The cerebral circulation.* San Rafael (CA): 2009.
53. Faraci FM, Heistad DD. Regulation of the cerebral circulation: Role of endothelium and potassium channels. *Physiol Rev.* 1998; 78:53–97. [PubMed: 9457169]
54. Fog M. The relationship between the blood pressure and the tonic regulation of the pial arteries. *J Neurol Psychiatry.* 1938; 1:187–197. [PubMed: 21610927]
55. Forbes HS. Regulation of the cerebral vessels; new aspects. *AMA Arch Neurol Psychiatry.* 1958; 80:689–695.
56. Vinall PE, Simeone FA. Cerebral autoregulation: An in vitro study. *Stroke.* 1981; 12:640–642. [PubMed: 7303050]
57. Koller A, Bagi Z. On the role of mechanosensitive mechanisms eliciting reactive hyperemia. *Am J Physiol Heart Circ Physiol.* 2002; 283:H2250–H2259. [PubMed: 12427591]
58. Koller A, Sun D, Huang A, Kaley G. Corelease of nitric oxide and prostaglandins mediates flow-dependent dilation of rat gracilis muscle arterioles. *Am J Physiol.* 1994; 267:H326–H332. [PubMed: 8048598]
59. Koller A, Sun D, Kaley G. Role of shear stress and endothelial prostaglandins in flow- and viscosity-induced dilation of arterioles in vitro. *Circ Res.* 1993; 72:1276–1284. [PubMed: 8495555]
60. de Wit C, Jahrbeck B, Schafer C, Bolz SS, Pohl U. Nitric oxide opposes myogenic pressure responses predominantly in large arterioles in vivo. *Hypertension.* 1998; 31:787–794. [PubMed: 9495262]
61. Bohlen HG, Harper SL. Evidence of myogenic vascular control in the rat cerebral cortex. *Circ Res.* 1984; 55:554–559. [PubMed: 6478557]

62. Meininger GA, Mack CA, Fehr KL, Bohlen HG. Myogenic vasoregulation overrides local metabolic control in resting rat skeletal muscle. *Circ Res.* 1987; 60:861–870. [PubMed: 3594758]
63. Paulson OB, Strandgaard S, Edvinsson L. Cerebral autoregulation. *Cerebrovasc Brain Metab Rev.* 1990; 2:161–192. [PubMed: 2201348]
64. Korzick DH, Laughlin MH, Bowles DK. Alterations in pkc signaling underlie enhanced myogenic tone in exercise-trained porcine coronary resistance arteries. *J Appl Physiol.* 2004; 96:1425–1432. [PubMed: 14672961]
65. Ping P, Johnson PC. Role of myogenic response in enhancing autoregulation of flow during sympathetic nerve stimulation. *Am J Physiol.* 1992; 263:H1177–H1184. [PubMed: 1415767]
66. Busija DW, Heistad DD. Factors involved in the physiological regulation of the cerebral circulation. *Rev Physiol Biochem Pharmacol.* 1984; 101:161–211. [PubMed: 6441228]
67. Cole WC, Welsh DG. Role of myosin light chain kinase and myosin light chain phosphatase in the resistance arterial myogenic response to intravascular pressure. *Arch Biochem Biophys.* 2011; 510:160–173. [PubMed: 21392499]
68. Hill MA, Davis MJ, Meininger GA, Potocnik SJ, Murphy TV. Arteriolar myogenic signalling mechanisms: Implications for local vascular function. *Clin Hemorheol Microcirc.* 2006; 34:67–79. [PubMed: 16543619]
69. Schubert R, Lidington D, Bolz SS. The emerging role of ca<sup>2+</sup> sensitivity regulation in promoting myogenic vasoconstriction. *Cardiovasc Res.* 2008; 77:8–18. [PubMed: 17764667]
70. Harder DR, Narayanan J, Gebremedhin D. Pressure-induced myogenic tone and role of 20-hete in mediating autoregulation of cerebral blood flow. *American journal of physiology Heart and circulatory physiology.* 2011; 300:H1557–H1565. [PubMed: 21257913]
71. Somlyo AP, Somlyo AV. Ca<sup>2+</sup> sensitivity of smooth muscle and nonmuscle myosin ii: Modulated by g proteins, kinases, and myosin phosphatase. *Physiol Rev.* 2003; 83:1325–1358. [PubMed: 14506307]
72. Johnson PC. The myogenic response in the microcirculation and its interaction with other control systems. *J Hypertens Suppl.* 1989; 7:S33–S39. discussion S40. [PubMed: 2681595]
73. D'Angelo G, Mogford JE, Davis GE, Davis MJ, Meininger GA. Integrin-mediated reduction in vascular smooth muscle [ca<sup>2+</sup>]<sub>i</sub> induced by rgd-containing peptide. *Am J Physiol.* 1997; 272:H2065–H2070. [PubMed: 9139994]
74. Davis MJ, Donovitz JA, Hood JD. Stretch-activated single-channel and whole cell currents in vascular smooth muscle cells. *Am J Physiol.* 1992; 262:C1083–C1088. [PubMed: 1373561]
75. Lucchesi PA, Sabri A, Belmadani S, Matrougui K. Involvement of metalloproteinases 2/9 in epidermal growth factor receptor transactivation in pressure-induced myogenic tone in mouse mesenteric resistance arteries. *Circulation.* 2004; 110:3587–3593. [PubMed: 15557365]
76. Martinez-Lemus LA, Crow T, Davis MJ, Meininger GA. Alpha<sub>v</sub>beta<sub>3</sub>- and alpha<sub>5</sub>beta<sub>1</sub>-integrin blockade inhibits myogenic constriction of skeletal muscle resistance arterioles. *American journal of physiology Heart and circulatory physiology.* 2005; 289:H322–H329. [PubMed: 15722407]
77. Knot HJ, Nelson MT. Regulation of arterial diameter and wall [ca<sup>2+</sup>] in cerebral arteries of rat by membrane potential and intravascular pressure. *J Physiol.* 1998; 508(Pt 1):199–209. [PubMed: 9490839]
78. Welsh DG, Nelson MT, Eckman DM, Brayden JE. Swelling-activated cation channels mediate depolarization of rat cerebrovascular smooth muscle by hyposmolarity and intravascular pressure. *J Physiol.* 2000; 527(Pt 1):139–148. [PubMed: 10944177]
79. Welsh DG, Morielli AD, Nelson MT, Brayden JE. Transient receptor potential channels regulate myogenic tone of resistance arteries. *Circ Res.* 2002; 90:248–250. [PubMed: 11861411]
80. Earley S, Waldron BJ, Brayden JE. Critical role for transient receptor potential channel trpm4 in myogenic constriction of cerebral arteries. *Circ Res.* 2004; 95:922–929. [PubMed: 15472118]
81. Drummond HA, Grifoni SC, Jernigan NL. A new trick for an old dogma: Enac proteins as mechanotransducers in vascular smooth muscle. *Physiology (Bethesda).* 2008; 23:23–31. [PubMed: 18268362]
82. Dopico AM, Kirber MT, Singer JJ, Walsh JV Jr. Membrane stretch directly activates large conductance ca<sup>2+</sup>-activated k<sup>+</sup> channels in mesenteric artery smooth muscle cells. *Am J Hypertens.* 1994; 7:82–89. [PubMed: 8136116]

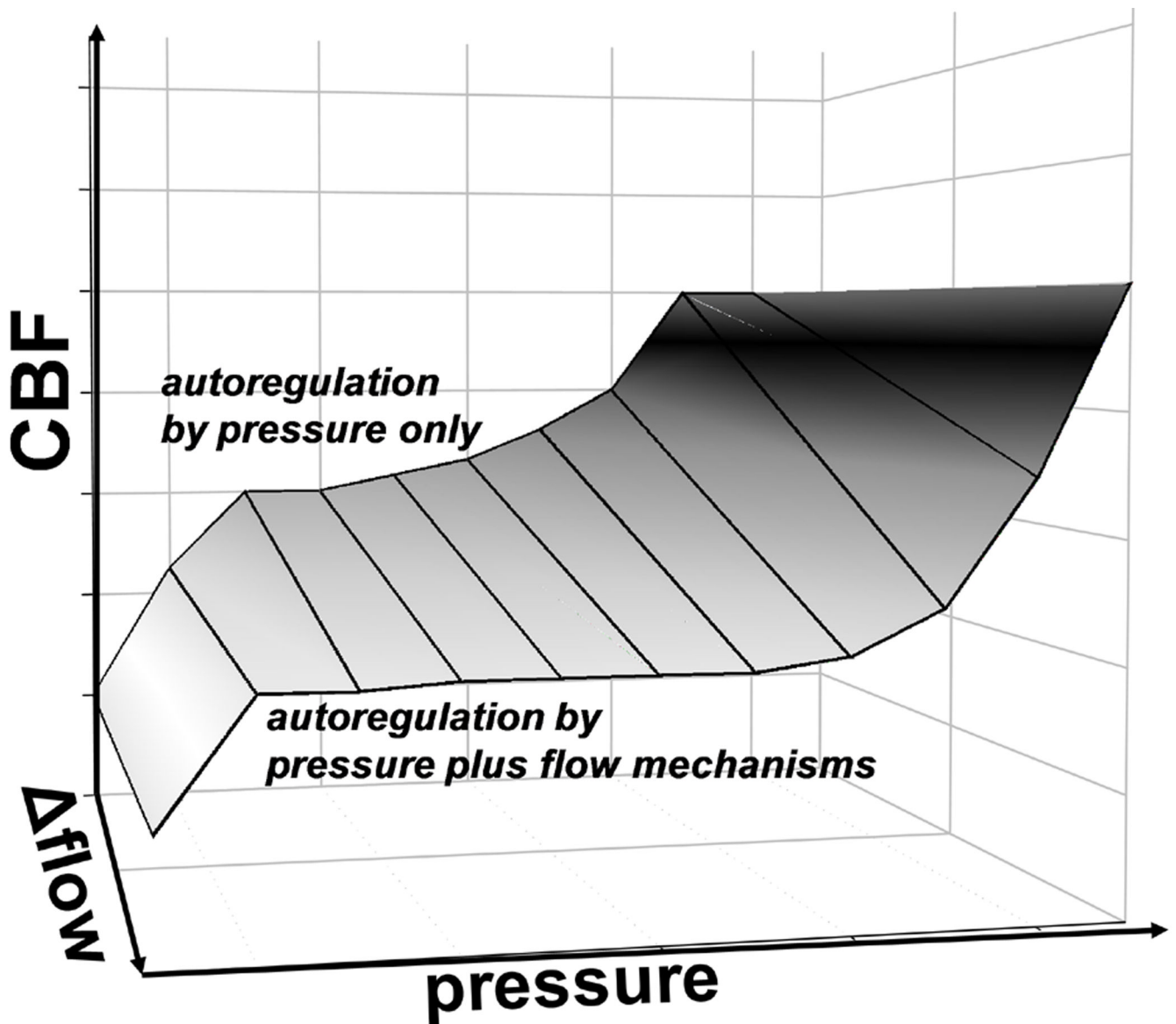
83. Nelson MT, Conway MA, Knot HJ, Brayden JE. Chloride channel blockers inhibit myogenic tone in rat cerebral arteries. *J Physiol.* 1997; 502((Pt 2)):259–264. [PubMed: 9263908]
84. Knot HJ, Nelson MT. Regulation of membrane potential and diameter by voltage-dependent k<sup>+</sup> channels in rabbit myogenic cerebral arteries. *Am J Physiol.* 1995; 269:H348–H355. [PubMed: 7631867]
85. McCarron JG, Crichton CA, Langton PD, MacKenzie A, Smith GL. Myogenic contraction by modulation of voltage-dependent calcium currents in isolated rat cerebral arteries. *J Physiol.* 1997; 498((Pt 2)):371–379. [PubMed: 9032685]
86. Moosmang S, Schulla V, Welling A, Feil R, Feil S, Wegener JW, Hofmann F, Klugbauer N. Dominant role of smooth muscle l-type calcium channel *cav1.2* for blood pressure regulation. *Embo J.* 2003; 22:6027–6034. [PubMed: 14609949]
87. Moosmang S, Haider N, Bruderl B, Welling A, Hofmann F. Antihypertensive effects of the putative t-type calcium channel antagonist mibefradil are mediated by the l-type calcium channel *cav1.2*. *Circ Res.* 2006; 98:105–110. [PubMed: 16306443]
88. Watanabe J, Horiguchi S, Karibe A, Keitoku M, Takeuchi M, Satoh S, Takishima T, Shirato K. Effects of ryanodine on development of myogenic response in rat small skeletal muscle arteries. *Cardiovasc Res.* 1994; 28:480–484. [PubMed: 8181034]
89. Davis MJ, Hill MA. Signaling mechanisms underlying the vascular myogenic response. *Physiol Rev.* 1999; 79:387–423. [PubMed: 10221985]
90. Schubert R, Mulvany MJ. The myogenic response: Established facts and attractive hypotheses. *Clin Sci (Lond).* 1999; 96:313–326. [PubMed: 10087237]
91. Hill MA, Zou H, Potocnik SJ, Meininger GA, Davis MJ. Invited review: Arteriolar smooth muscle mechanotransduction: Ca<sup>2+</sup> signaling pathways underlying myogenic reactivity. *J Appl Physiol.* 2001; 91:973–983. [PubMed: 11457816]
92. Pfitzer G. Invited review: Regulation of myosin phosphorylation in smooth muscle. *J Appl Physiol.* 2001; 91:497–503. [PubMed: 11408468]
93. Lagaud G, Gaudreault N, Moore ED, Van Breemen C, Laher I. Pressure-dependent myogenic constriction of cerebral arteries occurs independently of voltage-dependent activation. *American journal of physiology Heart and circulatory physiology.* 2002; 283:H2187–H2195. [PubMed: 12388215]
94. Narayanan J, Imig M, Roman RJ, Harder DR. Pressurization of isolated renal arteries increases inositol trisphosphate and diacylglycerol. *Am J Physiol.* 1994; 266:H1840–H1845. [PubMed: 8203583]
95. Gebremedhin D, Lange AR, Lowry TF, Taheri MR, Birks EK, Hudetz AG, Narayanan J, Falck JR, Okamoto H, Roman RJ, Nithipatikom K, Campbell WB, Harder DR. Production of 20-hete and its role in autoregulation of cerebral blood flow. *Circ Res.* 2000; 87:60–65. [PubMed: 10884373]
96. Gebremedhin D, Lange AR, Narayanan J, Aebly MR, Jacobs ER, Harder DR. Cat cerebral arterial smooth muscle cells express cytochrome p450 4a2 enzyme and produce the vasoconstrictor 20-hete which enhances l-type ca<sup>2+</sup> current. *J Physiol.* 1998; 507((Pt 3)):771–781. [PubMed: 9508838]
97. Frisbee JC, Roman RJ, Falck JR, Krishna UM, Lombard JH. 20-hete contributes to myogenic activation of skeletal muscle resistance arteries in brown norway and sprague-dawley rats. *Microcirculation.* 2001; 8:45–55. [PubMed: 11296852]
98. Lange A, Gebremedhin D, Narayanan J, Harder D. 20-hydroxyeicosatetraenoic acid-induced vasoconstriction and inhibition of potassium current in cerebral vascular smooth muscle is dependent on activation of protein kinase c. *J Biol Chem.* 1997; 272:27345–27352. [PubMed: 9341185]
99. Laher I, Bevan JA. Protein kinase c activation selectively augments a stretch-induced, calcium-dependent tone in vascular smooth muscle. *J Pharmacol Exp Ther.* 1987; 242:566–572. [PubMed: 3612552]
100. Fujii K, Heistad DD, Faraci FM. Flow-mediated dilatation of the basilar artery in vivo. *Circ Res.* 1991; 69:697–705. [PubMed: 1873864]
101. Fujii K, Heistad DD, Faraci FM. Effect of diabetes mellitus on flow-mediated and endothelium-dependent dilatation of the rat basilar artery. *Stroke.* 1992; 23:1494–1498. [PubMed: 1412587]

102. Paravicini TM, Miller AA, Drummond GR, Sobey CG. Flow-induced cerebral vasodilatation in vivo involves activation of phosphatidylinositol-3 kinase, nadph-oxidase, and nitric oxide synthase. *J Cereb Blood Flow Metab.* 2006; 26:836–845. [PubMed: 16222243]
103. Gaw AJ, Bevan JA. Flow-induced relaxation of the rabbit middle cerebral artery is composed of both endothelium-dependent and -independent components. *Stroke.* 1993; 24:105–109. discussion 109–110. [PubMed: 8418532]
104. Drouin A, Bolduc V, Thorin-Trescases N, Belanger E, Fernandes P, Baraghis E, Lesage F, Gillis MA, Villeneuve L, Hamel E, Ferland G, Thorin E. Catechin treatment improves cerebrovascular flow-mediated dilation and learning abilities in atherosclerotic mice. *American journal of physiology Heart and circulatory physiology.* 2011; 300:H1032–H1043. [PubMed: 21186270]
105. Drouin A, Thorin E. Flow-induced dilation is mediated by akt-dependent activation of endothelial nitric oxide synthase-derived hydrogen peroxide in mouse cerebral arteries. *Stroke.* 2009; 40:1827–1833. [PubMed: 19286591]
106. Garcia-Roldan JL, Bevan JA. Flow-induced constriction and dilation of cerebral resistance arteries. *Circ Res.* 1990; 66:1445–1448. [PubMed: 2335036]
107. Garcia-Roldan JL, Bevan JA. Augmentation of endothelium-independent flow constriction in pial arteries at high intravascular pressures. *Hypertension.* 1991; 17:870–874. [PubMed: 2045168]
108. Thorin-Trescases N, Bevan JA. High levels of myogenic tone antagonize the dilator response to flow of small rabbit cerebral arteries. *Stroke.* 1998; 29:1194–1200. discussion 1200–1191. [PubMed: 9626294]
109. Ward ME, Yan L, Kelly S, Angle MR. Flow modulation of pressure-sensitive tone in rat pial arterioles: Role of the endothelium. *Anesthesiology.* 2000; 93:1456–1464. [PubMed: 11149441]
110. Ngai AC, Winn HR. Modulation of cerebral arteriolar diameter by intraluminal flow and pressure. *Circ Res.* 1995; 77:832–840. [PubMed: 7554130]
111. Shimoda LA, Norins NA, Jeutter DC, Madden JA. Flow-induced responses in piglet isolated cerebral arteries. *Pediatr Res.* 1996; 39:574–583. [PubMed: 8848328]
112. Madden JA, Christman NJ. Integrin signaling, free radicals, and tyrosine kinase mediate flow constriction in isolated cerebral arteries. *Am J Physiol.* 1999; 277:H2264–H2271. [PubMed: 10600845]
113. Sipkema P, van der Linden PJ, Fanton J, Latham RD. Responses to mechanical stimuli of isolated basilar and femoral arteries of the rhesus monkey are different. *Heart Vessels.* 1996; 11:18–26. [PubMed: 9119801]
114. Bryan RM Jr, Marrelli SP, Steenberg ML, Schildmeyer LA, Johnson TD. Effects of luminal shear stress on cerebral arteries and arterioles. *Am J Physiol Heart Circ Physiol.* 2001; 280:H2011–H2022. [PubMed: 11299201]
115. Kim YS, Bogert LW, Immink RV, Harms MP, Colier WN, van Lieshout JJ. Effects of aging on the cerebrovascular orthostatic response. *Neurobiol Aging.* 2011; 32:344–353. [PubMed: 19356825]
116. Fang X, Faraci FM, Kaduce TL, Harmon S, Modrick ML, Hu S, Moore SA, Falck JR, Weintraub NL, Spector AA. 20-hydroxyeicosatetraenoic acid is a potent dilator of mouse basilar artery: Role of cyclooxygenase. *American journal of physiology Heart and circulatory physiology.* 2006; 291:H2301–H2307. [PubMed: 16782846]
117. Bryan RM Jr, Steenberg ML, Marrelli SP. Role of endothelium in shear stress-induced constrictions in rat middle cerebral artery. *Stroke.* 2001; 32:1394–1400. [PubMed: 11387504]
118. Andresen J, Shafi NI, Bryan RM Jr. Endothelial influences on cerebrovascular tone. *J Appl Physiol.* 2006; 100:318–327. [PubMed: 16357085]
119. Shi ZD, Tarbell JM. Fluid flow mechanotransduction in vascular smooth muscle cells and fibroblasts. *Ann Biomed Eng.* 2011; 39:1608–1619. [PubMed: 21479754]
120. Lee RM. Morphology of cerebral arteries. *Pharmacol Ther.* 1995; 66:149–173. [PubMed: 7630927]
121. Majack RA, Bhalla RC. Ultrastructural characteristics of endothelial permeability in chronic hypertension. *Hypertension.* 1981; 3:586–595. [PubMed: 7298113]
122. Roman RJ. P-450 metabolites of arachidonic acid in the control of cardiovascular function. *Physiol Rev.* 2002; 82:131–185. [PubMed: 11773611]



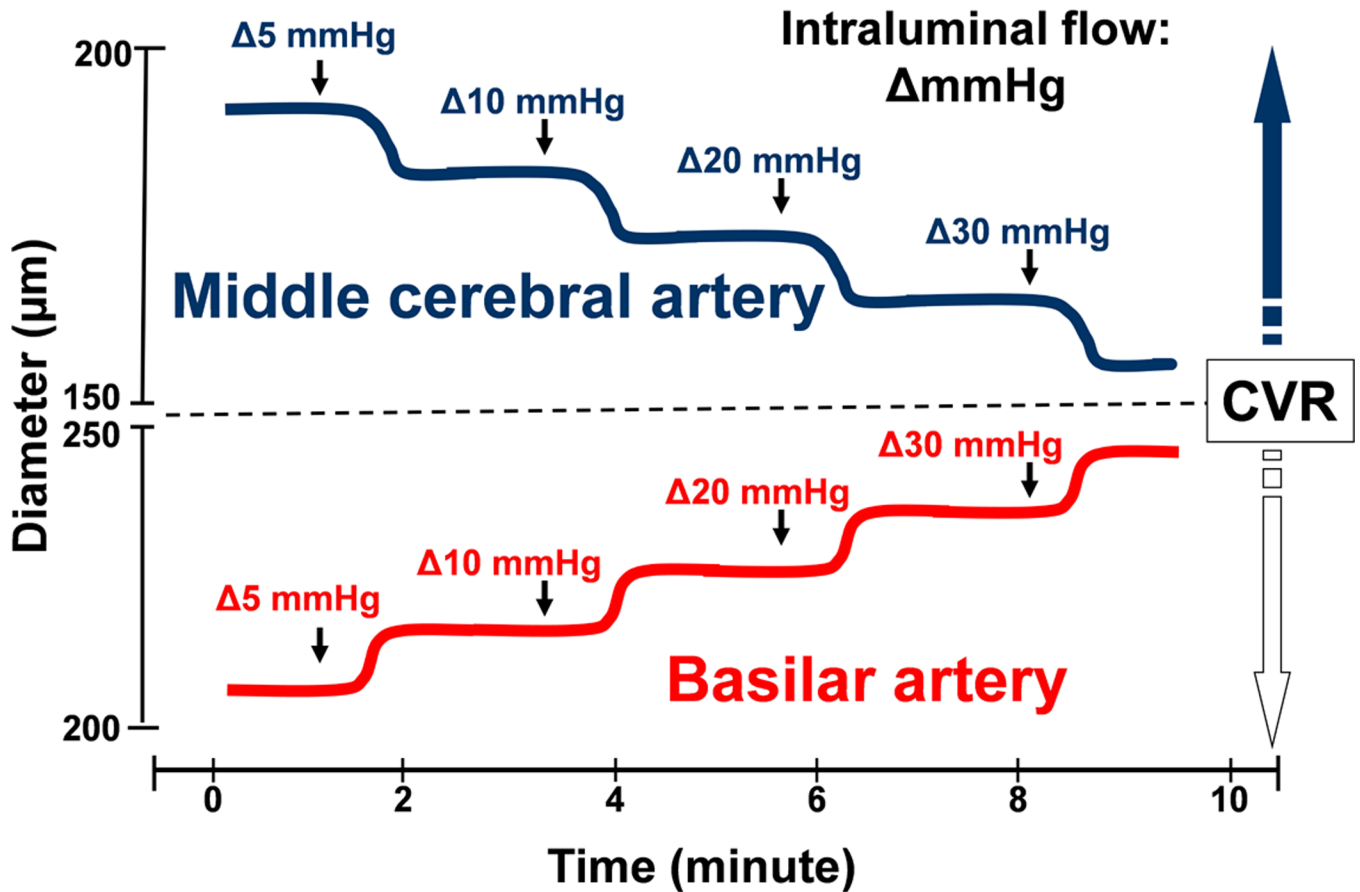
123. Campbell WB, Fleming I. Epoxyeicosatrienoic acids and endothelium-dependent responses. *Pflugers Arch.* 2010; 459:881–895. [PubMed: 20224870]
124. Behm DJ, Ogbonna A, Wu C, Burns-Kurtis CL, Douglas SA. Epoxyeicosatrienoic acids function as selective, endogenous antagonists of native thromboxane receptors: Identification of a novel mechanism of vasodilation. *J Pharmacol Exp Ther.* 2009; 328:231–239. [PubMed: 18836067]
125. Fujii K, Heistad DD, Faraci FM. Role of the basilar artery in regulation of blood flow to the brain stem in rats. *Stroke.* 1991; 22:763–767. [PubMed: 1905428]
126. Raisis JE, Kindt GW, McGillicuddy JE, Giannotta SL. The effects of primary elevation of cerebral venous pressure on cerebral hemodynamics and intracranial pressure. *J Surg Res.* 1979; 26:101–107. [PubMed: 106185]
127. Gordon GR, Choi HB, Rungta RL, Ellis-Davies GC, MacVicar BA. Brain metabolism dictates the polarity of astrocyte control over arterioles. *Nature.* 2008; 456:745–749. [PubMed: 18971930]
128. Golding EM, Robertson CS, Bryan RM Jr. Comparison of the myogenic response in rat cerebral arteries of different calibers. *Brain Res.* 1998; 785:293–298. [PubMed: 9518656]
129. Bohlen HG. Enhanced cerebral vascular regulation occurs by age 4 to 5 weeks in spontaneously hypertensive rats. *Hypertension.* 1987; 9:325–331. [PubMed: 3557598]
130. Strandgaard S. Autoregulation of cerebral blood flow in hypertensive patients. The modifying influence of prolonged antihypertensive treatment on the tolerance to acute, drug-induced hypotension. *Circulation.* 1976; 53:720–727. [PubMed: 815061]
131. New DI, Chesser AM, Thuraingham RC, Yaqoob MM. Cerebral artery responses to pressure and flow in uremic hypertensive and spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol.* 2003; 284:H1212–H1216. [PubMed: 12595297]
132. Smeda JS. Cerebral vascular changes associated with hemorrhagic stroke in hypertension. *Can J Physiol Pharmacol.* 1992; 70:552–564. [PubMed: 1498721]
133. Smeda JS, VanVliet BN, King SR. Stroke-prone spontaneously hypertensive rats lose their ability to auto-regulate cerebral blood flow prior to stroke. *J Hypertens.* 1999; 17:1697–1705. [PubMed: 10658935]
134. Dunn KM, Renic M, Flasch AK, Harder DR, Falck J, Roman RJ. Elevated production of 20-hete in the cerebral vasculature contributes to severity of ischemic stroke and oxidative stress in spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol.* 2008; 295:H2455–H2465. [PubMed: 18952718]
135. Poloyac SM, Zhang Y, Bies RR, Kochanek PM, Graham SH. Protective effect of the 20-hete inhibitor het0016 on brain damage after temporary focal ischemia. *J Cereb Blood Flow Metab.* 2006; 26:1551–1561. [PubMed: 16570075]
136. Renic M, Klaus JA, Omura T, Kawashima N, Onishi M, Miyata N, Koehler RC, Harder DR, Roman RJ. Effect of 20-hete inhibition on infarct volume and cerebral blood flow after transient middle cerebral artery occlusion. *J Cereb Blood Flow Metab.* 2009; 29:629–639. [PubMed: 19107134]





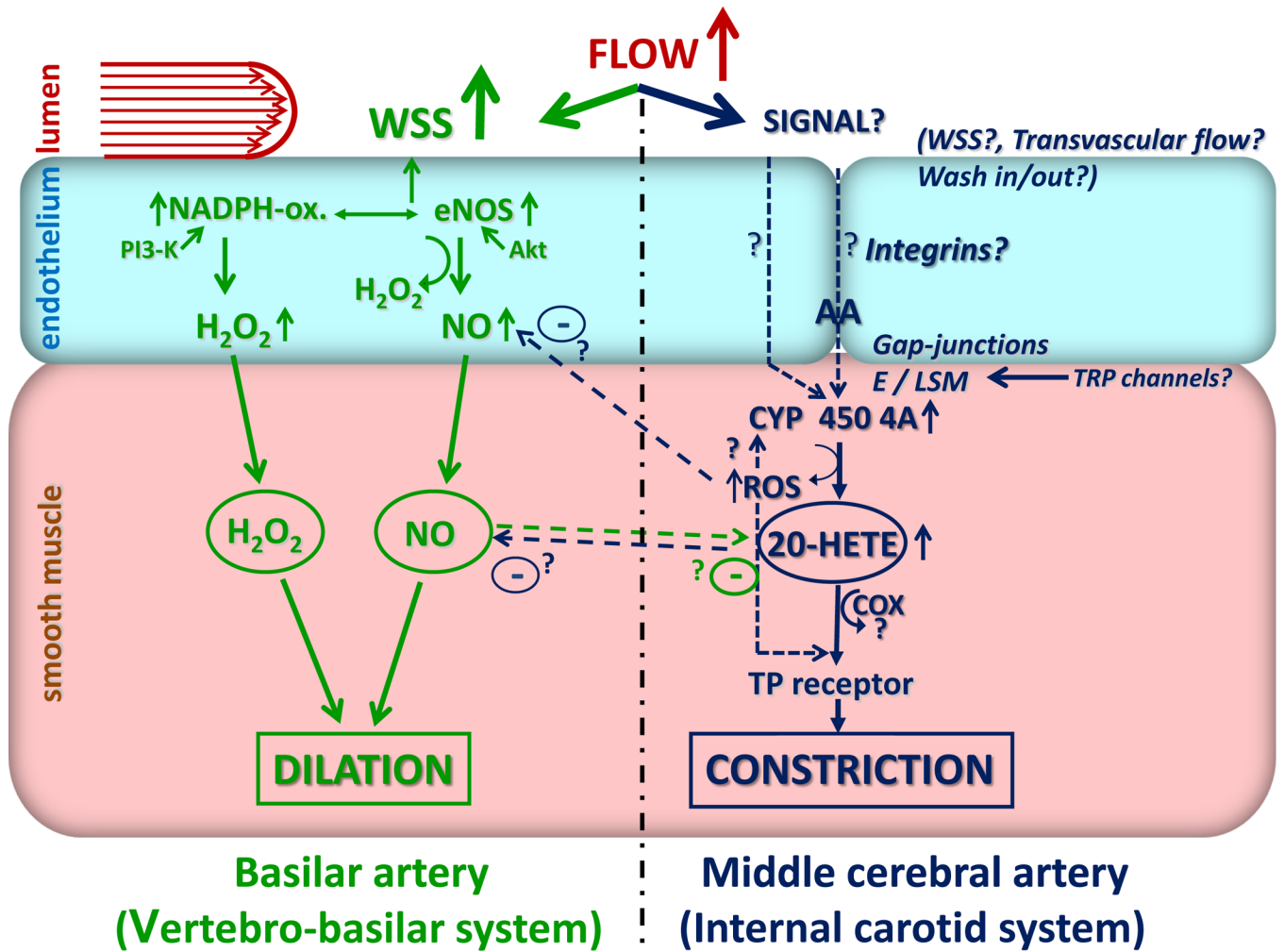
**Figure 1.**

Changes in diameter ( $\mu\text{m}$ ) of an isolated middle cerebral artery and a basilar artery to changes in intraluminal flow as a function of time, redrawn from original traces (P. Toth). In this experimental conditions flow is generated by increasing intraluminal pressure differences ( $\Delta 5$ ,  $\Delta 10$ , etc, mmHg) via the vessel by changing the inflow and outflow pressures with the same amount, but in the opposite directions (see details of methods in Toth, P. at al [23]). The opposite diameter responses indicate important regional differences in the nature of flow-induced responses of cerebral arteries. Constriction of middle cerebral artery by increasing cerebrovascular resistance (CVR) contributes to the autoregulation of cerebral blood flow by the “pressure-flow” mechanisms and thereby to the overall regulation of intracranial volume and pressure. In contrast, dilation of the basilar artery to increases in flow, similar to other peripheral vessels contributes to the development of functional hyperemia. See detailed discussion in the text.



**Figure 2.**

Schematic diagram showing the proposed physiological role of flow-induced constriction of cerebral arteries in autoregulation of cerebral blood flow. Combined effect of changes of intraluminal pressure and intraluminal flow ( $\Delta$  flow) achieves a more effective autoregulation of cerebral blood flow (CBF), whereas only pressure-induced diameter responses still allow substantial increases in CBF, thus inefficient autoregulation. It is likely that in vivo the “plateau” of autoregulation is not perfectly flat, i.e. the gain is less than 1, thus there is a slight increase in CBF as systemic pressure increases. Nevertheless, it is effective enough to prevent an exponential increase of CBF. Also, the range and shape of autoregulatory curve is likely to be more “rounded” at low and high pressure values.



**Figure 3.**

Proposed intracellular and molecular mechanisms eliciting flow-induced responses of cerebral vessels. Flow induces dilation, biphasic responses, or constriction of cerebral vessels depending on the regional and segmental localization of the vessels. We propose that in the internal carotid system larger arteries (such as the middle cerebral artery) constrict to increases in flow. The flow-induced constriction is mediated by 20-hydroxyeicosatetraenoic acid (20-HETE) (a metabolite of arachidonic acid (AA) produced by cytochrome P450 4A enzymes (CYP450 4A) acting via thromboxane A<sub>2</sub>/prostaglandin H<sub>2</sub> (TP) receptors and requires COX activity. CYP450 4A also produces reactive oxygen species (ROS), which contribute to the constriction. Whereas, in the brain stem supplied by the vertebro-basilar system larger arteries, such as basilar artery, dilate to flow. Dilation is mediated by NADPH-oxidase (activated by phosphatidylinositol3-kinase (PI3-K) derived H<sub>2</sub>O<sub>2</sub> and/or eNOS derived nitric oxide (NO). eNOS is activated in an Akt-dependent pathway, and probably generates H<sub>2</sub>O<sub>2</sub>, as well (ref number here!! based on Toth at. al, Paravicini at al. and Drouin at al.). The exact nature of signals and sensors are still unknown (see detailed description in the text).